

Electronic supplementary information for

**A broad-applicable method for mercury speciation in various seafood using  
microwave-assisted extraction and ion chromatography-inductively coupled  
plasma mass spectrometry**

Ruiling Li<sup>1</sup>, Yuquan Pan<sup>1</sup>, Chaochen Sun<sup>1</sup>, Chen Lin<sup>1</sup>, Shilong Chen<sup>1</sup>, Yongning Wu<sup>1</sup>,  
FengFu Fu<sup>1\*</sup>

<sup>1</sup>Key Laboratory for Analytical Science of Food Safety and Biology of MOE, Fujian  
Provincial Key Lab of Analysis and Detection for Food Safety, College of Chemistry,  
Fuzhou University, Fuzhou, Fujian 350116, China.

<sup>2</sup>NHC Key Lab of Food Safety Risk Assessment, Food Safety Research Unit  
(2019RU014) of China Academy of Medical Science, China National Center for Food  
Safety Risk Assessment, Beijing 100021, China

---

\* Corresponding author. Tel./Fax: +86-591-22866135; E-mail address: [fengfu@fzu.edu.cn](mailto:fengfu@fzu.edu.cn) (F.-F. Fu)

## 1. Chemicals, materials and apparatus used in the experiment

The standard solutions of  $\text{Hg}^{2+}$  (mercuric chloride, 1000  $\mu\text{g}/\text{mL}$ ),  $\text{CH}_3\text{Hg}^+$  (methylmercury chloride, 100  $\mu\text{g}/\text{mL}$ ) and  $\text{C}_2\text{H}_5\text{Hg}^+$  (ethylmercury chloride, 100  $\mu\text{g}/\text{mL}$ ) were all purchased from National Institute of Metrology, China (Beijing, China). The standards of each mercury species with different concentrations used in the experiments were prepared by directly diluted above standards with pure water. Ultra-pure  $\text{HNO}_3$  and analytical grade of L-Cysteine (Cys) were obtained from Sangon Biotech Co. Ltd (Shanghai, China). The water used in the experiment is Milli-Q water (18.2  $\text{M}\Omega\cdot\text{cm}$ ), which was prepared with Millipore pure system. Strong cation exchange guard column (Zorbax 300SCX, 4.6 $\times$ 12.5 mm) was purchased from Agilent Technology (Shanghai, China).

The IC-ICP-MS system, which couples an Agilent 7800 inductively coupled plasma mass spectrometry (ICP-MS, Agilent of USA) with a Dionex ICS-900 ion chromatography (ThermoFisher Scientific, China), was used in this study.

**Table S1:** Optimal running parameters of IC-ICP-MS system

Parameter	Value
<b>IC system</b>	
IC separation column	Two consecutive strong cation guard column (Zorbax 300SCX, 4.6×12.5 mm)
IC eluent	5.0 mmol/L HNO <sub>3</sub> -1.0 mmol/L Cys
IC elution mode	iso-elution
Sample injection volume	10 µL
Flow rate of IC eluent	1.0 mL/min
Temperature of IC separation column	30 °C
<b>ICP-MS system</b>	
ICP-MS RF power	1350 W
ICP-MS cool gas flow rate	15 L/min
ICP-MS auxiliary gas flow rate	0.90 L/min
ICP-MS nebulizer gas flow rate	0.80 L/min
ICP-MS makeup gas flow rate	0.20 L/min
Monitored isotope (m/z)	<sup>202</sup> Hg

**Table S2:** The effect of extracting solvent volume and extraction mode on mercury extraction efficiency of shellfish (*Morula uva*) sample

