

## Supporting Information

# Study on the Kinetics and Mechanism of Ferrocene-Triptide Inhibiting Insulin Aggregation

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## 1. Supplementary tables

**Table S1.** Effects of Fc-peptides on the kinetic parameters of insulin fibrillation in 20% (v/v) acetic acid solution. The concentration of insulin was 2 mg/mL.

	Fibril Formation		
	Lag time (h)	Fluorescence Intensity (a.u.)	Amyloidogenic extent (%) <sup>a</sup>
Insulin	62.9	138.0	100
Insulin + Fc-FFY			
100 $\mu$ M	89.9	58.8	42.6
200 $\mu$ M	135.1	23.7	17.2
300 $\mu$ M	n.a. <sup>b</sup>	0.8	0.6
Insulin + Fc-FFF			
100 $\mu$ M	98.4	52.5	38
200 $\mu$ M	120.0	28.2	20.4
300 $\mu$ M	n.a.	0.96	0.7
Insulin + Fc-FFD			
100 $\mu$ M	90.0	67.13	48.6
200 $\mu$ M	113.2	42.4	30.7
300 $\mu$ M	122.0	37.4	27.1
Insulin + Fc-FFK			
100 $\mu$ M	90.3	89.0	64.5
200 $\mu$ M	86.0	64.9	47
300 $\mu$ M	91.1	47.7	34.6

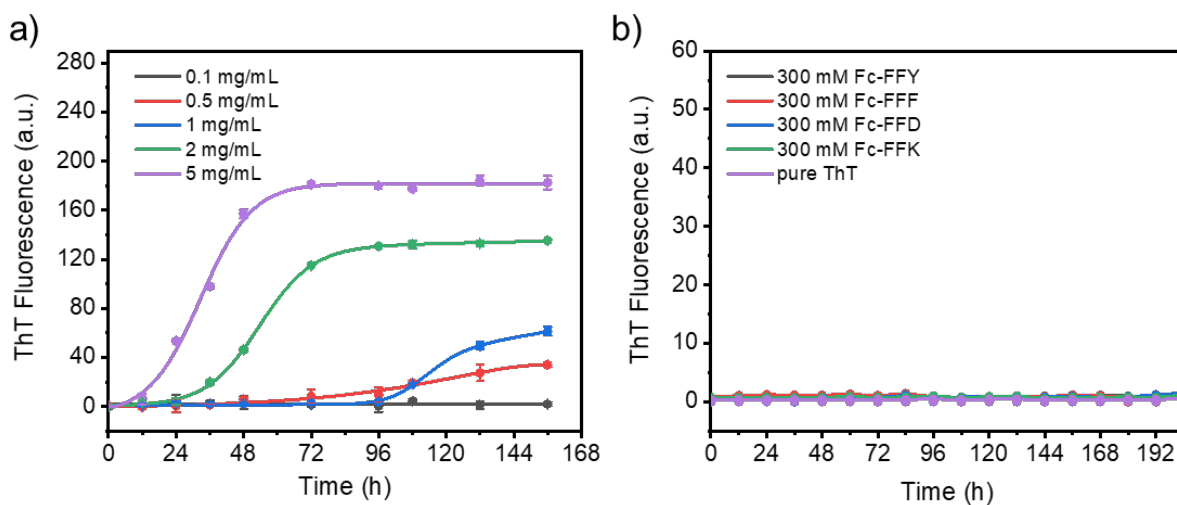
<sup>a</sup> The fluorescence intensity of insulin was set as a reference value. Amyloidogenic extent of a system was calculated by the final fluorescence intensity of the system divided by that of insulin.

<sup>b</sup> n.a. is short for not available.

