Supporting information for

Rational engineering of biomimetic flavylium fluorophores for regulating the lysosomal and mitochondrial localization behavior by pH-induced structure switch and application to fluorescence imaging

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Table of Contents

Pages

Materials and instruments	3
Determination of the fluorescence quantum yield	3
Cytotoxicity azzssay	3
Cell Culture and Imaging	3-4
Synthesis	4-8
Scheme S1	4
Figures S1-5	9-11
Table S1-5	
Figures S6-7	12-13
Scheme S2	13
Figures S8-45	14-35

Materials and instruments. Unless otherwise stated, all reagents were purchased from commercial suppliers and used without further purification. Solvents used were purified by standard methods prior to use. Twice-distilled water was used throughout all experiments. Mass spectra were performed using an LCQ Advantage ion trap mass spectrometer from Thermo Finnigan or Agilent 1100 HPLC/MSD spectrometer. ¹H NMR and ¹³C NMR spectra were recorded at ambient temperature using a Bruker AVANCE NEO 400 MHz and 600 MHz spectrometers, using TMS as an internal standard. Electronic absorption spectra were recorded at room temperature with a HITACHI F4600 fluorescence spectrophotometer with the excitation and emission slit widths at 10.0 and 10.0 nm respectively. The fluorescence imaging of cells was performed with Two-photon Confocal Scanning Laser Microscope (TCS SP8 DIVE). TLC analysis was performed on silica gel plates and column chromatography was conducted over silica gel (mesh 200-300), both of which were obtained from the Qingdao Ocean Chemicals.

Determination of the fluorescence quantum yield: Fluorescence quantum yields for the probes were determined by using Fluorescein ($\Phi_f = 0.79$ in 0.1 M NaOH, for FL-1, FL-2), Rh6G ($\Phi_f = 0.95$ in EtOH, for FL-5) and cresol purple ($\Phi_f = 0.58$ in ethanol, for other FL dyes) as a fluorescence standard. The quantum yield was calculated using the following equation:

$$\Phi_{\mathrm{F}(\mathrm{X})} = \Phi_{\mathrm{F}(\mathrm{S})} \left(A_{S} F_{X} / A_{X} F_{S} \right) \left(n_{X} / n_{S} \right)^{2}$$

Where Φ_F is the fluorescence quantum yield, *A* is the absorbance at the excitation wavelength, *F* is the area under the corrected emission curve, and *n* is the refractive index of the solvents used. Subscripts S and X refer to the standard and to the unknown, respectively.

Cytotoxicity azzssays. The toxicity of the probes towards living HeLa cells was determined by MTT (3-(4,5-dimethyl-2-thiazolyl)-2,5-diphenyl-2-H-tetrazolium bromide) assays. HeLa cells were grown in the Eagle's medium (1640) supplemented with 10% FBS (fetal bovine serum) in an atmosphere of 5% CO_2 and 95% air at 37 °C. Immediately before the experiments, the cells were placed in a 96-well plate, followed

by addition of increasing concentrations of the probes (99%1640 and 1% DMSO). The final concentrations of the probes was 0, 5, 10, 15, 20, 25 μ M (n = 5) respectively. The cells were then incubated at 37 °C in an atmosphere of 5% CO₂ and 95% air at 37 °C for 24 h, followed by MTT assays. Untreated assay with 1640 (n = 5) was also conducted under the same conditions.

HeLa cells, T24 cells and HL-7702 cells Culture and Imaging Using FL dyes.

