

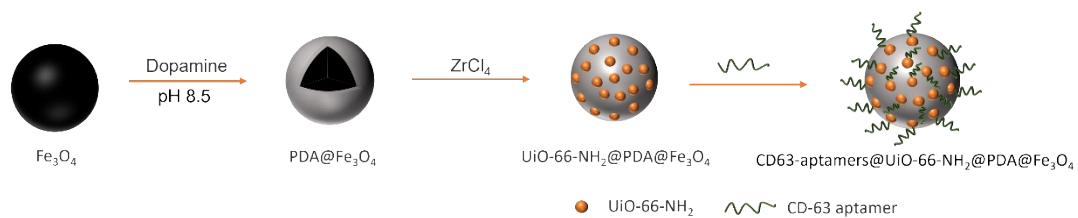
Aptasensors with palladium nanoparticles-modified hemin-containing metal-organic frameworks as signal marker for detection of exosomes

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Scheme S1. Schematic illustration of the synthesis of the CD63-aptamers@UiO-66-NH₂@PDA@Fe₃O₄ capture probe.

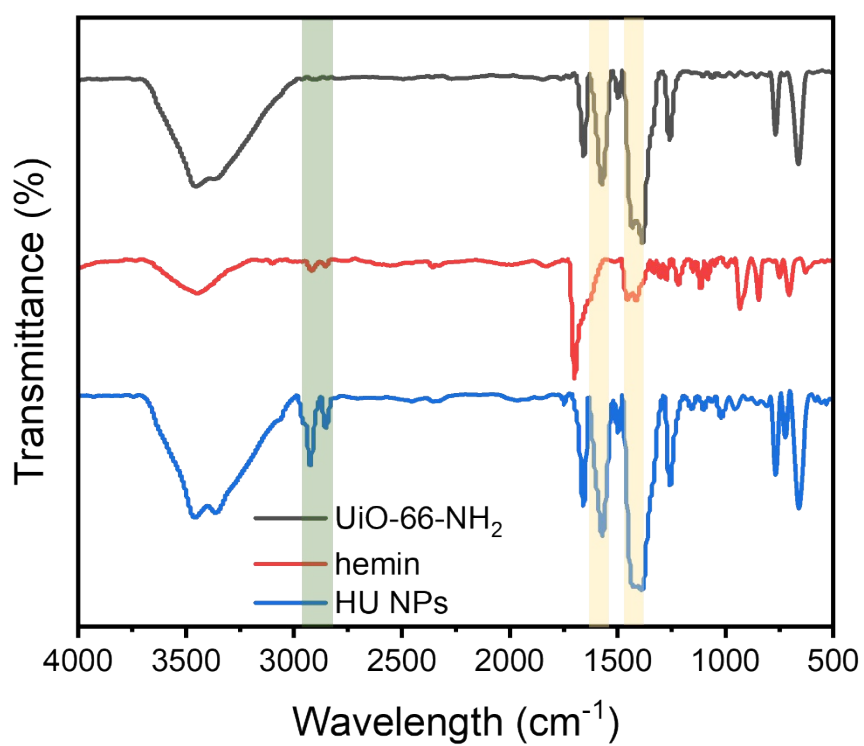


Fig. S1. FT-IR spectra of UiO-66-NH₂ (black line), hemin (red line) and the HU NPs (blue line).

