

**Artificial Light-Harvesting System with Sequential Energy Transfer for  
Information Dual Encryption and Anticounterfeiting**

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201620, China

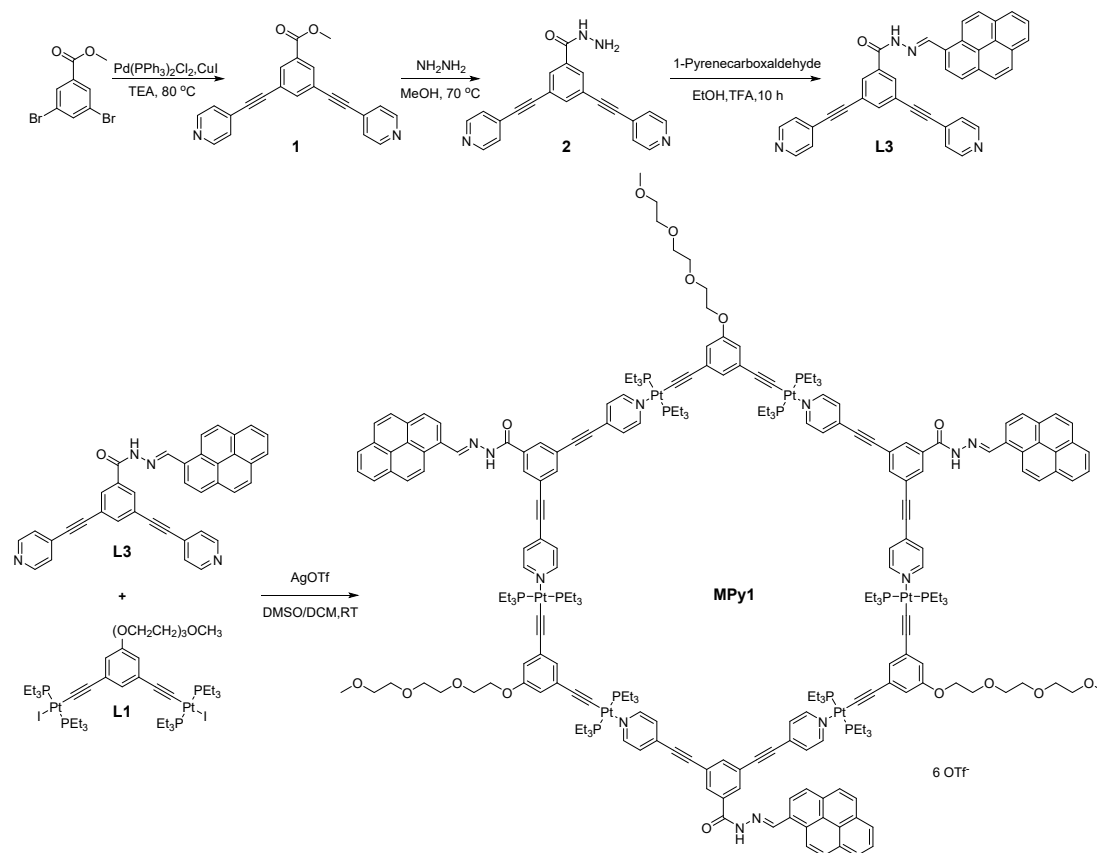
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## 1. General information

All reagents and solvents were commercially available and used without further purification. The compounds **L1**<sup>1</sup>, **L2**<sup>2</sup> and **1**<sup>3</sup> were prepared according to the published procedures. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded in CDCl<sub>3</sub> or DMSO-*d*<sub>6</sub> on Bruker Model Avance DMX 400 (400 MHz). Absorption spectra were recorded on a PERSEE model TU-1901 spectrophotometer. Fluorescence spectra were recorded on a F-7000 FL Spectrophotometer. The fluorescence lifetimes were measured employing time correlated single photon counting on a FLS980 instrument. The fluorescence quantum yields were measured on a PTI QM 40 instrument with the integrating sphere. Transmission electron microscopy (TEM) images were carried out on a JEOL JEM-2100 instrument. Scanning electron microscopy (SEM) images were recorded with a Hitachi S-4800 or SU8010 instruments. Dynamic light scattering (DLS) measurements were performed at a Malvern Instrument. Zeta-potential measurements were performed on a Zetasizer Nano Z apparatus at 25 °C. ESI-TOF-mass spectrum was recorded on a Micromass Quattro II triple-quadrupole mass spectrometer using electrospray ionization with a MassLynx operating system.

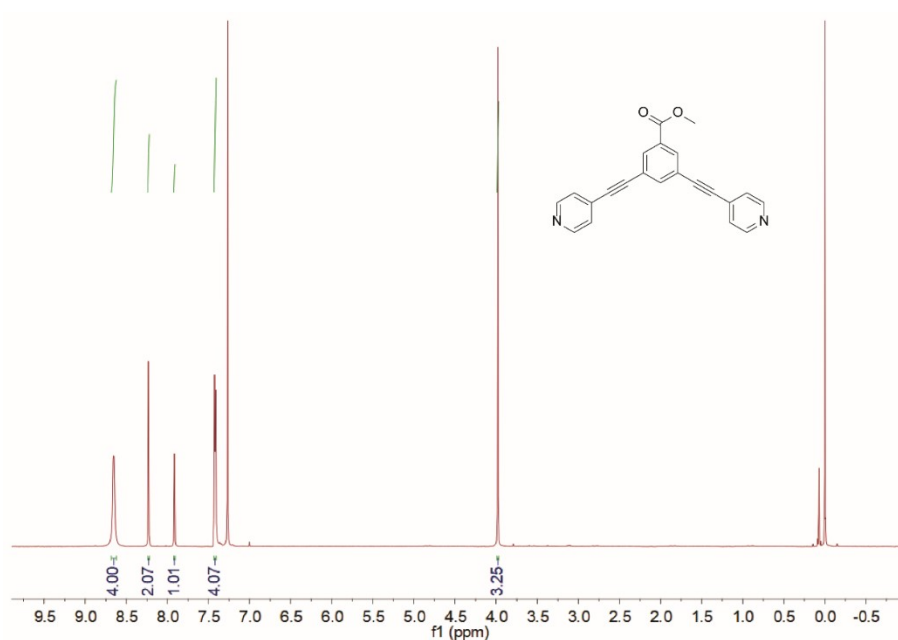
## 2. Synthesis of MPy1



**Figure S1.** Synthesis route of **MPy1**.

### Synthesis of **1**<sup>3</sup>

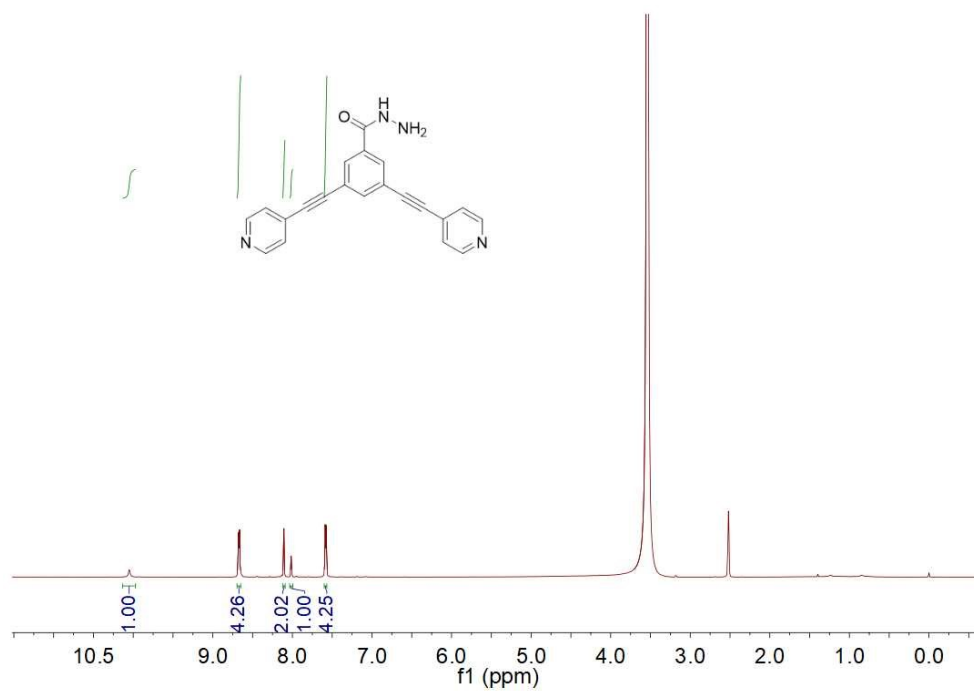
In a dry Schlenk tube, Pd(PPh<sub>3</sub>)Cl<sub>2</sub> (140 mg, 0.2 mmol, 0.1 eq) was solubilized in freshly distilled and degassed triethylamine (13 mL) at 80 °C. Methyl 3,5-dibromobenzoate (250 mg, 0.85 mmol), 3-ethynylpyridine (263 mg, 2.55 mmol) and CuI (32.4 mg, 0.17 mmol) were successively added, and the resulting mixture was heated to 80 °C under argon overnight. The solvent was removed in vacuo and the crude brown powder was purified by column chromatography on silica gel (PE: EA = 1:1) to give white solid (88 mg, 65 %). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.65 (d, *J* = 3.2 Hz, 4H), 8.23 (d, *J* = 1.4 Hz, 2H), 7.92 (d, *J* = 1.4 Hz, 1H), 7.42 (d, *J* = 5.6 Hz, 4H), 3.98 (s, 3H).



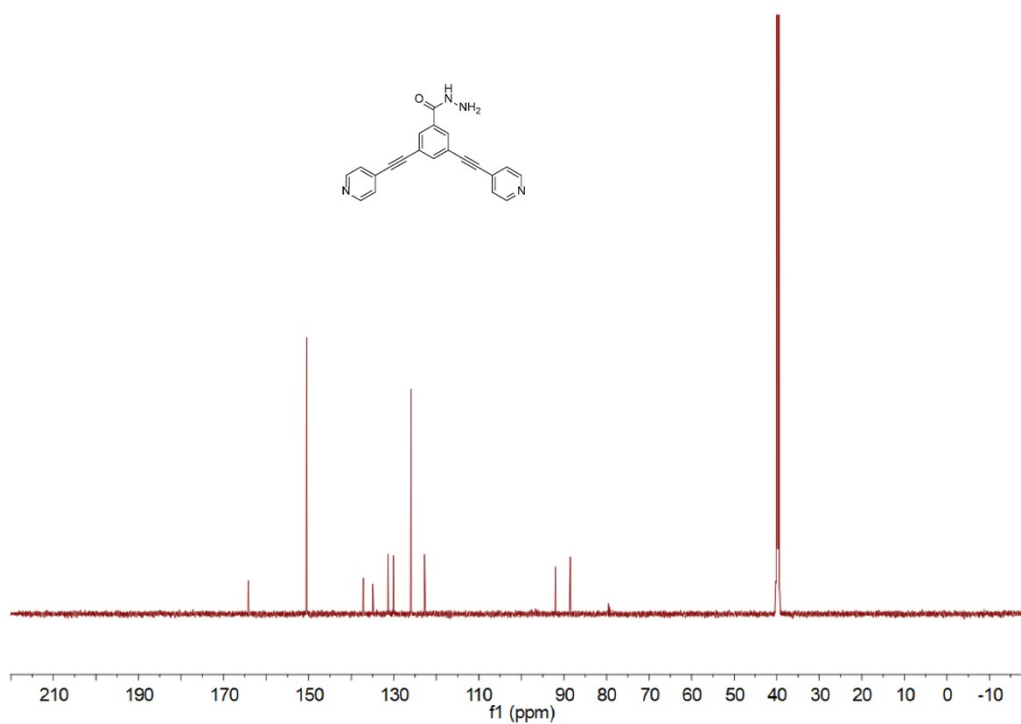
**Figure S2.**  $^1\text{H}$  NMR spectrum (400 MHz,  $\text{CDCl}_3$ , 298K) of compound **1**.

### Synthesis of **2**

Compound **1** (120 mg, 0.3546 mmol, 1 eq), hydrazine monohydrate (1 mL) and MeOH (10 mL) were mixed together and refluxed for 12 h at 70 °C under inert atmosphere. Then reaction was stopped and cooled to room temperature and methanol was evaporated. The product was extracted with dichloromethane (30 mL) and washed with water (3×50 mL) and brine solution (20 mL). The organic layer was dried over anhydrous  $\text{Na}_2\text{SO}_4$  and solvent was evaporated to get the crude product as light brown oil (400 mg, 80 %).  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO}-d_6$ )  $\delta$  10.02 (s, 1H), 8.68 (d,  $J=4.9\text{Hz}$ , 4H), 8.12 (s, 2H), 8.01 (s, 1H), 7.58 (d,  $J=4.9\text{Hz}$ , 4H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO}-d_6$ )  $\delta$  164.22, 150.48, 137.12, 134.94, 131.38, 130.11, 125.95, 122.77, 92.00, 88.48. ESI-MS:  $m/z$  Calcd for  $\text{C}_{21}\text{H}_{14}\text{N}_4\text{O}$   $[\text{M}+\text{H}]^+$  339.1, found 339.1.

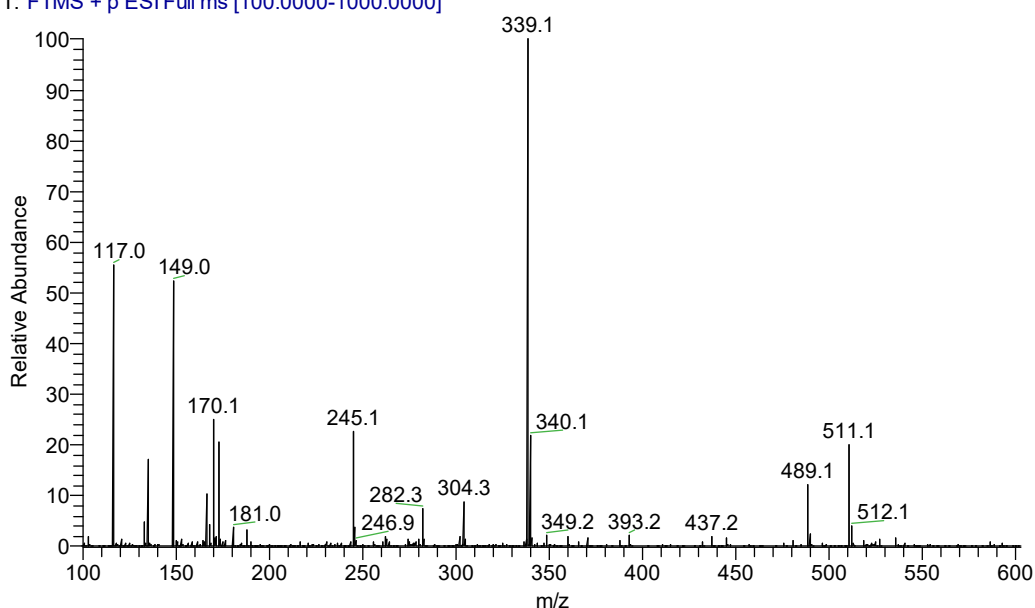


**Figure S3.**  $^1\text{H}$  NMR spectrum of **2** in  $\text{DMSO-}d_6$ .



**Figure S4.**  $^{13}\text{C}$  NMR spectrum of **2** in  $\text{DMSO-}d_6$ .

3 #10 RT: 0.08 AV: 1 NL: 4.76E7  
T: FTMS + p ESI Full ms [100.0000-1000.0000]



**Figure S5.** ESI spectrum of compound **2**.

### Synthesis of L3

To an ethanol solution of **2** (0.4064 mmol, 137 mg in 5 mL dry ethanol) was added a solution of 1-pyrenecarboxaldehyde (0.4064 mmol, 100 mg in 10 mL ethanol). The reaction mixture was stirred for 16 h. Then the mixture was allowed to cool to room temperature to produce a yellow precipitate. The crude product was purified by recrystallization with ethanol to afford compound **L3** as a yellow-green solid (160 mg). Yield: 64.8%.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  12.28 (s, 1H), 9.56 (s, 1H), 8.84 (d,  $J = 9.3$  Hz, 1H), 8.72 (d,  $J = 4.4$  Hz, 4H), 8.62 (d,  $J = 7.9$  Hz, 1H), 8.41 (d,  $J = 7.2$  Hz, 4H), 8.34 – 8.25 (m, 4H), 8.16 (s, 2H), 7.65 (d,  $J = 4.7$  Hz, 4H).  $^{13}\text{C}$  NMR (101 MHz,  $\text{DMSO-}d_6$ )  $\delta$  164.05, 150.09, 138.55, 136.87, 131.66, 131.22, 130.94, 130.42, 129.68, 127.83, 127.70, 127.60, 126.97, 126.09, 125.91, 125.71, 125.51, 125.40, 124.83, 124.23, 123.40, 123.20, 86.30, 80.10. ESI-MS: m/z Calcd for  $\text{C}_{38}\text{H}_{22}\text{N}_4\text{O}$   $[\text{M}+\text{H}]^+$  551.2, found 551.2.

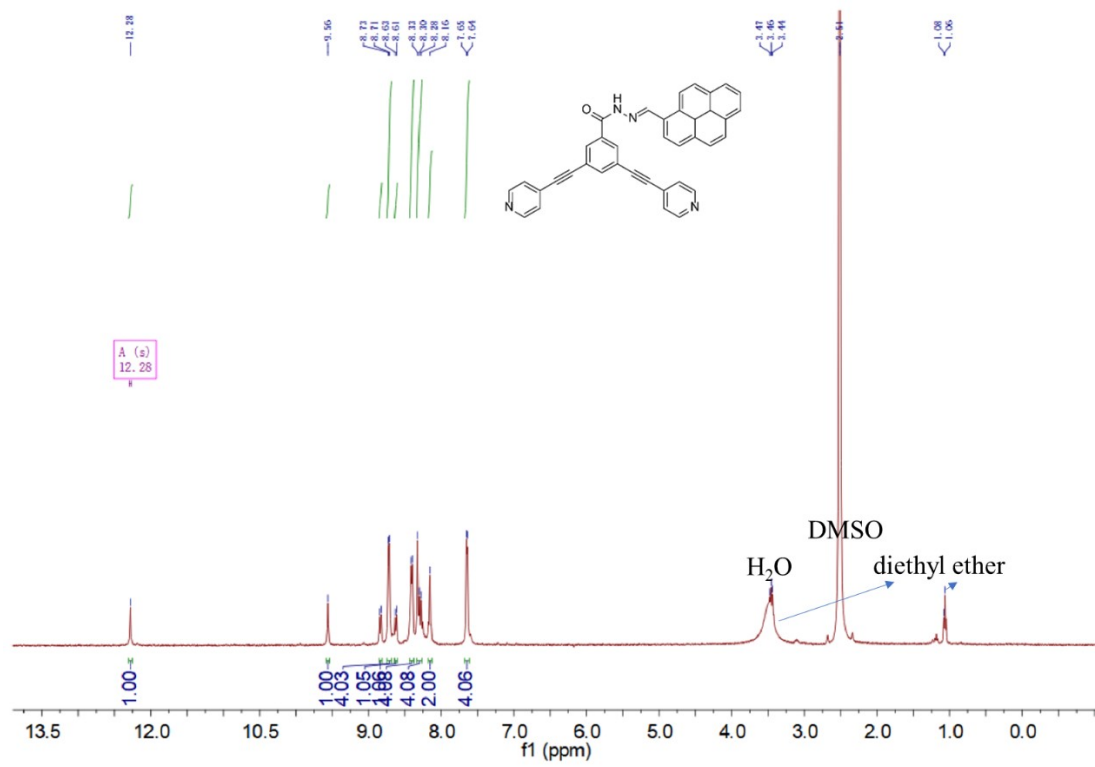


Figure S6. <sup>1</sup>H NMR spectrum of L3 in DMSO-*d*<sub>6</sub>.