T24 cells, HL-7702 cells were seeded in Dulbecco's modified Eagle'smedium (DMEM) supplemented with 10% fetal bovine serum for 24 h. HeLa cells were seeded in RPMI 1640 Media supplemented with 10% fetal bovine serum for 24 h. T24 cells, HL-7702 cells, and HeLa cells were incubated with 10.0 μ M FL dyes for 30 min, and then treated with Lyso-Trackers Green, Lyso-Trackers Red, Mito-Trackers Green, or Mito-Trackers Deep Red for another 15 min. The fluorescence imaging of cells was performed with with Two-photon Confocal Scanning Laser Microscope (TCS SP8 DIVE). For FL-1: $\lambda_{ex} = 488 \text{ nm}$, $\lambda_{em} = 498-550 \text{ nm}$; For FL-3: $\lambda_{ex} = 552 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-4: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-7: $\lambda_{ex} = 552 \text{ nm}$, $\lambda_{em} = 562-650 \text{ nm}$; For FL-8: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-8: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$; For FL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 650-750 \text{ nm}$; For FL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 650-780 \text{ nm}$; For FL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 650-780 \text{ nm}$; For FL-9: $\lambda_{ex} = 488 \text{ nm}$, $\lambda_{em} = 498-550 \text{ nm}$; For SL-9: $\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 580-610 \text{ nm}$; For Mito-Tracher Green: $\lambda_{ex} = 488 \text{ nm}$, $\lambda_{em} = 498-550 \text{ nm}$; For Lyso-Tracher Red: $\lambda_{ex} = 552 \text{ nm}$, $\lambda_{em} = 580-610 \text{ nm}$; For Mito-Tracher Green: $\lambda_{ex} = 488 \text{ nm}$, $\lambda_{em} = 498-550 \text{ nm}$.

Fluorescent imaging in living mice using probe FL-9.

Female BALB/C mice aged about 5 weeks were purchased from Hunan Slack Jingda Experimental Animal Co., Ltd. The tumors were developed by subcutaneously implanting 4×10^6 4T1 cells into the left hind leg injections of female BALB/c mice. Tumor volume was calculated using the following formula: volume = (tumor length)×(tumor width)² /2. When the tumor volume reached 200 mm³, the mice were administered with **FL-9** (20 µM). After the probe post-injection, the mice were imaged using a Kodak in vivo FX Pro imaging system (Bruker). With an excitation filter of 650 nm and the emission filter of 700 nm.

Synthesis of compound B1.

POC1₃ (2.0 mL) was added dropwisely by a separatory funnel to a flask containing DMF (5 mL) with stirring at 0 °C over 30 min. Subsequently, compounds A1 (1 g) in DMF (0.5 mL) was added slowly with stirring and the mixture was heated for 5 h at 60 °C. Then the mixture was poured to ice water and the resulting precipitate was filtered off and washed with cold water (200 mL) to afford the light yellow solid B1, which was utilized in the next reaction without purification again. ¹H NMR (400 MHz, CDCl₃) δ 10.39 (s, 1H), 7.87 (d, *J* = 7.5 Hz, 1H), 7.39 -7.32 (m, 2H), 7.22 (d, *J* = 7.1 Hz, 1H), 2.85 (t, *J* = 8.0 Hz, 2H), 2.64 (t, *J* = 8.0 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 190.71, 145.91, 138.99, 132.03, 131.41, 127.74, 127.11, 126.34, 27.04, 21.60.

Synthesis of compound B2.

POC1₃ (2.0 mL) was added dropwisely by a separatory funnel to a flask containing DMF (5 mL) with stirring at 0 °C over 30 min. Subsequently, compounds A2 (1 g) in DMF (0.5 mL) was added slowly with stirring and the mixture was heated for 5 h at 60 °C. Then the mixture was poured to ice water and the resulting precipitate was filtered off and washed with cold water (200 mL) to afford the light yellow solid B2,

which was utilized in the next reaction without purification again. ¹H NMR (400 MHz, CDCl₃) δ 10.33 (s, 1H), 7.80 (d, *J* = 8.7 Hz, 1H), 6.84 (dd, *J* = 8.7, 1.9 Hz, 1H), 6.75 (s, 1H), 3.86 (s, 3H), 2.82 (t, *J* = 7.9 Hz, 2H), 2.63 (t, *J* = 7.9 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 190.46, 162.19, 146.09, 141.31, 129.82, 128.37, 124.90, 113.56, 112.11, 55.49, 27.58, 21.54.

Synthesis of compound B3.

POC1₃ (2.0 mL) was added dropwisely by a separatory funnel to a flask containing DMF (5 mL) with stirring at 0 °C over 30 min. Subsequently, compounds A3 (1 g) in DMF (0.5 mL) was added slowly with stirring and the mixture was heated for 5 h at 60 °C. Then the mixture was poured to ice water and the resulting precipitate was filtered off and washed with cold water (200 mL) to afford the light yellow solid B3, which was utilized in the next reaction without purification again. ¹H NMR (400 MHz, CDCl₃) δ 10.27 (s, 1H), 7.72 (d, *J* = 8.8 Hz, 1H), 6.60 (d, *J* = 8.8 Hz, 1H), 6.49 (s, 1H), 3.06 (s, 6H), 2.79 (t, *J* = 7.9 Hz, 2H), 2.62 (t, *J* = 7.9 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 190.15, 141.19, 128.30, 110.51, 109.90, 40.15, 28.22, 21.76.

