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Electronic Supplementary Information

Thiadiazole-functionalized metal–organic frameworks for

photocatalytic C-N and C-C coupling reactions: Tuning ROS

generation efficiency via cobalt introduction

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Section 1. General

1.1. Materials and physical measurements

All reagents and solvents were purchased from commercial sources and used as received. $Co(NO_3)_2 \cdot 6H_2O$, solvents and organic substrates were purchased from Energy Chemical, Shanghai tengzhun Biotechnology Co., Ltd and Sigma-Aldrich Co., Inc. All heating reactions were heated by metal sand bath (WATTCAS, LAB-500, https://www.wattcas.com). The ligand 4,7-di(1H-pyrazol-4yl)benzo[c][1,2,5]thiadiazole (H₂PBT) was synthesized using our previously reported procedure. Fourier transform infrared (FT-IR) spectra were collected on a Thermo Scientific Nicolet iS10 spectrophotometer in the range of 4000–400 cm⁻¹. Powder Xray diffraction (PXRD) was performed on Rigaku Ultima IV diffractometer (Cu Ka radiation, $\lambda = 1.5406$ Å). Single-crystal X-ray diffraction (SCXRD) data were collected at 100 K, via an Oxford Cryo stream system on a XtaLAB PRO MM007-DW diffractometer system equipped with a RA-Micro7HF-MR-DW(Cu/Mo) X-ray generator and HyPix-6000HE Hybrid Photon Counting (HPC) X-ray detector (Rigaku, Japan, Cu K α , graphite monochromator, $\lambda = 1.54$ Å). CCDC-2173247 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Centre Data www.ccdc.cam.ac.uk/structures. Thermogravimetric analysis (TGA) curves were obtained on Mettler-Toledo (TGA/DSC) thermal analyzer from 40 °C to 800 °C with a heating rate of 10 °C min⁻¹ under a nitrogen gas atmosphere (20 mL·min⁻¹). Nitrogen gas adsorption/desorption measurements were performed on Micromeritics ASAP 2020 Plus adsorption instrument UV-Visible diffuse reflection spectra were recorded on Agilent Cary 4000 with BaSO₄ as the reference. X-ray photoelectron spectroscopy (XPS) data were collected on a Thermo ESCALAB 250XI system. Solid- and solutionstate luminescence spectra were measured on Horiba FluoroLog-3 spectrofluorometer. Decay curve was measured on Horiba FluoroMax-4 fluorometer with a NanoLED-455 flash lamp.

1.2. Synthesis of ligand H₂PBT

1,4-bis(1H-pyrazol-4-yl) benzothiadiazole (H₂PBT) was synthesized according to our previously reported method.¹

To a solution of 4,7-dibromobenzo[c][1,2,5]thiadiazole (1.0 g, 3.4 mmol) K₂CO₃ (4.37 g, 31.64 mmol) and Pd(PPh₃)₄ (110.4 mg, 0.078 mmol) in 80 mL of 1,2-Dimethoxyethane (DME). After being stirred and refluxed for 24 h at 80 °C under Ar. After the reaction is complete, the solvent was removed in vacuo. The obtained orange solid was separated and purified by column chromatography (EtOAc : PE = 1: 3) to obtain a large amount of orange yellow solid, which was the intermediate 4,7-bis(1-(tetrahydro-2H-pyran-2-yl)-1H-pyrazol-4-yl)benzo[c][1,2,5]thiadiazole with yield of about 88%.

¹H NMR (400MHz, CDCl₃): δ = 1.75 (m, 6H), 2.19 (m, 6H), 3.78 (m, 2H), 4.15 (dd, 2H), 5.51 (dd, 2H), 7.79 (s, 2H), 8.26 (s, 2H), 8.73 (s, 2H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 22.45, 24.98, 30.58, 67.92, 87.92, 119.04, 123.39, 124.59, 127.87, 137.8, 153.14 ppm.

To a solution of intermediate (1.22 g, 4.25mmol) in EtOH (50 mL), add HCl (25 mL, 1M), the resulting mixture was refluxed for 10 h at 80 °C. After the reaction is complete, the precipitate was filtered and washed with EtOH. Pour the obtained solid into a beaker containing 100 mL of distilled water, adjust the pH to 5–7 with NaOH (0.5 M), filter, and wash with distilled water to neutral. Then rinse with a small amount of ethanol and ether to afford H₂PBT as a dark-yellow solid H₂PBT with yield of about 95%. ¹H NMR (400 MHz, *d*₆-DMSO): δ = 8.53 (s, 4H), 8.01 (s, 2H). ¹³C NMR (100 MHz, *d*₆-DMSO): δ = 153.53, 133.21, 125.31, 123.50, 117.76 ppm. IR (KBr, cm⁻¹): 3133(m), 3045(m), 2950(w), 2741(w), 1685(w), 1590(s), 1576(w), 1560(w), 1519(s), 1396(w), 1357(m), 1335(m), 1276(m), 1243(m), 1152(m), 1045(m), 1005(s), 958(s), 884(s), 853(m), 835(s), 672(w), 605(s), 553(m), 540 (w), 505(w).