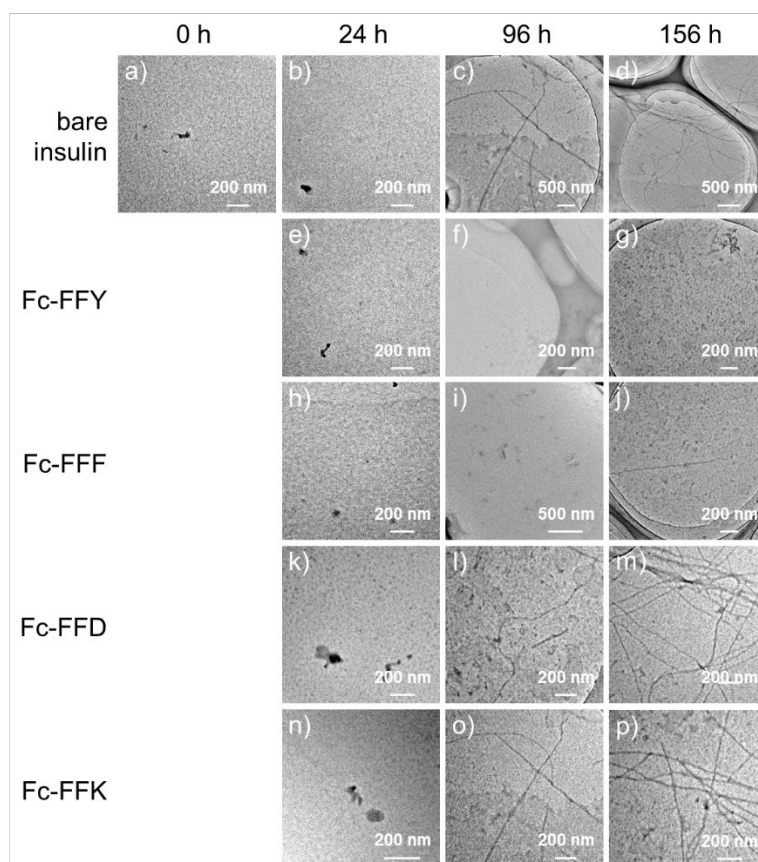
**Table S2.** Sizes of insulin fibrils incubated with Fc-peptides for 156 h.

	Peptide inhibitors concentration ( $\mu\text{M}$ )		
	100	200	300
Insulin + Fc-FFY			
Hydrodynamic diameter (nm)	490.47	248.43	181.60
Full Width at Half Maximum (nm)	473.66	215.66	101.85
Insulin + Fc-FFF			
Hydrodynamic diameter (nm)	583.37	344.55	284.45
Full Width at Half Maximum (nm)	313.09	369.26	118.68
Insulin + Fc-FFD			
Hydrodynamic diameter (nm)	523.49	442.41	328.68
Full Width at Half Maximum (nm)	431.13	416.98	102.45
Insulin + Fc-FFK			
Hydrodynamic diameter (nm)	713.88	483.25	349.96
Full Width at Half Maximum (nm)	529.91	370.14	329.60

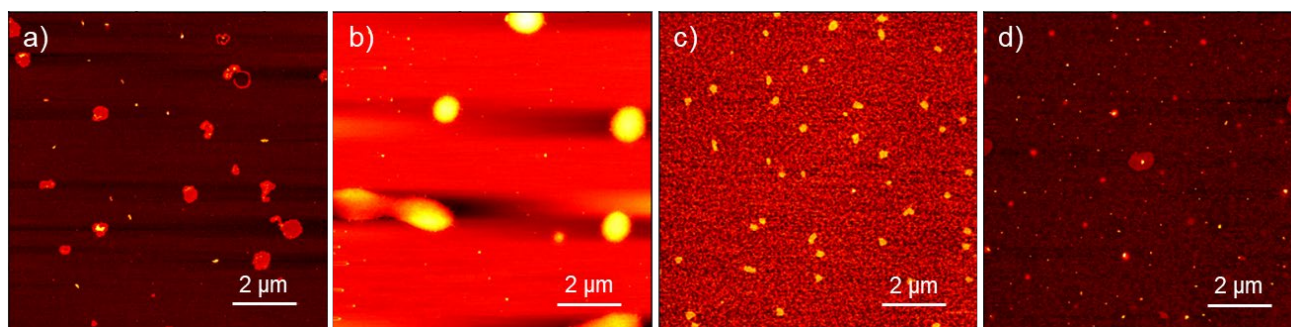
## 2. Supporting figures



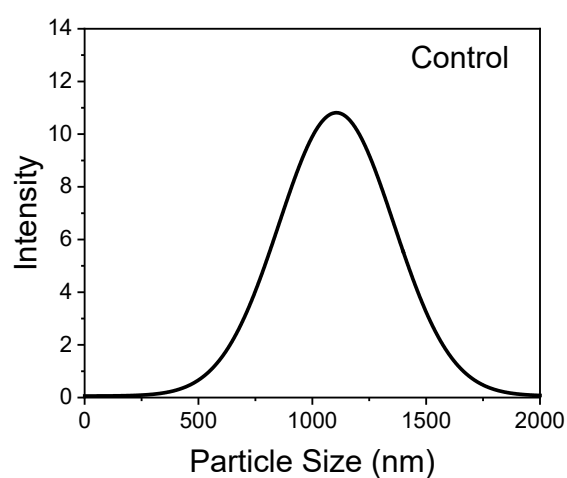
**Figure S1.** a) Fluorescence intensity of different concentrations of insulin incubated in 20% acetic acid solution at 60 °C. b) Fluorescence intensity of Fc-tripeptides and ThT incubated in 20% acetic acid solution at 60 °C.