General procedure for synthesis of FL1-9.

Compound A (0.5 mmol) and B (0.5 mmol) were dissolved with a mixture of AcOH (8 mL) and HClO₄ (2 mL). After refluxing and stirring for 4 h, the mixture was poured into ice water (200 mL), and then the aqueous phase was extracted with dichloromethane (3 \times 100 mL). The organic layers were collected and evaporated under reduced pressure. Finally, the crude products were purified by column chromatography on silica gel flash chromatography to give compounds FL1-9.

Synthesis of compound FL-1

Yellow solid; yield 62.9%; ¹H NMR (400 MHz, DMSO- d_6) δ 8.83 (s, 1H), 8.38 (d, J = 7.8 Hz, 2H), 7.73 (t, J = 7.5 Hz, 2H), 7.65-7.53 (m, 4H), 3.21-3.14 (m, 8H). ¹³C

NMR (100 MHz, DMSO-*d*₆) δ 165.42, 153.99, 142.38, 135.61, 132.06, 129.72, 128.82, 126.56, 125.92, 25.95, 25.32. HRMS (ESI) Calcd for C₂₁H₁₇O⁺ ([M]⁺): 285.1274, Found, 285.1274.

Synthesis of compound FL-2

Yellow solid; yield 65.1%; ¹H NMR (400 MHz, DMSO- d_6) δ 8.68 (s, 1H), 8.34 (dd, J = 16.9, 8.2 Hz, 2H), 7.69 (t, J = 7.3 Hz, 1H), 7.61-7.51 (m, 2H), 7.15 (d, J = 7.9 Hz, 2H), 3.95 (s, 3H), 3.18 -3.07 (m, 8H). ¹³C NMR (100 MHz, DMSO- d_6) δ 166.19, 165.74, 163.47, 152.59, 146.05, 141.65, 134.92, 130.99, 130.00, 129.58, 129.37, 128.65, 126.03, 126.00, 118.63, 115.60, 114.55, 56.64, 26.62, 26.21, 25.42, 25.12. HRMS (ESI) Calcd for C₂₂H₁₉O₂⁺ ([M]⁺): 315.1380, Found, 315.1372.

Synthesis of compound FL-3

Yellow solid; yield 63.2%; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.22 (s, 1H), 8.13 (d, *J* = 3.4 Hz, 1H), 7.58 (t, *J* = 7.3 Hz, 1H), 7.51 (t, *J* = 7.4 Hz, 1H), 7.46 -7.41 (m, 2H), 6.80-6.69 (m, 1H), 6.56 (d, *J* = 6.7 Hz, 1H), 3.06-2.91 (m, 8H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 166.62, 158.89, 157.90, 147.84, 147.06, 140.13, 133.38, 130.54, 129.96, 129.27, 128.32, 126.43, 125.50, 124.68, 115.04, 112.70, 26.87, 26.39, 25.83, 24.78. HRMS (ESI) Calcd for C₂₁H₁₈NO⁺ ([M]⁺): 300.1383, Found, 300.1379.

Synthesis of compound FL-4

Red solid; yield 68.8%; ¹H NMR (400 MHz, DMSO- d_6) δ 8.25 (s, 1H), 8.15 (d, J = 4.4 Hz, 1H), 7.73-7.65 (m, 1H), 7.61-7.51 (m, 2H), 7.46 (d, J = 7.3 Hz, 1H), 6.92 (dd, J = 9.1, 2.3 Hz, 1H), 6.84 -6.72 (m, 1H), 3.20 (s, 6H), 3.06-3.00 (m, 8H). ¹³C NMR (100 MHz, DMSO- d_6) δ 166.58, 159.30, 155.82, 148.06, 146.12, 140.26, 133.50, 132.00, 130.24, 129.62, 129.23, 128.39, 126.45, 125.92, 124.75, 112.92, 111.29, 65.50, 30.48, 27.11, 26.39, 24.83. HRMS (ESI) Calcd for C₂₃H₂₂NO ⁺ ([M]⁺): 328.1696, Found, 328.1690.

