

Supporting Information

Adhesive SERS substrate based on stretched silver nanowire-tape for in-situ multicomponent analysis of pesticide residues

Xing Dai^a, Danni Xue^a, Xiaohan Liu^a, Chenjie Gu^a, Tao Jiang^{a,*}

^aDepartment of Microelectronic Science and Engineering, School of Physical Science and Technology, Ningbo University, Ningbo 315211, Zhejiang, P. R. China

*Corresponding author: Tao Jiang, E-mail: jiangtao@nbu.edu.cn

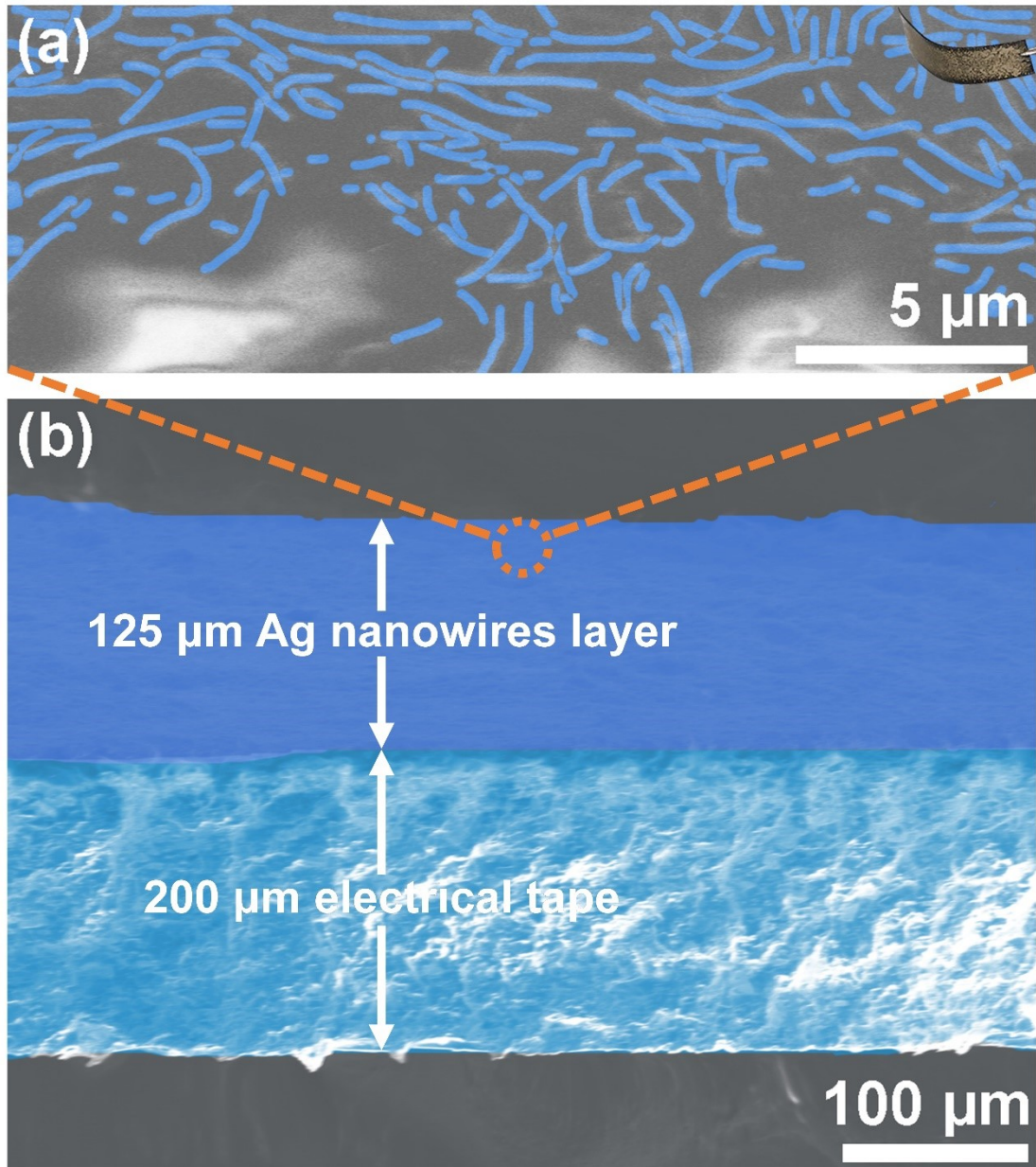


Fig. S1 (a and b) Cross-sectional SEM image of the Ag NW-tape. (b) is the magnified image of the orange region in (a).