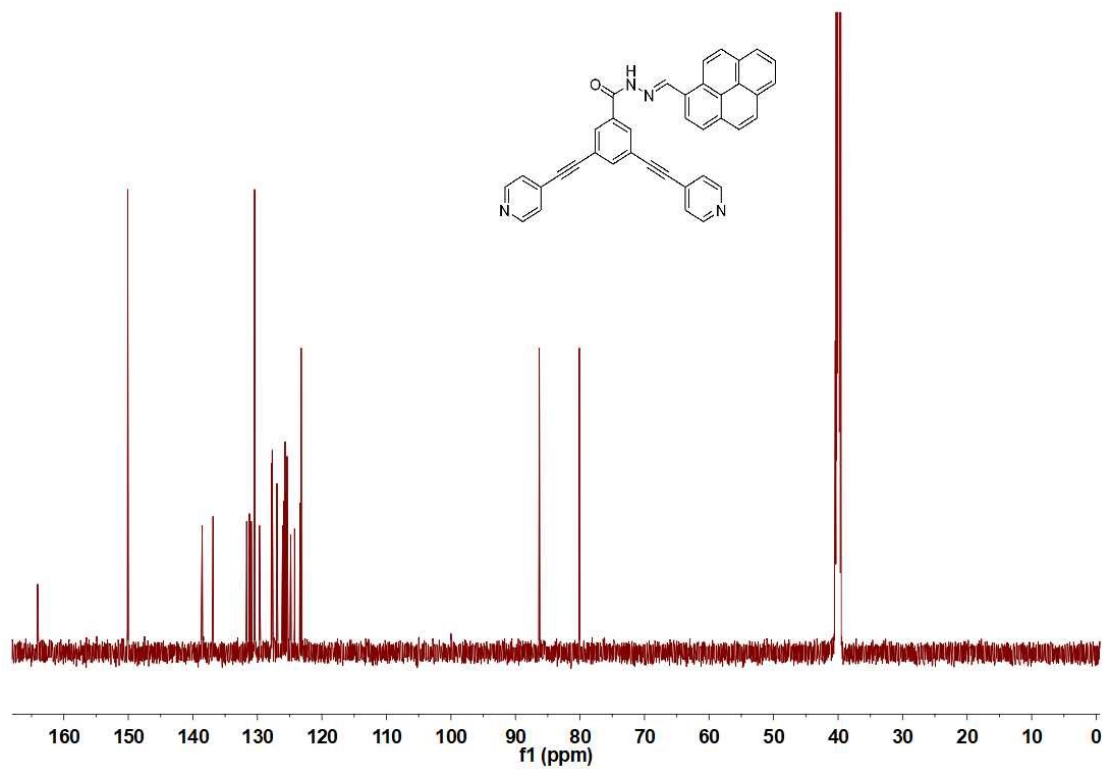
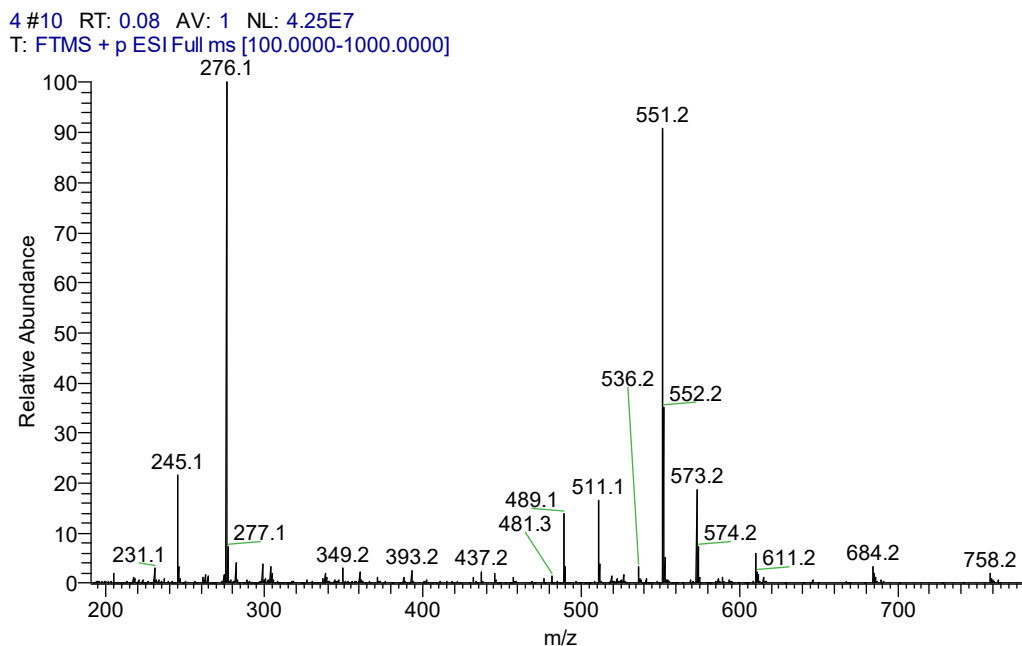


Figure S7. <sup>13</sup>C NMR spectrum of L3 in DMSO-*d*<sub>6</sub> at 323 K.

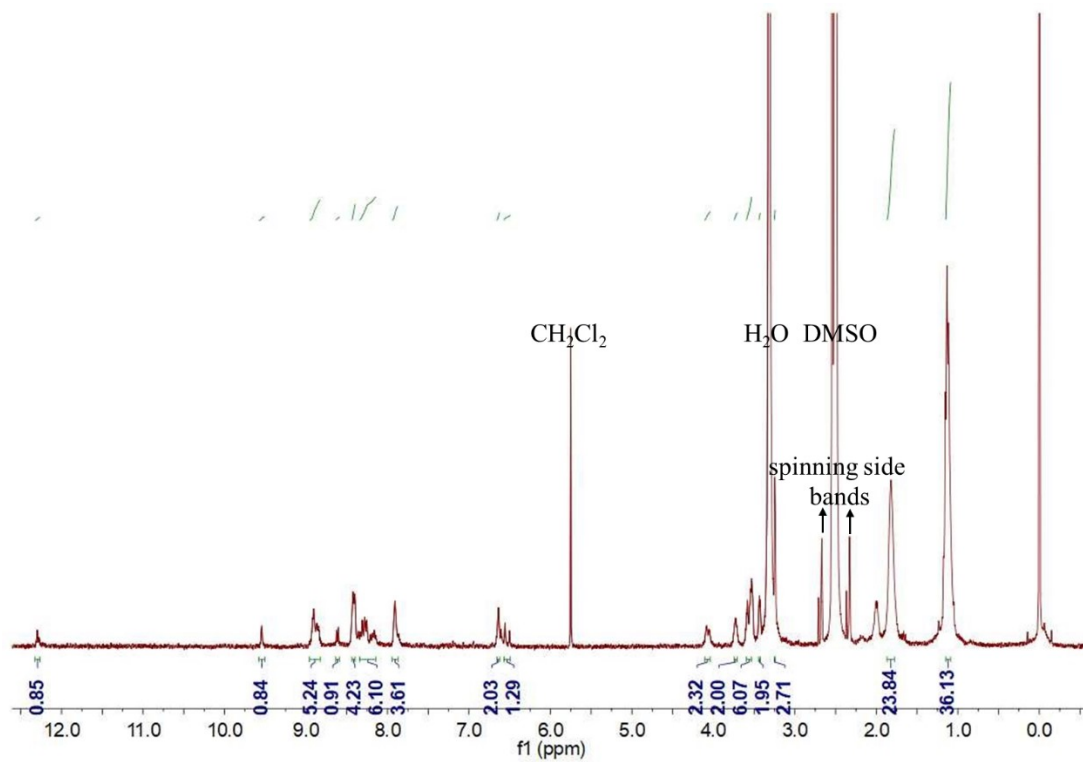




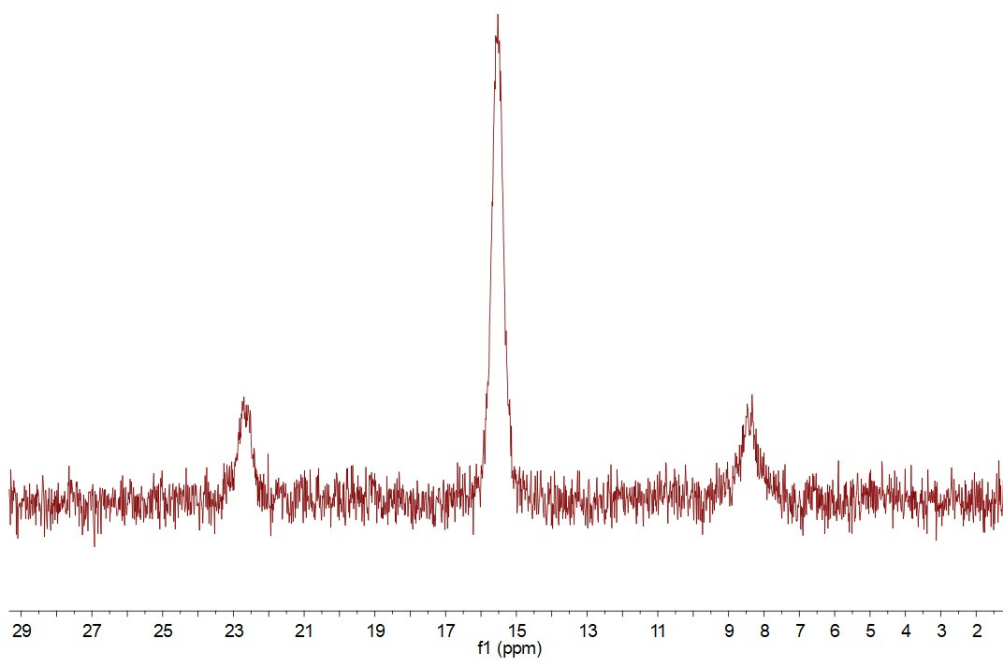
**Figure S8.** ESI spectrum of compound **L3**.

### Synthesis of MPy1

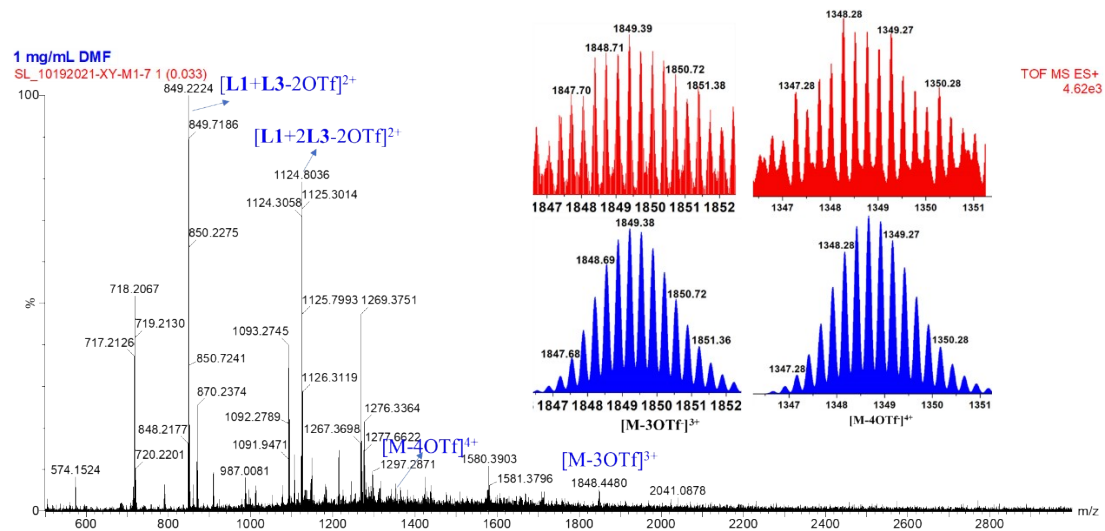
**L1** (10 mg, 7.0  $\mu\text{mol}$ ), AgOTf (5.5 mg, 21.0  $\mu\text{mol}$ ) and **L3** (4 mg, 7.0  $\mu\text{mol}$ ) were dissolved in the mixture of DMSO and DCM (2.0 mL,  $V_{\text{DCM}}:V_{\text{DMSO}} = 10:1$ ) in a 20 mL glass vial. The reaction mixture was allowed to stir for 6 h at room temperature, and the reaction mixture was centrifuged at 3400 r/h for 20 min. To the resulting homogeneous solution, diethyl ether was added to precipitate the product, which was then isolated and dried under reduced pressure and re-dissolved in DMSO- $d_6$  for characterization.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  12.30 (s, 1H), 9.54 (s, 1H), 8.95 – 8.82 (m, 5H), 8.61 (d,  $J = 8.7$  Hz, 1H), 8.41 (d,  $J = 9.4$  Hz, 4H), 8.34 – 8.15 (m, 6H), 7.91 (s, 4H), 6.64 (s, 2H), 6.53 (s, d,  $J = 22.3$  Hz, 1H), 4.08 (d,  $J = 5.8$  Hz, 2H), 3.73 (s, 2H), 3.60 – 3.50 (m, 6H), 3.44 – 3.42 (m, 2H), 3.25 (s, 3H), 1.82 (s, 24H), 1.19 – 1.06 (m, 36H).  $^{31}\text{P}$  NMR (121.4 MHz, DMSO- $d_6$ )  $\delta$  (ppm) 15.52 (s,  $^{195}\text{Pt}$  satellites,  $^1J_{\text{Pt-P}} = 1729.95$  Hz). ESI-TOF-MS:  $\text{C}_{243}\text{H}_{300}\text{F}_{18}\text{N}_{12}\text{O}_{33}\text{P}_{12}\text{Pt}_6\text{S}_6$  m/z [**M**-3OTf] $^{3+}$  1849.39; [**M**-4OTf] $^{4+}$  1348.28.



**Figure S9.**  $^1\text{H}$  NMR spectrum of MPy1 in DMSO- $d_6$ .

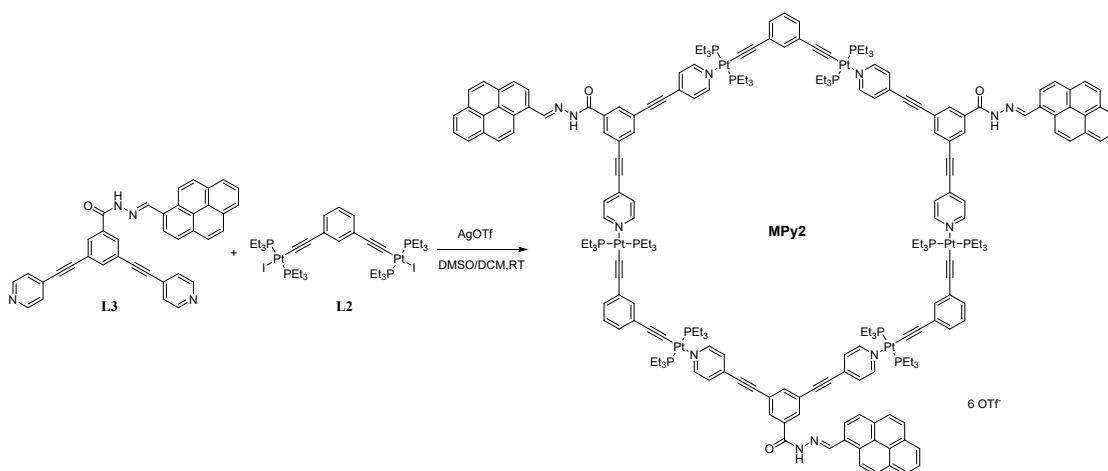


**Figure S10.**  $^{31}\text{P}$  { $^1\text{H}$ } NMR spectrum of MPy1 in DMSO- $d_6$ .



**Figure S11.** Experimental (red) and calculated (blue) electrospray ionization mass spectra of MPy1.

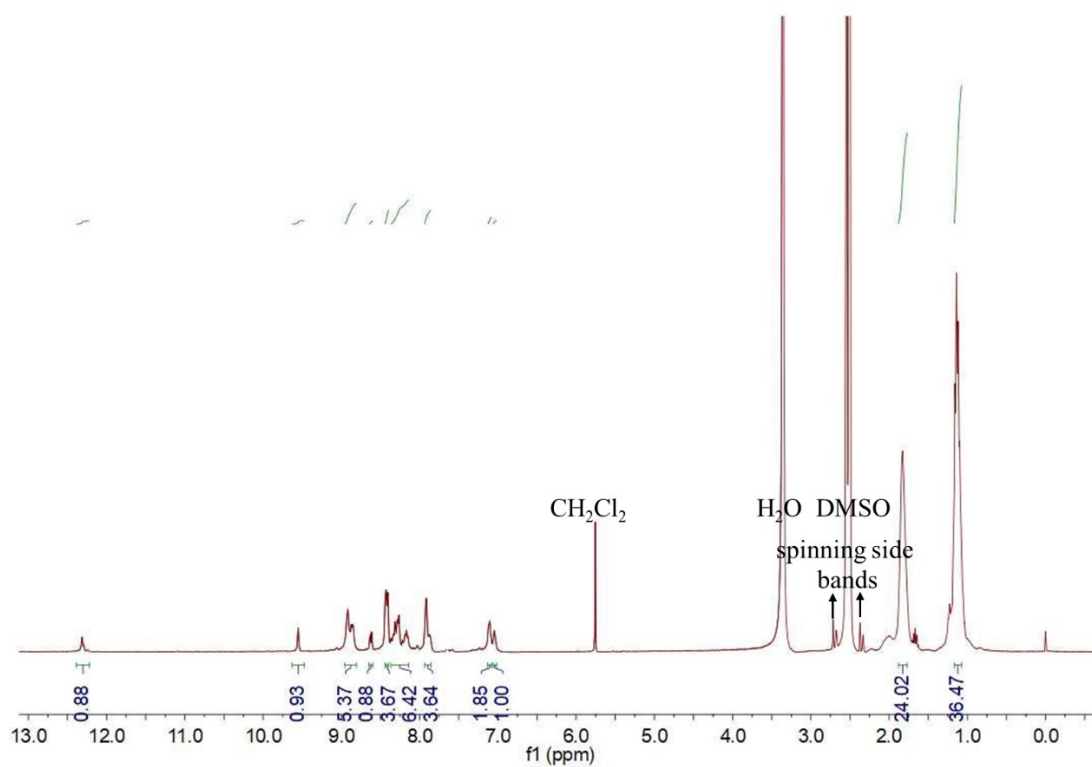
### 3. Synthesis of MPy2



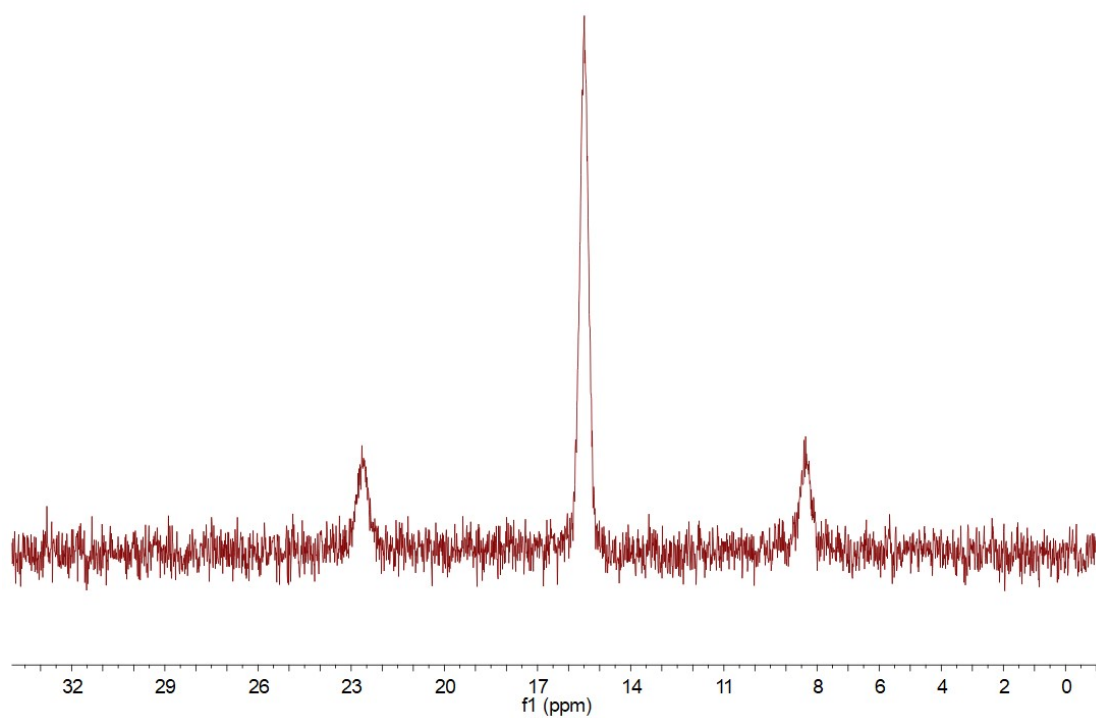
**Figure S12.** Synthesis route of MPy2.