Synthesis of compound FL-5

Yellow solid; yield 65.7%; ¹H NMR (400 MHz, DMSO- d_6) δ 8.55 (s, 1H), 8.25 (d, J = 9.0 Hz, 2H), 7.10 (d, J = 7.5 Hz, 4H), 3.92 (s, 6H), 3.08 (s, 8H). ¹³C NMR (400 MHz, DMSO- d_6) δ 165.15, 164.43, 151.66, 145.19, 128.94, 128.69, 118.69, 115.25, 114.51, 56.51, 26.65, 25.21. HRMS (ESI) Calcd for C₂₃H₂₁O₃⁺ ([M]⁺): 345.1485, Found, 345.1479.

Synthesis of compound FL-6

Red solid; yield 61.8%; ¹H NMR (400 MHz, DMSO- d_6) δ 8.19 (s, 1H), 8.11 (d, J = 8.4 Hz, 1H), 8.05 (d, J = 8.8 Hz, 1H), 7.22 (s, 1H), 7.06 (s, 1H), 6.73 (dd, J = 8.8, 2.0 Hz, 1H), 6.55 (s, 1H), 3.89 (s, 3H), 3.02-2.89 (m, 8H). ¹³C NMR (400 MHz, DMSO- d_6) δ 165.72, 163.89, 157.28, 148.03, 146.26, 143.41, 129.87, 128.09, 127.19, 124.51, 119.13, 114.68, 114.60, 114.43, 112.75, 56.30, 29.48, 26.93, 26.83, 25.66, 24.89. HRMS (ESI) Calcd for C₂₂H₂₀NO₂⁺ ([M]⁺): 330.1489, Found, 330.1488.

Synthesis of compound FL-7

Red solid; yield 66.2%; ¹H NMR (400 MHz, DMSO-*d*₆) δ 8.22 (s, 1H), 8.11 (d, *J* = 4.1 Hz, 1H), 7.72 (dt, *J* = 6.9, 3.6 Hz, 1H), 7.66 (dd, *J* = 5.8, 3.2 Hz, 1H), 7.09-7.04 (m, 2H), 6.77 (d, *J* = 2.2 Hz, 1H), 3.90 (s, 3H), 3.18 (s, 6H), 3.04 -2.96 (m, 8H). ¹³C NMR (100 MHz, DMSO-*d*₆) δ 166.56, 159.28, 155.80, 148.04, 146.10, 140.25, 133.49, 131.99, 130.22, 129.60, 129.22, 128.38 (s), 126.43, 125.91, 124.74, 112.91, 111.28, 65.49, 27.11, 26.39, 25.89, 24.83. HRMS (ESI) Calcd for C₂₄H₂₄NO₂⁺ ([M]⁺): 358.1802, Found, 358.1796.

Synthesis of compound FL-8

Purple solid; yield 69.3%; ¹H NMR (400 MHz, DMSO- d_6) δ 8.04 (s, 1H), 7.99 (d, J = 9.0 Hz, 1H), 7.92 (d, J = 8.7 Hz, 1H), 6.88 (s, 1H), 6.81 (dd, J = 9.0, 2.5 Hz, 1H), 6.73-6.70 (m, 1H), 6.53 (d, J = 2.0 Hz, 1H), 3.12 (s, 6H), 2.97-2.85 (m, 8H). ¹³C NMR (400 MHz, DMSO- d_6) δ 163.26, 162.64, 155.94, 154.39, 147.01, 144.62,

143.67, 128.42, 127.55, 124.55, 124.30, 114.04, 113.17, 112.78, 111.84, 111.21, 31.62, 30.30, 27.32, 27.06, 25.36. HRMS (ESI) Calcd for $C_{23}H_{23}N_2O^+$ ([M]⁺): 343.1805, Found, 343.1802.

Synthesis of compound FL-9

Purple solid; yield 70.4%; ¹H NMR (400 MHz, DMSO- d_6) δ 8.05 (s, 1H), 7.97 (d, J = 8.9 Hz, 2H), 6.83 (d, J = 8.4 Hz, 2H), 6.73 (s, 2H), 3.13 (s, 12H), 2.94 (s, 8H). ¹³C NMR (400 MHz, DMSO- d_6) δ 162.94, 154.48, 147.01, 143.84, 127.62, 124.74, 113.13, 111.97, 111.21, 27.30, 25.40. HRMS (ESI) Calcd for C₂₅H₂₇N₂O⁺ ([M]⁺): 371.2118, Found, 371.2130.



