

## Electronic supplementary information

### **Rapid detection of *Saccharomyces cerevisiae* with Boronic Acid-Decorated Multivariate Metal-Organic Frameworks and Aptamer**

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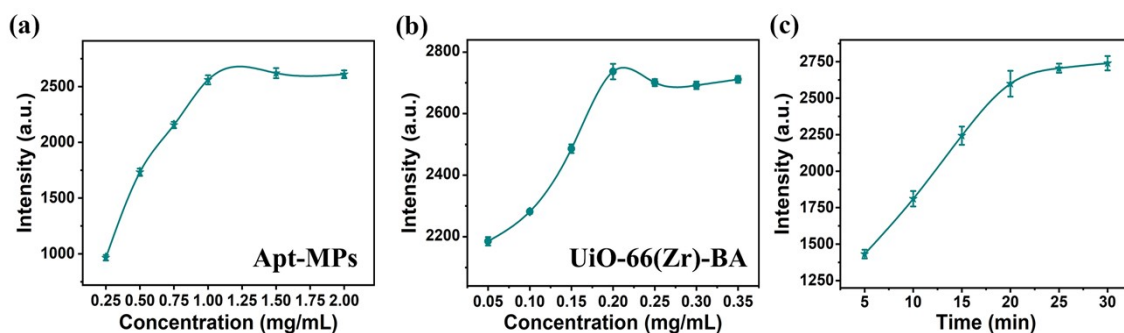
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### **Aptamer screening**

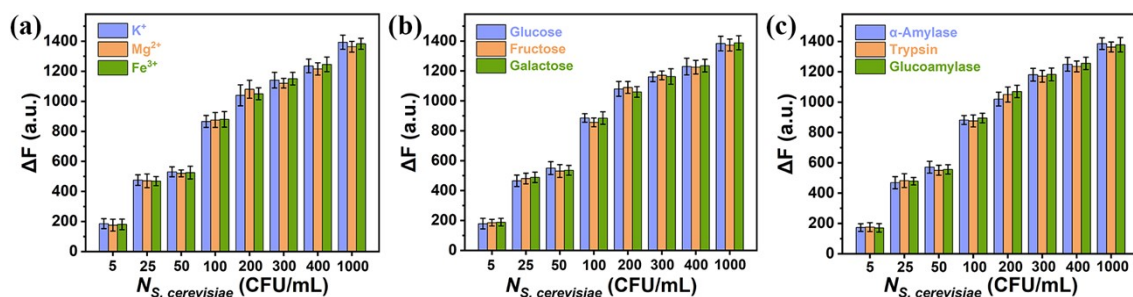
A toggle-cell systematic evolution of ligands by exponential enrichment (SELEX) method was used to select aptamer for *S. cerevisiae*. The specific steps include microbial isolation, DNA extraction and Sanger sequencing, pangenome analysis, sequential toggle-cell SELEX operation, PCR amplification and determination of binding affinity. The dissociation constants ( $K_d$ ) and maximum binding intensity ( $B_{max}$ ) of the aptamer used in this study were  $4.936 \pm 0.121$  nM and  $0.993 \pm 0.048$  nM, respectively, according to fluorometer analysis.

### **Direct detection of *S. cerevisiae* in distilled yeast samples**

Distilled yeast (10 g) was added to a triangular flask containing glass beads and 90 mL of sterile physiological saline and incubated for 30 min (160 rpm/min). The supernatant was gradiently diluted and the amount of *S. cerevisiae* detected using the proposed fluorescence method. For the counting method, the diluted microbial solution was placed on the sterile plate, mixed with Bengal red agar medium (cooled to 50°C), and cultivated at 28°C for 1 day. The counting range was 30-300.



**Fig. S1** Optimization of the assay. (a) Concentration of Apt-MPs; (b) concentration of UiO-66(Zr)-BA; (c) incubation time.



**Fig. S2** Susceptibility fluorescence tests with different concentrations of *S. cerevisiae* and metal ions ( $K^+$ ,  $Mg^{2+}$ , and  $Fe^{3+}$ , 100 mM) (a), small molecules (glucose, fructose, and galactose, 100 mM) (b), and proteins ( $\alpha$ -amylase, trypsin, and glucoamylase, 5 mg/mL) (c).

**Table S1** Comparison of the current approach to reported methods for *S. cerevisiae* analysis.

Detection method	Recognition component	Linear range (CFU/mL)	LOD (CFU/mL)	Test time (h)	Ref.
PCR	Specific primers	$3.8-3.8 \times 10^5$	3.8	> 2	[1]
Optics	Silver sensitized by lectins	$3.2 \times 10^3-7 \times 10^7$	-	> 1.5	[2]
qPCR	Taqman	$3.4 \times 10^2-3.4 \times 10^7$	78	> 4	[3]
Cyclic voltammetry	TTCC/GCE	$2 \times 10^4-10^8$	$10^4$	~ 1	[4]
Impedance spectroscopy	MPA(SAM)/AuE	$10^2-10^8$	$10^2$	~ 1	[5]
Fluorescence	Ionic liquid mediated carbon dots	$5 \times 10^2-1 \times 10^6$	$5 \times 10^2$	~ 1.5	[6]
Fluorescence	Fluorescence	$10-10^6$	3	~ 0.5	This work

**Table S2** The intra-batch and inter-batch precision of the proposed biosensor for *S. cerevisiae* detection.

<i>S. cerevisiae</i> (CFU/mL)	Intra-batch		Inter-batch	
	Mean $\pm$ SD	CV (%; n=6)	Mean $\pm$ SD	CV (%; n=6)
20	$18 \pm 1.0$	5.6	$22 \pm 1.0$	4.5
$10^2$	$9.7 \times 10 \pm 5.9$	6.1	$1.1 \times 10^2 \pm 6.5$	5.9
$10^4$	$1.0 \times 10^4 \pm 3.3 \times 10^2$	3.3	$9.9 \times 10^3 \pm 1.8 \times 10^2$	1.8
$10^6$	$9.8 \times 10^5 \pm 5.0 \times 10^4$	5.1	$9.6 \times 10^5 \pm 7.1 \times 10^4$	7.4

**Table S3** Direct detection of *S. cerevisiae* in distilled yeasts.

	Proposed method		Counting method	
	Mean	RSD	Mean	RSD
	( $\times 10^3$ CFU/mL, n=5)	(%)	( $\times 10^3$ CFU/mL, n=5)	(%)
Distilled yeast 1	2.45	4.2	4.42	12
Distilled yeast 2	1.38	5.5	3.91	8.2
Distilled yeast 3	3.24	5.0	5.37	9.6

## REFERENCES

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