

**Design of a label-free aptasensor for electrochemical determination of hemoglobin;
Investigation of the peroxidase-like activity of hemoglobin for the sensing of different
substrates**

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Supplementary Material

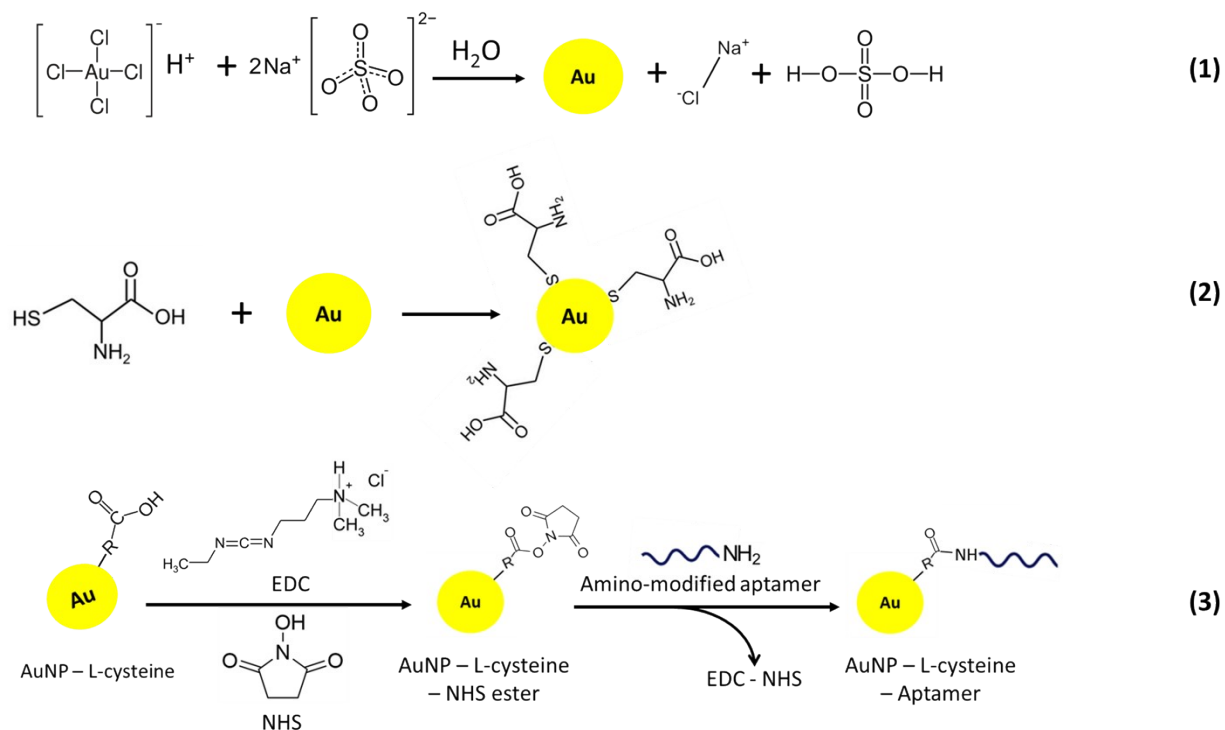


Fig. S1 Schematic representation of (1) the formation of gold nanoparticles by the interaction of Hydrogen tetrachloroaurate (HAuCl_4) and Sodium sulfate (Na_2SO_4); (2) the formation of cysteine modified gold nanoparticles via Au-thiol group (SH) interaction; (3) the immobilization of Aptamer on L-cysteine modified gold nanoparticles via EDC/NHS interaction.