L2 (10 mg, 7.0  $\mu\text{mol}$ ), AgOTf (5.5 mg, 21.0  $\mu\text{mol}$ ) and L3 (4 mg, 7.0  $\mu\text{mol}$ ) were dissolved in the mixture of DMSO and DCM (2.0 mL,  $V_{\text{DCM}}:V_{\text{DMSO}} = 10:1$ ) in a 20 mL glass vial. The reaction mixture was allowed to stir for 6 h at room temperature, and the reaction mixture was centrifuged at 3400 r/h for 20 min. To the resulting homogeneous solution, diethyl ether was added to precipitate the product, which was then isolated and dried under reduced pressure and re-dissolved in DMSO- $d_6$  for

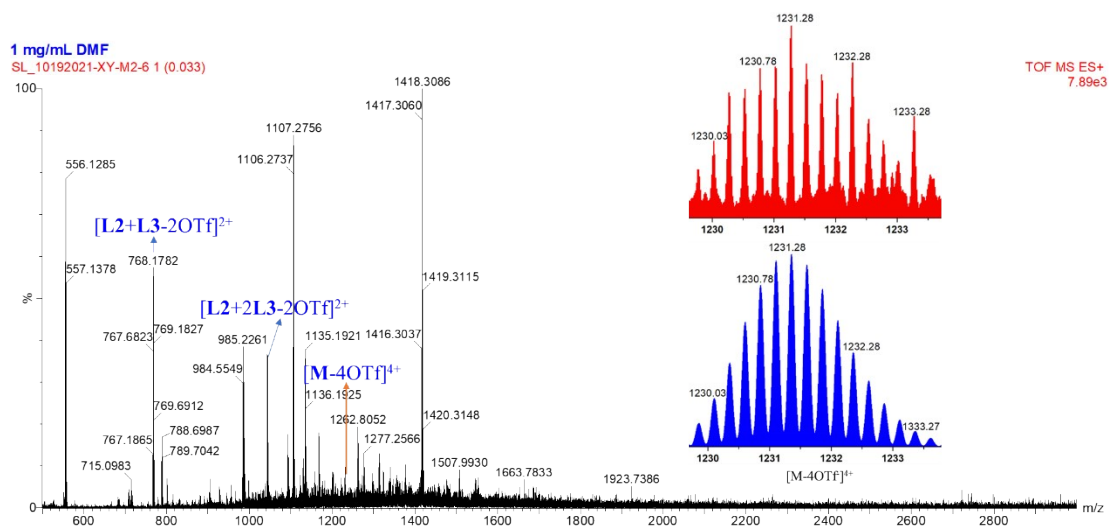
characterization.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  12.31 (s, 1H), 9.55 (s, 1H), 8.96 – 8.81 (m, 5H), 8.62 (d,  $J = 8.1$  Hz, 1H), 8.42 (d,  $J = 9.1$  Hz, 4H), 8.37 – 8.14 (m, 6H), 7.89 (d,  $J = 15.6$  Hz, 4H), 7.10 (s, 2H), 7.04 (s, 1H), 1.83 (s, 24H), 1.18 – 1.07 (m, 36H).  $^{31}\text{P}$  NMR (121.4 MHz,  $\text{DMSO-}d_6$ )  $\delta$  (ppm) 15.50 (s,  $^{195}\text{Pt}$  satellites,  $^1J_{\text{Pt-P}} = 1728.74$  Hz). ESI-TOF-MS:  $\text{C}_{222}\text{H}_{258}\text{F}_{18}\text{N}_{12}\text{O}_{21}\text{P}_{12}\text{Pt}_6\text{S}_6$   $m/z$   $[\text{M-4OTf}]^{4+}$  1231.28;  $[\text{L1}+2\text{L3-2OTf}]^{2+}$  1043.77;  $[\text{L1}+\text{L3-2OTf}]^{2+}$  768.18. (Where M represents the intact assembly).



**Figure S13.**  $^1\text{H}$  NMR spectrum of **MPy2** in  $\text{DMSO-}d_6$ .

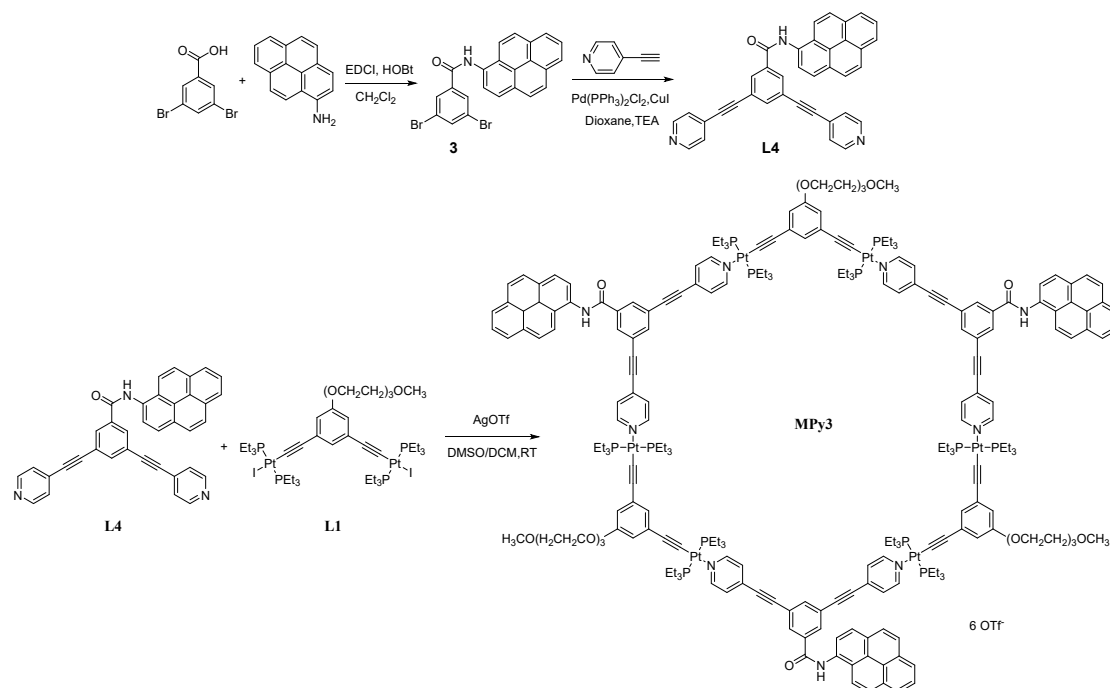


**Figure. S14.**  $^{31}\text{P}\{^1\text{H}\}$  NMR spectrum of **MPy2** in  $\text{DMSO-}d_6$ .



**Figure S15.** Experimental (red) and calculated (blue) electrospray ionization mass spectra of **MPy2**.

## 4. Synthesis of MPy3

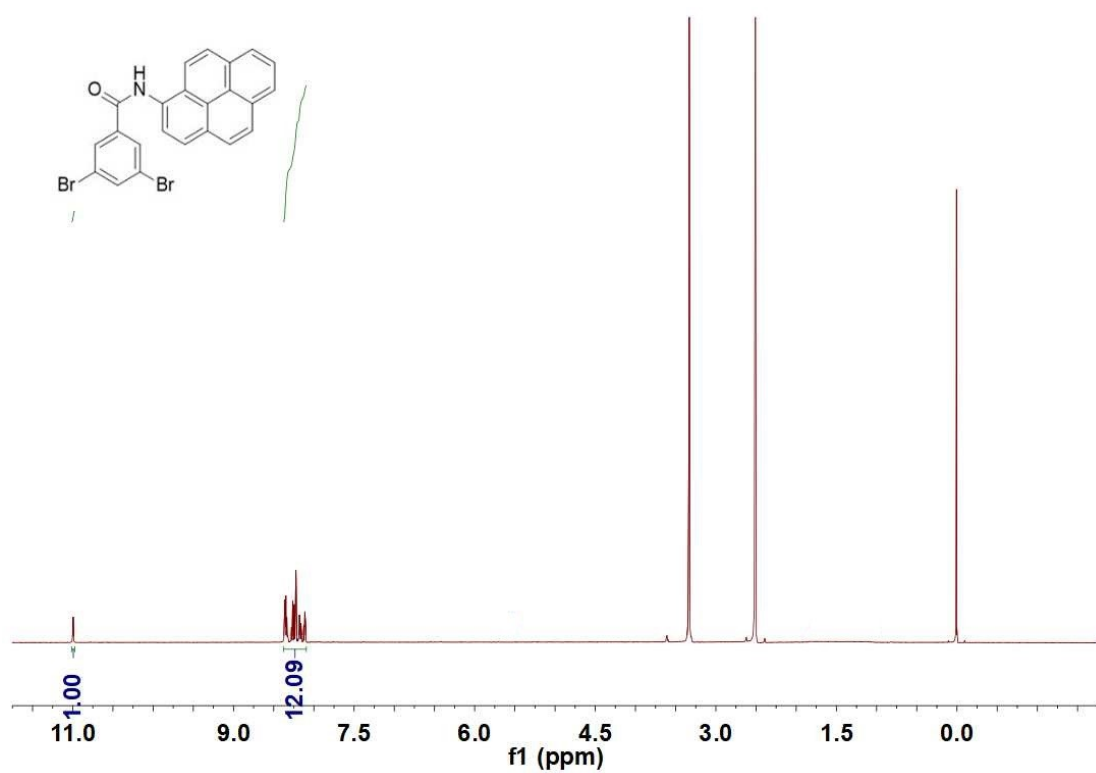


**Figure S16.** Synthesis route of **MPy3**.

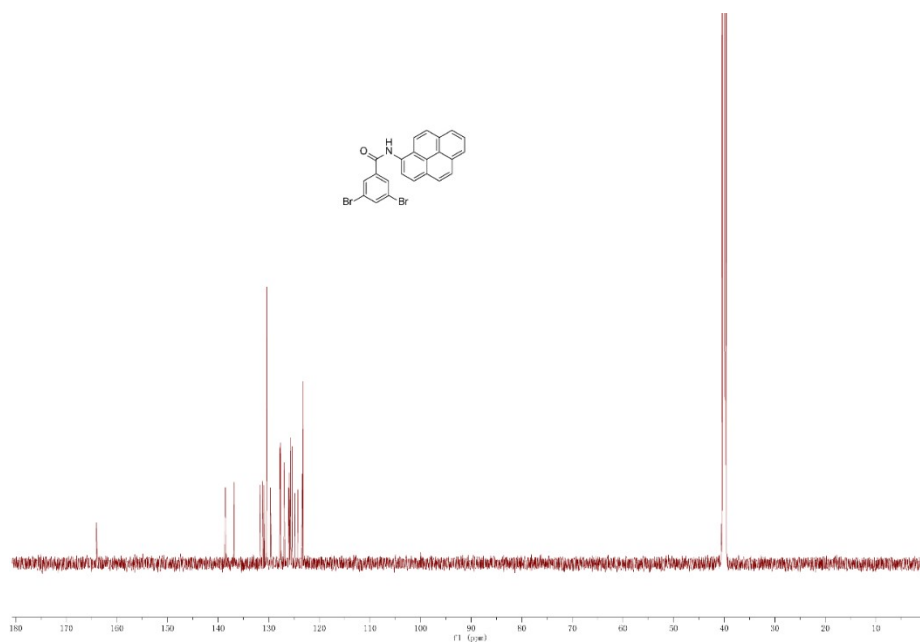
### Synthesis of **3**

3,5-dibromobenzoic acid (140 mg, 1 eq), HOBT (148.6 mg, 1.1 eq), 1-ethyl- (3-dimethylaminopropyl) carbonyl diimidehydrochloride (EDCI) (210 mg, 1.1 eq) and triethylamine (300 mg, 3 eq) were added into a 50 mL two-necked flask and pumped under the protection of argon for three times. Then the newly steamed methylene chloride (20 mL) was added at 0 °C. After stirring for 30 min in ice bath, 1-aminopyrene (93 mg, 0.8 mmol, 0.8 eq) dichloromethane solution was added drop by drop. After dropping, the solution was removed from the ice bath and reacted at room temperature for 24 h. The solvent was removed by reduced pressure, and the resulting mixture was dissolved with ethyl acetate and washed with water to remove water-soluble impurities. The residue was purified by column chromatography (silica, petroleum:ethyl acetate = 1:2 as the eluent) to obtain brown oily compound. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 10.99 (s, 1H), 8.36-8.10 (m, 12H). <sup>13</sup>C NMR (101 MHz, DMSO-*d*<sub>6</sub>) δ 164.05, 138.55, 136.87, 131.66, 131.22, 130.94, 130.43, 129.68, 127.83,

127.70, 127.60, 126.97, 126.09, 125.92, 125.71, 125.51, 125.40, 124.83, 124.23, 123.40, 123.20. ESI-MS:  $m/z$  Calcd for  $C_{23}H_{13}Br_2NO$   $[M+H]^+$  477.9, found 477.9.

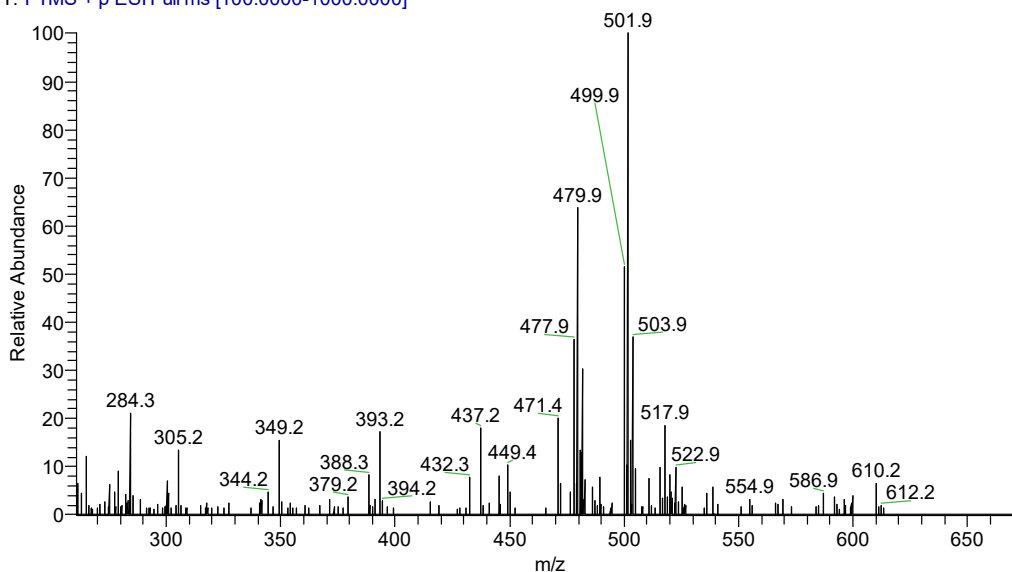


**Figure S17.**  $^1H$  NMR spectrum of **3** in  $DMSO-d_6$ .



**Figure S18.**  $^{13}C$  NMR spectrum of **3** in  $DMSO-d_6$ .

1 #14 RT: 0.12 AV: 1 NL: 5.69E6  
T: FTMS + p ESI Full ms [100.0000-1000.0000]

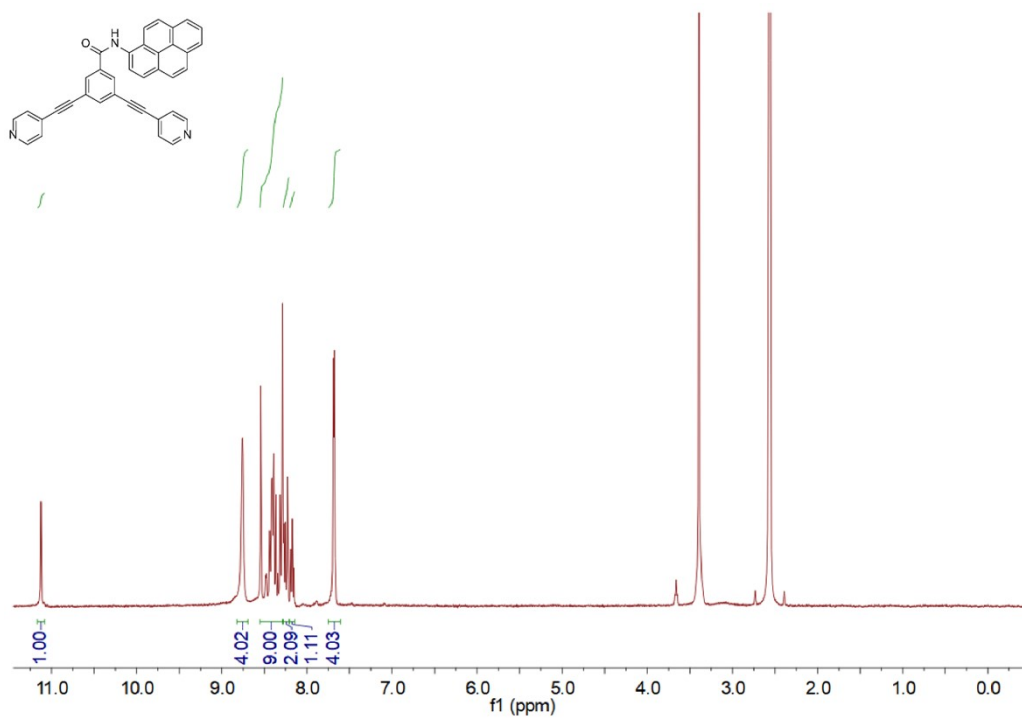


**Figure S19.** ESI spectrum of compound **3**.

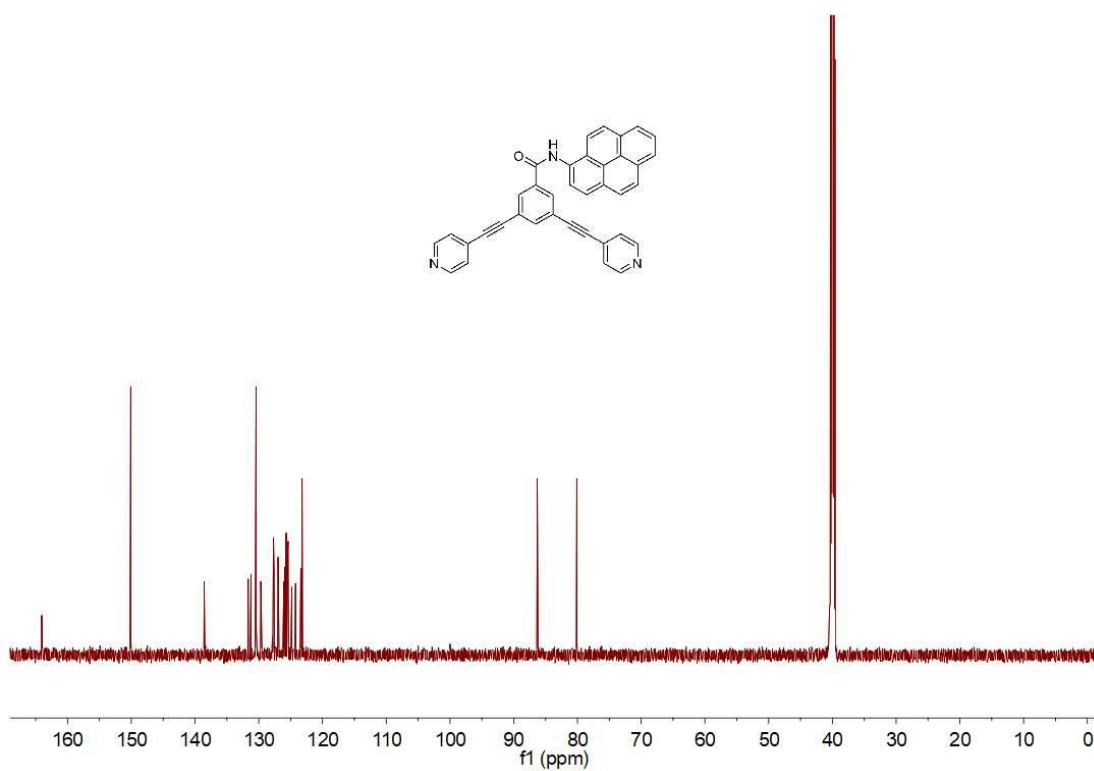
### Synthesis of L4

A Schlenk tube was charged with **3** (250 mg, 0.85 mmol), 4-ethynylpyridine hydrochloride (263 mg, 2.55 mmol), Pd(PPh<sub>3</sub>)Cl<sub>2</sub> (140 mg, 0.2 mmol), and CuI (32.4 mg, 0.17 mmol). The mixture was placed under nitrogen atmosphere, and anhydrous triethylamine (15 mL) and 1,4-dioxane (10 mL) were added by syringe. The reaction was heated at 85 °C for 48 h under nitrogen, then cooled down to room temperature and the solvents were removed under reduced pressure. The residue was dissolved in dichloromethane (200 mL) and washed by water (150 mL). After drying over Na<sub>2</sub>SO<sub>4</sub>, the solvent was removed and the residue was purified by column chromatography on silica gel (EA/petroleum ether = 1:1) to afford **L4** as a yellow powder. Yield: 187 mg, 59%. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 11.12 (s, 1H), 8.76 (s, 4H), 8.55-8.29 (m, 9H), 8.27-8.21 (m, 2H), 8.18 (d, *J* = 7.6 Hz, 1H), 7.68 (d, *J* = 5.0 Hz, 4H). <sup>13</sup>C NMR (400 MHz, DMSO-*d*<sub>6</sub>) δ 164.05, 150.09, 138.55, 131.66, 131.22, 130.42, 129.68, 127.83, 127.70, 127.60, 126.97, 126.09, 125.91, 125.71, 125.51, 125.40, 124.83, 124.23, 123.40, 123.20, 86.30, 80.10. ESI-MS: *m/z* Calcd for C<sub>37</sub>H<sub>21</sub>N<sub>3</sub>O [M+H]<sup>+</sup> 524.2, found 524.2.