Fig. S1. The synthesis procedure of ligand H₂PBT.

1.3. Synthesis of JNU-207

A mixture of H₂PBT (4,7-di(1H-pyrazol-4-yl)benzo[*c*][1,2,5]thiadiazole, 6 mg, 0.02 mmol), Co(NO₃)₃·6H₂O (5.8 mg, 0.02 mmol), *N*, *N*-dimethylformamide (DMF, 3 mL), and HNO₃/H₂O (12/50 mL, 130 μ L) was sealed in a 10 mL glass vial and placed in an oven at 120 °C for 72 h. After cooling to room temperature, the black prismatic crystals of **JNU-207** were collected (46% yield, based on H₂PBT). The fresh crystalline samples were solvent exchanged with DMF and EtOH for three days and then heated up to 100 °C overnight before the next use. FT-IR (cm⁻¹): *v* = 3351.34 (w), 1664 (s), 1587 (s), 1546 (s), 1406 (w), 1376 (m), 1295 (w), 1248 (s), 1172 (w), 1052 (s), 1016 (m), 880 (m), 834 (s), 643 (w), 610 (s), 544 (s), 470 (m), 463 (m).

Note that **JNU-207** can also be obtained using a thick-walled pressure flask heated in a sand bath and this method can be scaled up appropriately. H₂PBT (0.11 mmol, 29.5 mg) and Co(OAc)₂·4H₂O (0.19 mmol, 48 mg) were dissolved in 8.0 mL DMF and 4.0 mL water in a 25 mL thick-walled pressure flask. The flask was then tightly sealed and the resulting suspension was heated in a sand bath at 150 °C for 12 h. After cooling naturally to room temperature, and the target product was obtained.



Fig. S2. All heating reactions were heated by metal sand bath (WATTCAS, LAB-500).

Section 2. Characterization of JNU-207

2.1 Crystallographic data

Single crystal structures of MOFs were measured by X-ray diffraction at 100 K on XtaLAB PRO MM007-DW diffractometer system equipped with a RA-Micro7HF-MR-DW(Cu/Mo) X-ray generator and HyPix-6000HE Hybrid Photon Counting (HPC) X-ray detector (Rigaku, Japan, Cu K α , graphite monochromator, $\lambda = 1.54178$ Å). The structure was solved by direct methods and refined by full-matrix least-squares refinements based on F². Anisotropic thermal parameters were applied to all non-hydrogen atoms. The crystallographic calculations were performed using Olex 2 with 'XL' plugins. CCDC-2173247 (JNU-207), contain the supplementary crystallographic data for this paper.

MOF	JNU-207
CCDC number	2173247
Empirical formula	$C_{12}H_6N_6SCo$
Formula weight	319.29
Crystal system	tetragonal
Space group	<i>I</i> 4 ₁ 22
a/Å	16.5978(2)
$b/{ m \AA}$	16.5978(2)
$c/{ m \AA}$	12.3112(2)
V/Å ³	3391.58(10)
$\alpha/^{\circ}$	90
$eta/^\circ$	90
γ/°	90
Z	8
$D_{\rm C}/{\rm g~cm^{-3}}$	1.274
μ/mm^{-1}	9.079
λ/Å	1.54184
T/K	100
Reflections collected	6012
Independent reflections	1758 [$R_{int} = 0.0331$]
Goodness-of-fit on F ²	1.089
$R_1^a, w R_2^b$ [I > 2 σ (I)]	$R_1 = 0.0795, wR_2 = 0.2213$
$R_1^{\rm a}$, $wR_2^{\rm b}$ (all data)	$R_1 = 0.0823, wR_2 = 0.2240$
Largest diff. peak and hole /e. $Å^{-3}$	0.62/-0.83
Flack parameter	0.037(14)

 Table S1. Crystal data and structure refinement for JNU-207.

 $aR_{1} = \Sigma ||F_{o}| - |F_{c}|| / \Sigma |F_{o}|$ $bWR_{2} = \{\Sigma[w (F_{o}^{2} - F_{c}^{2})^{2}] / [w (F_{o}^{2})^{2}]\} 1/2, [F_{o} > 4\sigma (F_{o})]$



Fig. S3. Views of **JNU-207** along the [001] direction with uniformed square 1D channels (colour code: C, gray; N, blue; S, yellow and Co, violet; H atoms are omitted for clarity).



