

Supporting information

Low molecular weight poly((d,l)-lactide-co-caprolactone) liquid resins for diluent-free DLP printing of cell culture platforms.

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¹H-NMR signal analysis of intermediate and methacrylated polymers

¹H NMR **2-arm-P(d,l)LA** (CDCl₃, 500 MHz): δ (ppm) = 1.58 (d, CH₃ PLA, 48H), δ = 3.51 (q, CH PLA, 2H), δ (ppm) = 3.65 (t, CH₂ TEG, 4H), δ = 4.33 (t, CH₂ TEG, 8H), δ = 4.97 (q, CH PLA, 14H). ¹H NMR **2-arm-PCL** (CDCl₃, 500 MHz): δ (ppm) = 1.25 (tt, CH₂ PCL, 48H), δ = 1.56 (tt, CH₂ PCL, 48H), δ = 1.66 (tt, CH₂ PCL, 48H), δ (ppm) = 2.23 (t, CH₂ PCL, 48H), δ = 3.64 (t, CH₂ PCL, 48H), δ (ppm) = 3.65 (t, CH₂ TEG, 4H), δ = 4.33 (t, CH₂ TEG, 8H). ¹H NMR **2-arm-P((d,l)LA-co-CL)** (CDCl₃, 500 MHz): δ (ppm) = 1.25 (tt, CH₂ PCL, 36H), δ = 1.56 (tt, CH₂ PCL, 36H), δ (ppm) = 1.58 (d, CH₃ PLA, 36H), δ = 1.66 (tt, CH₂ PCL, 36H), δ (ppm) = 2.23 (t, CH₂ PCL, 36H), δ = 3.51 (q, CH PLA, 2H), δ = 3.64 (t, CH₂ PCL, 36H), δ (ppm) = 3.65 (t, CH₂ TEG, 4H), δ = 4.33 (t, CH₂ TEG, 8H), δ = 4.97 (q, CH PLA, 10H). ¹H NMR **3-arm-P(d,l)LA** (CDCl₃, 500 MHz): δ (ppm) = 0.91 (t, CH₃ TMP, 3H), δ (ppm) = 1.58 (d, CH₃ PLA, 72H), δ = 3.51 (q, CH PLA, 3H), δ = 4.02 (s, CH₂ TMP, 6H), δ = 4.97 (q, CH PLA, 21H). ¹H NMR **3-arm-PCL** (CDCl₃, 500 MHz): δ (ppm) = 0.91 (t, CH₃ TMP, 3H), δ (ppm) = 1.25 (tt, CH₂ PCL, 72H), δ = 1.56 (tt, CH₂ PCL, 72H), δ = 1.66 (tt, CH₂ PCL, 72H), δ (ppm) = 2.23 (t, CH₂ PCL, 72H), δ = 3.64 (t, CH₂ PCL, 72H), δ = 4.02 (s, CH₂ TMP, 6H). ¹H NMR **3-arm-P((d,l)LA-co-CL)** (CDCl₃, 500 MHz): δ (ppm) = 0.91 (t, CH₃ TMP, 3H), δ (ppm) = 1.25 (tt, CH₂ PCL, 54H), δ = 1.56 (tt, CH₂ PCL, 54H), δ (ppm) = 1.58 (d, CH₃ PLA, 54H), δ = 1.66 (tt, CH₂ PCL, 54H), δ (ppm) = 2.23 (t, CH₂ PCL, 54H), δ = 3.51 (q, CH PLA, 2H), δ = 3.64 (t, CH₂ PCL, 54H), δ = 4.02 (s, CH₂ TMP, 6H), δ = 4.97 (q, CH PLA, 16H).