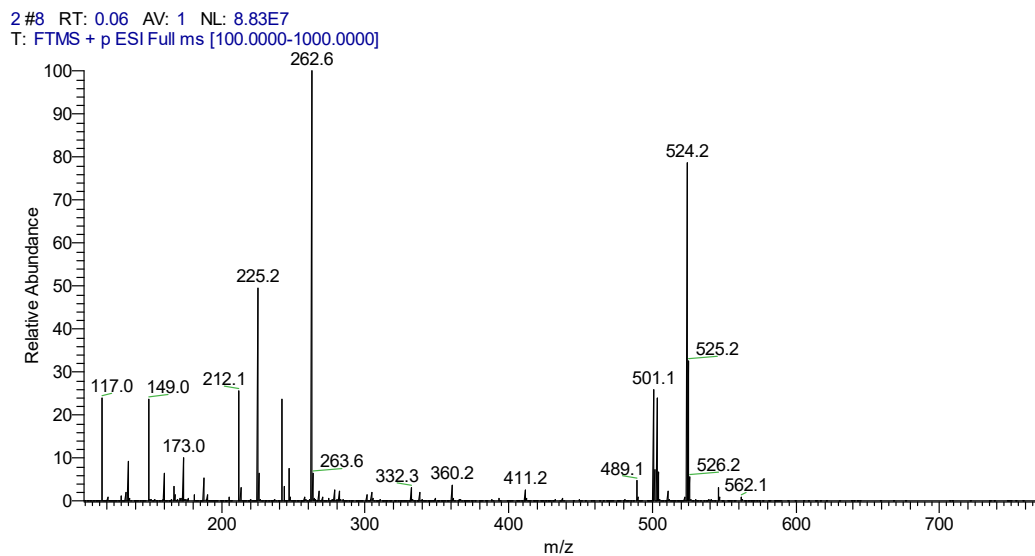




**Figure S20.** <sup>1</sup>H NMR spectrum of L4 in DMSO-*d*<sub>6</sub>.



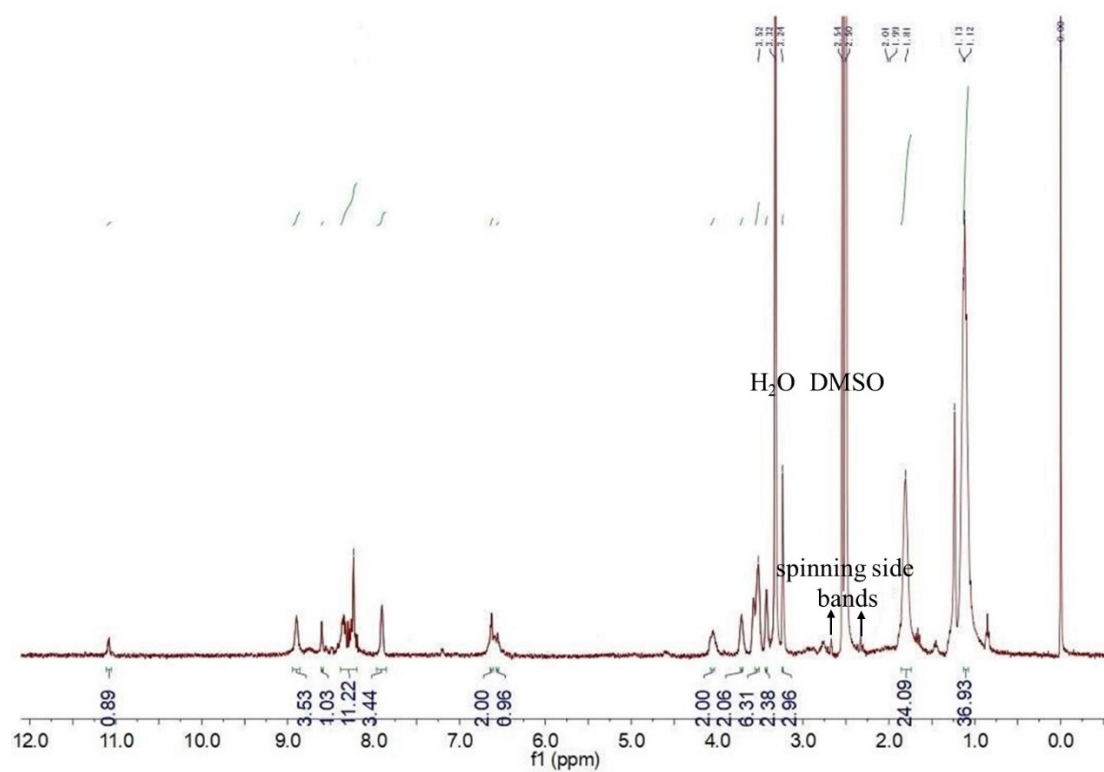
**Figure S21.** <sup>13</sup>C NMR spectrum of L4 in DMSO-*d*<sub>6</sub>.



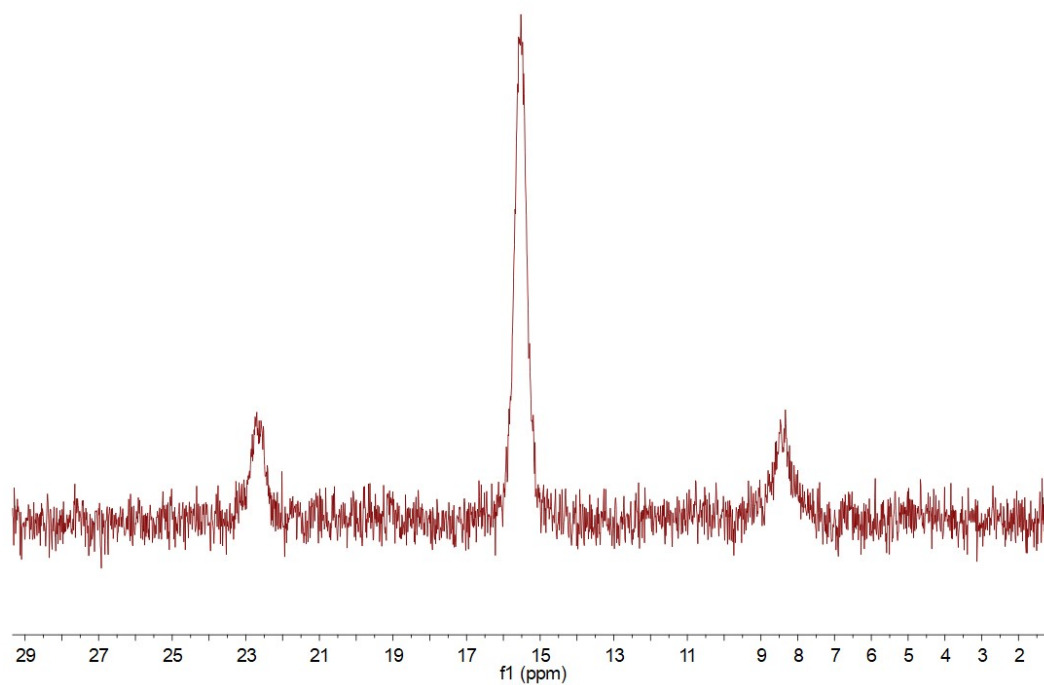
**Figure S22.** ESI spectrum of compound **L4**.

### Synthesis of MPy3

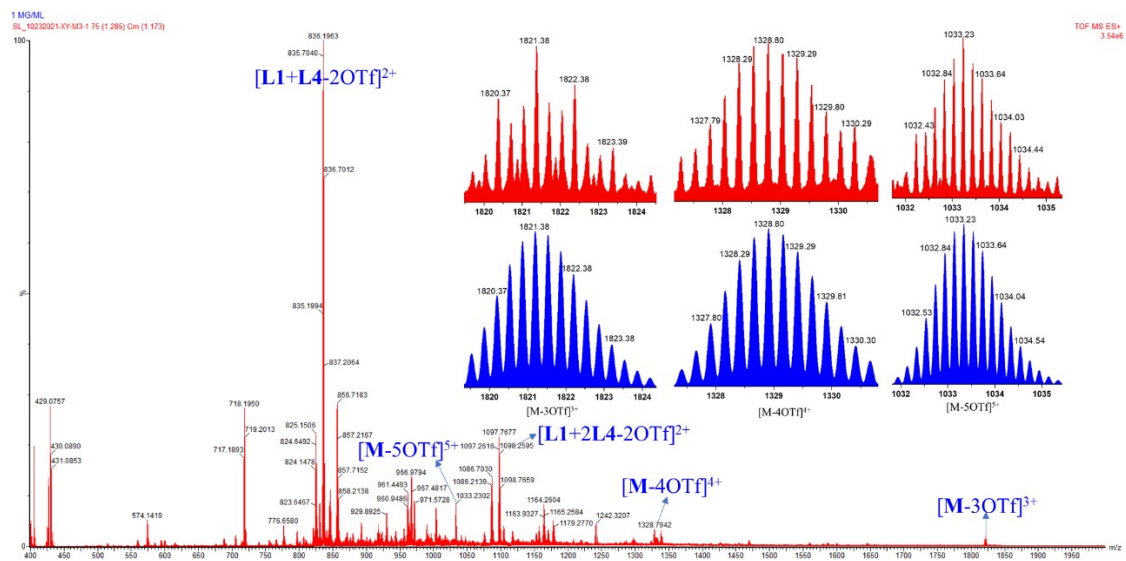
**L1** (10 mg, 7.0  $\mu\text{mol}$ ), AgOTf (5.5 mg, 21.0  $\mu\text{mol}$ ) and **L4** (4 mg, 7.0  $\mu\text{mol}$ ) were dissolved in the mixture of DMSO and DCM (2.0 mL,  $V_{\text{DCM}}:V_{\text{DMSO}}=10:1$ ) in a 20 mL glass vial. The reaction mixture was allowed to stir for 6 h at room temperature, and the reaction mixture was centrifuged at 3400 r/h for 20 min. To the resulting homogeneous solution, diethyl ether was added to precipitate the product, which was then isolated and dried under reduced pressure and re-dissolved in DMSO- $d_6$  for characterization.  $^1\text{H}$  NMR (400 MHz, DMSO- $d_6$ )  $\delta$  11.08 (s, 1H), 8.90 (s, 4H), 8.60 (s, 1H), 8.4 – 8.2 (m, 11H), 7.90 (s, 4H), 6.63 (s, 2H), 6.56 (s, 1H), 4.05 (s, 2H), 3.72 (s, 2H), 3.61 – 3.49 (m, 6H), 3.43 (s, 2H), 3.24 (s, 3H), 1.81 (s, 24H), 1.15 – 1.05 (m, 36H).  $^{31}\text{P}$  NMR (121.4 MHz, DMSO- $d_6$ )  $\delta$  (ppm) 15.52 (s,  $^{195}\text{Pt}$  satellites,  $^1J_{\text{Pt-P}} = 1731.16$  Hz).  $\text{C}_{240}\text{H}_{297}\text{F}_{18}\text{N}_9\text{O}_{33}\text{P}_{12}\text{Pt}_6\text{S}_6$  m/z  $[\text{M}-3\text{OTf}]^{3+}$  1821.38;  $[\text{M}-4\text{OTf}]^{4+}$  1328.80;  $[\text{L1}+2\text{L4}-2\text{OTf}]^{2+}$  1097.77;  $[\text{M}-5\text{OTf}]^{5+}$  1033.23;  $[\text{L1}+\text{L4}-2\text{OTf}]^{2+}$  835.70. (Where M represents the intact assembly).



**Figure S23.**  $^1\text{H}$  NMR spectrum of MPy3 in  $\text{DMSO-}d_6$ .

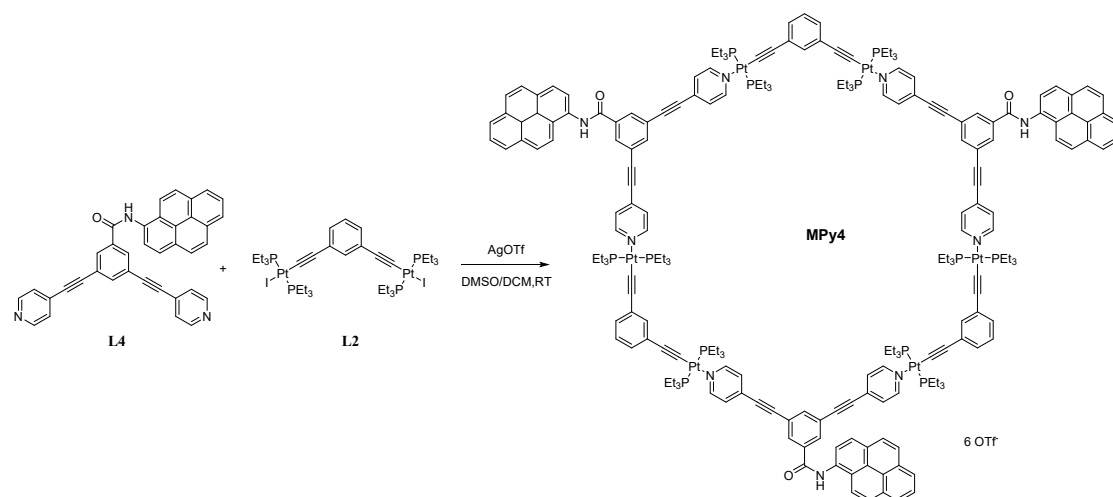


**Figure S24.**  $^{31}\text{P}\{^1\text{H}\}$  NMR spectrum of MPy3 in  $\text{DMSO-}d_6$ .



**Figure S25.** Experimental (red) and calculated (blue) electrospray ionization mass spectra of MPy3.

## 5. Synthesis of MPy4

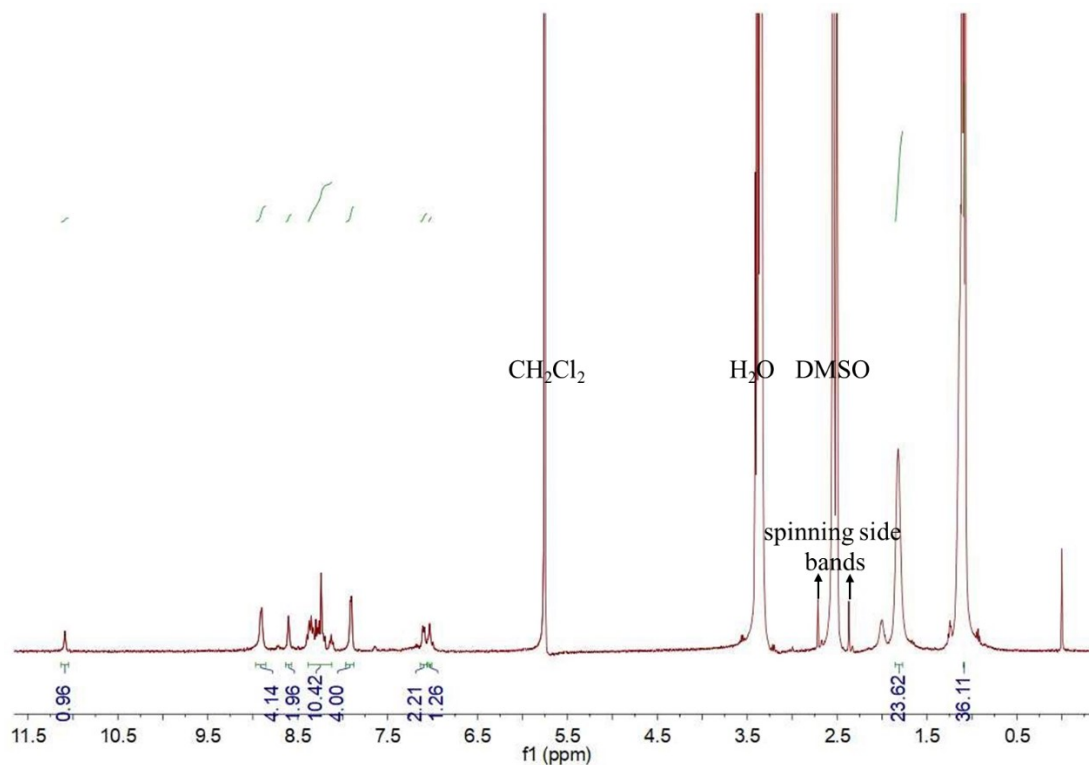


**Figure S26.** Synthesis route of MPy4.

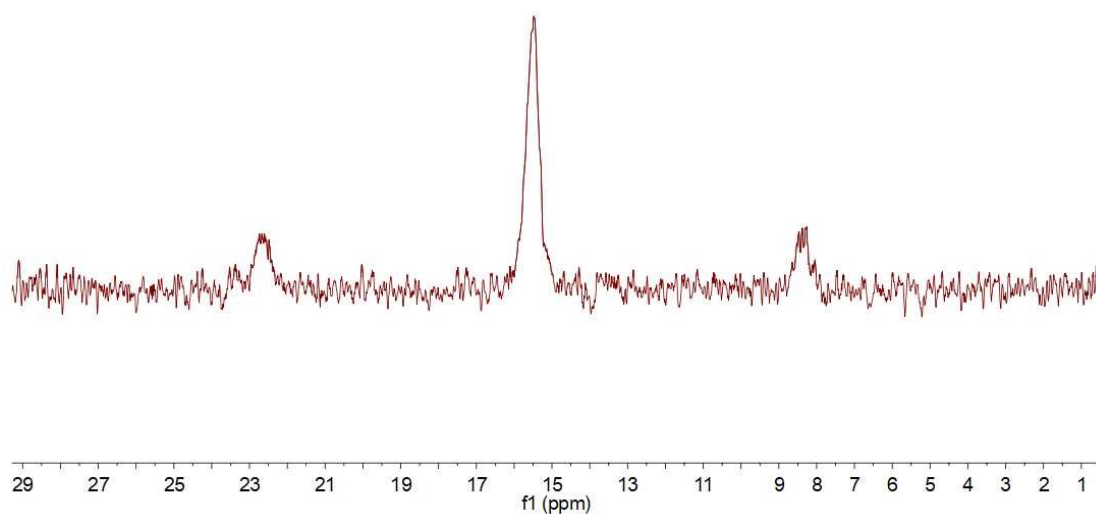
### Synthesis of MPy4

L2 (10 mg, 7.0  $\mu\text{mol}$ ), AgOTf (5.5 mg, 21.0  $\mu\text{mol}$ ) and L4 (4 mg, 7.0  $\mu\text{mol}$ )