Fig. S4. (a) CoN_4 and (b) ZnN_4 clusters are connected through edge-sharing pyrazoles to form helical rod SBUs. Linker distortion between the pyrazole plane and the benzothiadiazole plane (c) **JNU-207** and (d) **JNU-204** (Co, violet; Zn, sky blue; C, gray; N, blue; H atoms are omitted for clarity).

2.2. Additional characterization



Fig. S5. FT-IR spectra of H₂PBT and JNU-207.



Fig. S6. High resolution XPS spectra of Co elements in JNU-207.



Fig. S7. High resolution XPS spectra of Zn elements in JNU-204.



Fig. S8. SEM image of JNU-207.



Fig. S9. SEM image of JNU-204.



Fig. S10. SEM-EDS mapping and EDS spectra profiles of JNU-207.



Fig. S11. SEM-EDS mapping and EDS spectra profiles of JNU-204.



Fig. S12. Comparison of the PXRD patterns of JNU-207 and JNU-204.



Fig. S13. Comparison of the PXRD patterns of JNU-207 after being treated with different aqueous solutions at different pH values.



Fig. S14. Thermogravimetric analysis (TGA) of JNU-207.

2.3. Electrochemical measurements.

10 mg of the MOF was ground and added into 100 μ L of ethanol and 10 μ L of Nafion (5 wt%), sonicated for 30 min to afford a finely dispersed suspension. Then, the obtained suspension was coated on the surface of fluorine-doped tin oxide (FTO) glass, scraped into a uniform film and dried for 0.5 hours. The electrochemical measurements were performed in a conventional three-electrode cell on a CHI-760E electrochemical workstation (Shanghai Chenhua Instrument Co., Ltd, China), with the MOF-coated FTO glass as working electrode, Pt wire as the counter electrode, and Ag/AgCl as the 300W Xe reference electrode. lamp (China Education Au-light, CEL-HXF300/CEL-HXUV300) coupled with a UV cutoff filter ($\lambda \ge 420$ nm) was used to simulate the visible light for the photocurrent measurements. The photoresponsive signals were measured under visible light irradiation at 0.4 V in a 0.5 M Na₂SO₄ solution, and Mott Schottky plots were made by using three different frequencies, 1000, 2000, and 3000 Hz. Electrochemical impedance spectroscopy (EIS) measurements were carried out in a 0.5 M Na₂SO₄ solution, the amplitude of the sinusoidal wave was 5 mV with frequency ranging from 100 kHz to 0.05 Hz.



Fig. S15. Schematic depiction of the optical band gap of JNU-207 and JNU-204.

Table S2. Fitting data of the photoluminescence decay of JNU-207 and JNU-204.

	Al	A2	A3	τ_1	τ_2	τ ₃	$\tau_{\rm av}$	Chi2
JNU-207	0.6253725	2.551819E-	5.612912E-05	1.827734E-	1.840305E-	6.459449E-08	0.25 ns	1.54
		02		10 sec	09 sec	sec		
JNU-204	0.2242236	5.310204E-	1.372023E-04	1.152505E-	2.495059E-	9.16784E-08 sec	1.45 ns	1.33
		02		09 sec	09 sec			

 $\tau_{av} = (A1 \times \tau_1^2 + A2 \times \tau_2^2 + A3 \times \tau_3^2)/(A1 \times \tau_1 + A2 \times \tau_2 + A3 \times \tau_3).$

Section 3. Photocatalytic Application

3.1 photocatalytic synthesis of imines

General procedure A:

A solution of primary amine (1) (1.001 mmol) and JNU-207 (1 mol%) in CH₃CN (5 mL) was stirred under blue LED for 24 h at room temperature. After evaporation of solvent, the crude residue was dissolved in CDCl₃ along with 20 μ L CHCl₂CHCl₂ as internal standard for crude NMR.



A solution of benzylamine (1a) (126.4 μ L, 1.001 mmol), JNU-207 (1 mol%) in CH₃CN (5 mL) was stirred under blue LED for 24 h at room temperature. After

evaporation of solvent, the crude residue was dissolved in CDCl₃ along with 20 μ L CHCl₂CHCl₂ as internal standard for crude NMR. Based on crude ¹H NMR, the desired product **2a** was detected in 94% yield.