¹H NMR **2-arm-P(d,l)LA-MA** (CDCl₃, 500 MHz): δ (ppm) = 1.58 (d, CH₃ PLA, 48H), δ (ppm) = 1.99 (s, CH₃ MA, 6H), δ = 3.51 (q, CH PLA, 2H), δ (ppm) = 3.65 (t, CH₂ TEG, 4H), δ = 4.33 (t, CH₂ TEG, 8H), δ = 4.97 (q, CH PLA, 14H), δ = 5.58 (dd, CH_{trans}=C MA, 2H), δ = 6.15 (dd, CH_{cis}=C MA, 2H). ¹H NMR **2-arm-PCL-MA** (CDCl₃, 500 MHz): δ (ppm) = 1.25 (tt, CH₂ PCL, 48H), δ = 1.56 (tt, CH₂ PCL, 48H), δ = 1.66 (tt, CH₂ PCL, 48H), δ (ppm) = 1.99 (s, CH₃ MA, 6H), δ (ppm) = 2.23 (t, CH₂ PCL, 48H), δ = 3.64 (t, CH₂ PCL, 48H), δ (ppm) = 3.65 (t, CH₂ TEG, 4H), δ = 4.33 (t, CH₂ TEG, 8H), δ = 5.58 (dd, CH_{trans}=C MA, 2H), δ = 6.15 (dd, CH_{cis}=C MA, 2H). ¹H NMR **2-arm-P((d,l)LA-co-CL)-MA** (CDCl₃, 500 MHz): δ (ppm) = 1.25 (tt, CH₂ PCL, 36H), δ = 1.56 (tt, CH₂ PCL, 36H), δ (ppm) = 1.58 (d, CH₃ PLA, 36H), δ = 1.66 (tt, CH₂ PCL, 36H), δ (ppm) = 1.99 (s, CH₃ MA, 6H), δ (ppm) = 2.23 (t, CH₂ PCL, 36H), δ = 3.51 (q, CH PLA, 2H), δ = 3.64 (t, CH₂ PCL, 36H), δ (ppm) = 3.65 (t, CH₂ TEG, 4H), δ = 4.33 (t, CH₂ TEG, 8H), δ = 4.97 (q, CH PLA, 10H), δ = 5.58 (dd, CH_{trans}=C MA, 2H), δ = 6.15 (dd, CH_{cis}=C MA, 2H). ¹H NMR **3-arm-P(d,l)LA-MA** (CDCl₃, 500 MHz): δ (ppm) = 0.91 (t, CH₃ TMP, 3H), δ (ppm) = 1.58 (d, CH₃ PLA, 72H), δ (ppm) = 1.99 (s, CH₃ MA, 9H), δ = 3.51 (q, CH PLA, 3H), δ = 4.02 (s, CH₂ TMP, 6H), δ = 4.97 (q, CH PLA, 21H), δ = 5.58 (dd, CH_{trans}=C MA, 3H), δ = 6.15 (dd, CH_{cis}=C MA, 3H). ¹H NMR **3-arm-PCL-MA** (CDCl₃, 500 MHz): δ (ppm) = 0.91 (t, CH₃ TMP, 3H), δ (ppm) = 1.25 (tt, CH₂ PCL, 72H), δ = 1.56 (tt, CH₂ PCL, 72H), δ = 1.66 (tt, CH₂ PCL, 72H), δ (ppm) = 1.99 (s, CH₃ MA, 9H), δ (ppm) = 2.23 (t, CH₂ PCL, 72H), δ = 3.64 (t, CH₂ PCL, 72H), δ = 4.02 (s, CH₂ TMP, 6H), δ = 5.58 (dd, CH_{trans}=C MA, 3H), δ = 6.15 (dd, CH_{cis}=C MA, 3H). ¹H NMR **3-arm-P((d,l)LA-co-CL)-MA** (CDCl₃, 500 MHz): δ (ppm) = 0.91 (t, CH₃ TMP, 3H), δ (ppm) = 1.25 (tt, CH₂ PCL, 54H), δ = 1.56 (tt, CH₂ PCL, 54H), δ (ppm) = 1.58 (d, CH₃ PLA, 54H), δ = 1.66 (tt, CH₂ PCL, 54H), δ (ppm) = 1.99 (s, CH₃ MA, 9H), δ (ppm) = 2.23 (t, CH₂ PCL, 54H), δ = 3.51 (q, CH PLA, 2H), δ = 3.64 (t, CH₂ PCL, 54H), δ = 4.02 (s, CH₂ TMP, 6H), δ = 4.97 (q, CH PLA, 16H), δ = 5.58 (dd, CH_{trans}=C MA, 3H), δ = 6.15 (dd, CH_{cis}=C MA, 3H).