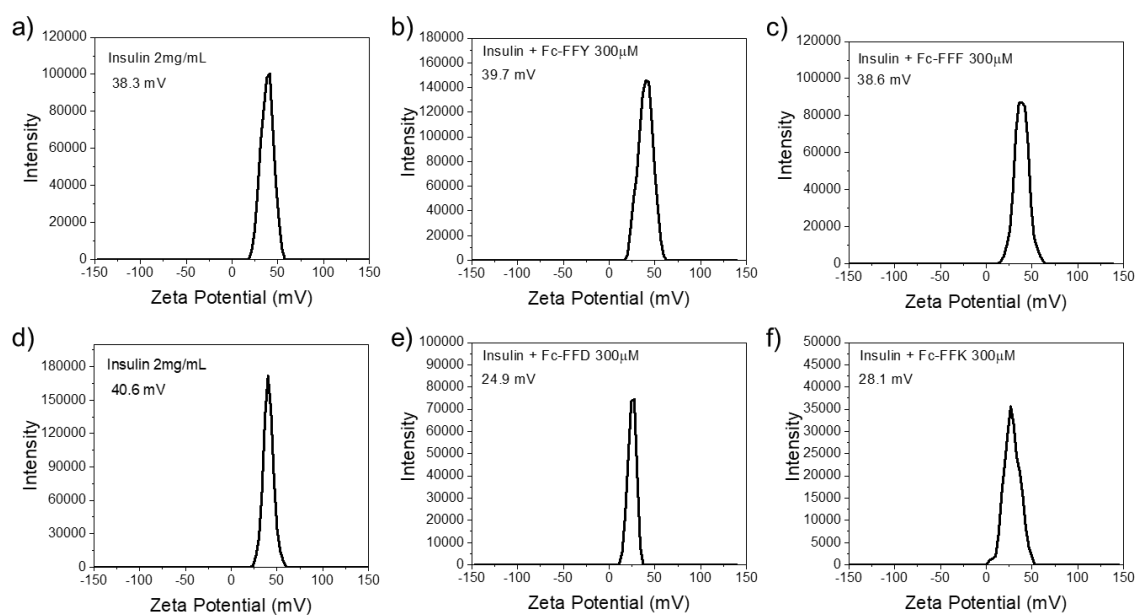
**Figure S2.** TEM images of insulin and insulin/Fc-tripeptides incubated for 156 h.



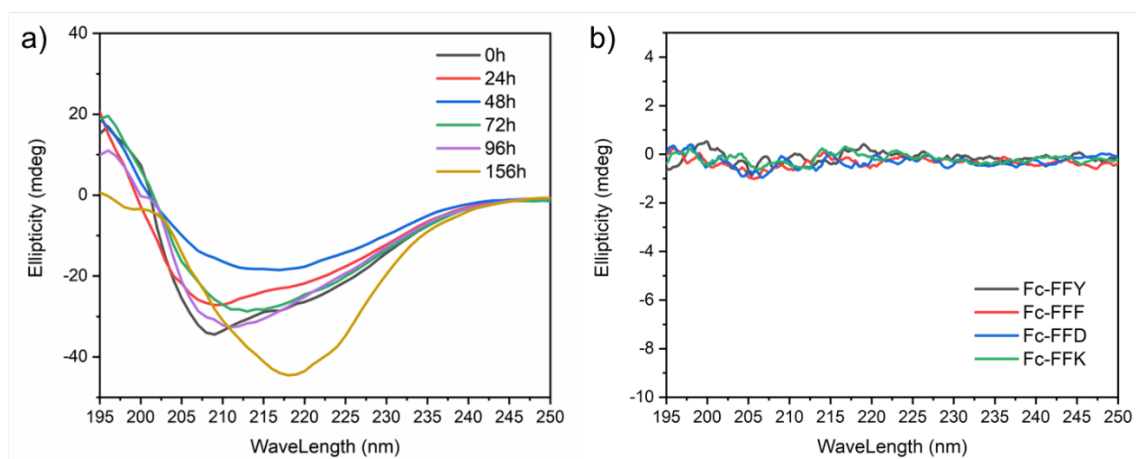
**Figure S3.** AFM images of Fc-tripeptides incubated for 156 h. (a) Fc-FFY, (b) Fc-FFF, (c) Fc-FFD or (d) Fc-FFK.