(E)-N-benzyl-1-phenylmethanimine (2a)



NMR yield: 94%. ¹H NMR (400 MHz, CDCl₃) 4.81 (2H, s), 7.26-7.33 (8H, m), 7.68-7.87 (2H, m), 8.38 (1H, s).

(E)-N-(4-methylbenzyl)-1-(p-tolyl)methanimine (2b)



NMR yield: 80%. ¹H NMR (400 MHz, CDCl₃) 2.24 (3H, s), 2.28 (3H, s), 4.67 (2H, s), 7.04-7.14 (6H, m), 7.57 (2H, d, *J* = 8.0 Hz), 8.24 (1H, s).

(E)-N-(4-butylbenzyl)-1-(4-butylphenyl)methanimine (2c)



NMR yield: 95%. ¹H NMR (400 MHz, CDCl₃) 0.97-1.01 (6H, m), 1.37-1.46 (4H, m), 1.61-1.71 (4H, m), 2.64-2.71 (4H, m), 4.83 (2H, s), 7.21 (2H, d, *J* = 8.0 Hz), 7.26-7.31 (4H, m), 7.75 (2H, d, *J* = 8.4 Hz), 8.40 (1H, s).

(E)-N-(4-methoxybenzyl)-1-(4-methoxyphenyl)methanimine (2d)



NMR yield: 88%. ¹H NMR (400 MHz, CDCl₃) 3.70 (3H, s), 3.74 (3H, s), 4.64 (2H, s), 6.79 (2H, d, *J* = 8.4 Hz), 6.84 (2H, d, *J* = 8.8 Hz), 7.26 (2H, d, *J* = 8.8 Hz), 7.63 (2H, d, *J* = 8.8 Hz), 8.21 (1H, s).

(E)-N-(4-chlorobenzyl)-1-(4-chlorophenyl)methanimine (2e)



NMR yield: 88%. ¹H NMR (400 MHz, CDCl₃) 4.75 (2H, s), 7.21-7.31 (6H, m), 7.37 (2H, d, *J* = 8.8 Hz), 7.69 (2H, d, *J* = 8.4 Hz), 8.32 (1H, s).

(E)-N-(4-fluorobenzyl)-1-(4-fluorophenyl)methanimine (2f)



NMR yield: 96%. ¹H NMR (400 MHz, CDCl₃) 4.67 (2H, s), 6.94 (2H, dddd, *J* = 2.0, 2.8, 8.4, 8.8 Hz), 7.01 (2H, dddd, *J* = 2.0, 2.8, 8.4, 8.8 Hz), 7.20 (2H, dddd, *J* = 2.0, 2.8, 5.2, 8.4 Hz), 7.68 (2H, dddd, *J* = 2.0, 2.8, 5.2, 8.4 Hz), 8.25 (1H, s).

(E)-N-(3-methylbenzyl)-1-(m-tolyl)methanimine (2g)



NMR yield: 89%. ¹H NMR (400 MHz, CDCl₃) 2.33 (3H, s), 2.36 (3H, s), 4.76 (2H, s), 7.05-7.13 (3H, m), 7.19-7.23 (2H, m), 7.29 (1H, dd, *J* = 7.2, 7.6 Hz), 7.53 (1H, d, *J* = 7.6 Hz), 7.63 (1H, s), 8.33 (1H, s).

(E)-N-(2-methylbenzyl)-1-(o-tolyl)methanimine (2h)



NMR yield: 81%. ¹H NMR (400 MHz, CDCl₃) 2.38 (3H, s), 2.49 (3H, s), 4.81 (2H, s), 7.15-7.28 (7H, m), 7.92 (1H, d, *J* = 7.6), 8.65 (1H, s).

Table S3. Representative heterogeneous porous catalysts for the oxidative coupling of benzylamines.

Catalyst	Reaction conditions	Yield	Reference
MOF-6 (Ru(bpy) ₃ ²⁺	38μl [B], 1 mol % [P], 3 mL CH ₃ CN, 300W Xe lamp O ₂ , 60 °C 1h	83%	2
NH ₂ -MIL125(Ti)	0.1 mmol [B], 5 mg [P], 2 mL CH ₃ CN, 300W Xe lamp, O ₂ , 12h	73%	3
PCN-222 (Zr)	0.1 mmol [B], 5 mg [P], 3 mL CH ₃ CN, 300W Xe lamp, air, 1h	100%	4
UNLPF-12 (Zr) (SnIVporphyrin)	0.27 mmol [B], 1.0 μmol [P], 1 mL dry CH ₃ CN, 14 W CFL, air, 2h	99%	5
Zn-PDI	1 mmol [B], 1 mol% [P], 5 mL CH ₃ CN, 500W Xe lamp air, 4h	74%	6
[In(OH)(ADBEB)]·DMF	0.2 mmol [B], 4 mg [P], 1 mL DMSO, visible light, O ₂ , 2.67 h	99%	7
Cd-MOF	0.48 mmol [B], 10 mg [P], 5 mL DMF, 300 W Xe lamp air, 7 h	99.1%	8
ZIF-8	11 μL [B], 10 mg [P], 1 mL anhydrous DMF, 450 nm LED, O ₂ , 7h	3.5%	9
UiO-66	11 μL [B], 10 mg [P], 1 mL dry DMF, 450 nm LED, O ₂ , 7h	3.0%	9
MIL-125	11 μL [B], 10 mg [P], 1 mL dry DMF, 450 nm LED, O ₂ , 7h	3.3%	9
NP5-DM-COF	0.2 mmol [B], 3 mg [P], 2 mL CH ₃ CN, 30W LED, r.t.,	99%	10