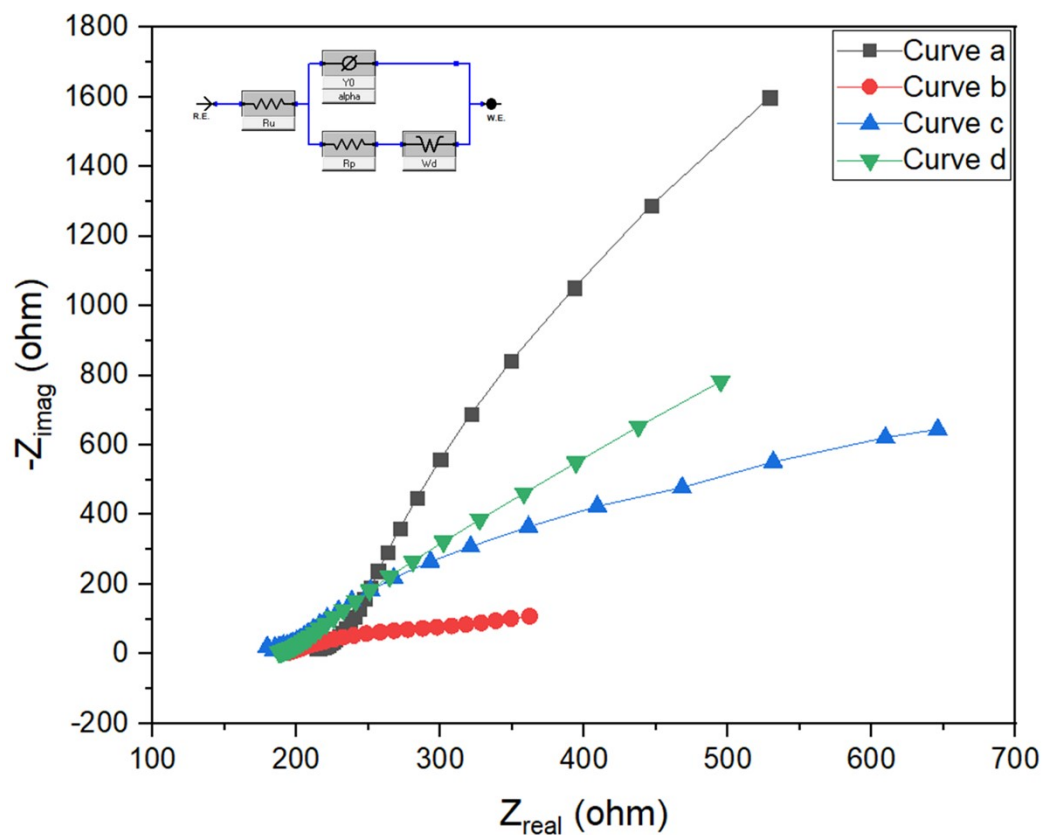


Fig. S2 Nyquist plots of 5 mM $[\text{Fe}(\text{CN})_6]^{4-/3-}$ redox couple in PBS, pH 7.4, for bare SPCE (curve a), AuNPs /SPCE(curve b), L-cysteine/ AuNPs /SPCE (Curve c), and Aptamer/ L-cysteine/ AuNPs/ SPCE (curve d) using a frequency range of 100000 Hz - 0.1 Hz, 10 point/decade and an AC voltage of 10 mv ms. Inset: Equivalent circuit (CPE with diffusion) $\text{Ru}(\emptyset[\text{RpWd}])$ used to fit the frequency scans along with the impedance spectra.