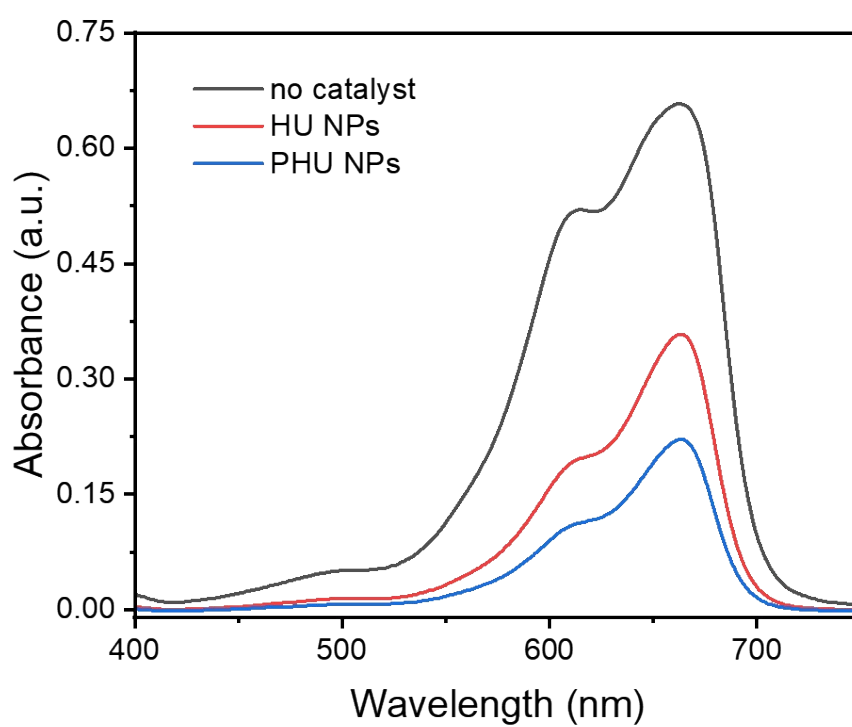


Fig. S2. UV-vis absorption spectra of MB (black line) with its oxidation to indicate $\cdot\text{OH}$ production via H_2O_2 decomposition by the HU NPs (red line) and the PHU NPs (blue line), respectively.

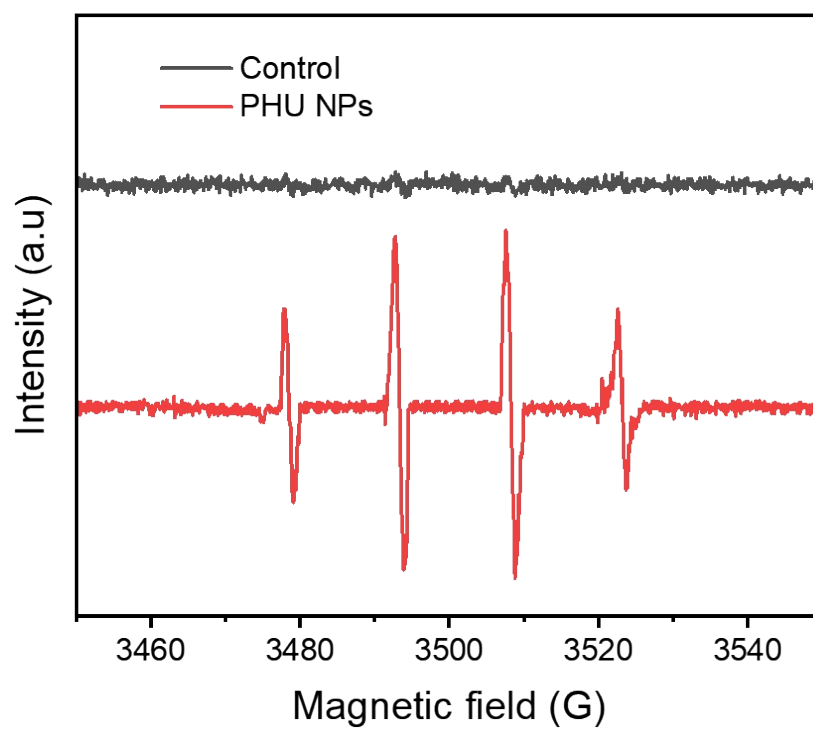


Fig. S3. Typical spin-adduct EPR of $\cdot\text{OH}$ of DMPO in phosphate buffer at pH 7.4.