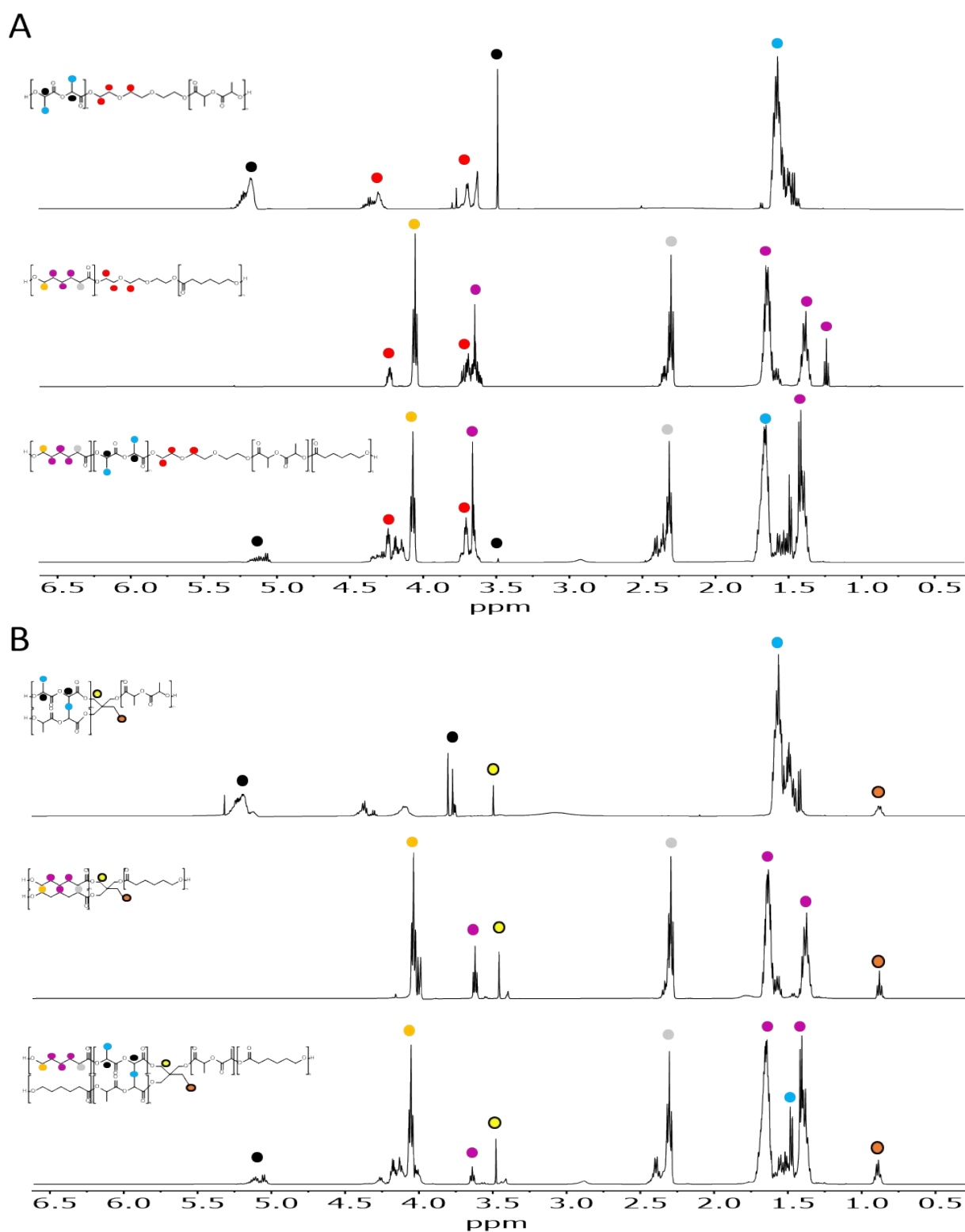


Fig. S1 $^1\text{H-NMR}$ spectrum analysis of 2- (A) and 3-arm (B) intermediate polymers in CDCl_3 : (top) P(d,l)LA, (middle) PCL and (bottom) P(d,l)LA-co-CL. The specific proton signals are related to the molecule ones using colour dots.