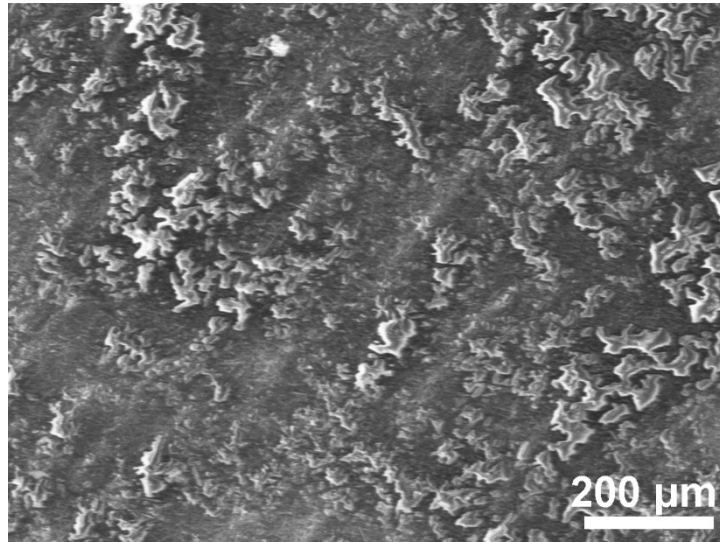


Fig. S2 SEM images of the stretched Ag NW-tape at the strain of 45%.