Scheme S1 Synthesis of flavylium fluorophores FL-1~9.



Figure S1 Normalized absorption (a) and fluorescence emission (b) spectra of 10 μ M compounds FL-1~9 in DMF.

Compound	λ _{abs} (nm)	ε _{max} (10 ⁴ M ⁻¹ cm ⁻¹)	E _x (nm)	λ _{em} (nm)	$\Phi_{\rm f}$	Stokes Shift (nm)
FL-1	445	0.25	432	500	0.14	55
FL-2	475	0.99	476	550	0.36	75
FL-3	542	0.78	556	643	0.02	101
FL-4	563	0.53	583	682	0.02	119
FL-5	499	0.27	489	566	0.53	67
FL-6	554	1.89	556	643	0.06	89
FL-7	574	0.96	580	681	0.03	107
FL-8	610	0.32	600	693	0.02	83
FL-9	624	1.40	630	717	0.02	93

Table S1 Photophysical properties of the compounds FL-1~9 in DMF.



Figure S2 Normalized absorption (a) and fluorescence emission (b) spectra of 10 μ M compounds FL-1~9 in DMSO.

Compound	λ _{abs} (nm)	ε _{max} (10 ⁴ M ⁻¹ cm ⁻¹)	E _x (nm)	λ _{em} (nm)	$\Phi_{\rm f}$	Stokes Shift (nm)
FL-1	449	2.39	445	502	0.04	53
FL-2	459	1.44	456	552	0.06	93
FL-3	544	1.07	543	645	0.02	101
FL-4	564	1.66	560	686	0.01	122
FL-5	499	0.31	492	569	0.51	70
FL-6	557	1.89	534	645	0.05	88
FL-7	575	1.01	573	686	0.02	111
FL-8	607	0.52	598	702	0.01	95
FL-9	625	1.91	630	721	0.02	96

Table S2. Photophysical properties of the compounds FL-1~9 in DMSO.



Figure S3 Normalized absorption (a) and fluorescence emission (b) spectra of 10 μ M compounds FL-1~9 in EtOH.

Compoun d	λ _{abs} (nm)	ε _{max} (10 ⁴ M ⁻¹ cm ⁻¹)	E _x (nm)	λ _{em} (nm)	$\Phi_{ m f}$	Stokes Shift (nm)
FL-1	448	0.73	443	495	0.99	47
FL-2	459	1.30	455	547	0.73	88
FL-3	544	1.04	546	636	0.01	92
FL-4	561	1.61	576	667	0.02	106
FL-5	502	4.17	482	561	0.07	59
FL-6	558	2.01	550	640	0.08	82
FL-7	575	0.93	573	667	0.05	92
FL-8	613	0.57	580	683	0.04	70
FL-9	624	2.06	600	702	0.04	78

Table S3. Photophysical properties of the compounds FL-1~9 in EtOH.





Figure S4 Normalized absorption (a) and fluorescence emission (b) spectra of 10 μ M compounds FL-1~9 in MeCN.

Compound	λ _{abs} (nm)	ε _{max} (10 ⁴ M ⁻¹ cm ⁻¹)	E _x (nm)	λ _{em} (nm)	$\Phi_{ m f}$	Stokes Shift (nm)
FL-1	445	2.48	443	495	0.99	50
FL-2	456	1.11	446	528	0.89	72
FL-3	525	0.93	520	627	0.06	102
FL-4	559	1.44	555	671	0.02	112
FL-5	497	0.24	485	558	0.68	61
FL-6	538	1.86	532	627	0.02	89
FL-7	568	0.97	582	671	0.05	103
FL-8	594	0.51	600	681	0.03	87
FL-9	617	1.62	630	708	0.04	91

Table S4 Photophysical properties of the compounds FL-1~9 in MeCN.



Figure S5 Normalized absorption (a) and fluorescence emission (b) spectra of 10 μ M compounds FL-1~9 in PBS.