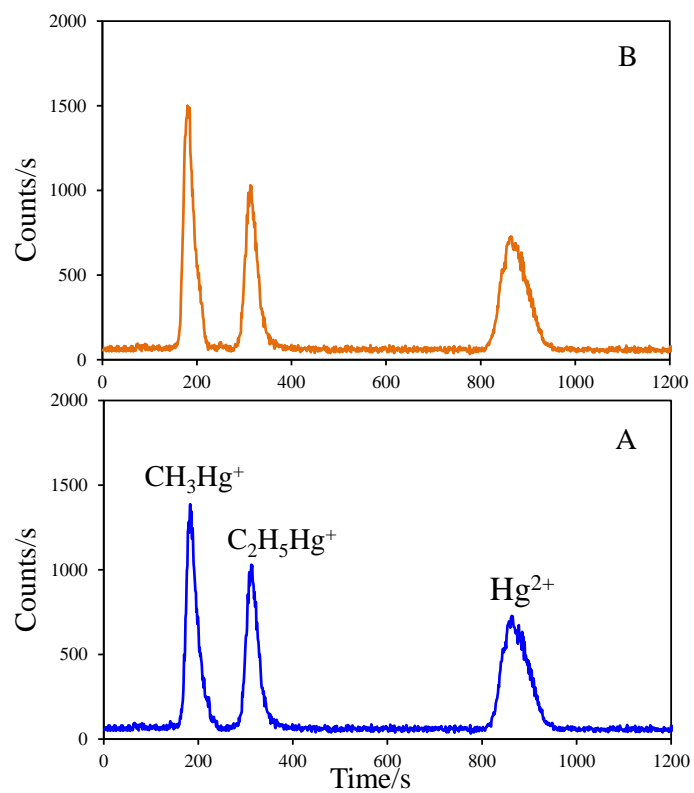
Sample	0.10 g shellfish ( <i>Morula uva</i> )	
Extracting solvent volume	Extraction mode	Extraction efficiency of Hg
8 mL	4.0 mL for previous soaking and first extraction, 4.0 mL for second extraction	72.4%
10 mL	5.0 mL for previous soaking and first extraction, 5.0 mL for second extraction	83.6%
12 mL	6.0 mL for previous soaking and first extraction, 6.0 mL for second extraction	89.8%
	7.0 mL for previous soaking and first extraction, 5.0 mL for second extraction	91.2%
14 mL	7.0 mL for previous soaking and first extraction, 7.0 mL for second extraction	90.8%
	8.0 mL for previous soaking and first extraction, 6.0 mL for second extraction	91.5%

**Table S3:** The comparison on analytical performance among previous ICP-MS-based methods for mercury speciation in seafood and our method

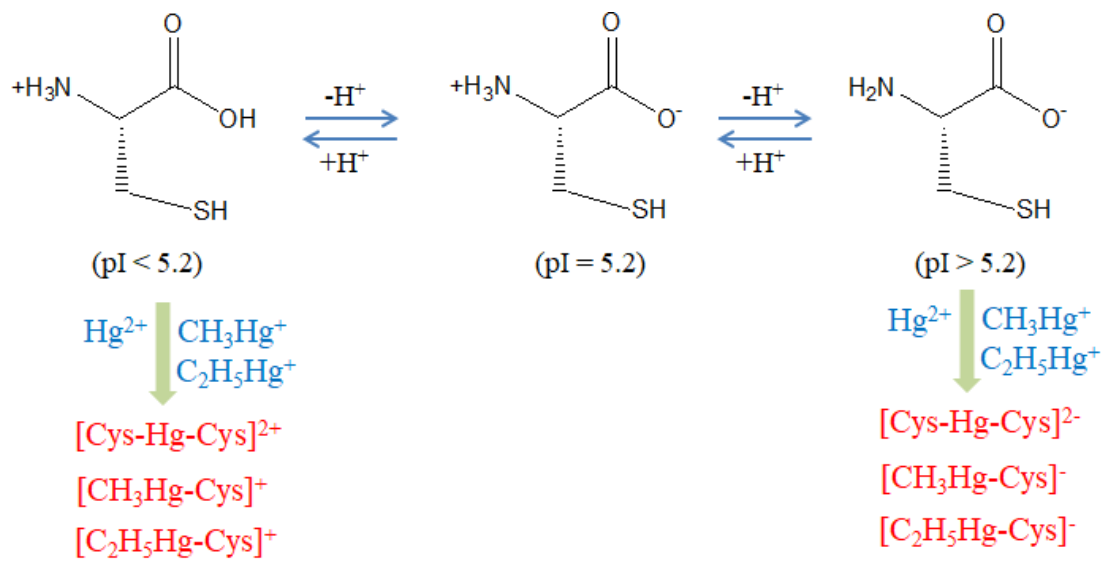
Method	Separation column	Analyte	Extraction method	Applicable sample	Instrument LOD	Method LOD in seafood	Ref.
HPLC-ICP-MS	C8 column	Hg <sup>2+</sup> and CH <sub>3</sub> Hg <sup>+</sup>	Enzymatic extraction and microwave-assisted extraction	Fish	0.4 ng/mL	10 ng/g dried weight	1
Ion chromatography-ICP-MS	Anion guard column	Hg <sup>2+</sup> , CH <sub>3</sub> Hg <sup>+</sup> and C <sub>2</sub> H <sub>5</sub> Hg <sup>+</sup>	Ultrasonic extraction	Fish	0.008-0.029 ng/mL	Not provided	2
Ion-pairing reversed-phase HPLC-ICP-MS	C18 guard columns	Hg <sup>2+</sup> , CH <sub>3</sub> Hg <sup>+</sup> and C <sub>2</sub> H <sub>5</sub> Hg <sup>+</sup>	Ultrasonic extraction	Fish	0.014-0.028 ng/mL	0.18-0.33 ng/g fresh weight	3
HPLC-ICP-MS	C18 column	Hg <sup>2+</sup> , CH <sub>3</sub> Hg <sup>+</sup> and C <sub>2</sub> H <sub>5</sub> Hg <sup>+</sup>	Ultrasonic extraction together with magnetic solid phase pre-concentration	Water and fish	0.49-0.76 ng/L	Not provided	4
Ion chromatography-ICP-MS	Cation guard columns	Hg <sup>2+</sup> , CH <sub>3</sub> Hg <sup>+</sup> and C <sub>2</sub> H <sub>5</sub> Hg <sup>+</sup>	Ultrasonic extraction	Water and fish	Not provided	1.9-3.1 ng/g fresh weight	5
HPLC-ICP-MS	ZORBAX SB-Aq C18 column	Hg <sup>2+</sup> and CH <sub>3</sub> Hg <sup>+</sup>	Microwave-assisted extraction with protease	Fish	0.013–0.015 ng/mL	Not provided	6
HPLC-isotope dilution ICP-MS	Symmetry Shield RP C18 column	Hg <sup>2+</sup> and CH <sub>3</sub> Hg <sup>+</sup>	Microwave-assisted extraction	Marine biota	Not provided	Not provided	7
IC-ICP-MS	Strong cation guard column	Hg <sup>2+</sup> , CH <sub>3</sub> Hg <sup>+</sup> and C <sub>2</sub> H <sub>5</sub> Hg <sup>+</sup>	Microwave-assisted extraction without protease	Various seafood (seaweeds, fishes and shellfishes)	0.02-0.05 ng/mL	2.4-6.0 ng/g dried weight (about 0.48 – 1.2 ng/g fresh weight)	This study