Table S1 Comparison of the apparent Michaelis-Menten constant (K_m) and maximum reaction rate (V_{max}).

| Catalysts | K_m (mM) | | V_{max} (10^{-8} M/s) | |
|--|------------|-------------------------------|----------------------------|-------------------------------|
| | TMB | H ₂ O ₂ | TMB | H ₂ O ₂ |
| Fe ₃ O ₄ ¹ | 0.098 | 154.0 | 3.440 | 9.780 |
| HRP ¹ | 0.434 | 3.700 | 10.00 | 8.710 |
| Hemin@MIL-101(Al)-NH ₂ ² | 0.068 | 10.90 | 6.070 | 8.980 |
| PHU NPs, this work | 0.11 | 0.09 | 12.05 | 7.65 |

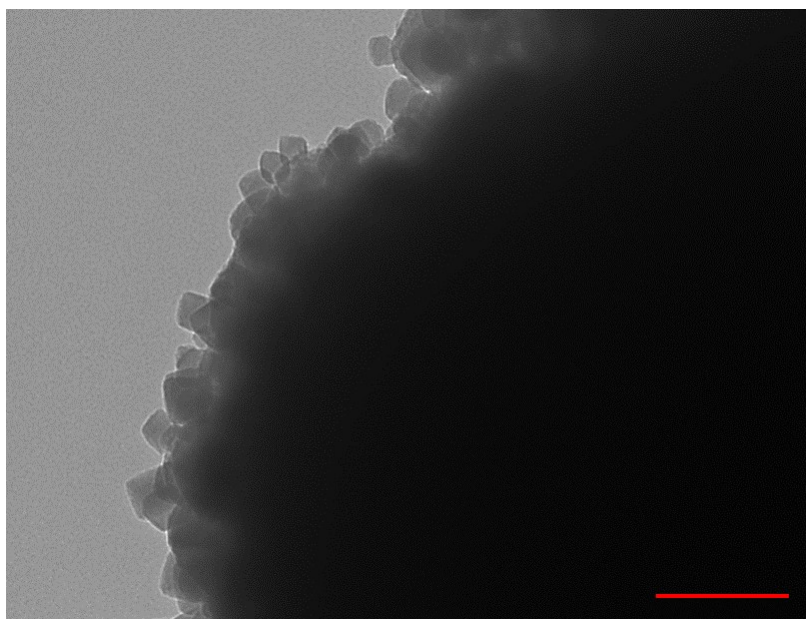


Fig. S4. Typical TEM image of the aptamer@UiO-66-NH₂@PDA@Fe₃O₄ particles. Scale bar: 200 nm.

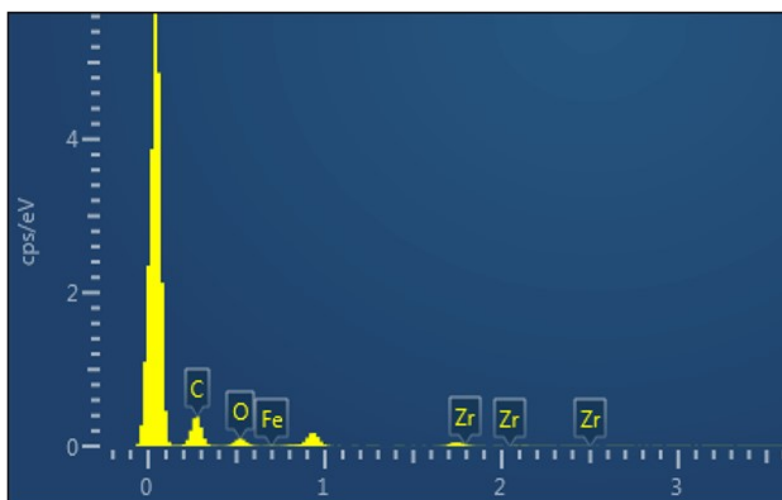


Fig. S5. Energy-dispersive X-ray (EDX) elemental mapping analysis of the aptamer@UiO-66-NH₂@PDA@Fe₃O₄ particles.