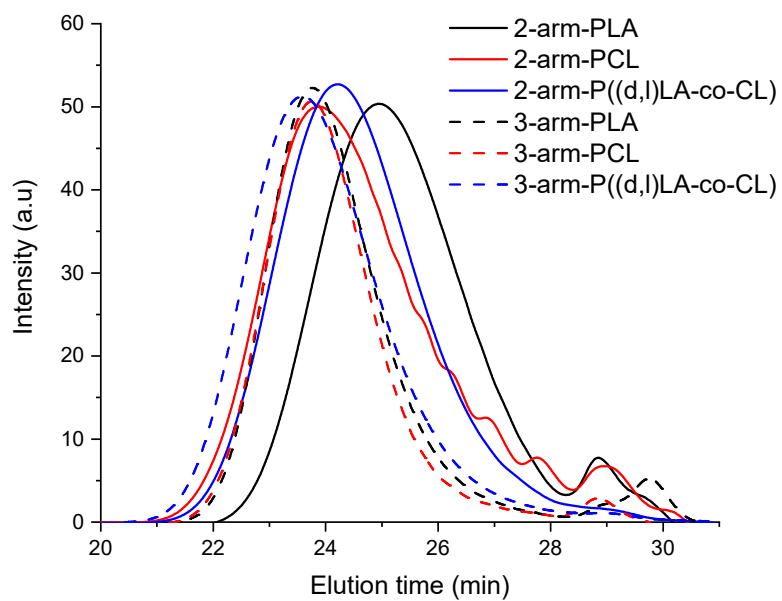


Fig. S2 GPC chromatogram of synthesized polymers.

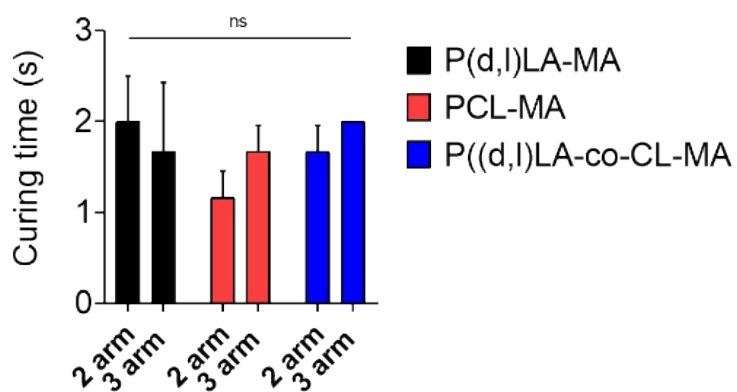


Fig. S3 Curing time of the methacrylated resins during photorheological experiments. Error bars denote standard deviation.