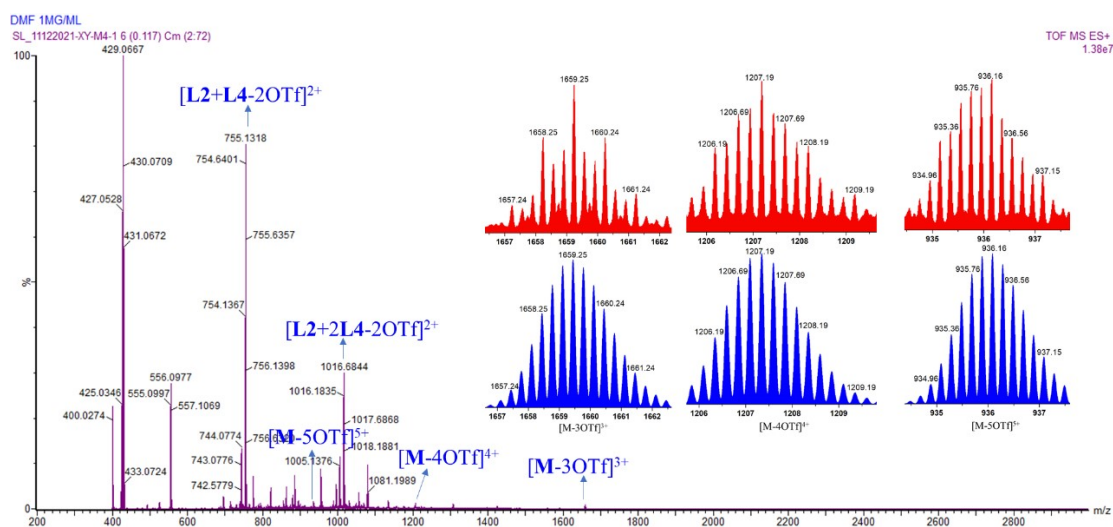
were dissolved in the mixture of DMSO and  $\text{CH}_2\text{Cl}_2$  (2.0 mL,  $V_{\text{CH}_2\text{Cl}_2}:V_{\text{DMSO}} = 10:1$ ) in a 20 mL glass vial. The reaction mixture was allowed to stir for 6 h at room temperature, and the reaction mixture was centrifuged at 3400 r/h for 20 min. To the resulting homogeneous solution, diethyl ether was added to precipitate the product, which was then isolated and dried under reduced pressure and re-dissolved in  $\text{DMSO-}d_6$  for characterization.  $^1\text{H}$  NMR (400 MHz,  $\text{DMSO-}d_6$ )  $\delta$  11.09 (s, 1H), 8.90 (s, 4H), 8.60 (s, 2H), 8.38 – 8.12 (m, 10H), 7.90 (s, 4H), 7.10 (d,  $J = 7.8$  Hz, 2H), 7.03 (s, 1H), 1.82 (s, 24H), 1.12 – 1.06 (m, 36H).  $^{31}\text{P}$  NMR (121.4 MHz,  $\text{DMSO-}d_6$ )  $\delta$  (ppm) 15.48 (s,  $^{195}\text{Pt}$  satellites,  $^1J_{\text{Pt-P}} = 1734.81$  Hz). ESI-TOF-MS:  $\text{C}_{219}\text{H}_{255}\text{F}_{18}\text{N}_9\text{O}_{21}\text{P}_{12}\text{Pt}_6\text{S}_6$  m/z  $[\text{M-3OTf}]^{3+}$  1659.25;  $[\text{M-4OTf}]^{4+}$  1207.19;  $[\text{L2+2L4-2OTf}]^{2+}$  1016.68;  $[\text{M-5OTf}]^{5+}$  936.16;  $[\text{L2+L4-2OTf}]^{2+}$  755.13. (Where M represents the intact assembly).



**Figure S27.**  $^1\text{H}$  NMR spectrum of **MPy4** in  $\text{DMSO-}d_6$ .



**Figure S28.**  $^{31}\text{P}\{^1\text{H}\}$  NMR spectrum of **MPy4** in  $\text{DMSO-}d_6$ .



**Figure S29.** Experimental (red) and calculated (blue) electrospray ionization mass spectra of **MPy4**.

## 6. Experimental procedure and methods

### 6.1 The preparation of the samples

The preparation process for the assembly: the stock solution of **MPy1** ( $2 \times 10^{-4}$  M, dissolved in DMSO), ESY ( $1 \times 10^{-5}$  M, dissolved in  $\text{H}_2\text{O}$ -DMSO, v/v; 3:2), SR101

( $1 \times 10^{-5}$  M, dissolved in H<sub>2</sub>O–DMSO, v/v; 3:2) were prepared, respectively. **MPy1** assembly was prepared as follows: 0.2 mL of **MPy1** stock solution was added into the mixture of water (2.4 mL) and DMSO (1.4 mL) under stirring to afford  $1 \times 10^{-5}$  M solution (H<sub>2</sub>O–DMSO, v/v; 3:2). **MPy1-ESY** assembly was prepared as follows: 20  $\mu$ L of ESY stock solution was added into 2 mL of **MPy1** assembly. **MPy1-SR101** assembly was prepared as follows: 20  $\mu$ L of SR101 stock solution was added into 2 mL of **MPy1** assembly. **MPy1-ESY-SR101** assembly was prepared as follows: 10  $\mu$ L of SR101 solution was added into 2 mL of **MPy1-ESY** assembly (100:1).

The fluorescence experiments were conducted as follows: ESY and SR101 was dissolved in H<sub>2</sub>O–DMSO (3:2; v/v) and added into the H<sub>2</sub>O–DMSO mixture of **MPy1** or **MPy1-ESY** assembly (100:1), respectively.

## 6.2 The measurement of the Tyndall effect

As shown in 6.1, the solution of **MPy1**, **MPy1-ESY** (100:1) and **MPy1-ESY-SR101** (200:2:1) in H<sub>2</sub>O–DMSO (3:2; v/v) was prepared, [**MPy1**] =  $1.0 \times 10^{-6}$  M, [ESY] =  $1.0 \times 10^{-8}$  M, [SR101] =  $5.0 \times 10^{-9}$  M, using a laser pointer to send a beam of light through cuvette, which were photographed under 365 nm UV light.

## 6.3 Preparation of anti-counterfeiting inks

The luminescent printing was performed with a modified HP Deskjet 1112 inkjet printer. The conventional inkjet office printer cartridge (HP 803) was refitted first. After removal of ink, the cartridge was thoroughly cleared with water and ethanol until it was clean, and dried by blowing with N<sub>2</sub> at room temperature. The fluorescent ink (**MPy1-ESY-SR101** system at different donor/acceptor ratios) was injected into the cartridge, and then different patterns were printed as designed on unmodified copy paper.

## 6.4 Energy transfer efficiency ( $\Phi_{ET}$ )

The energy-transfer efficiency ( $\Phi_{ET}$ ) was calculated using equation S1.

$$\Phi_{ET} = 1 - \frac{I_{DA, (\lambda_{ex}=\text{donor})}}{I_{D, (\lambda_{ex}=\text{donor})}} \quad (\text{eq. S1})$$

For **MPy1**–**ESY** system, where  $I_{DA}$  and  $I_D$  are the fluorescence intensity of **MPy1**–**ESY** assembly (donor and acceptor) and **MPy1** assembly (donor) at 460 nm when excited at 384 nm, respectively. The energy-transfer efficiency ( $\Phi_{ET}$ ) was calculated as 12.2 % in H<sub>2</sub>O–DMSO (3:2; v/v), measured under the condition of  $[MPy1] = 1.0 \times 10^{-5}$  M,  $[ESY] = 1.0 \times 10^{-7}$  M, and  $\lambda_{ex} = 384$  nm.

For **MPy1**–**SR101** system, where  $I_{DA}$  and  $I_D$  are the fluorescence intensity of **MPy1**–**SR101** assembly (donor and acceptor) and **MPy1** assembly (donor) at 460 nm when excited at 384 nm, respectively. The energy-transfer efficiency ( $\Phi_{ET}$ ) was calculated as 2.3 % in H<sub>2</sub>O–DMSO (3:2; v/v), measured under the condition of  $[MPy1] = 1.0 \times 10^{-5}$  M,  $[SR101] = 1.0 \times 10^{-7}$  M, and  $\lambda_{ex} = 384$  nm.

For **MPy1**–**ESY**–**SR101** system, where  $I_{DA}$  and  $I_D$  are the fluorescence intensity of **MPy1**–**ESY**–**SR101** assembly (donor and acceptor) and **MPy1**–**ESY** assembly (donor) at 550 nm when excited at 384 nm, respectively. The energy-transfer efficiency ( $\Phi_{ET}$ ) was calculated as 20.1 % in H<sub>2</sub>O–DMSO (3:2; v/v), measured under the condition of  $[MPy1] = 1.0 \times 10^{-5}$  M,  $[ESY] = 1.0 \times 10^{-7}$  M,  $[SR101] = 5.0 \times 10^{-8}$  M, and  $\lambda_{ex} = 384$  nm.

## 6.5 Antenna effect (AE)

The antenna effect (AE) was calculated using equation S2.

$$\text{Antenna effect} = (I_{DA, (\lambda_{ex}=\text{donor})} - I_{D, (\lambda_{ex}=\text{donor})}) / I_{DA, (\lambda_{ex}=\text{acceptor})} \quad (\text{eq. S2})$$

Where  $I_{DA, (\lambda_{ex}=\text{donor})}$  and  $I_{DA, (\lambda_{ex}=\text{acceptor})}$  are the fluorescence intensity of the system with excitation of donor and direct excitation of acceptor, respectively.  $I_{D, (\lambda_{ex}=\text{donor})}$  is the fluorescence intensity of the donor.

For **MPy1**–**ESY** system, where  $I_{DA, (\lambda_{ex}=\text{donor})}$  and  $I_{DA, (\lambda_{ex}=\text{acceptor})}$  are the fluorescence intensity at 550 nm with the excitation of the donor at 384 nm and the direct excitation of the acceptor at 500 nm, respectively.  $I_{D, (\lambda_{ex}=\text{donor})}$  is the fluorescence intensity at 550 nm of the **MPy1** assembly, which was normalized with the **MPy1**–**ESY** assembly at 460 nm. The antenna effect value was calculated as 14.3 in H<sub>2</sub>O–DMSO (3:2; v/v), measured under the condition of  $[MPy1] = 1.0 \times 10^{-5}$  M,  $[ESY] = 1.0 \times 10^{-7}$  M, and  $\lambda_{ex} = 384$  nm (Figure S56).



For **MPy1**–SR101 system, where  $I_{DA, (\lambda_{ex}=donor)}$  and  $I_{DA, (\lambda_{ex}=acceptor)}$  are the fluorescence intensity at 610 nm with the excitation of the donor at 384 nm and the direct excitation of the acceptor at 580 nm, respectively.  $I_{D, (\lambda_{ex}=donor)}$  is the fluorescence intensity at 610 nm of the **MPy1** assembly, which was normalized with the **MPy1**–SR101 assembly at 460 nm. The antenna effect value was calculated as 5.8 in H<sub>2</sub>O–DMSO (3:2; v/v), measured under the condition of  $[MPy1] = 1.0 \times 10^{-5}$  M,  $[SR101] = 1.0 \times 10^{-7}$  M, and  $\lambda_{ex} = 384$  nm (Figure S56).

For **MPy1**–ESY–SR101 system, where  $I_{DA, (\lambda_{ex}=donor)}$  and  $I_{DA, (\lambda_{ex}=acceptor)}$  are the fluorescence intensities at 610 nm with the excitation of the donor at 384 nm and the direct excitation of the acceptor at 580 nm, respectively.  $I_{D, (\lambda_{ex}=donor)}$  is the fluorescence intensities at 610 nm of the **MPy1**–ESY assembly, which was normalized with the **MPy1**–ESY–SR101 assembly at 550 nm. The antenna effect value was calculated as 9.3 in H<sub>2</sub>O–DMSO (3:2; v/v), measured under the condition of  $[MPy1] = 1.0 \times 10^{-5}$  M,  $[ESY] = 1.0 \times 10^{-7}$  M,  $[SR101] = 5.0 \times 10^{-8}$  M, and  $\lambda_{ex} = 384$  nm (Figure S58).

## 7. Additional tables

**Table S1.** Fluorescence lifetimes of **MPy1**, **MPy1**–ESY and **MPy1**–ESY–SR101 assemblies in H<sub>2</sub>O–DMSO (3:2; v/v).

Sample	$\tau_1/ns$	Rel %	$\tau_2/ns$	Rel %	A	$\chi^2$
<b>MPy1</b> (solid)	1.64	41.45	5.39	58.55	4.118	1.237
<b>MPy1</b> <sup>a</sup>	1.64	39.07	5.35	60.93	4.308	1.208
<b>MPy1</b> –ESY (300:1) <sup>a</sup>	1.47	31.23	4.90	68.77	3.210	1.212
<b>MPy1</b> –ESY (100:1) <sup>a</sup>	1.30	29.50	4.56	70.50	1.794	1.158
<b>MPy1</b> –ESY (100:1) <sup>b</sup>	1.28	71.32	4.78	28.68	1.482	1.283
<b>MPy1</b> –ESY–SR101 (200:2:0.5) <sup>b</sup>	1.19	58.21	4.34	41.79	2.115	1.278
<b>MPy1</b> –ESY–SR101	1.19	65.28	3.73	34.72	4.087	1.287

(200:2:1) <sup>b</sup>						
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a: monitored at 460 nm upon excitation at 375 nm; [MPy1] = 1.0×10<sup>-5</sup> M, [ESY] = 1.0×10<sup>-7</sup> M.

b: monitored at 550 nm upon excitation at 375 nm; [MPy1] = 1.0×10<sup>-5</sup> M, [ESY] = 1.0×10<sup>-7</sup> M, [SR101] = 5.0×10<sup>-8</sup> M, (Figures S31, S47-S52).

**Table S2.** Fluorescence quantum yields of ESY in H<sub>2</sub>O–DMSO (3:2; v/v), SR101 in H<sub>2</sub>O–DMSO (3:2; v/v), MPy1 (in DMSO solution), MPy1 (solid), MPy1–ESY and MPy1–ESY–SR101 in H<sub>2</sub>O–DMSO (3:2; v/v).

Sample	Concentration	Fluorescence quantum yields ( $\Phi_f$ )
ESY	[ESY] = 1.0×10 <sup>-5</sup> M	38.86%
SR101	[SR101] = 1.0×10 <sup>-5</sup> M	81.58%
MPy1 in DMSO solution	[MPy1] = 1.0×10 <sup>-5</sup> M	0.98%
MPy1 in the solid state	-	0.02%
MPy1	[MPy1] = 1.0×10 <sup>-5</sup> M	13.2 %
MPy1–ESY	[MPy1] = 1.0×10 <sup>-5</sup> M [ESY] = 1.0×10 <sup>-7</sup> M	17.4 %
MPy1–SR101	[MPy1] = 1.0×10 <sup>-5</sup> M [SR101] = 1.0×10 <sup>-7</sup> M	9.11 %
MPy1–ESY–SR101	[MPy1] = 1.0×10 <sup>-5</sup> M [ESY] = 1.0×10 <sup>-7</sup> M [SR101] = 5.0×10 <sup>-8</sup> M	21.3 %

**Table S3.** The energy transfer efficiency in H<sub>2</sub>O–DMSO (3:2; v/v).

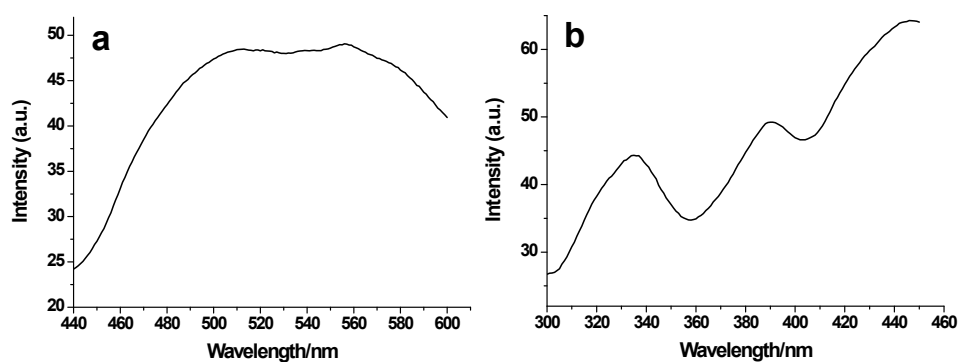
Sample	Concentration	energy transfer efficiency
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		$(\Phi_{ET})$
<b>MPy1-ESY</b> (100:1)	[MPy1] = $1.0 \times 10^{-5}$ M [ESY] = $1.0 \times 10^{-7}$ M	12.2 %
<b>MPy1-SR101</b> (100:1)	[MPy1] = $1.0 \times 10^{-5}$ M [SR101] = $1.0 \times 10^{-7}$ M	2.3 %
<b>MPy1-ESY-SR101</b> (200:2:1)	[MPy1] = $1.0 \times 10^{-5}$ M [ESY] = $1.0 \times 10^{-7}$ M [SR101] = $5.0 \times 10^{-8}$ M	20.1 %

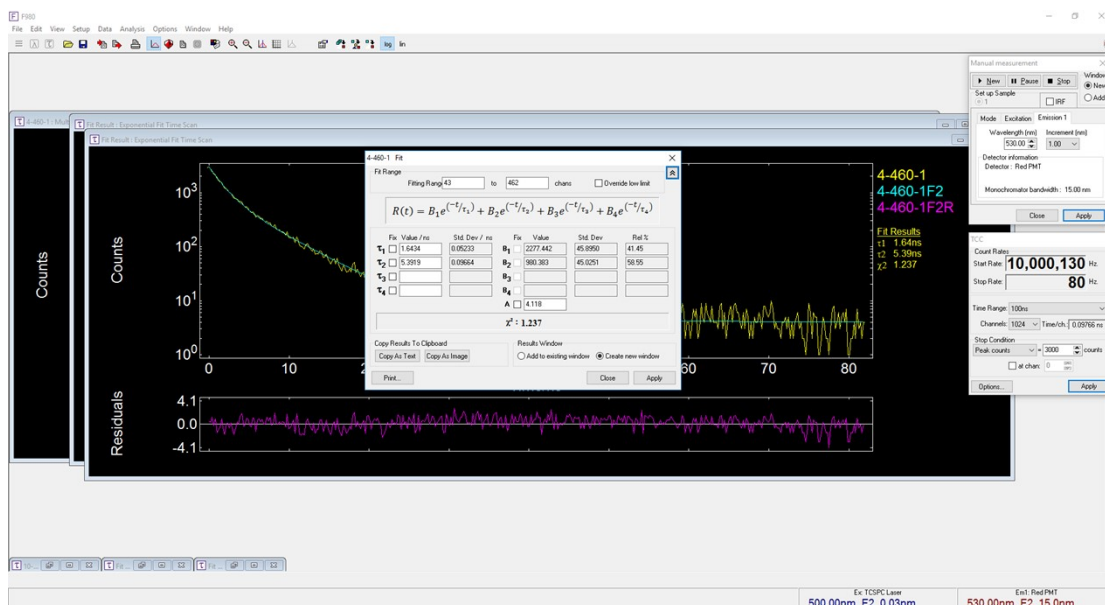
**Table S4.** Antenna effect

Sample	Concentration	Antenna effect
<b>MPy1-ESY</b> (100:1)	[MPy1] = $1.0 \times 10^{-5}$ M [ESY] = $1.0 \times 10^{-7}$ M	14.3
<b>MPy1-SR101</b> (100:1)	[MPy1] = $1.0 \times 10^{-5}$ M [SR101] = $1.0 \times 10^{-7}$ M	5.8
<b>MPy1-ESY-SR101</b> (200:2:1)	[MPy1] = $1.0 \times 10^{-5}$ M [ESY] = $1.0 \times 10^{-7}$ M [SR101] = $5.0 \times 10^{-8}$ M	9.3