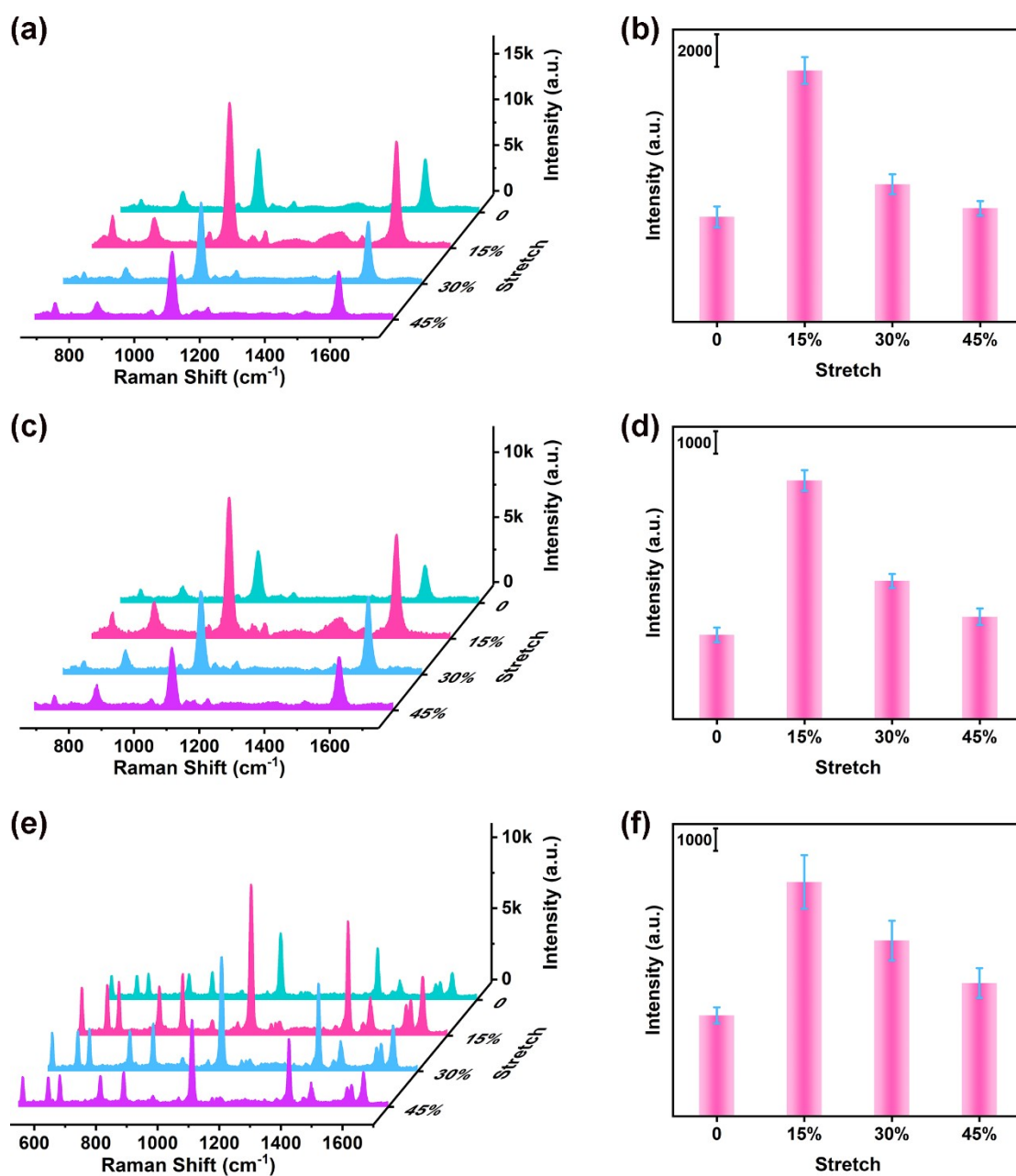


Fig. S3 SERS spectra of Ag NW-tape substrates under varied strains using diverse probe molecules with different concentrations: (a) 10^{-3} M 4-MBA, (c) 10^{-5} M 4-MBA, (e) 10^{-3} M 2-NAT. (b, d and f) The corresponding histogram of characteristic Raman peak intensity of each target analyte.

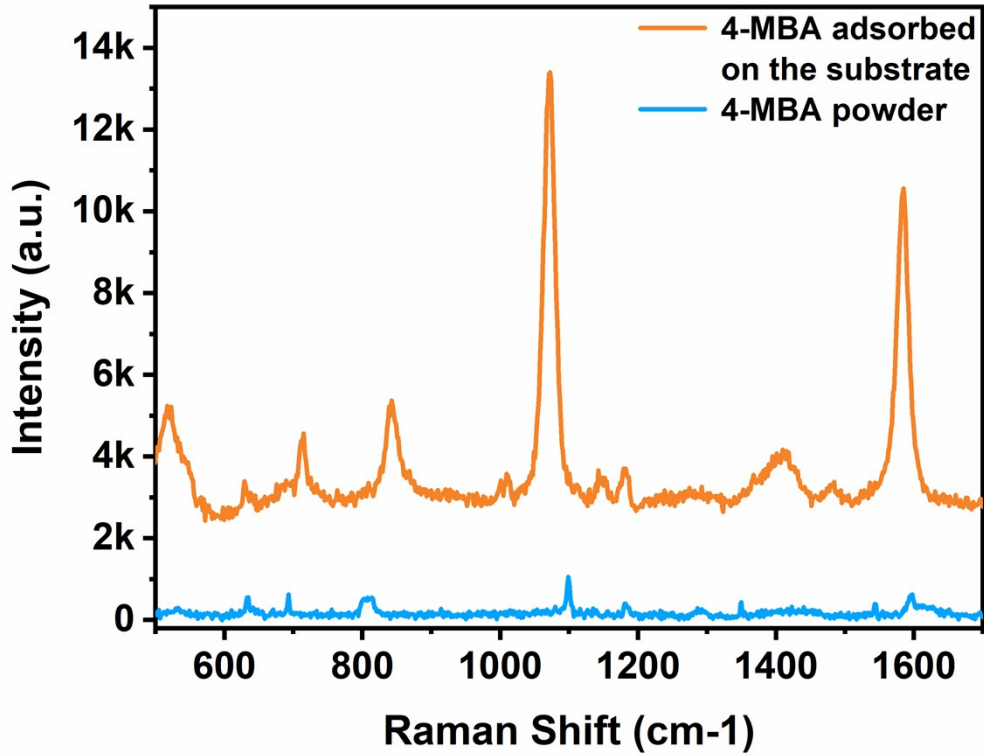


Fig. S4 SERS spectrum of 4-MBA on the SERS-active substrate and Raman spectrum of 4-MBA powder.

The EF was calculated by the representative equation: $EF = (I_{SERS}/I_{bulk}) / (N_{SERS}/N_{bulk})$.

Here, I_{SERS} and I_{bulk} are the integrated intensities of the peak at 1072 cm^{-1} for 4-MBA adsorbed on the substrate and 4-MBA powder, respectively. The required I_{SERS} and I_{bulk} were calculated to be 262198.28 and 24058.81 according to Figure S4, respectively.

