1	Supporting Information
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3	Magnetic molecularly imprinted polymers as a fluorescent sensor for
4	selective detection of Sudan I in chili powder
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23	Experimental
24	Materials

Ethylene glycol, sodium hydroxide (NaOH), acetic acid, glucose, sucrose, 25 lactose, urea, citric acid, sodium citrate, sodium citrate, sodium acetate, ferrous 26 chloride (FeCl₂·4H₂O) and other metallic salt ions were obtained from Beijing 27 Chemical Works (Beijing, China). Ferric trichloride (FeCl₃·6H₂O), ammonia, ethyl 28 alcohol, Sudan I, Sudan II, Sudan B, Sudan G, azophloxine, orange II, L-tryptophan, 29 L-cysteine, isoleucine (Ile), methionine (Met), threonine (Thr), phenylalanine (Phe), 30 valine (Val), and histidine (His) were obtained from Sinopharm Group Co. Ltd. 31 (Shanghai, China). Mmethyl alcohol, ascorbic acid, sodium chloride (NaCl), 32 potassium chloride (KCl), and tetraethoxysilane (TEOS, 40%) were purchased from 33 Tianjin Guangfu Institute of Fine Chemicals (Tianjin, China). 3-aminopropyl 34 triethoxysilane (APTES, 98%) was purchased from Shanghai McLean Biochemical 35 Technology Co. Ltd (Shanghai, China). All the other reagents in the present work 36 were analytical reagent without any purification. Deionized water used throughout the 37 experiments was acquired by the laboratory purification system. 38

39 Instruments

40 The morphology and microstructure of CDs@SiO₂ were characterized by 41 JEM-2100 Field-emission transmission electron microscope (TEM, JEOL, Japan). 42 The morphologies of MMIPs and MNIPs were characterized by SU8020 Field-43 emission scanning electron microscope (SEM, Hitachi, Japan). Elemental analysis 44 was investigated by energy dispersive X-ray spectroscopy (EDS, Thermo, USA). X-45 ray diffraction (XRD) measurements were performed using an Empyrean X-ray 46 diffractometer equipped with a Cu Kα radiation source (PANalytical B.V.,

Netherlands). Fourier-transform infrared spectroscopies (FT-IR) were recorded on a 47 Nicolet-iS5 FT-IR spectrophotometer (Thermo, USA) with KBr pellets. The water 48 contact angle was measured using a droplet angle measuring instrument 49 (Changchun, Jilin). Thermal gravimetric analysis (TGA) was carried out with a TGA 50 Q500 thermal analyzer instrument (TA, USA). X-ray photoelectron spectroscopy 51 (XPS) was performed on an ESCALAB 250 electron spectrometer (Thermo Electron, 52 USA). The magnetic hysteresis loops of samples were measured using a SQUID-VSM 53 Magnetic Property Measurement System (Quantum Design, USA). Ultraviolet-visible 54 (UV-Vis) absorption spectroscopies were collected on a Shimadzu UV-2550 55 spectrophotometer (Shimadzu, Japan). Fluorescence spectra were performed on an 56 F-2700 fluorescence spectrophotometer (Hitachi, Japan). Nanosecond fluorescence 57 lifetime was investigated by FLS 920 fluorescence spectrophotometer (Edinburgh, 58 UK). 59

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61 Results and discussion

62 Characterization of MMIPs

The elemental analysis of MMIPs was investigated through EDS mapping (Fig.
S1). The results revealed the presence of C, N, O, Si and Fe elements in the MMIPs
composites.



Fig. S1 Elemental mapping images of Combine, C, N, O, Si and Fe of MMIPs (A–F,
respectively)

The hydrophilic properties of MMIPs and MNIPs were investigated by water contact angle test, and the corresponding results were shown in **Fig. S2**. The water contact angles of MMIPs and MNIPs are 12.53° and 14.50° respectively, indicating that the prepared MMIPs exhibited good hydrophilicity.



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74 Fig. S2 Water contact angle test of MMIPs (A) and MNIPs (B)

76 Calculation of fluorescence lifetime

When a substance molecule is excited by the exciting light, it absorbs energy and transitions from the ground state to the excited state, and then emits fluorescence in the form of a radiation transition and returns to the ground state. The time required for the fluorescence intensity to decay to 1/e of the initial intensity is the fluorescence lifetime of the fluorescent substance under the measurement conditions.¹ **Fig. S3** showed the fluorescence decay curves of MMIPs in the presence and absence of

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Sudan I, and the fluorescence lifetime of MMIPs gradually decreased from 6.69 ns to
5.69 ns with the increase of Sudan I concentration. Combined with previously
reported literatures,¹⁻⁴ it was further confirmed that the fluorescence lifetime of donor
NS-CDs decreased after the addition of acceptor Sudan I.



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88 Fig. S3 Fluorescence decay curves of MMIPs in the absence and presence of Sudan I89

90 Different fluorescence methods for detection of Sudan I

91 In Table S1, the fluorescence detection methods of Sudan I was summarized,

92 including the linear range and limit of detection (LOD) of the methods.

93 Table S1 Comparison of different fluorescence detection methods of Sudan I with

94 this work

Method	Linear range (µM)	LOD (µM)	Ref.
Mn-ZnS quantum dots	0.68–7.41	0.10	6
carbon quantum dots	2.40–104.0	0.95	7
CsPbBr3 quantum dots	0.40-40.28	0.013	8

tire-derived carbon dots	0.5–60	0.17	9
N, P co-doped carbon dots	0.043–52.0	0.043	10
MMIPs	1–40	0.38	This work

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