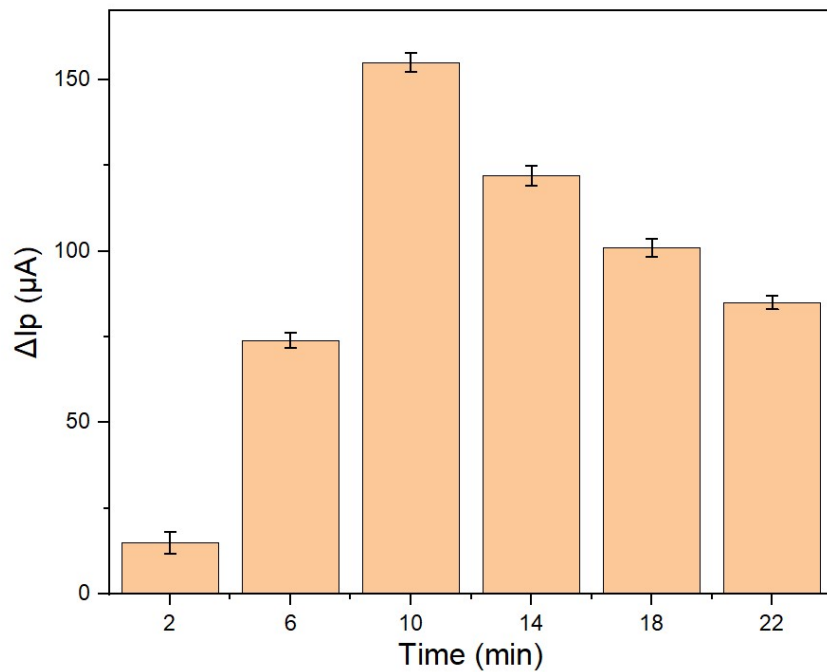


Fig. S3 Optimization of Hb-aptamer incubation time (2, 6, 10, 14, 18, 22 min). Plot of peak current difference vs. Hb incubation time obtaining from SWV in the potential range of -0.2 to +0.4 with a frequency of 7 Hz and an amplitude of 0.025V using 0.1 M PBS (pH 7.4) containing 5.0 mM $[\text{Fe}(\text{CN})_6]^{3-/4-}$.

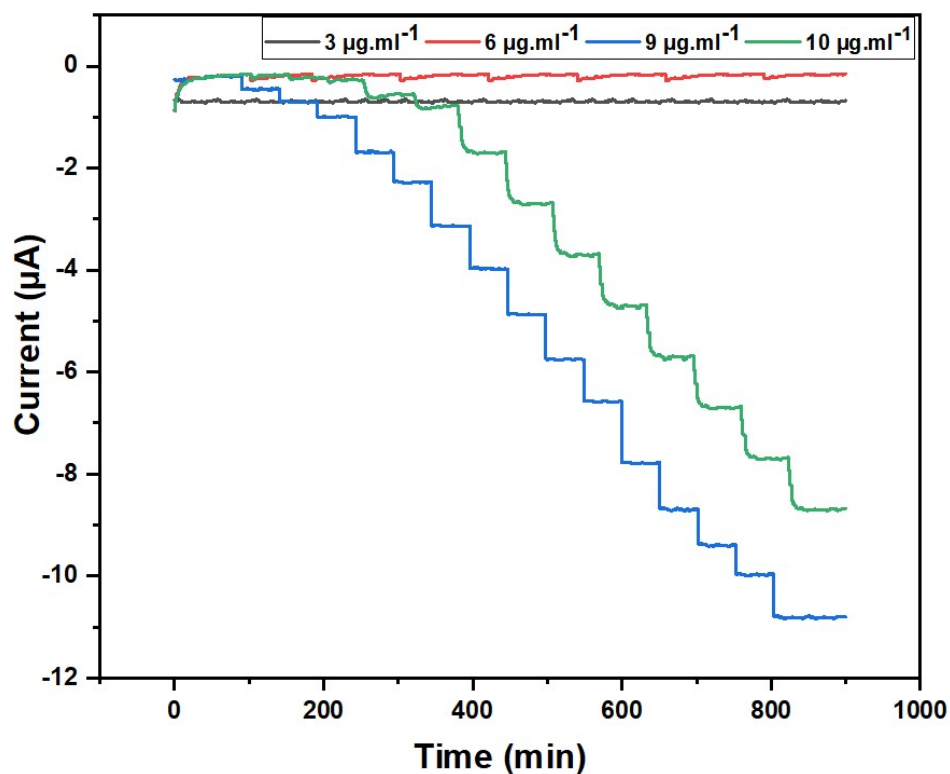


Fig. S4 Effect of Hb concentrations (3, 6, 9, 10 $\mu\text{g.ml}^{-1}$) on the H_2O_2 electrocatalytic activity using amperometry measurements toward successive additions of H_2O_2 in 0.1M PBS (pH 7) with an applied potential of -0.4V. Error bars correspond to the relative standard deviation of three independent experiments