Thereafter, N_{SERS} and N_{bulk} indicate the effective amount of 4-MBA molecules from the substrate and 4-MBA powder, respectively. The corresponding values were acquired

by the following equations:

$$N_{SERS} = N_A \times S_{illumination} \times \sigma$$

$$N_{bulk} = n_{powder} \times V_{illumination}$$

$$V_{illumination} = \frac{1}{3} \times S_{illumination} \times D_{depth}$$

$$D_{depth} = \frac{\lambda}{(NA)^2}$$

$$\frac{N_{SERS}}{N_{bulk}} = \frac{N_A \times \sigma}{\frac{1}{3} \times n_{powder} \times D_{depth}}$$

The entire molecule number of 100 mg 4-MBA (N_{powder}) was calculated to be 3.90×10^{20} . When 100 mg of solid 4-MBA powder was tightly compacted into a cube shape by two clean glasses, its volume (V_{powder}) was estimated as $2.5 (5 \times 5 \times 0.1) \text{ mm}^3$. Consequently, the volume density of 4-MBA molecules in the powder (n_{powder}) was estimated to be $1.56 \times 10^{20} \text{ mm}^{-3}$ ($n_{powder} = N_{powder}/V_{powder}$). In addition, the penetration depth of the focused beam into the 4-MBA powder (D_{depth}) was estimated to be $4.91 \mu\text{m}$ ($NA = 0.4$ for the $20 \times$ objective lens of the Raman spectrometer at the 785 nm wavelength). Subsequently, σ for the mole amount of 4-MBA molecules per mm^2 in a monolayer was calculated as $4.0 \times 10^{-12} \text{ mol} \cdot \text{mm}^{-2}$. Accordingly, the value of N_{SERS}/N_{bulk} was estimated to be 9.43×10^{-6} . Hence, the EF value of the Ag NW-tape substrate was calculated as 1.16×10^6 .

Table S1 Comparison of the performance in the detection of TMTD

Substrate	Detection range	LOD	Ref.
Ag NRs embedded PDMS	$10^{-2} \sim 10^{-7}$ M	2.4×10^{-9} g/cm ²	1
interfacial self-assembly Au NRs array	0.001 ~ 30 ppm	0.029 ng/cm ²	2
2D Au@Ag nanodot array	0.005 ~ 1 ppm	0.0011 ppm	3
interfacial self-ordered GNP arrays	19 ~ 1900 ng/cm ²	1 nM	4
CNF-Au NR based SERS platform	6 ~ 600 ng/cm ²	6 ng/cm ²	5
CNF/Au NP substrate	0 ~ 10 ppm	52 ppb	6
stretched Ag NW-tape substrate	$10^{-1} \sim 10^{-6}$ mg/mL	0.0102 ng/cm ²	This work

Table S2 Comparison of the performance in the Detection of TBZ

Substrate	Detection range	LOD	Ref.
interfacial self-assembly Au NRs array	0.01 ~ 200 ppm	0.76 ng/cm ²	2
2D Au@Ag nanodot array	0.05 ~ 10 ppm	0.051 ppm	3
Ag/NC jelly like substrate	5 ~ 10 ng/cm ²	5 ng/cm ²	7
Au@Ag/PMMA/qPCR-PET film	0.05 ~ 10 ppm	20 ppb	8

Biomimetic assembles GNPs	material	$10^{-5} \sim 10^{-9}$ M	10^{-9} M	9
Vertically aligned arrays	Au NRs	0 ~ 1000 $\mu\text{g/L}$	149 $\mu\text{g/L}$	10
stretched substrate	Ag NW-tape	1 ~ 10^{-5} mg/mL	0.0819 ng/cm^2	This work

The detail conversion process of the concentration was shown as below:

In this work, 10 μL standard solution of pesticide (1 mg/mL) was dropped on the Ag NW-tape substrate with an area of $1.5 \times 0.5 \text{ cm}^2$.

The mass of pesticide on the substrate was:

$$m = C \times V = 1 \text{ mg/mL} \times 10 \mu\text{L} = 10 \times 10^3 \text{ ng};$$

The mass-to-area ratio (R_m/a) of the pesticide on the Ag NW-tape substrate was:

$$R_m/a = \text{mass/area} = 10 \times 10^3 \text{ ng} / (1.5 \times 0.5) \text{ cm}^2 = 13.3 \times 10^3 \text{ ng/cm}^2;$$

Therefore, the limits of detection (LOD) of TMTD ($7.67 \times 10^{-7} \text{ mg/mL}$) and TBZ ($6.16 \times 10^{-6} \text{ mg/mL}$) could be evaluated to be:

$$7.67 \times 10^{-7} \times 13.3 \times 10^3 \text{ ng/cm}^2 = 1.02 \times 10^{-2} \text{ ng/cm}^2$$

$$6.16 \times 10^{-6} \times 13.3 \times 10^3 \text{ ng/cm}^2 = 8.19 \times 10^{-2} \text{ ng/cm}^2$$

For the in-situ detection of pesticide residues on food, 20 μL mixtures of pesticide (1 mg/mL) was spiked on the food surface. The contaminated area was about $1 \times 1 \text{ cm}^2$.