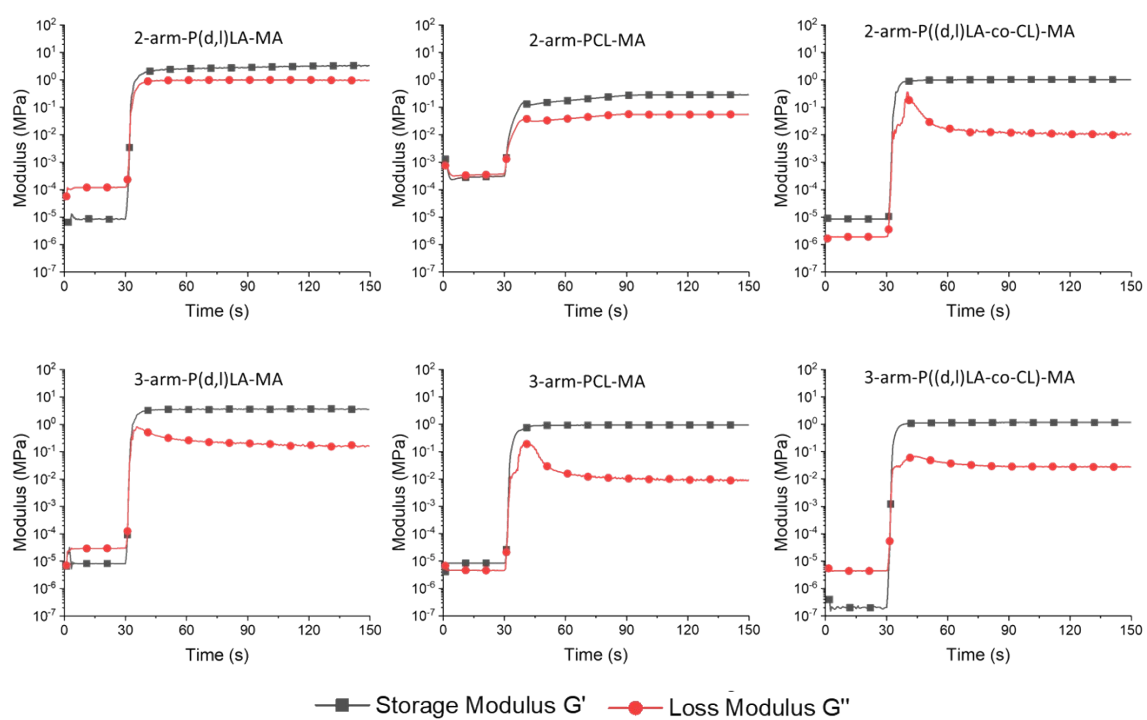


Fig. S4 Storage and Loss moduli before and after UV exposure of 2-arm (top row) and 3-arm (bottom row) P((d,l)LA-co-CL)-MA (left), PCL-MA (middle) and P((d,l)-LA-MA (left).