	O ₂ , 15h		
Zn ₂ (DPNDI)(W ₁₀ O ₃₂)(DMA) ₆	0.4 mmol [B], 1 mol% [P], 2 mL DMF,100 W white LED lamp, O ₂ , 24h	99%	11
RPF-30-Er	0.05 mmol [B], 10 mol% [P], 1 mL CH ₃ CN, 100 W warming lamp O ₂ , 18h	76%	12
ZJU-56-0.2	1 mmol [B], 1.0 mol % [P], 2 mL CH ₃ CN, 660 nm LEDs, 60 °C, O ₂ , 24h	62%	13
Ni MOF-74 nanosheets	0.1 mmol [B], 10 mg [P], 2 mL CH ₃ CN, 300 W Xenon Lamp, O ₂ , 4 h		14
FJI-Y10	1 mmol [B], 2 mmol % [P] basing on the L, 5 mL DMF, 300 W Xe lamp, 40 °C, O ₂ , 6 h.	100%	15
Zn2(diPyPI-Cl4)(NDC)2·3DMF	0.2 mmol [B], 0.5 mol% [P], 300W Xe lamp O ₂ , r.t., 6h	100%	16
JNU-204	1 mmol [B], 1mol% [P], 5mL CH ₃ CN, blue LED, air, 86% r.t., 24h		This work
JNU-207	1 mmol [B], 1mol% [P], 5mL CH ₃ CN, blue LED, air, r.t., 24h	94%	This work

[B] = benzylamine; [P] = photocatalyst; r.t. = room tempature.

3.2 photocatalytic synthesis of tetrahydroquinolines

General procedure B:

A solution of tertiary amine (3) (1.001 mmol), maleimide (4) (0.501 mmol), JNU-207 (1 mol%) in DMF (5 mL) was stirred under blue LED for 48 h at room temperature. After completion of the reaction, the reaction was extracted with ethyl acetate. The combined organic layer was washed with brine, dried over Na₂SO₄, filtered and concentrated. The crude residue was dissolved in CDCl₃ along with 20 μ L CHCl₂CHCl₂ as internal standard for crude NMR.



A solution of *N*,*N*-Dimethylaniline (3a) (126.4 μ L, 1.001 mmol), *N*-Phenylmaleimide (4a) (86.7 mg, 0.501 mmol), **JNU-207** (1 mol%)) in DMF (5 mL) was stirred under blue LED for 48 h at room temperature. After completion of the reaction, the reaction was extracted with ethyl acetate. The combined organic layer was washed with brine, dried over Na₂SO₄, filtered and concentrated. The crude residue was dissolved in CDCl₃ along with 20 μ L CHCl₂CHCl₂ as internal standard for crude NMR. Based on crude ¹H NMR, the desired cyclization product **5a** was detected in 78% yield.

5-Methyl-2-phenyl-3a,4,5,9b-tetrahydro-1*H*-pyrrolo[3,4-c]quinoline-1,3(2*H*)-dion





NMR Yield: 77%. ¹H NMR (400 MHz, CDCl₃) δ 2.75 (3H, s), 3.03 (1H, dd, J = 4.4, 11.6 Hz), 3.42-3.46 (1H, m), 3.52 (1H, dd, J = 2.4, 11.2 Hz), 4.06 (1H, d, J = 9.6 Hz), 6.67 (1H, d, J = 8.0 Hz), 6.83 (1H, dd, J = 7.2, 7.6 Hz), 7.13-7.18 (3H, m), 7.27 (1H, dd, J = 7.2, 7.2 Hz), 7.34 (2H, dd, J = 7.2, 8.0 Hz), 7.44 (1H, d, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃) δ = 39.4, 42.0, 43.4, 50.6, 112.6, 118.5, 119.7, 126.3, 128.5, 128.6, 128.9, 130.3, 131.9, 148.4, 175.7, 177.6.