**Reference:**

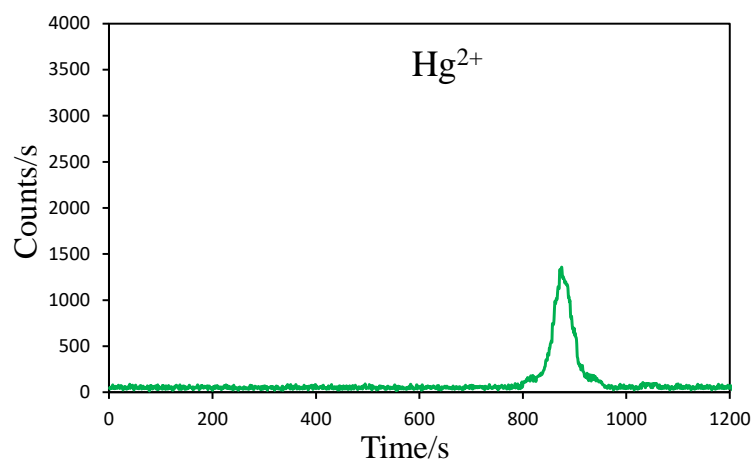
- (1) R. Jagtap, F. Krikowa, W. Maher, S. Foster, M. Ellwood, *Talanta*, 2011, 85, 49-55
- (2) X. P. Chen, C. Han, H. Y. Cheng, J. H. Liu, Z. G. Xu, X. F. Yin, *Anal. Chim. Acta*, 2013, 796, 7-13.
- (3) H. Y. Cheng, X. P. Chen, L. H. Shen, Y. C. Wang, Z. G. Xu, J. H. Liu, *J. Chromatogr. A*, 2018, 1531, 104-111.
- (4) S. Q. Zhu, B. B. Chen, M. He, T. Huang, B. Hu, *Talanta*, 2017, 171, 213-219.
- (5) X. P. Chen, C. Han, H. Y. Cheng, Y. C. Wang, J. H. Liu, Z. G. Xu, L. Hu, *J. Chromatogr. A*, 2013, 1314, 86-93.
- (6) Y.-C. Chen, S.-J. Jiang, *J. Anal. At. Spectrom.*, 2021, 36, 938-945.
- (7) A. A. Krata, E. Vassileva, *Talanta*, 2020, 217, 121113.



