- 1 Supplementary information
- 2 Improving design features and air bubble manipulation techniques for a single-step sandwich
- 3 electrochemical ELISA incorporating commercial electrodes into capillary-flow driven
- 4 immunoassay devices
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1 SI Results



Figure S1. The iceCaDI device design, A) the layout and details, B) the setting of the dashed line for the

4 foldable hinge.

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Figure S2. A) The iceCaDI device after the final assembly and B) the cross-section area at the electrode
 chamber.





Figure S3. The optimization of streptavidin concentration using 25 μg mL⁻¹ of HRP (biotin) and 0.5%
 BSA for blocking.



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Figure S4. The optimization of blocking using BSA with or without 25 µg mL⁻¹ of streptavidin (SV), A)
The optimization of BSA concentration at 30 min of blocking time, B) The optimization of blocking time.
All experiments were obtained using 25 µg mL⁻¹ of HRP (biotin).

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2 **Figure S5.** The optimization of CRP capture antibody (biotin) immobilization. A) The optimization of 3 biotin and streptavidin binding time obtained using 25 μ g mL⁻¹ of HRP (biotin), B) The optimization of 4 CRP capture antibody (biotin) concentration with or without 10 μ g/mL of CRP. All experiments were 5 performed with 25 μ g mL⁻¹ of streptavidin for 1 hr and 0.5% BSA blocking for 30 min.

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Α



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- 9 Figure S6. The flow in electrode chamber comparison, A) without the arrow-shaped obstacle, B) with the
- 10 arrow-shaped obstacle.

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Figure S7. The flow study of the waste pad used, A) the velocity driven by the waste pad (Whatman 3 No.4.) over distance from the inlet, B) the flow rate comparison of the waste pad using Whatman No.1 vs
Whatman No.4 filter paper.

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7 Figure S8. Velocity comparison of paper-based (nitrocellulose strip; CN 95) and laminated devices with

8 3-mm wide and 4-cm long channels. The velocity was measured at a 4-cm distance from the inlet.

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