Supporting Information for

## An Ultrasensitive Lipid Droplet-Targeted NIR Emission Fluorescent Probe for Polarity Detection and Its Application in Liver Disease Diagnosis

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Scheme S1. Synthesis process of probe SSR-LDs.



Reagents and conditions: (i) Absolute ethanol, 85 °C, 4 h; (ii) anhydrous dichloride, 0 °C, under the protection of nitrogen, 8 h; (iii) 4-(dimethylamino)-benzaldehyde, toluene, room temperature, 24 h.



**Fig. S1.** Calculated orbital energy levels and electron density contours of HOMOs and LUMOs for S<sub>0</sub>-optimized geometry and S<sub>1</sub>-optimized geometry of **SSR-LDs** in 1,4-dioxane (a) and water (b), respectively. Transition properties (the energy gap  $E_{gap}$ , wavelength  $\lambda$ , and oscillator strength *f* ) for absorption and emission were also given.



Fig. S2. The visual change of solution color of SSR-LDs in different solvents (10  $\mu$ M). The solvents in sequence: toluene, 1,4-dioxane, tetrahydrofuran (THF), ethanol (ETOH), N,N-dimethylformamide (DMF), dimethyl sulfoxide (DMSO).



Fig. S3. Fluorescence emission spectra of SSR-LDs (10  $\mu$ M) in different ratios of mixed solvents of PBS and Glycerol.



Fig. S4. The pH adaptability tests of SSR-LDs (10  $\mu M)$  at  $\lambda_{ex}{=}580$  nm



**Fig. S5.** The photo-stability performance test curves of the probe **SSR-LDs** (10  $\mu$ M) in two different polarities solutions (the PBS buffer (red and black), and the mixed solution (1,4-dioxane: PBS = 9:1, v: v) (blue and green)) with different wavelengths (580 nm and 365 nm) of light radiation. A) The photo-stability tests of **SSR-LDs** in the PBS buffer (10 mM, pH 7.4) and the mixed solution (1,4-dioxane: PBS = 9:1, v: v) under light irradiation of 580 nm. B) The photo-stability tests of **SSR-LDs** in the PBS buffer (10 mM, pH 7.4) (red) and the mixed solution (1,4-dioxane: PBS = 9:1, v: v) under light irradiation of 365 nm.



**Fig. S6.** Cytotoxicity tests of four kinds of cell lines (Hela cells, HL7702 cells, 4T1 cells, and 3T3 cells).



Fig. S7. A) Confocal imaging of four different cell lines treated with SSR-LDs (10  $\mu$ M) and OA (400  $\mu$ M). ( $\lambda_{ex} = 580$ ,  $\lambda_{em} = 610-750$ ), Scale bar = 30  $\mu$ m; B) The relative fluorescence intensities of the red channels of A). (\*p<0.05).



**Fig. S8.** A) The confocal fluorescence imaging of probe **SSR-LDs** (10  $\mu$ M) in living Hela cells stimulated by CCCP ( $\lambda_{ex} = 580$ ,  $\lambda_{em} = 610-750$ ), Scale bar = 30  $\mu$ m; B) The relative fluorescence intensities of the red channels of A) (\*p<0.05).



**Fig. S9.** Co-localization imaging of **SSR-LDs** (10  $\mu$ M) and bodipy (200 nM) for imaging three cell lines (Hela cells, 3T3 cells, and 4T1 cells) pre-treated, Scale bar: 30  $\mu$ M. A) The co-localization imaging of Hela cells treated with **SSR-LDs** (10  $\mu$ M) and bodipy (200 nM); B) The co-localization imaging of 3T3 cells treated with **SSR-LDs** (10  $\mu$ M) and bodipy (200 nM); C) The co-localization imaging of 4T1 cells treated with **SSR-LDs** (10  $\mu$ M) and bodipy (200 nM); C) The co-localization imaging of 4T1 cells treated with **SSR-LDs** (10  $\mu$ M) and bodipy (200 nM); C) The red channel of **SSR-LDs**; 2) the green channel of bodipy; 3) the bright field channel; 4) the merge of 1-3); 5) Intensity profile of **SSR-LDs** in the red channel and bodipy in green channel. Green channel:  $\lambda_{ex}$ : 580 nm, collected 610 nm-750 nm.



Fig. S10. Co-localization imaging of SSR-LDs (10  $\mu$ M) and different organelle localization dyes in Hela cells, Scale bar: 30  $\mu$ M. A) The co-localization imaging of Hela cells treated with SSR-LDs (10  $\mu$ M) and ER tracker blue (200 nM). B) The co-localization imaging of Hela cells treated with SSR-LDs (10  $\mu$ M) and Hoechst 33342 (200 nM). C) The co-localization imaging of Hela cells treated with SSR-LDs (10  $\mu$ M) and Lyso Tracker green (200 nM). D) The co-localization imaging of Hela cells treated with SSR-LDs (10  $\mu$ M) and Mito Tracker green (200 nM).



**Fig. S11.** Co-localization imaging of **SSR-LDs** (10  $\mu$ M) and different organelle localization dyes in Hela cells pre-treated with OA (400  $\mu$ M). Scale bar: 30  $\mu$ M. A) The co-localization imaging of Hela cells pre-treated with OA (400  $\mu$ M), then treated with **SSR-LDs** (10  $\mu$ M) and ER tracker blue (200 nM). B) The co-localization imaging of Hela cells pre-treated with OA (400  $\mu$ M), then treated with **SSR-LDs** (10  $\mu$ M) and Hoechst 33342 (200 nM). C) The co-localization imaging of Hela cells pre-treated with OA (400  $\mu$ M), then treated with **SSR-LDs** (10  $\mu$ M) and Hoechst 33342 (200 nM). C) The co-localization imaging of Hela cells pre-treated with OA (400  $\mu$ M), then treated with **SSR-LDs** (10  $\mu$ M) and Lyso Tracker green (200 nM). D) The co-localization imaging of Hela cells pre-treated with **SSR-LDs** (10  $\mu$ M) and Mito Tracker green (200 nM).



Fig. S12. A) Time-dependent confocal fluorescence imaging of the probe SSR-LDs (10  $\mu$ M) at different temperatures in living Hela cells treated with OA (28 °C and 35 °C) ( $\lambda_{ex}$  = 580,  $\lambda_{em}$  = 610-750), Scale bar = 30  $\mu$ m. B) The relative fluorescence intensities of the red channels of A). (black represents 28 °C and red represents 35 °C) (\*p<0.05).



Fig. S13. A) The confocal fluorescence imaging of probe SSR-LDs (10  $\mu$ M) in living Hela cells stimulated by monensin ( $\lambda_{ex} = 580$ ,  $\lambda_{em} = 610-750$ ), Scale bar = 30  $\mu$ m; B) The relative fluorescence intensities of the red channels of A) (\*p<0.05).



Fig. S14. <sup>1</sup>H NMR (DMSO- $d_6$ ) spectrum of SSR-LDs.



Fig. S15. <sup>13</sup>C NMR (DMSO- $d_6$ ) spectrum of SSR-LDs.



Fig. S16. <sup>19</sup>F NMR (DMSO- $d_6$ ) spectrum of SSR-LDs.



**Fig. S17.** <sup>1</sup>H NMR (CDCl<sub>3</sub>-*d*) spectrum of compound **3**.



Fig. S18. <sup>1</sup>H NMR (CDCl<sub>3</sub>-d) spectrum of compound 4.



Fig. S19. HR-MS (ESI) spectrum of SSR-LDs, (M+H)<sup>+</sup>, 481.2219.