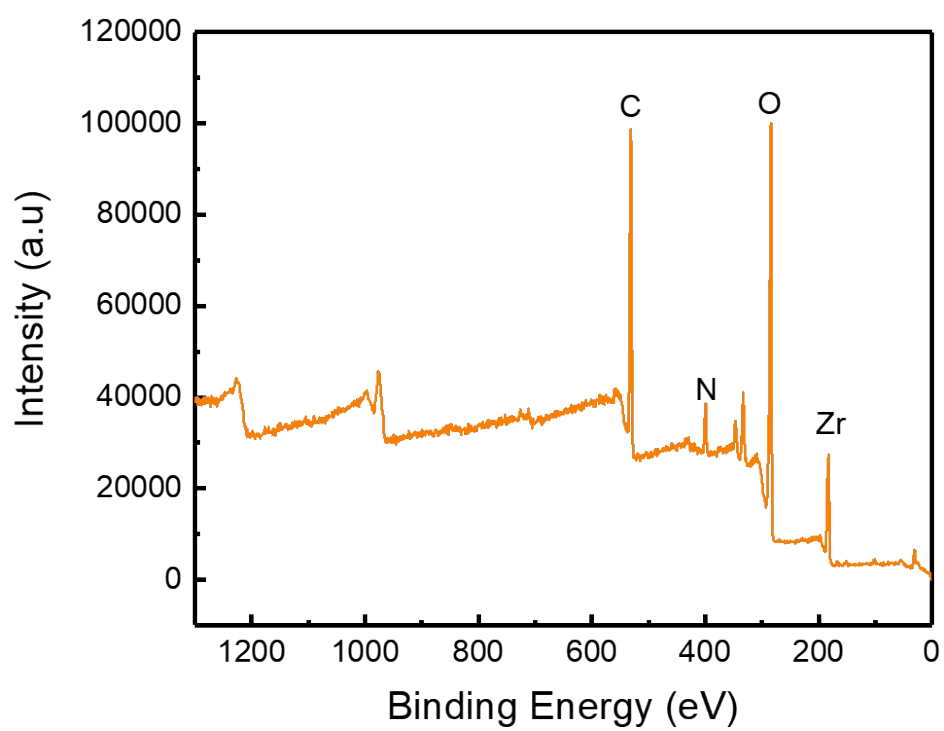


Fig. S6. XPS of the aptamer@UiO-66-NH₂@PDA@Fe₃O₄ particles.