Compound	λ _{abs} (nm)	ε _{max} (10 ⁴ M ⁻¹ cm ⁻¹)	E _x (nm)	λ _{em} (nm)	$\Phi_{\rm f}$	Stokes Shift (nm)
FL-1	444	0.31	435	492	0.99	48
FL-2	453	1.04	452	520	0.83	67
FL-3	523	0.91	523	627	0.02	104
FL-4	549	1.02	546	664	0.01	115
FL-5	510	0.29	489	552	0.73	42

Table S5 Photophysical properties of the compounds FL-1~9 in PBS.

FL-6	523	1.75	522	618	0.02	95
FL-7	557	1.75	556	664	0.01	107
FL-8	574	0.41	586	673	0.01	99
FL-9	596	1.46	615	705	0.05	109



Figure S6 Photostability test of the **FL-1~9** by Xenon lamp (150 W) irradiation within 30 min (normalized fluorescence intensity vs. time). The excitation wavelengths of the spectra obtained for dyes **FL-1~9** refer to the wavelengths in PBS and emission wavelengths of the spectra obtained for dyes FL-1~9 also refer to the wavelengths in PBS. Insets: Color map of representative **FL** dyes before and after irradiation.



Figure S7 (a-i) pH-dependence of the fluorescence intensity of compounds FL-1~9 (10 μ M) with the arrow indicating the change of the fluorescence intensities with pH increase from 3 to 10.



Figure S8 (a-i) pH-dependence of the UV of compounds FL-1~9 (10 μ M) with the arrow indicating the change of the UV absorbance with pH increase from 3 to 10.



Figure S9 (a-c) pH-dependence of the fluorescence intensity of compounds FL-1, FL-2, FL-5 (10 μ M) with the arrow indicating the change of the fluorescence intensities with pH increase from 3 to 10. FL-1 and FL-2 (λ_{ex} =330 nm); FL-5 (λ_{ex} =380 nm).



Scheme S2 pH-induced structure switch from flavylium cations to hemiketals of FL.



Figure S10 Charge distributions of flavylium rings, molecular orbital plots (LUMO and HOMO) of flavylium fluorophores and their corresponding pK_{cycl} values.



Figure S11 Color map of representative FL dyes that mimic lysosome and mitochondrial pH at the test-tube level.



Figure S12 Cytotoxicity of FL dyes against HeLa cells.



Figure S13 Cell viability of HeLa cells incubated with FL dyes (10 μ M) with or without laser irradiation (1.0 W cm⁻²).



Figure S14 Confocal fluorescence images of **FL-1~4** dyes by HL-7702 cells via TCS SP8 DIVE imaging. (a) HL-7702 cells were co-stained with **FL-1~4** dyes and

different Lyso Trackers; (b) The pK_{cycl} value of **FL-1~4** dyes; (c) The Pearson's correlation coefficient of **FL-1~4** dyes. All images were obtained at 63 oil magnification. Confocal fluorescence images of **FL-1** ($\lambda_{ex} = 488 \text{ nm}, \lambda_{em} = 498-550 \text{ nm}$); **FL-2** ($\lambda_{ex} = 488 \text{ nm}, \lambda_{em} = 498-550 \text{ nm}$); **FL-2** ($\lambda_{ex} = 488 \text{ nm}, \lambda_{em} = 498-550 \text{ nm}$); **FL-4** ($\lambda_{ex} = 638 \text{ nm}, \lambda_{em} = 650-750 \text{ nm}$); Lyso-Tracher Red ($\lambda_{ex} = 552 \text{ nm}, \lambda_{em} = 580-610 \text{ nm}$); Lyso-Tracher Green ($\lambda_{ex} = 488 \text{ nm}, \lambda_{em} = 500-550 \text{ nm}$). Scale bar=10 µm.