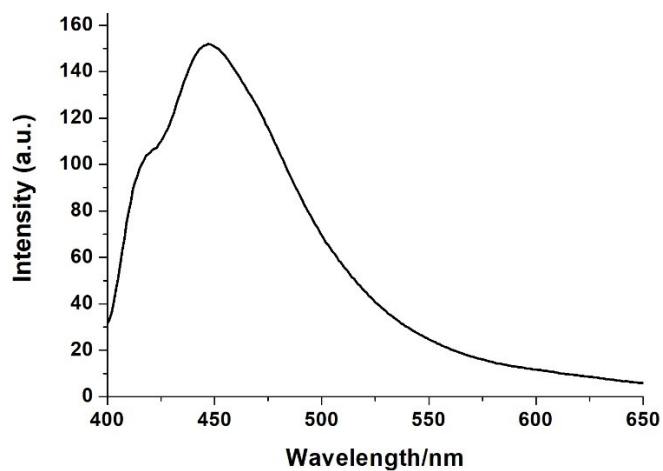
## 8. Additional Spectra and images



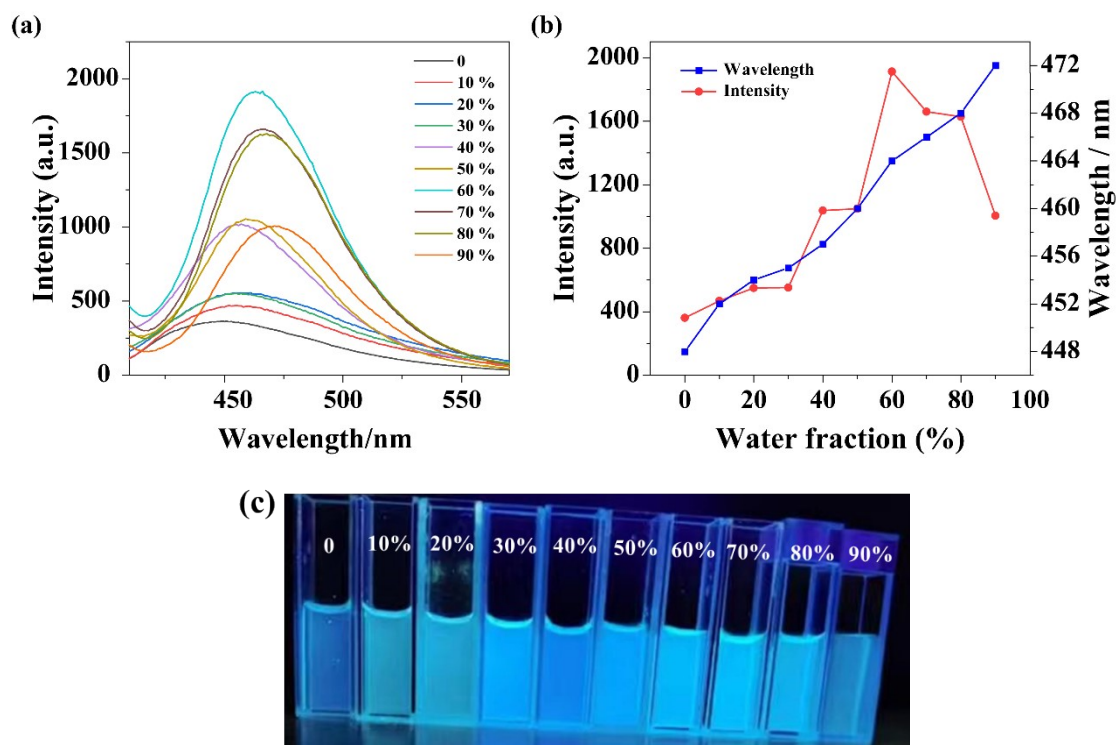
**Figure S30.** The emission of MPy1 on  $\lambda_{ex}$  390 nm (a) and excitation spectrum for  $\lambda_{em}$  530 nm (b) in the solid state.



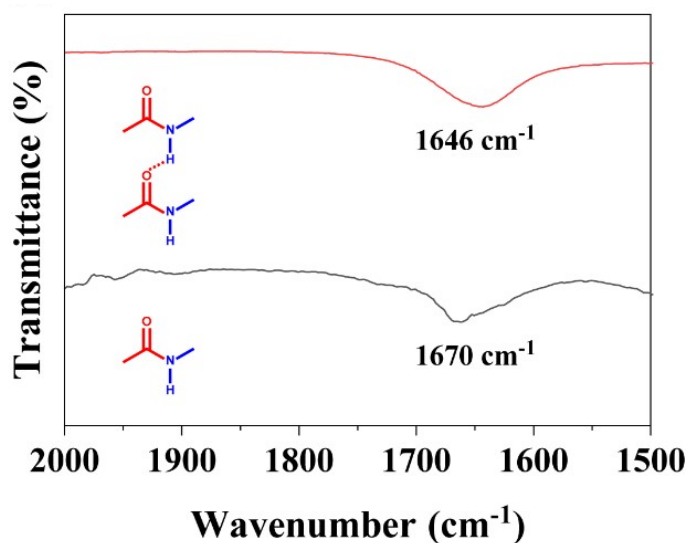
**Figure S31.** Fluorescence decay profiles of **MPy1** ( $\lambda_{\text{ex}} = 390$  nm,  $\lambda_{\text{collected}} = 530$  nm) in the solid state.



**Figure S32.** Fluorescence spectrum of **MPy1** in THF (1% DMSO) at 298 K ( $\lambda_{\text{ex}} = 390$  nm).

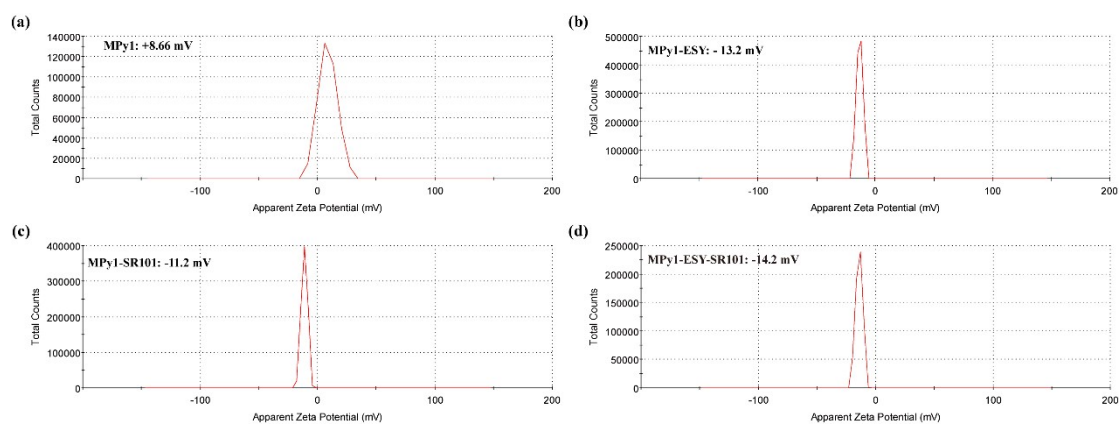


**Figure S33.** (a) Fluorescence spectra of **MPy1** in H<sub>2</sub>O–DMSO mixtures with different water volume fractions ( $f_w$ , vol %) ( $\lambda_{\text{ex}} = 384$  nm). (b) Fluorescence intensity and emission wavelength of **MPy1** with various fractions of water from 0 to 90 %. (c) Photographs of **MPy1** with various fractions of water under 365 nm irradiation.

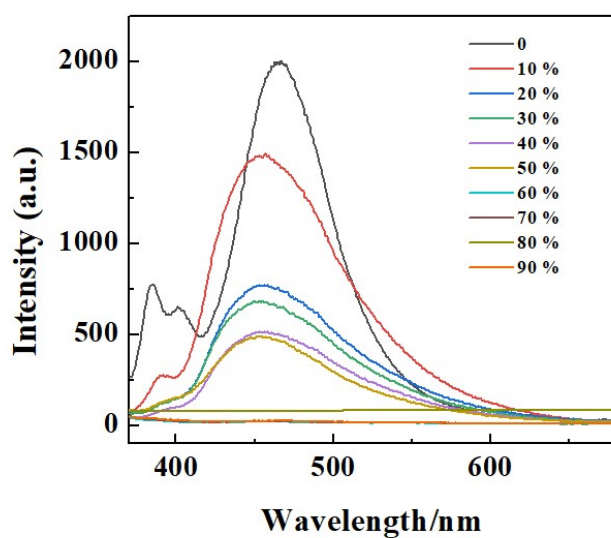


**Figure S34.** FTIR spectra of compound **MPy1** (10 mM) in DMSO (black line) and H<sub>2</sub>O–DMSO (3:2; v/v) (red line). The sharp amide I band shifted from 1670 cm<sup>-1</sup> in

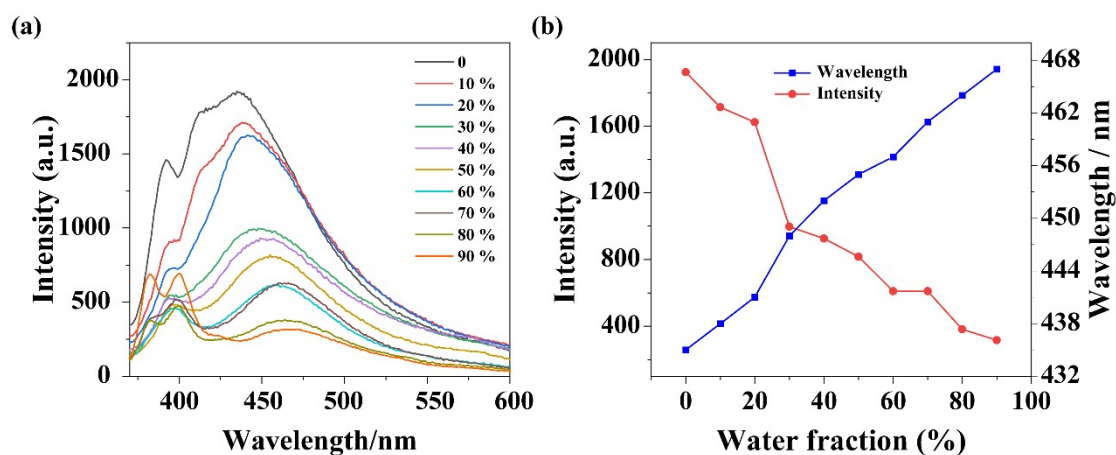
DMSO (monomers) to  $1646\text{ cm}^{-1}$  in  $\text{H}_2\text{O}$ –DMSO (3:2; v/v) (aggregation), suggesting the formation of intermolecular hydrogen bonds.



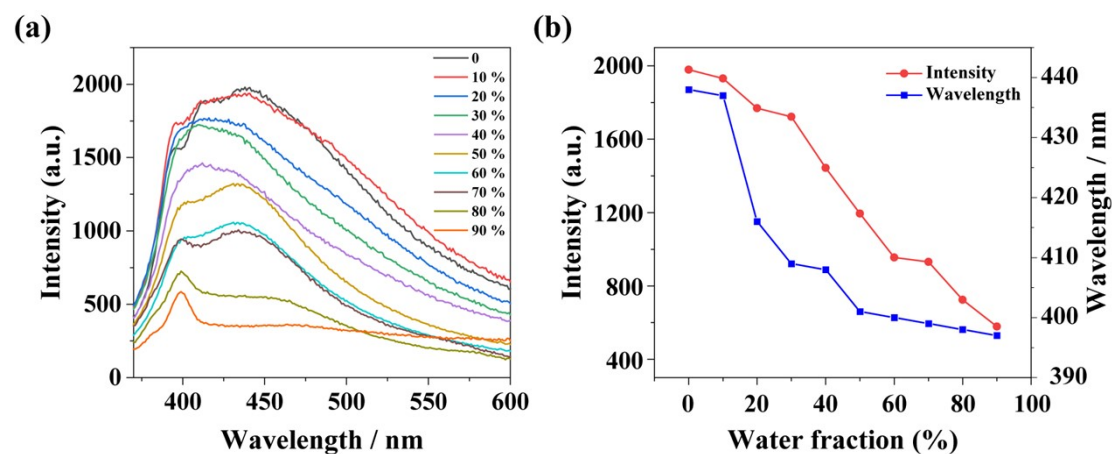
**Figure S35.** Zeta potentials of MPy1, MPy1–ESY (100:1), MPy1–SR101 (100:1) and MPy1–ESY–SR101 (200:2:1) in  $\text{H}_2\text{O}$ –DMSO (3:2; v/v).



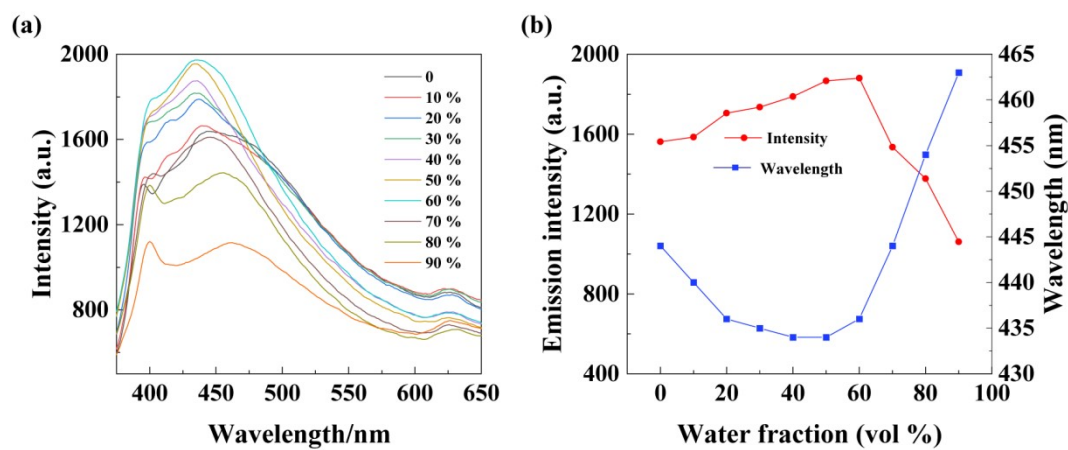
**Figure S36.** Fluorescence spectra of L3 in  $\text{H}_2\text{O}$ –DMSO mixtures with different water volume fractions ( $f_w$ , vol %) ( $\lambda_{\text{ex}} = 350\text{ nm}$ ).



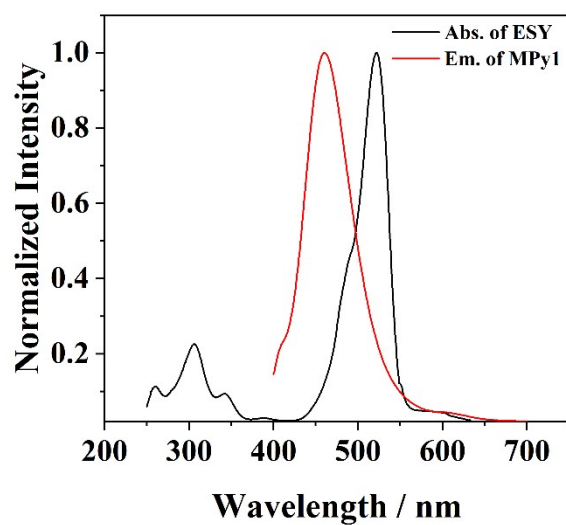
**Figure S37.** (a) Fluorescence spectra of **MPy2** in H<sub>2</sub>O–DMSO mixtures with different water volume fractions ( $f_w$ , vol %) ( $\lambda_{ex} = 350$  nm). (b) Fluorescence intensity of **MPy2** with various fractions of water from 0 to 90 %.



**Figure S38.** (a) Fluorescence spectra of **MPy4** in H<sub>2</sub>O–DMSO mixtures with different water volume fractions ( $f_w$ , vol %) ( $\lambda_{ex} = 350$  nm). (b) Fluorescence intensity of **MPy4** with various fractions of water from 0 to 90 %.

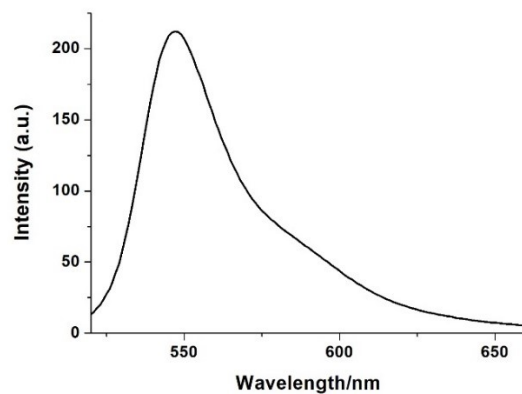


**Figure S39.** (a) Fluorescence spectra of **MPy3** in H<sub>2</sub>O–DMSO mixtures with different water volume fractions ( $f_w$ , vol %) ( $\lambda_{ex} = 350$  nm). (b) Fluorescence intensity of **MPy3** with various fractions of water from 0 to 90 %.

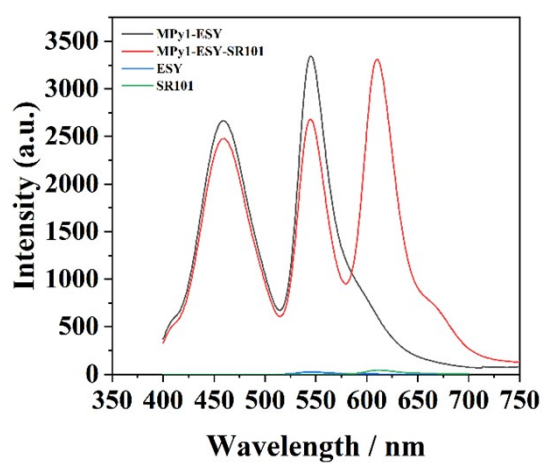


**Figure S40.** Normalized absorption spectrum of ESY and emission spectrum of **MPy1**.

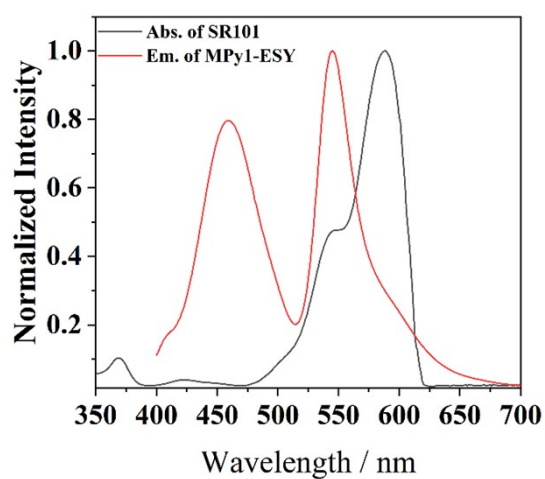




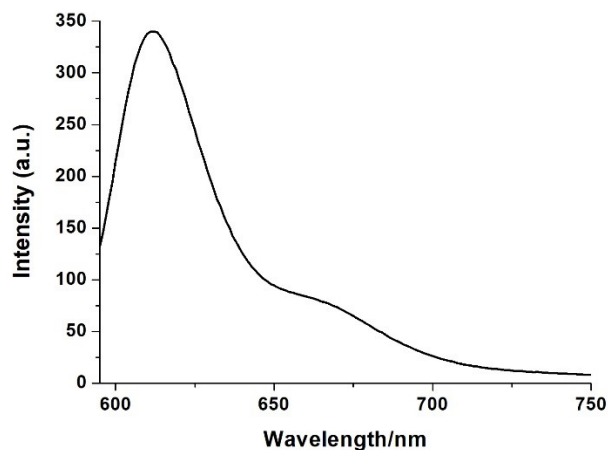
**Figure S41.** Fluorescence spectra of ESY in H<sub>2</sub>O–DMSO (3:2, v/v) ( $\lambda_{\text{ex}} = 500 \text{ nm}$ ).