Table S1. Comparison between this method and other reported electrochemical techniques for detection of H_2O_2 .

Method	Target	Principle	Linear range (μM)	LOD (μM)	Ref
Amperometry	H_2O_2	Hb-PCL/GCE	2 – 30	6.07	1
Amperometry		Hb-AgNPs-PAMAM/GCE	6 – 91	4.9	2
CV		Hb/IL/CILE	100 – 500	40	3

Linear square wave		NF-Hb-Cys- AuNPs/SPCE.	10 – 450	4.4	4
Amperometry		Hb/Ag NPs-BDDE	0.5 – 20	2.5	5
Amperometry		Hb-SWCNTs- CTAB/GCEs.	23.6 – 134	7.87	6
CV + Amperometry		Hb/CMC–TiO ₂ -NTs/GC	4 – 64	4.6	7
Amperometry		GE/Fe ₃ O ₄ /Hb/GCE	0.25 – 1700	06	8
Amperometry		Hb/Aptamer/L- cysteine/AuNPs/SPCE	0.2 – 2100	0.044	This work
Amperometry	Nitrite	Hb/AuNPsPTH/PtNPs/GC	70×10 ⁻³ – 1200	0.02	9
E					
		Hb/HS-CdS/GCE	0.3 – 182	0.08	10
		Hb/Au/GACS/GCE	0.05 -1000	0.01	11
		Nafion/Hb/MX-Ti ₃ C ₂ /GCE	0.5 - 11800	0.12μM	12
CV		AuNPs@MoS ₂ /GCE	10 – 1100	5	13
Amperometry		Hb/Apt/L- cysteine/AuNPs/SPCE	3.6 ×10 ⁻³ – 1300	0.00055	This work

Abbreviations: **SPCE**: screen printed carbon electrode; **GCE**: glassy carbon electrode; **PCL**: poly(ϵ -caprolactone); **AgNPs**: silver nanoparticles; **PAMAM**: poly(amidoamine) dendrimer; **IL**: ionic liquid; **CILE**: carbon ionic liquid electrode; **NF**: nafion; **Cys**: cysteine; **BDDE**: Boron doped diamond electrodes; **SWCNTs**: single walled carbon nanotubes; **CTAB**: cetyltrimethylammonium bromide; **CMC**: Carboxymethyl cellulose; **TiO₂-NTs**: Titanium oxide nanotubes; **GE**: Graphene. **GCE**: Glassy carbon electrode; **PTH**: Polythionine; **PtNPs**: Platinum nanoparticles, **AuNPs**: Gold nanoparticles, **HS-CdS**: CdS hollow nanospheres; **GACS**: Graphene with biocompatible chitosan; **MX-Ti₃C₂**: MXene- Ti₃C₂; **MoS₂**: Molybdenum disulfide; **Hb**: Hemoglobin; **APT**: Aptamer; **CV**: Cyclic voltammetry