Figure S15 Confocal fluorescence images of **FL-1~4** dyes by T24 cells via TCS SP8 DIVE imaging. (a) T24 cells were co-stained with **FL-1~4** dyes and different Lyso Trackers; (b) The pK_{cycl} value of **FL-1~4** dyes; (c) The Pearson's correlation coefficient of **FL-1~4** dyes. All images were obtained at 63 oil magnification. Confocal fluorescence images of **FL-1** ($\lambda_{ex} = 488 \text{ nm}$, $\lambda_{em} = 498-550 \text{ nm}$); **FL-2** ($\lambda_{ex} =$ 488 nm, $\lambda_{em} = 498-550 \text{ nm}$); **FL-3** ($\lambda_{ex} = 552 \text{ nm}$, $\lambda_{em} = 562-610 \text{ nm}$); **FL-4** ($\lambda_{ex} = 638 \text{ nm}$, $\lambda_{em} = 650-750 \text{ nm}$); Lyso-Tracher Red ($\lambda_{ex} = 552 \text{ nm}$, $\lambda_{em} = 580-610 \text{ nm}$); Lyso-Tracher Green ($\lambda_{ex} = 488 \text{ nm}$, $\lambda_{em} = 500-550 \text{ nm}$). Scale bar=10 µm.



Figure S16 Confocal fluorescence images of FL-5 dyes and different organelle trackers by HeLa cells via TCS SP8 DIVE imaging, as well as the corresponding in situ spectra in FL-5 and different organelle trackers. For FL-5: $\lambda_{ex} = 488$ nm, $\lambda_{em} = 498-550$ nm; For Lyso-Tracher Red: $\lambda_{ex} = 552$ nm, $\lambda_{em} = 580-610$ nm; For Mito-Tracher Deep Red: $\lambda_{ex} = 638$ nm, $\lambda_{em} = 650-750$ nm. Scale bar=10 µm.



Figure S17 Time-dependent fluorescence and absorption spectra of probe **FL-9** (10 μ M) in the presence of 100 μ M Na₂S in PBS buffer (pH=7.4). Excitation, 410 nm.



Figure S18 The fluorescence enhancement at 506 nm to various biologically relevant species in aqueous solution (pH 7.4, 25 mM PBS with 20% DMSO). 1. Blank, 2. HSO₃⁻ (25 μM), 3. CN⁻(25 μM), 4. CO₃²⁻(25 μM), 5. N₃⁻(25 μM), 6. NO₂⁻(25 μM), 7. H₂PO₄⁻(25 μM), 8. NO₃⁻(25 μM), 9. SO₄²⁻(25 μM), 10. SCN⁻(25 μM), 11. Na₂S(25 μM), 12. S₂O₃²⁻(25 μM), 13. Cys(1 mM), 14. GSH(1 mM), 15. I⁻(25 μM), 16. Cl⁻(25 μM). Excitation, 410 nm.







Figure S20 First row, HeLa cells incubated with only **FL-9** (10 μ M) for 30 minutes; Second row, the cells pretreated with Na₂S (50 μ M) for 30 min and then treated with **FL-9** (10 μ M) for 30 minutes; Third row, the cells pretreated with Na₂S (100 μ M) for 30 min and then treated with **FL-9** (10 μ M) for 30 minutes. The excitation wavelength was 488 nm, the emission collected at 500-610 nm was denoted as the green channel; the excitation wavelength was 638 nm, the emission collected at 650-780 nm was denoted as the green channel. Scale bar = 20 μ m.



Figure S21 In vivo fluorescence images of H₂S in living mice with **FL-9**. (a) Only saline and saline containing AOAA (400 μ M) were injected into normal issue (A) and the tumor tissues (B, C). After 30 min, **FL-9** (20 μ M) was injected into the tumor tissues (B, C) and normal issue (A); (b) quantifification of the A, B and C area fluorescence intensity in (a) (n = 3). $\lambda_{ex} = 650$ nm, 700 nm emission flter.







Figure S23 The ¹³C NMR spectra of B1.



Figure S25 The ¹³C NMR spectra of B2.



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 fl (ppm)

Figure S27 The ¹³C NMR spectra of B3.



Figure S28 The ¹H NMR spectra of FL-1.



Figure S29 The ¹³C NMR spectra of FL-1.



Figure S31 The ¹³CNMR spectra of FL-2.



210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 £1 (ppm)

Figure S33 The ¹³C NMR spectra of FL-3.



Figure S35 The ¹³C NMR spectra of FL-4.



Figure S37 The ¹³C NMR spectra of FL-5.





Figure S39 The ¹³C NMR spectra of FL-6.

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Figure S40 The ¹H NMR spectra of FL-7.



Figure S41 The ¹³C NMR spectra of FL-7.



Figure S43 The ¹³C NMR spectra of FL-8.



Figure S45 The ¹³CNMR spectra of FL-9.