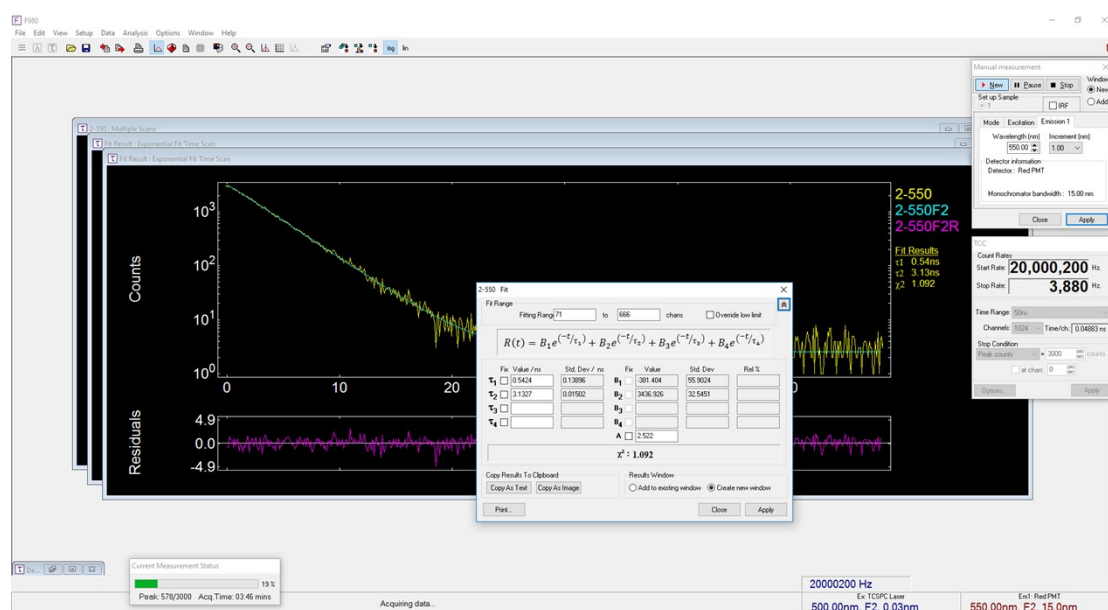
**Figure S42.** Fluorescence spectra of **MPy1-ESY**, **MPy1-ESY-SR101**, SR101 and ESY ( $\lambda_{\text{ex}} = 384 \text{ nm}$ ) in H<sub>2</sub>O–DMSO (3:2; v/v).



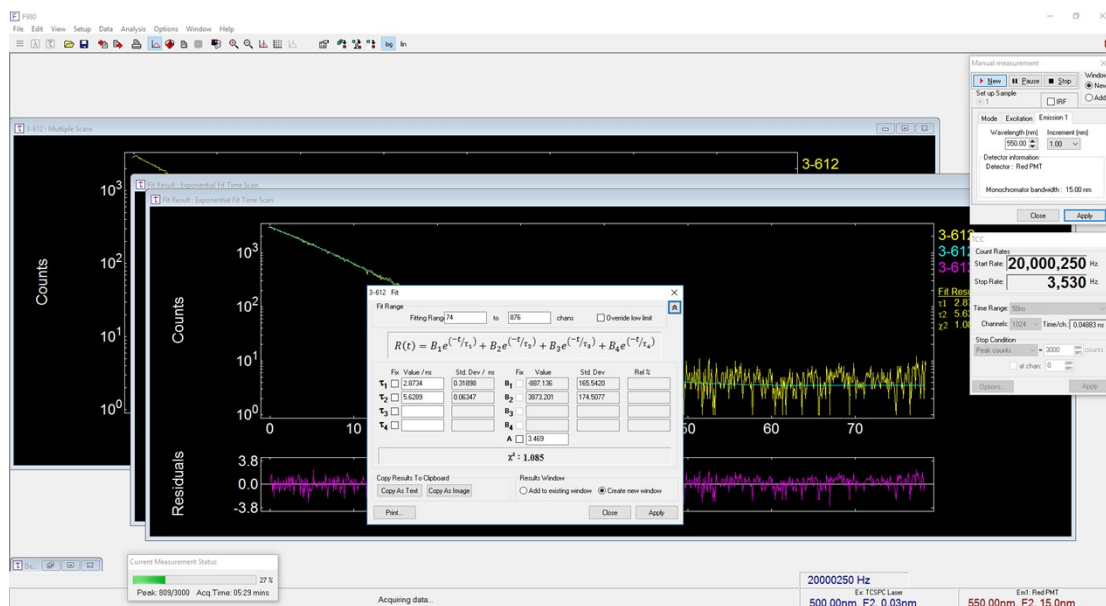
**Figure S43.** Normalized absorption spectrum of SR101 and emission spectrum of **MPy1-ESY** assembly.



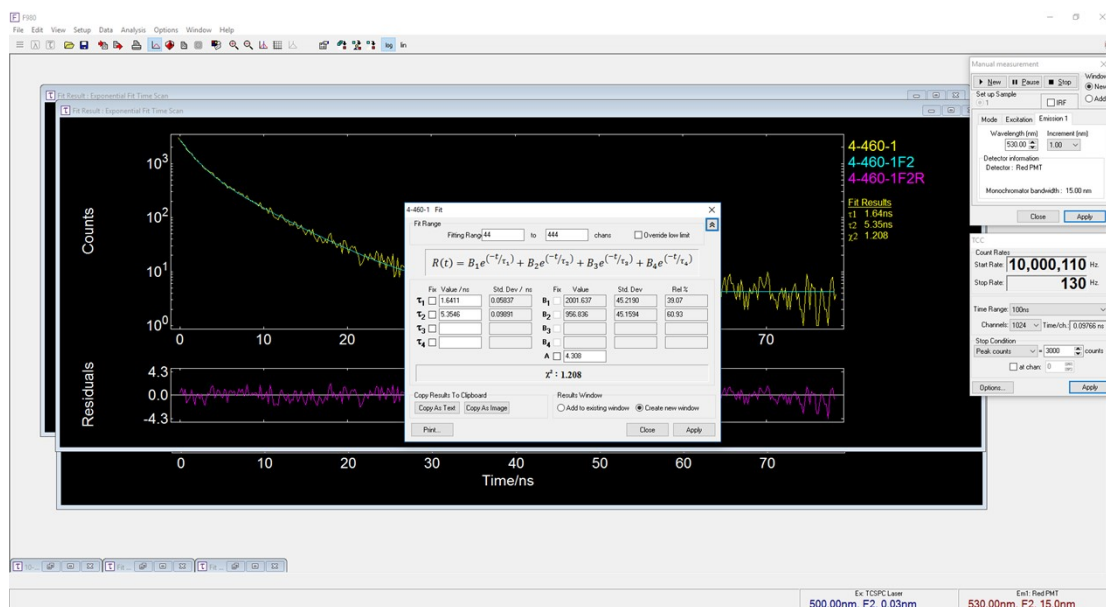
**Figure S44.** Fluorescence spectra of SR101 in H<sub>2</sub>O–DMSO (3:2, v/v) ( $\lambda_{\text{ex}} = 580 \text{ nm}$ ).



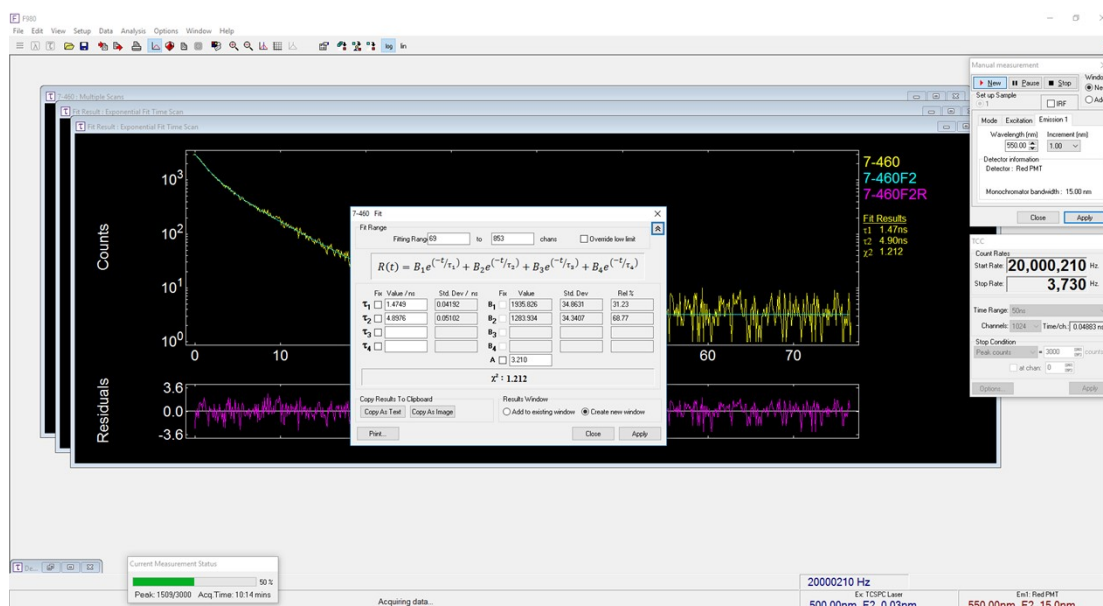
**Figure S45.** Fluorescence decay profile of ESY in H<sub>2</sub>O–DMSO (3:2, v/v) monitored at 550 nm.



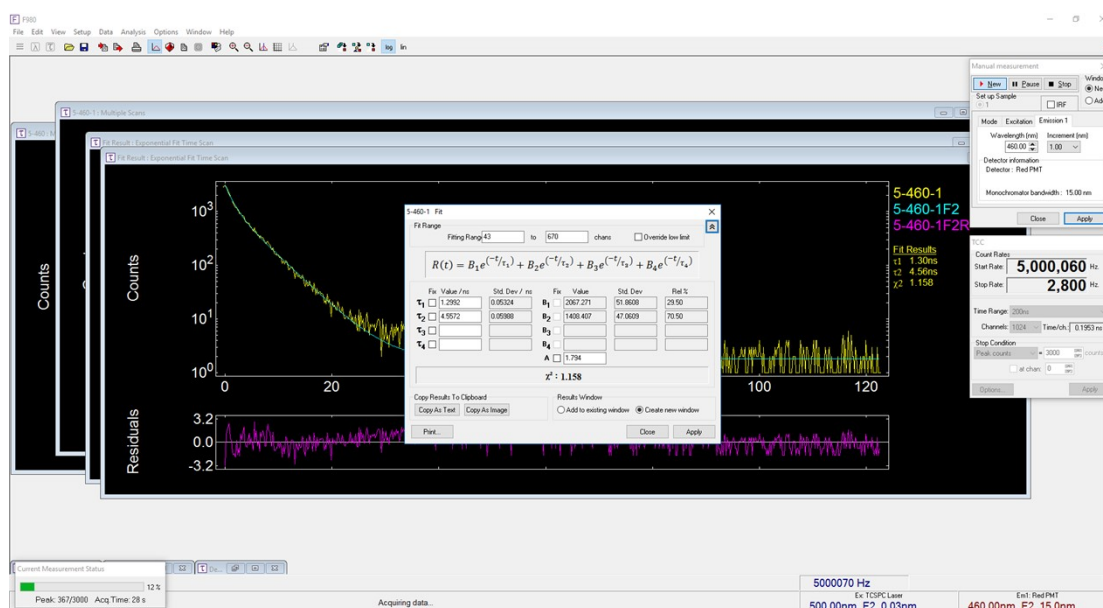
**Figure S46.** Fluorescence decay profile of SR101 in H<sub>2</sub>O–DMSO (3:2, v/v) monitored at 612 nm.



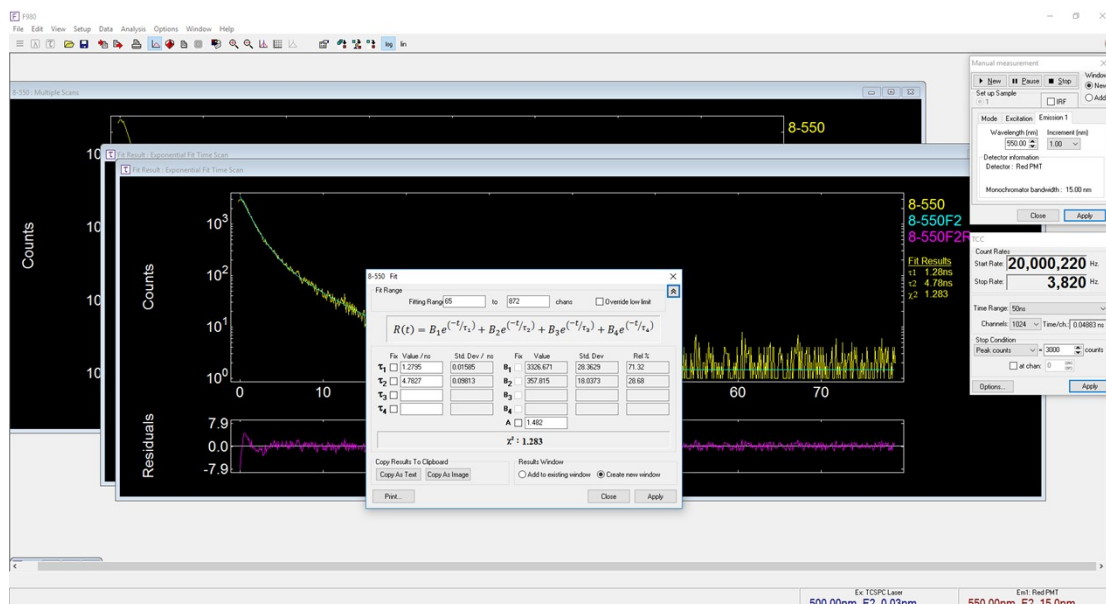
**Figure S47.** Fluorescence decay profile of MPy1 assembly in H<sub>2</sub>O–DMSO (3:2; v/v) monitored at 460 nm upon excitation at 375 nm.



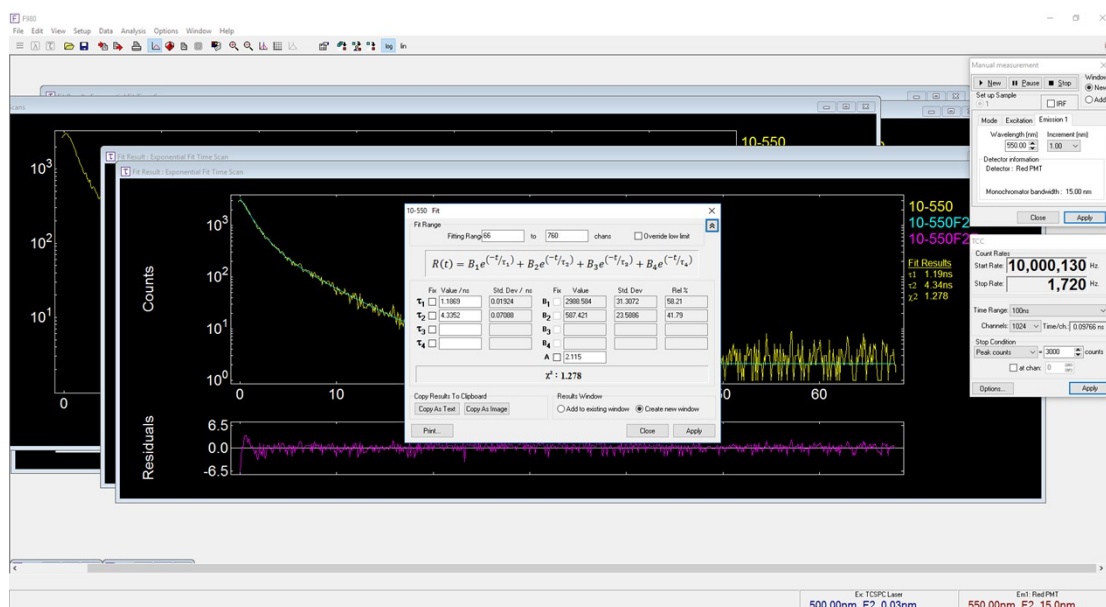
**Figure S48.** Fluorescence decay profile of MPy1-ESY (300:1) assembly in H<sub>2</sub>O-DMSO (3:2; v/v) monitored at 460 nm upon excitation at 375 nm.



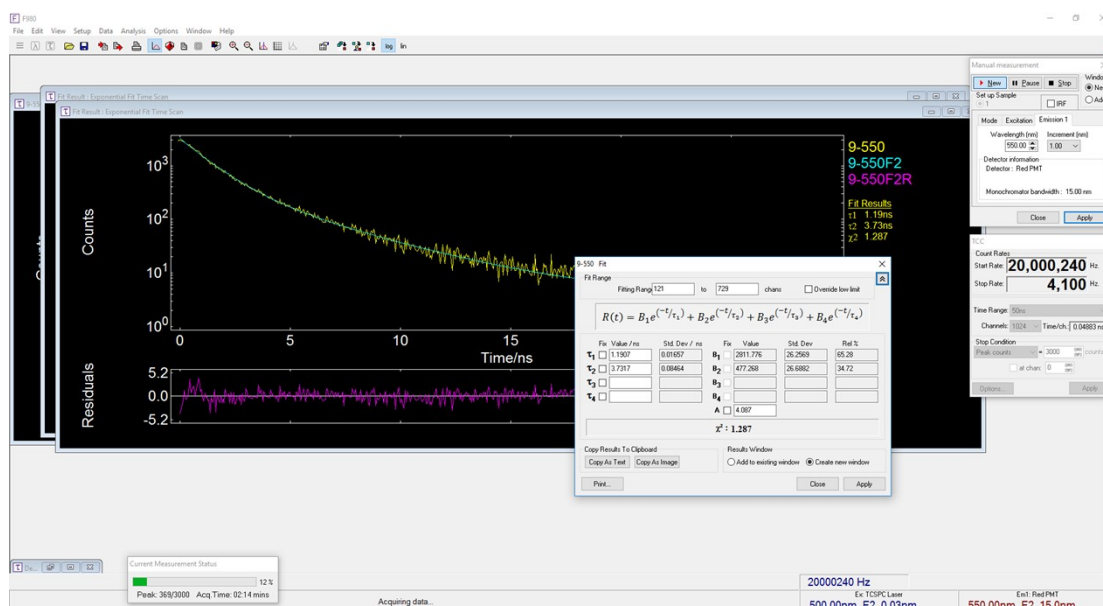
**Figure S49.** Fluorescence decay profile of MPy1-ESY (100:1) assembly in H<sub>2</sub>O-DMSO (3:2; v/v) monitored at 460 nm upon excitation at 375 nm.