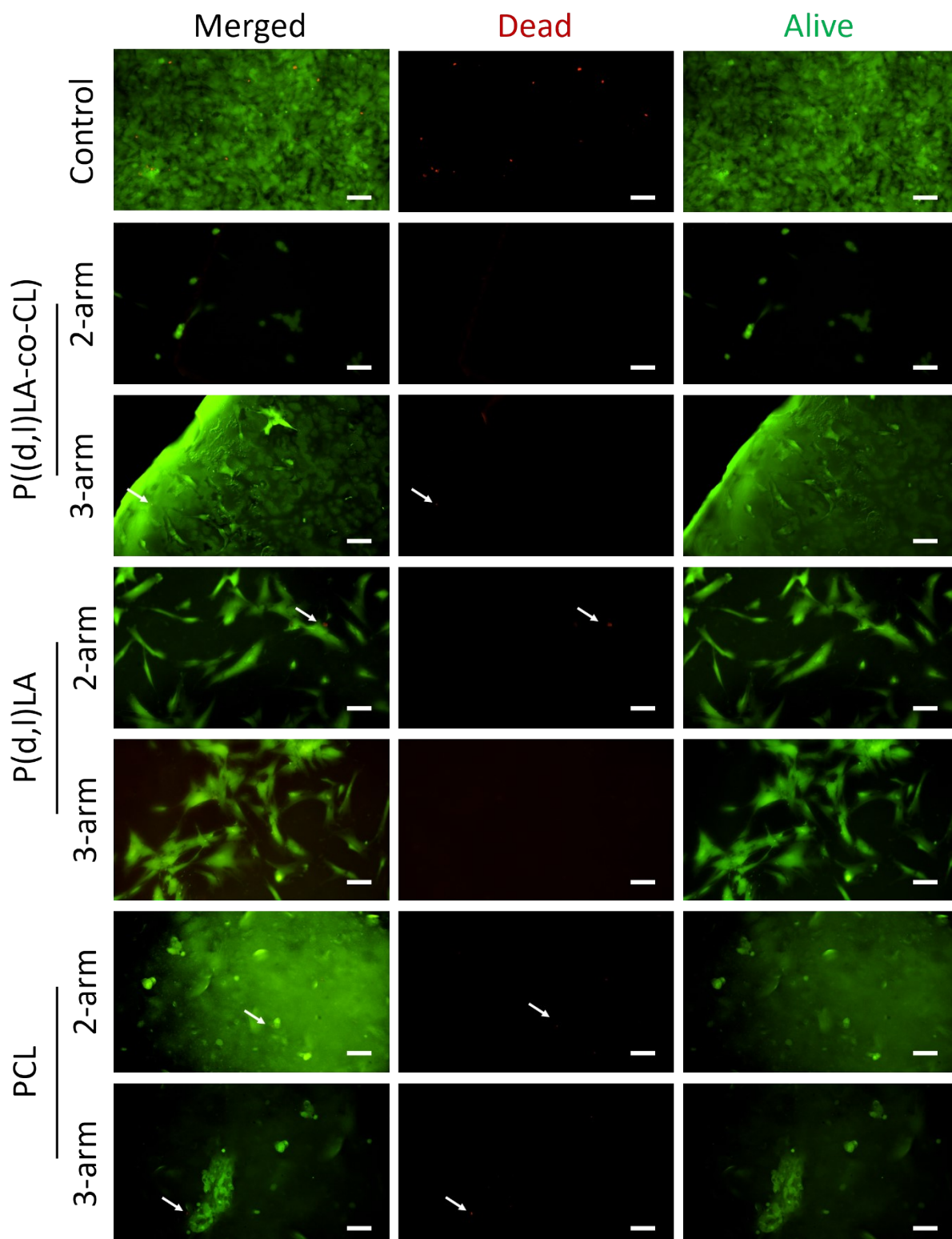


Fig. S5 Split channels and merged images for the LIVE/DEAD assay of hMSCs after 24 hours seeded onto the films of the different (2- and 3-arms) homo- and copolymers. Dead cells are stained red (ethidium homodimer) and alive cells appear stained with calcein (green). Scale bars are 100 μm .

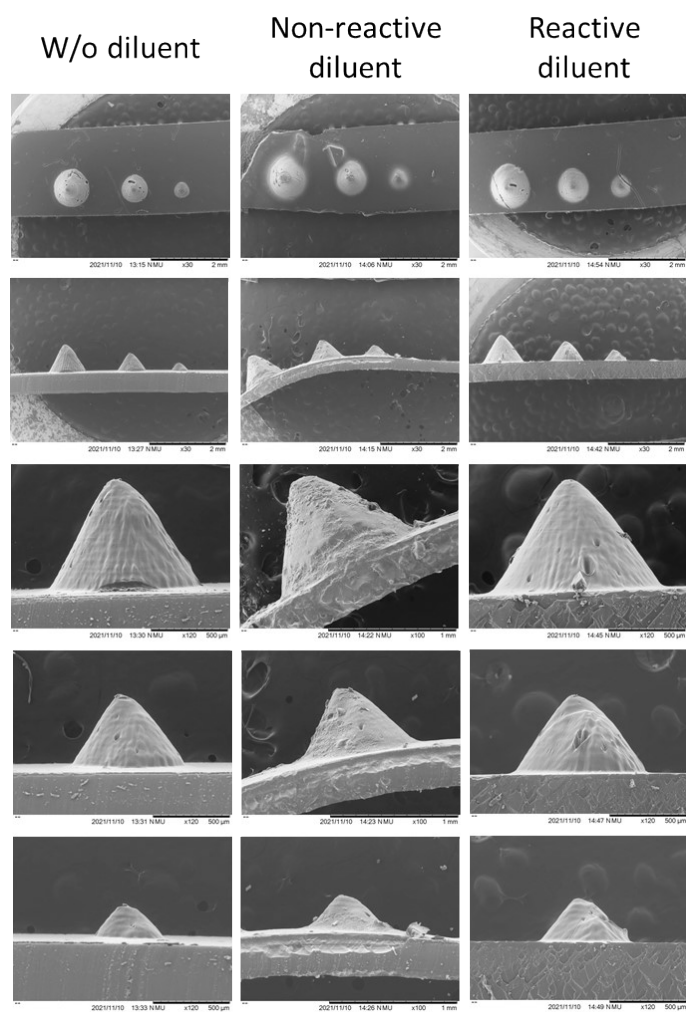


Fig. S6 SEM images of the models with a gradient of cones in the surface using the copolymer P((d,l)LA-co-CL) of 2 arms without diluent (left), with non-reactive diluent (middle) and reactive diluent (right). Higher magnification of the cones with different sizes are shown from the bigger one to the smallest at the bottom.

Table S1 **Micro-topography printing error of the gradients of cubes.** Average error in the diameter (ϵ_ϕ) and the average error in height (ϵ_h). The theoretical dimensions of the 3 cones respectively was (Diameter x height): 1 mm x 1 mm, 0.75 mm x 0.75 mm and 0.5 mm x 0.5 mm.

2-arm-P((d,l)LA-co-CL)-MA				
	Cone position	W/o diluent	W/ non-reactive diluent	W/ reactive diluent
ϵ_ϕ (%) \pm SD	1	2 ± 0.7	10 ± 2	3 ± 2
	2	5 ± 0.7	1 ± 0.5	7 ± 0.2
	3	19 ± 7	24 ± 3	11 ± 3
ϵ_h (%) \pm SD	1	35 ± 0.4	33 ± 2	37 ± 2
	2	57 ± 1	39 ± 2	42 ± 2
	3	126 ± 3	71 ± 11	79 ± 1