Therefore, the mass of pesticide on the food surface was:

$$m = C \times V = 1 \text{ mg/mL} \times 20 \mu\text{L} = 20 \times 10^3 \text{ ng};$$

The mass-to-area ratio (R_m/a) of the pesticide on the fruit surface was:

$$R_m/a = \text{mass/area} = 20 \times 10^3 \text{ ng} / 1 \text{ cm}^2 = 20 \times 10^3 \text{ ng/cm}^2;$$

After being dried, the pesticide was extracted through the “paste and peel off” method before SERS detection. Therefore, the detected amounts of TMTD (2×10^{-3} and $2 \times 10^{-5} \text{ mg/mL}$) and TBZ (5×10^{-1} and $5 \times 10^{-5} \text{ mg/mL}$) on the surface of the vegetables and fruits could be estimated to be:

$$2 \times 10^{-3} \times 20 \times 10^3 \text{ ng/cm}^2 = 40 \text{ ng/cm}^2$$

$$2 \times 10^{-5} \times 20 \times 10^3 \text{ ng/cm}^2 = 0.4 \text{ ng/cm}^2$$

$$5 \times 10^{-1} \times 20 \times 10^3 \text{ ng/cm}^2 = 10^4 \text{ ng/cm}^2$$

$$5 \times 10^{-5} \times 20 \times 10^3 \text{ ng/cm}^2 = 1 \text{ ng/cm}^2$$

References

- 1 S. Kumar, P. Goel and J. P. Singh, Flexible and Robust SERS Active Substrates for Conformal Rapid Detection of Pesticide Residues from Fruits, *Sensor. Actuat. B-Chem.*, 2017, **241**, 577-583.
- 2 B. Hu, D.-W. Sun, H. Pu and Q. Wei, Rapid Nondestructive Detection of Mixed Pesticides Residues on Fruit Surface Using SERS Combined with Self-Modeling Mixture Analysis Method, *Talanta*, 2020, **217**, 120998.
- 3 K. Wang, D.-W. Sun, H. Pu and Q. Wei, Two-Dimensional Au@Ag Nanodot Array for Sensing Dual-Fungicides in Fruit Juices with Surface-Enhanced Raman Spectroscopy Technique, *Food Chem.*, 2019, **310**, 125923.
- 4 F. Yu, M. Su, L. Tian, H. Wang and H. Liu, Organic Solvent as Internal Standards for Quantitative and High-Throughput Liquid Interfacial SERS Analysis in Complex Media, *Anal. Chem.*, 2018, **90**, 5232-5238.
- 5 G. Kwon, J. Kim, D. Kim, Y. Ko, Y. Yamauchi and J. You, Nanoporous Cellulose Paper-Based SERS Platform for Multiplex Detection of Hazardous Pesticide, *Cellulose*, 2019, **26**, 4935-4944.
- 6 Z. Xiong, M. Lin, H. Lin and M. Huang, Facile Synthesis of Cellulose Nanofiber Nanocomposite as a SERS Substrate for Detection of Thiram in Juice, *Carbohydr. Polym.*, 2018, **189**, 79-86.
- 7 J. Chen, M. Huang, L. Kong and M. Lin, Jellylike Flexible Nanocellulose SERS Substrate for Rapid In-Situ Non-Invasive Pesticide Detection in Fruits/Vegetables, *Carbohydr. Polym.*, 2019, **205**, 596-600.

- 8 K. Wang, D.-W. Sun, H. Pu, Q. Wei and L. Huang, Stable, Flexible, and High-Performance SERS Chip Enabled by a Ternary Film-Packaged Plasmonic Nanoparticle Array, *ACS Appl. Mater. Inter.*, 2019, **11**, 29177-29186.
- 9 V. Sharma and V. Krishnan, Fabrication of Highly Sensitive Biomimetic SERS Substrates for Detection of Herbicides in Trace Concentration, *Sensor. Actuat. B-Chem.*, 2018, **262**, 710-719.
- 10 F. K. Alsammarraie, M. Lin, A. Mustapha, H. Lin, X. Chen, Y. Chen, H. Wang and M. Huang, Rapid Determination of Thiabendazole in Juice by SERS Coupled with Novel Gold Nanosubstrates, *Food Chem.*, 2018, **259**, 219-225.