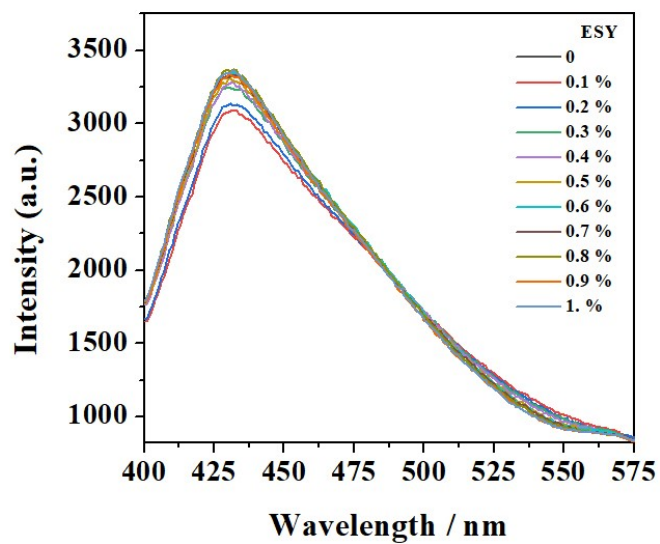
**Figure S50.** Fluorescence decay profile of **MPy1-ESY (100:1)** assembly monitored at 550 nm upon excitation at 375 nm.



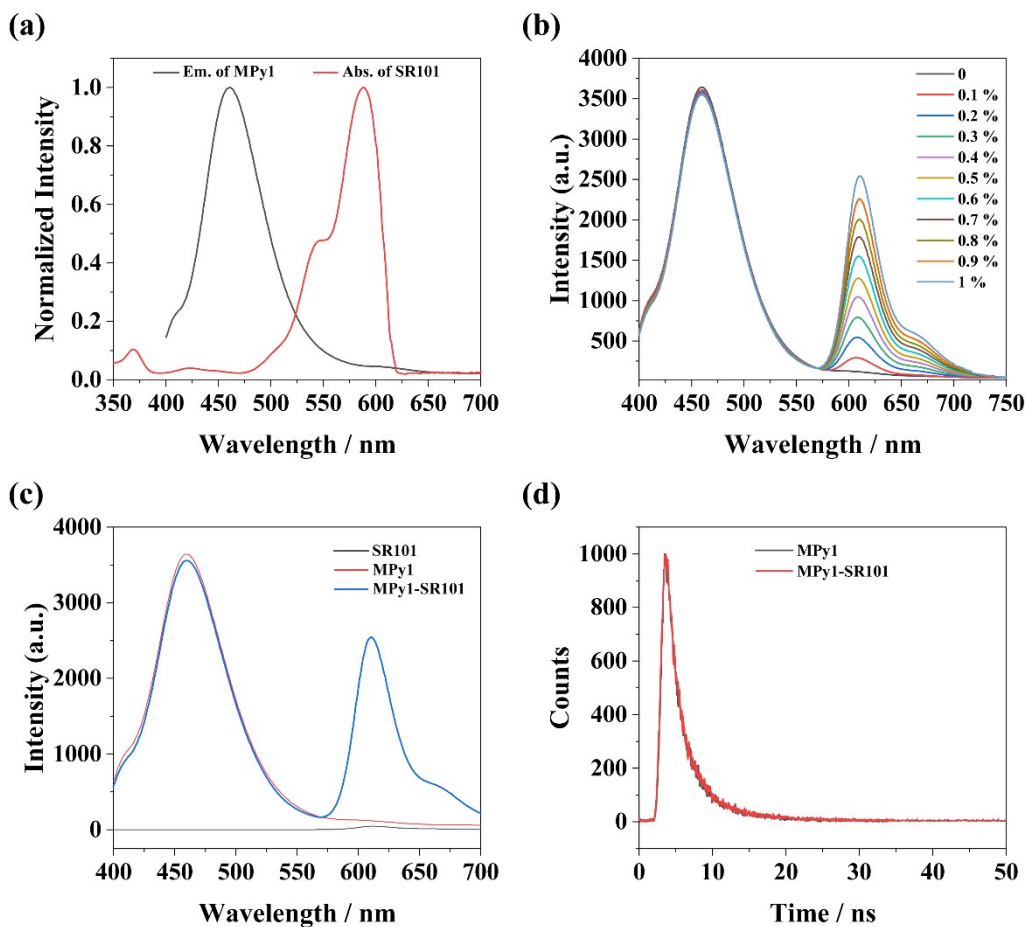
**Figure S51.** Fluorescence decay profile of **MPy1-ESY-SR101 (200:2:0.5)** assembly monitored at 550 nm upon excitation at 375 nm.



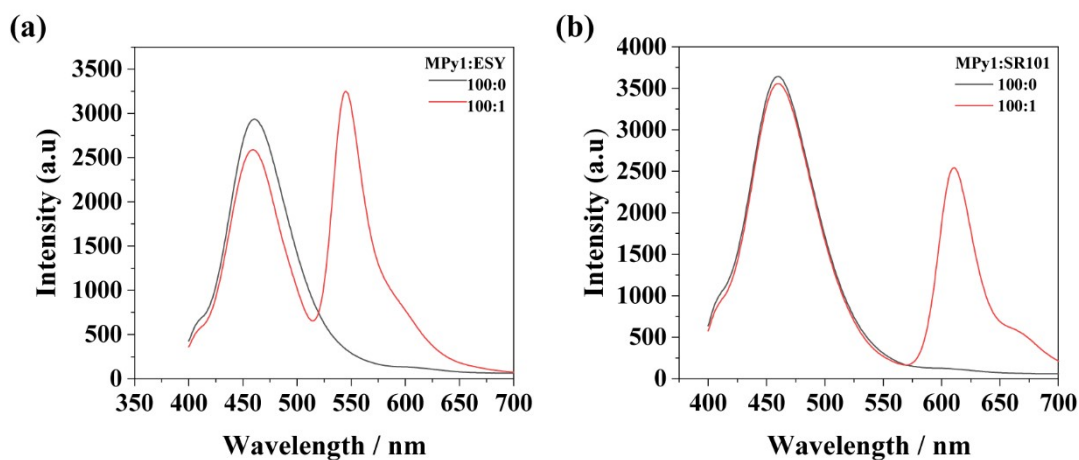
**Figure S52.** Fluorescence decay profile of MPy1-ESY-SR101 (200:2:1) assembly monitored at 550 nm upon excitation at 375 nm.



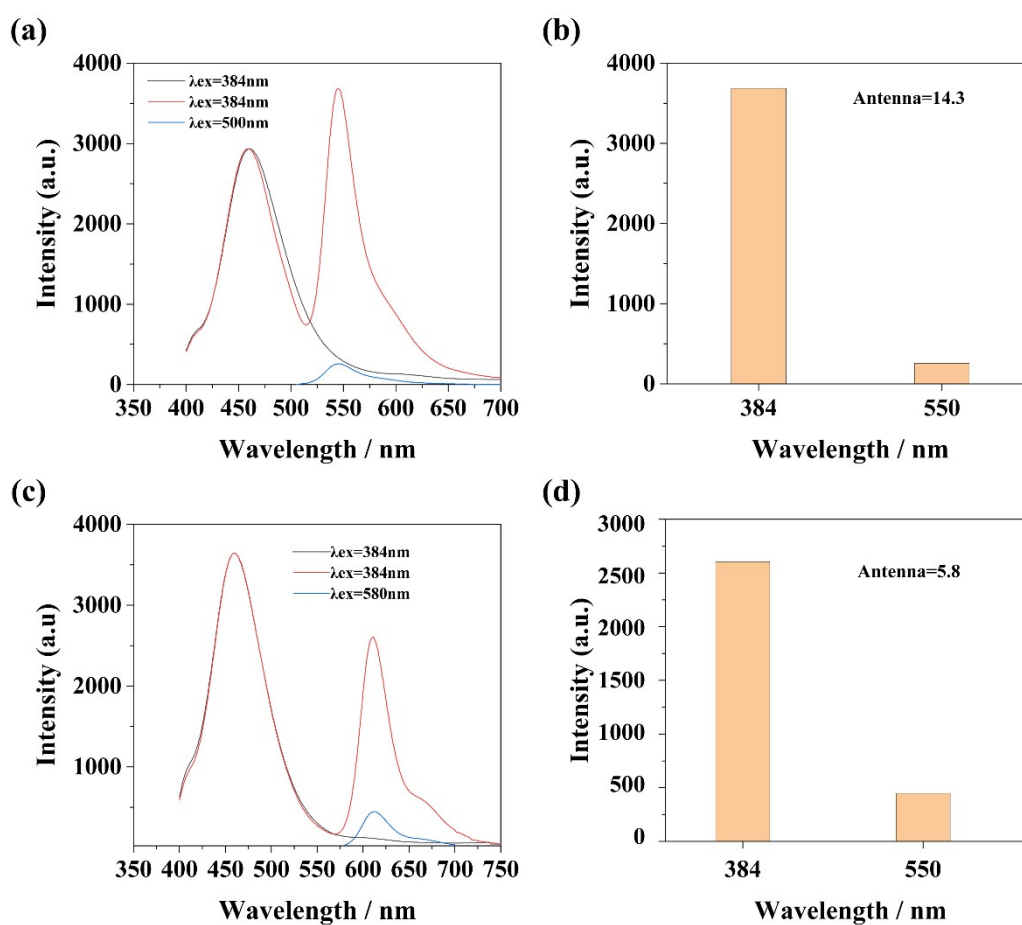
**Figure S53.** Fluorescence spectra of MPy3 with different concentrations of ESY in H<sub>2</sub>O-DMSO (3:2; v/v).



**Figure S54.** (a) Normalized absorption spectrum of SR101 and emission spectrum of **MPy1**. (b) Fluorescence spectra of **MPy1** with different concentrations of SR101 ( $\lambda_{\text{ex}} = 384 \text{ nm}$ ) in  $\text{H}_2\text{O}$ -DMSO (3:2; v/v). (c) Fluorescence spectra of **MPy1**, **MPy1**-SR101, SR101 ( $\lambda_{\text{ex}} = 384 \text{ nm}$ ) in  $\text{H}_2\text{O}$ -DMSO (3:2; v/v). (d) Fluorescence decay profiles of **MPy1** nanoparticles (black line) and **MPy1**-SR101 nanoparticles (**MPy1**:SR101 = 100:1) in  $\text{H}_2\text{O}$ -DMSO (3:2; v/v).



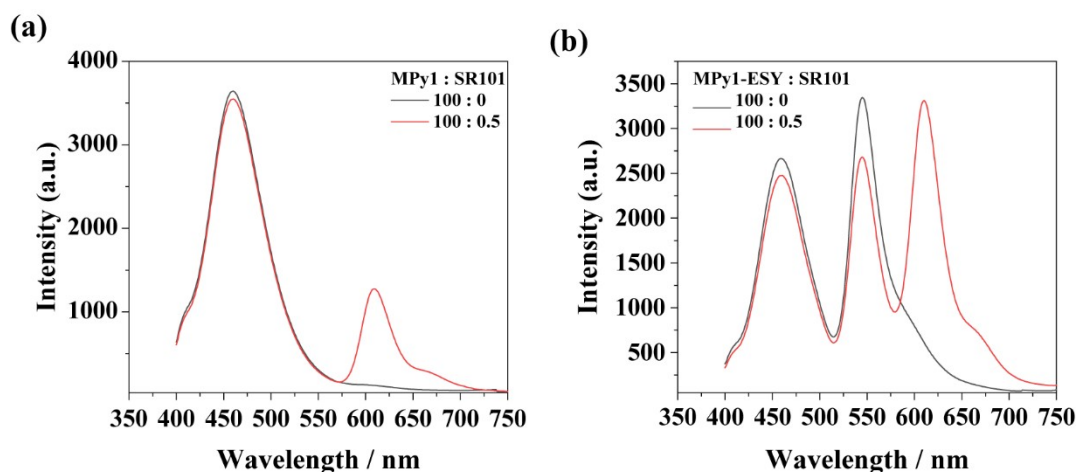
**Figure S55.** Fluorescence spectra of (a) MPy1 and MPy1-ESY assembly and (b) MPy1 and MPy1-SR101 assembly.  $[\text{MPy1}] = 1.0 \times 10^{-5} \text{ M}$ .



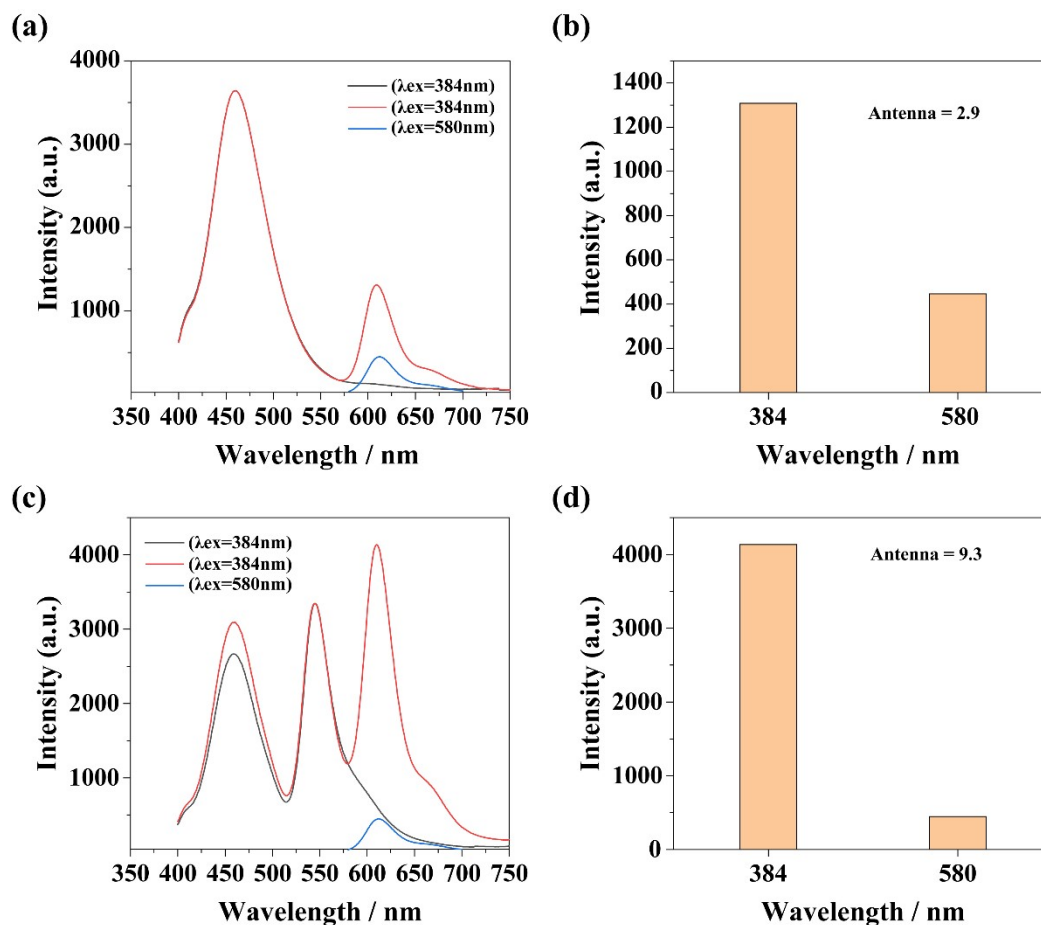
**Figure S56.** (a) Fluorescence spectra of MPy1-ESY (black line,  $[\text{MPy1}] = 1.0 \times 10^{-5} \text{ M}$ ,  $[\text{ESY}] = 1.0 \times 10^{-7} \text{ M}$ ,  $\lambda_{\text{ex}} = 384 \text{ nm}$ ), the donor (red line,  $\lambda_{\text{ex}} = 384 \text{ nm}$ ) and



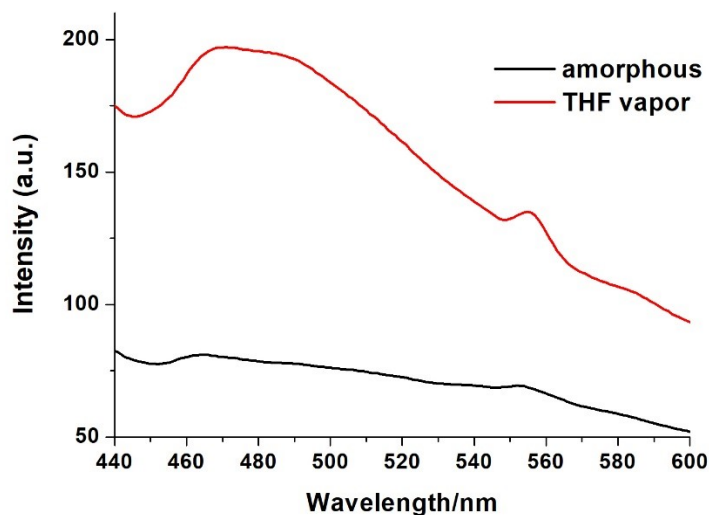
acceptor (blue line,  $\lambda_{\text{ex}} = 500$  nm). The fluorescence intensity of **MPy1**–ESY at 460 nm was normalized according to the fluorescence intensity of the donor; (b) The histogram of the emission intensity of **MPy1**–ESY at 550 nm on the excitation of the donor ( $\lambda_{\text{ex}} = 384$  nm) and acceptor ( $\lambda_{\text{ex}} = 500$  nm). (c) Fluorescence spectra of **MPy1**–SR101 (black line,  $[\text{MPy1}] = 1.0 \times 10^{-5}$  M,  $[\text{SR101}] = 1.0 \times 10^{-7}$  M,  $\lambda_{\text{ex}} = 384$  nm), the donor (red line,  $\lambda_{\text{ex}} = 384$  nm) and the acceptor (blue line,  $\lambda_{\text{ex}} = 580$  nm). The fluorescence spectrum of **MPy1**–SR101 was normalized according to the fluorescence intensity at 460 nm; (d) The histogram of the emission intensity of **MPy1**–SR101 at 610 nm on the excitation of the donor ( $\lambda_{\text{ex}} = 384$  nm) and acceptor ( $\lambda_{\text{ex}} = 580$  nm).



**Figure S57.** Fluorescence spectra of a) **MPy1** and **MPy1**–SR101 assembly, b) **MPy1**–ESY and **MPy1**–ESY–SR101 assembly.  $[\text{MPy1}] = 1.0 \times 10^{-5}$  M.



**Figure S58.** (a) Fluorescence spectra of **MPy1**–SR101 (black line,  $[\text{MPy1}] = 1.0 \times 10^{-5} \text{ M}$ ,  $[\text{SR101}] = 5.0 \times 10^{-8} \text{ M}$ ,  $\lambda_{\text{ex}} = 384 \text{ nm}$ ), the donor (red line,  $\lambda_{\text{ex}} = 384 \text{ nm}$ ) and acceptor (blue line,  $\lambda_{\text{ex}} = 580 \text{ nm}$ ). The fluorescence intensity of **MPy1**–SR101 at 460 nm was normalized according to the fluorescence intensity of the donor; (b) The histogram of the emission intensity of **MPy1**-SR101 at 610 nm on the excitation of the donor ( $\lambda_{\text{ex}} = 384 \text{ nm}$ ) and acceptor ( $\lambda_{\text{ex}} = 580 \text{ nm}$ ). (c) Fluorescence spectra of **MPy1**–ESY–SR101 (black line,  $[\text{MPy1}] = 1.0 \times 10^{-5} \text{ M}$ ,  $[\text{ESY}] = 1.0 \times 10^{-7} \text{ M}$ ,  $[\text{SR101}] = 5.0 \times 10^{-8} \text{ M}$ ,  $\lambda_{\text{ex}} = 384 \text{ nm}$ ), the donor (red line,  $\lambda_{\text{ex}} = 384 \text{ nm}$ ) and the acceptor (blue line,  $\lambda_{\text{ex}} = 580 \text{ nm}$ ). The fluorescence spectrum of **MPy1**–ESY–SR101 was normalized according to the fluorescence intensity at 550 nm; (d) The histogram of the emission intensity of **MPy1**–ESY–SR101 at 610 nm on the excitation of the donor ( $\lambda_{\text{ex}} = 384 \text{ nm}$ ) and acceptor ( $\lambda_{\text{ex}} = 580 \text{ nm}$ ).



**Figure S59.** Fluorescence spectra of **MPy1** in different solid states: amorphous and fuming with THF vapor for 5 min ( $\lambda_{\text{ex}} = 384$  nm).

## 9. References

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