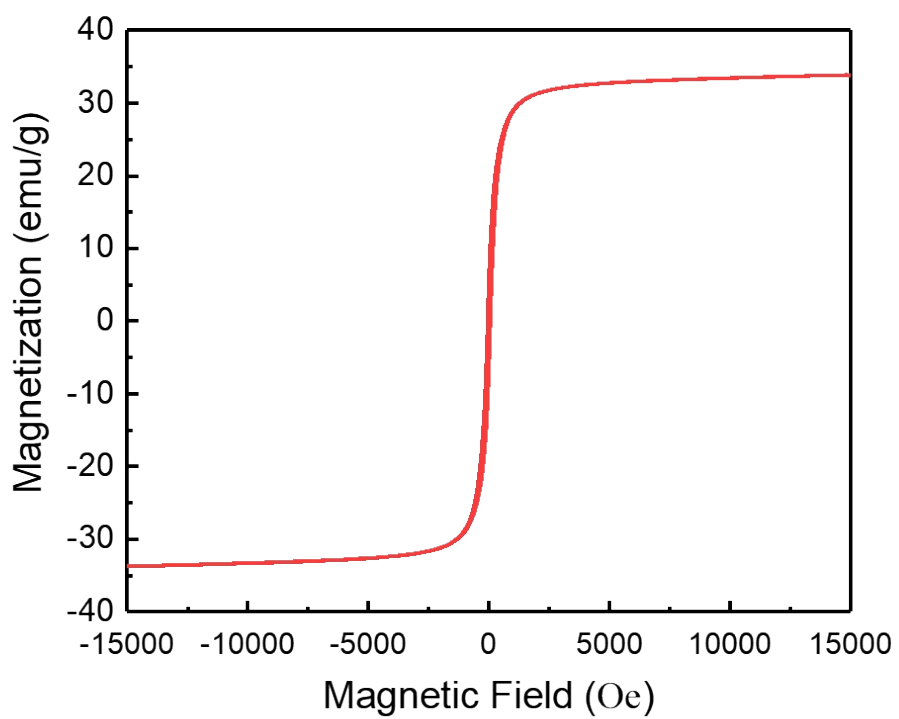


Fig. S7. Magnetic hysteresis curves of the aptamer@UiO-66-NH₂@PDA@Fe₃O₄ particles.

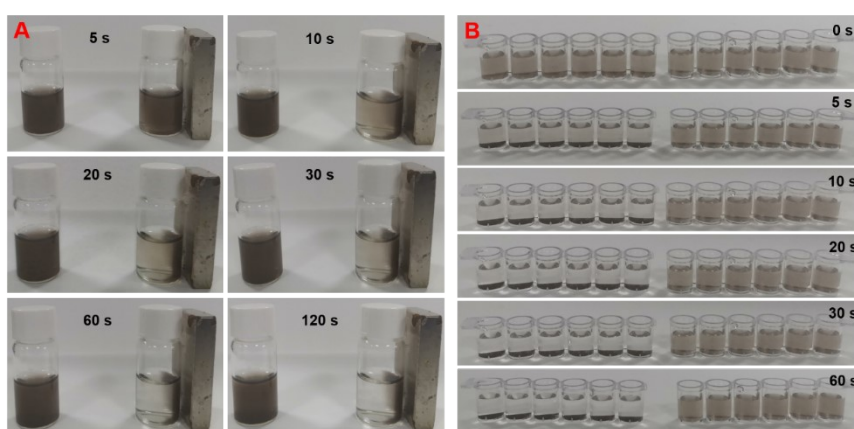


Fig. S8. Separation performance of the aptamer@UiO-66-NH₂@PDA@Fe₃O₄ particles under a magnet in a glass bottle (A) and in 96-well plate (B).

Table S2. Comparison of LOD and detection time of currently available methods for the colorimetric detection of exosomes.

| Analytical method | LOD (particles/ μL) | Linear range (particles/ μL) | References |
|---|---------------------------------|--|------------|
| g-C ₃ N ₄ nanozyme single-walled | 1.352 $\times 10^6$ | 0.19 $\times 10^7$ -3.38 $\times 10^7$ | 3 |
| carbon nanotubes nanozyme | 5.2 $\times 10^5$ | 1.84 $\times 10^6$ -2.21 $\times 10^7$ | 4 |
| TdT-aided signal amplification | 6.7 $\times 10^3$ | 9.75 $\times 10^3$ -1.95 $\times 10^6$ | 5 |
| HRP-accelerated dopamine polymerization | 7.7 | 1-10 ⁸ | 6 |
| CuCo ₂ O ₄ nanozyme | 4.5 $\times 10^3$ | 5.6 $\times 10^4$ - 8.9 $\times 10^5$ | 7 |
| HRP-GNPs | 100 | | 8 |
| Dual functional Zr- MOF nanozyme | 82.6 | 4.28 $\times 10^2$ - 4.28 $\times 10^5$ | This work |

TdT: terminal deoxynucleotidyl transferase. HRP-GNPs: HRP modified gold nanoparticles.

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Table S3. Comparison with commercial ELISA kits

| Products | | | | | LOD (particales/ μ L) | Linear range (particales/ μ L) |
|--|--|--|--|--|-------------------------------|---------------------------------------|
| Human Total Exosome Capture & Quantification Kit | | | | | 1×10^5 | $6.09 \times 10^5 - 1.95 \times 10^6$ |
| This work | | | | | 86.2 | $4.28 \times 10^2 - 4.28 \times 10^5$ |

- not found