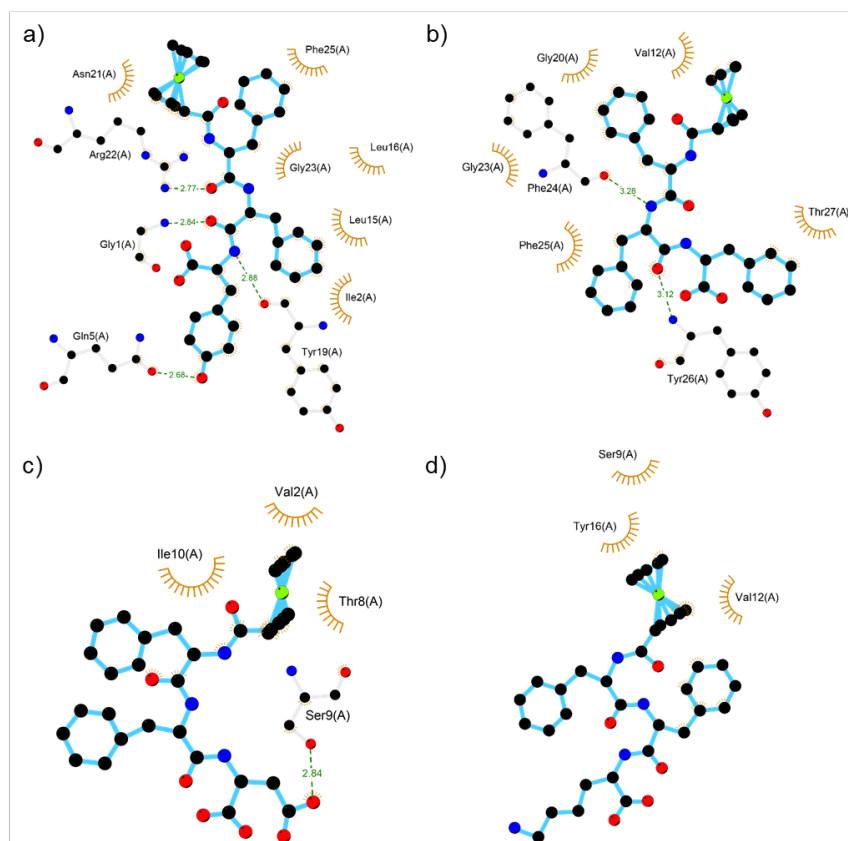
**Figure S4.** Size distribution of insulin incubated for 156 h.



**Figure S5.** Zeta potential values of (a, d) insulin, (b) 300 μM Fc-FFY, (c) 300 μM Fc-FFF, (e) 300 μM Fc-FFD and (f) 300 μM Fc-FFK in water solution.



**Figure S6.** (a) CD spectrum of insulin in 20% (v/v) acetic acid solution after incubation for up to 156 h. (b) CD spectrum of the Fc-tripeptides.



**Figure S7.** Intermolecular interactions between insulin and (a) Fc-FFY, (b) Fc-FFF, (c) Fc-FFD or (d) Fc-FFK were drawn with Ligplot+ program<sup>1</sup>. The atoms were color-coded as follows: C, black; O, red; N, blue. Bonds in Fc-tripeptides were colored cyan and those in insulin residues were colored gray. Hydrophobic interactions between Fc-tripeptides and insulins were shown in arc-shaped brown lines.

### **3. Supplementary methods**

#### **Transmission electron microscopy (TEM)**

The morphology of insulin samples incubated in the absence and presence of Fc-tripeptides were represented on a JEOL 100CX-II transmission electron microscope (JEOL Ltd, Japan) operated at an accelerating voltage of 80 kV. Each sample was diluted and dropped 10  $\mu$ L onto a carbon-coated copper grid and air-dried at room temperature. Then the grids with samples were negatively stained with 1% phosphotungstic acid solution for 4 min and examined.

#### **References**

1. R. A. Laskowski and M. B. Swindells, *J. Chem. Inf. Model.*, 2011, **51**, 2778-2786.