2-(4-Methoxyphenyl)-5-methyl-3a,4,5,9b-tetrahydro-1*H*-pyrrolo[3,4-*c*]quinoline-1,3(2*H*)-dione (5b)



NMR Yield: 76%. ¹H NMR (400 MHz, CDCl₃) δ 2.87 (3H, s), 3.16 (1H, dd, J = 4.4, 11.6 Hz), 3.53-3.58 (1H, m), 3.63 (1H, dd, J = 2.8, 11.2 Hz), 3.83 (3H, s), 4.17 (1H, d, J = 9.6 Hz), 6.78 (1H, d, J = 8.0 Hz), 6.92-6.97 (3H, m), 7.20 (2H, ddd, J = 3.0, 3.2, 9.2 Hz), 7.26 (1H, dd, J = 7.2, 8.4 Hz), 7.56 (1H, d, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃) δ = 39.5, 42.0, 43.4, 50.6, 55.5, 112.7, 114.3, 118.7, 119.8, 124.6, 127.6, 128.7, 130.3, 148.3, 159.4, 175.9, 177.9.

2-(4-Chlorophenyl)-5-methyl-3a,4,5,9b-tetrahydro-1*H*-pyrrolo[3,4-*c*]quinoline-1, 3(2*H*)-dione (5c)



NMR Yield: 77%. ¹H NMR (400 MHz, CDCl₃) δ 2.77 (3H, s), 3.05 (1H, dd, J = 4.4, 11.2 Hz), 3.45-3.49 (1H, m), 3.54 (1H, dd, J = 2.8, 11.6 Hz), 4.09 (1H, d, J = 9.6 Hz), 6.68 (1H, d, J = 8.0 Hz), 6.84 (1H, ddd, J = 0.8, 7.2, 7.6 Hz), 7.15-7.19 (3H, m), 7.32 (2H, ddd, J = 2.0, 2.8, 8.8 Hz), 7.45 (2H, d, J = 7.2 Hz); ¹³C NMR (100 MHz, CDCl₃) $\delta = 39.5$, 42.0, 43.4, 50.6, 55.5, 112.7, 114.3, 118.7, 119.8, 124.6, 127.6, 128.7, 130.3, 148.3, 159.4, 175.9, 177.9.

5-Methyl-2-(*m*-tolyl)-3a,4,5,9b-tetrahydro-1*H*-pyrrolo[3,4-*c*]quinoline-1,3(2*H*)dione (5d)



NMR Yield: 72%. ¹H NMR (400 MHz, CDCl₃) δ 2.27 (3H, s), 2.77 (3H, s), 3.06 (1H, dd, J = 4.4, 11.6 Hz), 3.44-3.48 (1H, m), 3.54 (1H, dd, J = 2.4, 11.2 Hz), 4.08 (1H, d, J = 9.6 Hz), 6.69 (1H, d, J = 8.4 Hz), 6.84 (1H, dd, J = 7.2, 7.6 Hz), 6.97-6.98 (2H, m), 7.09 (1H, d, J = 7.6 Hz), 7.17 (1H, dd, J = 5.6, 8.0 Hz), 7.24 (1H, dd, J = 7.6, 8.4 Hz), 7.46 (1H, d, J = 7.6 Hz); ¹³C NMR (100 MHz, CDCl₃) $\delta = 21.3$, 39.5, 42.1, 43.4, 50.6, 112.7, 118.6, 119.8, 123.4, 126.9, 128.7, 128.8, 129.4, 130.3, 131.8, 139.1, 148.3, 175.8, 177.7.

5-Methyl-2-(*o*-tolyl)-3a,4,5,9b-tetrahydro-1*H*-pyrrolo[3,4-*c*]quinoline-1,3(2*H*)dione (5e)



NMR Yield: 67%, a mixture of diastereomers (dr = 2:1) was obtained. ¹H NMR (400 MHz, CDCl₃) δ 1.83 (3H, s), 2.20 (3H, s), 2.85 (3H, s), 3.09-3.16 (1H, m), 3.55-3.65 (1H, m), 4.17-4.19 (1H, m), 6.74-6.78 (1H, m), 6.89-7.14 (2H, m), 7.17-7.31 (2H, m), 7.49-7.53 (2H, m); ¹³C NMR (100 MHz, CDCl₃) δ = 17.0, 17.8, 39.2, 39.5, 42.4, 42.8, 43.7, 44.3, 50.8, 51.1, 112.3, 112.5, 118.6, 118.9, 119.7, 119.8, 126.77, 126.81, 127.8, 128.2, 128.7, 129.4, 129.5, 129.6, 130.3, 131.0, 131.1, 135.3, 135.9, 148.56, 148.60, 175.7, 175.8, 177.6, 177.9.