**Figure S1:** The IC-ICP-MS chromatograms of the mixed standard of mercury species. The concentrations of each mercury species are all 10.0 ng/mL. (A): Original mixed standard; (B) The mixed standard pretreated with pre-soaking microwave-assisted extraction.

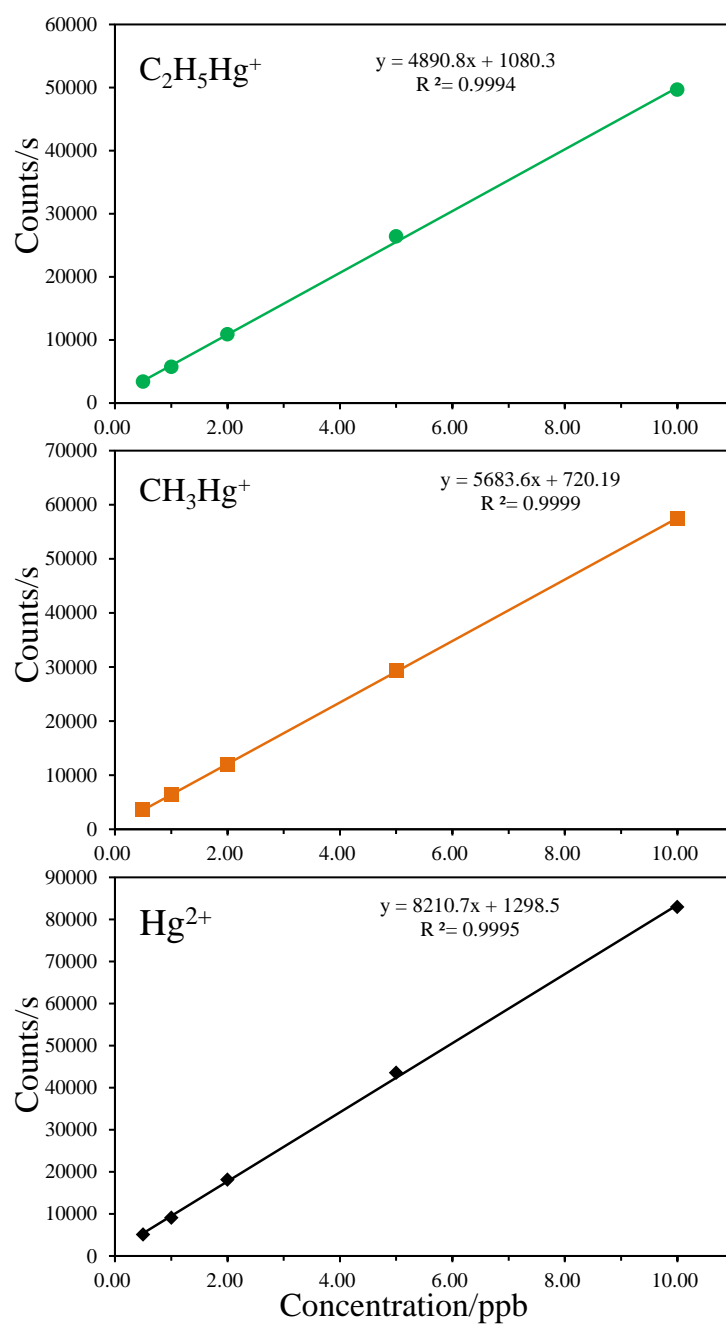


**Figure S2:** The structure and protonation state of Cys and the charge state of the Cys-mercury species complexes.



**Figure S3:** The IC-ICP-MS chromatogram for determining Hg<sup>2+</sup> after Hg<sup>2+</sup> was pre-chelated with 6.0 mM of L-Cys. The concentrations of Hg<sup>2+</sup> is 10 ng/mL, and the IC-ICP-MS chromatogram was obtained under Table S1 conditions.





**Figure S4:** The linear relationship between IC-ICP-MS signal (peak area) and the concentrations of mercury species (calibration curves) for determining Hg<sup>2+</sup>, CH<sub>3</sub>Hg<sup>+</sup> and C<sub>2</sub>H<sub>5</sub>Hg<sup>+</sup> with IC-ICP-MS under Table S1 conditions.