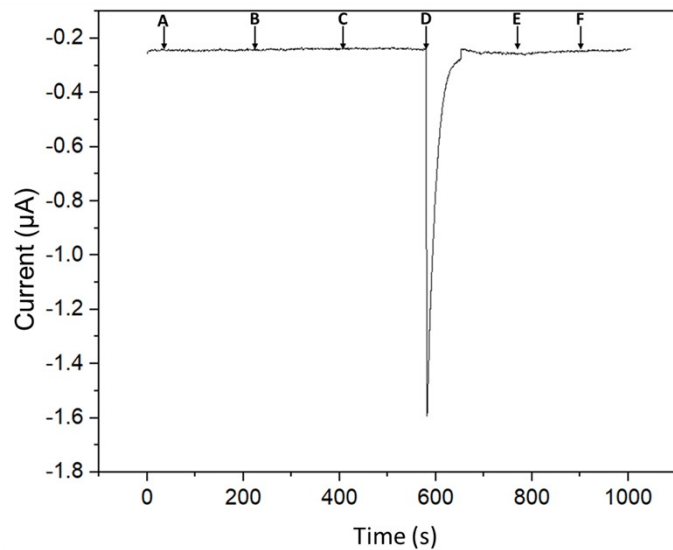


Fig. S5 Amperometric response at constant potential of -0.4 V for Hb/Aptamer/L-cysteine/AuNPs/SPCE in phosphate buffer with successive addition of 35 μM H_2O_2 (D), 100 μM ascorbic acid (A), 500 μM dopamine (B), 100 μM Glucose (C), 100 μM Fructose (E), and 100 μM Arginine (F)

Table S2. Determination and recovery results corresponding to H_2O_2 and nitrite determination in rain water and river water samples, respectively (n = 3).

Sample	Added (μM)	Founded (μM)	RSD (%)	Recovery (%)
	4.5	4.3	0.9	95.55
Rain water	100	103	2.01	103
	800	793	1.56	99.12
	1500	1496	1.3	99.73

	5	5.18	1.5	103.6
River water	80	82.63	3.01	103.28
	500	497.23	1.26	99.44
	1000	1000.96	0.98	100.09

References

1. W. Zheng, J. Li and Y. Zheng, *Biosensors and Bioelectronics*, 2008, **23**, 1562-1566.
2. M. Baccarin, B. C. Janegitz, R. Berté, F. C. Vicentini, C. E. Banks, O. Fatibello-Filho and V. Zucolotto, *Materials Science and Engineering: C*, 2016, **58**, 97-102.
3. A. Safavi, N. Maleki, O. Moradlou and M. Sorouri, *Electrochemistry Communications*, 2008, **10**, 420-423.
4. A. S. Elewi, S. A. W. Al-Shammaree and A. K. M. A. Al Sammarraie, *Sensing and Bio-Sensing Research*, 2020, **28**, 100340.
5. L. Jiang, J. Hu and J. S. Foord, *Electrochimica Acta*, 2015, **176**, 488-496.
6. S. Wang, F. Xie and G. Liu, *Talanta*, 2009, **77**, 1343-1350.
7. W. Zheng, Y. Zheng, K. Jin and N. Wang, *Talanta*, 2008, **74**, 1414-1419.
8. Y. Wang, H. Zhang, D. Yao, J. Pu, Y. Zhang, X. Gao and Y. Sun, *Journal of Solid State Electrochemistry*, 2013, **17**, 881-887.
9. Y. Zhang, R. Yuan, Y. Chai, J. Wang and H. Zhong, *Journal of Chemical Technology, Biotechnology and Applied Biochemistry*, 2012, **87**, 570-574.
10. Z. Dai, H. Bai, M. Hong, Y. Zhu, J. Bao and J. Shen, *Biosensors and Bioelectronics*, 2008, **23**, 1869-1873.
11. J. Jiang, W. Fan and X. Du, *Biosensors and Bioelectronics*, 2014, **51**, 343-348.
12. H. Liu, C. Duan, C. Yang, W. Shen, F. Wang and Z. Zhu, *Sensors and Actuators B: Chemical*, 2015, **218**, 60-66.
13. J. Chao, M. Zou, C. Zhang, H. Sun, D. Pan, H. Pei, S. Su, L. Yuwen, C. Fan and L. Wang, *Nanotechnology*, 2015, **26**, 274005.