dione (5f)



NMR Yield: 78%. ¹H NMR (400 MHz, CDCl₃) δ 1.12-1.33 (3H, m), 1.50-1.64 (3H, m), 1.76-1.80 (2H, m), 2.01-2.17 (2H, m), 2.79 (3H, s), 3.02 (1H, dd, *J* = 4.4, 11.6 Hz), 3.25-3.29 (1H, m), 3.45 (1H, dd, *J* = 3.2, 11.6 Hz), 3.88-3.98 (2H, m), 6.69 (1H, d, *J* = 8.0 Hz), 6.88 (1H, dd, *J* = 7.2, 7.6 Hz), 7.20 (1H, ddd, *J* = 1.2, 6.8, 8.4 Hz), 7.46 (1H, d, *J* = 7.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ = 25.0, 25.69, 25.74, 28.7, 28.8, 39.3, 41.7, 43.0, 50.8, 52.1, 112.3, 119.0, 119.4, 128.3, 130.1, 148.3, 176.8, 178.6.





NMR Yield: 67%. ¹H NMR (400 MHz, CDCl₃) δ 2.22 (3H, s), 2.72 (3H, s), 2.97 (1H, dd, J = 4.4, 11.2 Hz), 3.40-3.44 (1H, m), 3.50 (1H, dd, J = 2.8, 11.6 Hz), 4.02 (1H, d, J = 9.6 Hz), 6.57 (1H, d, J = 8.4 Hz), 6.95 (1H, dd, J = 1.6, 8.4 Hz), 7.17-7.20 (2H, m), 7.32-7.36 (2H, m); ¹³C NMR (100 MHz, CDCl₃) $\delta = 20.4$, 39.6, 42.1, 43.5, 50.9, 112.6, 118.5, 126.3, 128.4, 128.9, 129.0, 129.2, 130.8, 132.0, 146.2, 175.8, 177.8.

8-Fluoro-5-methyl-2-phenyl-3a,4,5,9b-tetrahydro-1*H*-pyrrolo[3,4-*c*]quinoline-1,3 (2*H*)-dione (5h)



NMR Yield: 73%. ¹H NMR (400 MHz, CDCl₃) δ 2.84 (3H, s), 3.11 (1H, dd, J = 4.4, 11.2 Hz), 3.54-3.63 (2H, m), 4.14 (1H, d, J = 9.6 Hz), 6.71 (1H, dd, J = 4.8, 8.8 Hz), 6.97 (1H, ddd, J = 2.8, 6.0, 14.4 Hz), 7.11-7.18 (1H, m), 7.28-7.32 (3H, m), 7.44-7.48 (2H, m); ¹³C NMR (100 MHz, CDCl₃) $\delta = 39.8$, 42.1, 43.3, 51.0, 113.5 (d, J = 7.5 Hz), 115.1 (d, J = 21.9 Hz), 116.5 (d, J = 22.6 Hz), 117.0 (d, J = 23.3 Hz), 120.1 (d, J = 7.7 Hz), 126.3, 128.6, 129.0, 131.8, 144.8 (d, J = 1.9 Hz), 156.7 (d, J = 237.7 Hz), 177.4.

5-Ethyl-2-phenyl-3a,4,5,9b-tetrahydro-1*H*-pyrrolo[3,4-*c*]quinoline-1,3(2*H*)-dione (5i)



NMR Yield: 73%. ¹H NMR (400 MHz, CDCl₃) δ 1.11 (3H, t, *J* = 7.2 Hz), 3.11 (1H, dd, *J* = 4.0, 11.6 Hz), 3.15-3.31 (2H, m), 3.46-3.50 (1H, m), 3.58 (1H, dd, *J* = 2.8, 11.6 Hz), 4.07 (1H, d, *J* = 9.6 Hz), 6.71 (1H, d, *J* = 8.0 Hz), 6.79 (1H, t, *J* = 7.2 Hz), 7.12-7.19 (3H, m), 7.28 (1H, dd, *J* = 7.2, 7.6 Hz), 7.36 (2H, dd, *J* = 7.2, 8.0 Hz), 7.45 (1H, d, *J* = 7.6 Hz); ¹³C NMR (100 MHz, CDCl₃) δ = 10.8, 42.3, 43.8, 44.9, 47.1, 112.8, 118.6, 119.2, 126.3, 128.5, 128.6, 129.0, 130.7, 132.0, 146.9, 175.8, 177.7.

Table S4. Representative heterogeneous porous catalysts for the oxidative coupling of tertiary anilines with maleimides.

Catalyst	Reaction conditions	Yield	Reference

Ru(bpy) ₃ @NKMOF-	0.5 mmol [T], 0.25 mmol [M], 3 mol % [P], 3.0		17
4	mL DMF, two 23 W CFL, r.t., 36 h		1 /
D BBT 10	0.5 mmol [T], 0.25 mmol [M], 10 mg [P], 3.0 mL	67%	18
F-DD1-10	DMF, white LED, r.t., air, 24 h.	0770	10
U.D. Dub COF	0.2 mmol [T], 0.1 mmol [M], 8 mol% [P], 3 mL	700/	10
II2F-Bpil-COF	CHCl ₃ , blue LEDs (λ = 450 nm), r.t., O ₂ , 14 h		19
0.2 mmol [T], 0.1 mmol [M], 4 mg [P], 1 mL		e20/	20
CH ₃ CN, blue LED, air, r.t., 12 h		82%	20
JNU-204	0.5 mmol [T], 0.25 mmol [M], 1mol% [P], 5mL DMF, blue LED, air, r.t., 48h		This work
JNU-207	0.5 mmol [T], 0.25 mmol [M], 1mol% [P], 5mL DMF, blue LED, air, r.t., 48h	77%	This work

[T] = tertiary anilines; [M] = maleimides; [P] = photocatalyst; r.t. = room temparture.

3.3 Photocatalytic reaction mechanisms verification

We performed the photocatalytic synthesis of imines in the presence of a known O_2^{-} quencher, *p*-benzoquinone (BQ). As shown below, the significantly reduced yield of **2a** indicates that O_2^{-} is key intermediate in this reaction.



Fig. S16. Mechanistic studies of the photocatalytic synthesis of imines.

We performed the photocatalytic synthesis of tetrahydroquinoline derivatives in the presence of a known O_2^{\bullet} quencher, *p*-benzoquinone (BQ). As shown below, the significantly reduced yield of **5a** indicates that O_2^{\bullet} is key intermediate in this reaction.



Fig. S17. Mechanistic studies of the photocatalytic synthesis of tetrahydroquinoline

derivatives.

Section 4. Recycling experiments



According to the general procedure **A**, 10 parallel reactions were carried out at the same time. After the reaction completion, **JNU-207** was recovered by centrifugation, one of them was used to detect the yield by NMR, and the rest were all centrifuged, washed with acetonitrile and dried. Following the above reaction condition, the second cycle and third cycle of this reaction was performed, and **2a** could be assembled in 94% yield and 92% yield, respectively. The resulting powder JNU-207 was used for other characterizations.



According to the general procedure **B**, 10 parallel reactions were carried out at the same time. After the reaction completion, **JNU-207** was recovered by centrifugation, one of them was used to detect the yield by NMR, and the rest were all centrifuged, washed with acetonitrile and dried. Following the above reaction condition, the second cycle and third cycle of this reaction was performed, and **5a** could be assembled in 76% yield and 76% yield, respectively.



Fig. S18. Recycling experiments with **JNU-207** as the photocatalyst for synthesis of imines and synthesis of tetrahydroquinolines (the three cycles are light red, dark yellow, and olive, respectively).



Fig. S19. Comparison of PXRD patterns of the pristine **JNU-207** and the recovered **JNU-207** after three runs of each experiment (three reactions)



Fig. S20. FT-IR spectra of **JNU-207** before and after photocatalytic synthesis of (E)-N-benzyl-1-phenylmethanimine.

Table S5. ICP-AES results^[a] for JNU-207 after photocatalytic oxidation of benzylamine.

	$ ho({ m Co})$ / ppm
1	2.149
2	1.098
3	2.253

Section 5. Reference

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Appendix

¹H and ¹³C NMR spectra for new compounds.

¹H NMR (400 MHz, CDCl₃) spectrum of **2a**







4.811



¹H NMR (400 MHz, CDCl₃) spectrum of **2c**







 1 H NMR (400 MHz, CDCl₃) spectrum of **2e**





 1 H NMR (400 MHz, CDCl₃) spectrum of **2f**









¹H NMR (400 MHz, CDCl₃) spectrum of **5a**





¹H NMR (400 MHz, CDCl₃) spectrum of **5b**





¹H NMR (400 MHz, CDCl₃) spectrum of **5c**





























 ^{13}C NMR (100 MHz, CDCl₃) spectrum of **5h**





