

Supporting information

## Enhanced biophysical screening strategy to investigate the affinity of ASOs for their target RNA.

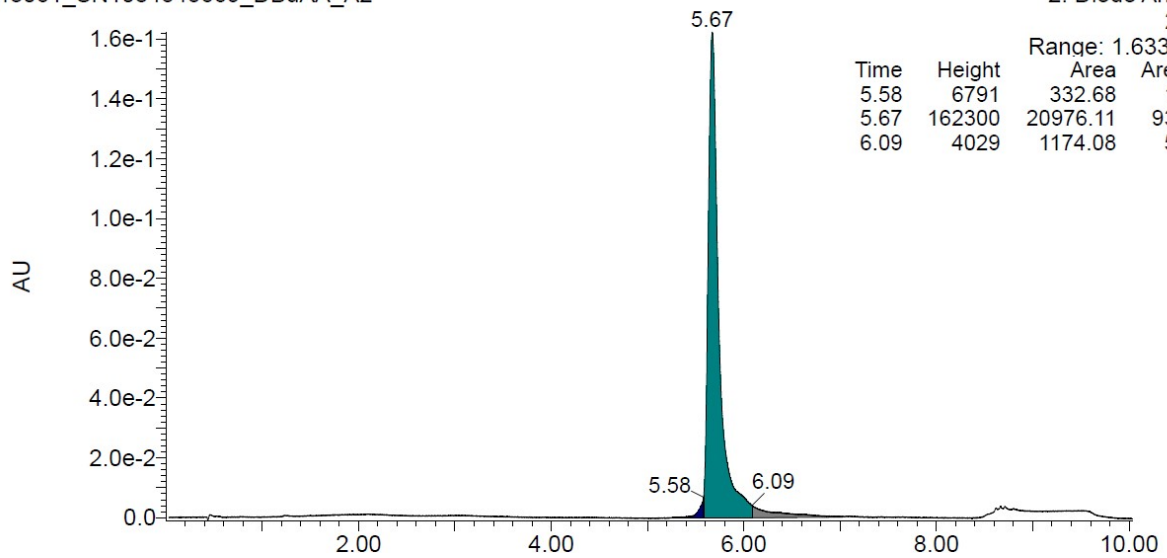
Table S1. Table of ASO sequences used in this study and correspondent properties. Bold letters denote LNA nucleotides, italic letters denote MOE nucleotides <sup>m</sup>C denotes 5-methylcytidine, † denotes 2'-O-Methyl nucleotides. All sequences were fully phosphorothioate modified.

ASO	Sequence	Modifications included	Length	Average Molecular Weight (g/mol)	Molar Extinction Coefficient (L/mol·cm)	Calc. HRMS (3-)	Measured HRMS (3-)
1	<b>TG<sup>m</sup>CTACAAAAC<sup>m</sup>C<sup>m</sup>CA</b>	LNA PS gapmers 3-8-3	14	4619.76	132100	1537.82 13	1537.8464
2	<i>TG<sup>m</sup>CTACAAAAC<sup>m</sup>C<sup>m</sup>CA</i>	MOE PS gapmers 3-8-3	14	4896.17	133200	1629.90 50	1629.9308
3	†U†G†C†U†A†C†A†A†A†A†C† C†C†A	2'OMe PS	14	4801.93	138400	1598.52 14	1598.5465
4	<b>TG<sup>m</sup>CT†ACAAAAC<sup>m</sup>C<sup>m</sup>CA</b>	LNA PS gapmers 3-8-3, 2'-OMe in position 2	14	4649.79	132600	1547.82 48	1547.8496
5	<b>TG<sup>m</sup>CTA†CAAAAAC<sup>m</sup>C<sup>m</sup>CA</b>	LNA PS gapmers 3-8-3, 2'-OMe in position 3	14	4649.79	132100	1547.82 48	1547.8503
6	<b>AUUAAUAAAAAUGC</b>	LNA PS gapmers 3-8-3	14	4633.73	133800	1542.46 94	1542.4938
7	<b>AACCCAGAAUAAAU</b>	LNA PS gapmers 3-8-3	14	4651.70	131600	1548.46 08	1548.4880
8	<b>TGATACAAAAC<sup>m</sup>C<sup>m</sup>CA</b>	LNA PS gapmers 3-8-3	14	4629.76	141000	1541.15 32	1541.1784
9	<i>TGATACAAAAC<sup>m</sup>C<sup>m</sup>CA</i>	MOE PS gapmers 3-8-3	14	4906.17	142200	1633.23 69	1633.2643
10	<b>TG<sup>m</sup>CTA<sup>A</sup>AAAAC<sup>m</sup>C<sup>m</sup>CA</b>	LNA PS gapmers 3-8-3	14	4643.79	136500	1545.82 51	1545.8483
11	<i>TG<sup>m</sup>CTA<sup>A</sup>AAAAC<sup>m</sup>C<sup>m</sup>CA</i>	MOE PS gapmers 3-8-3	14	4920.20	137600	1637.90 88	1637.9323
12	<b>GTCTGTGGAAG<sup>m</sup>CG</b>	LNA PS gapmers 2-8-3	13	4377.46	123100	1457.13 27	1457.1570
13	<b><sup>m</sup>CGTCA<sup>G</sup>TATGCGAATC</b>	LNA PS gapmers 3-8-3-1	16	5304.28	152200	1324.11 64 (4-)	1324.1384 (4-)

### ASO1

SN1084545669\_DBuAA\_5\_50

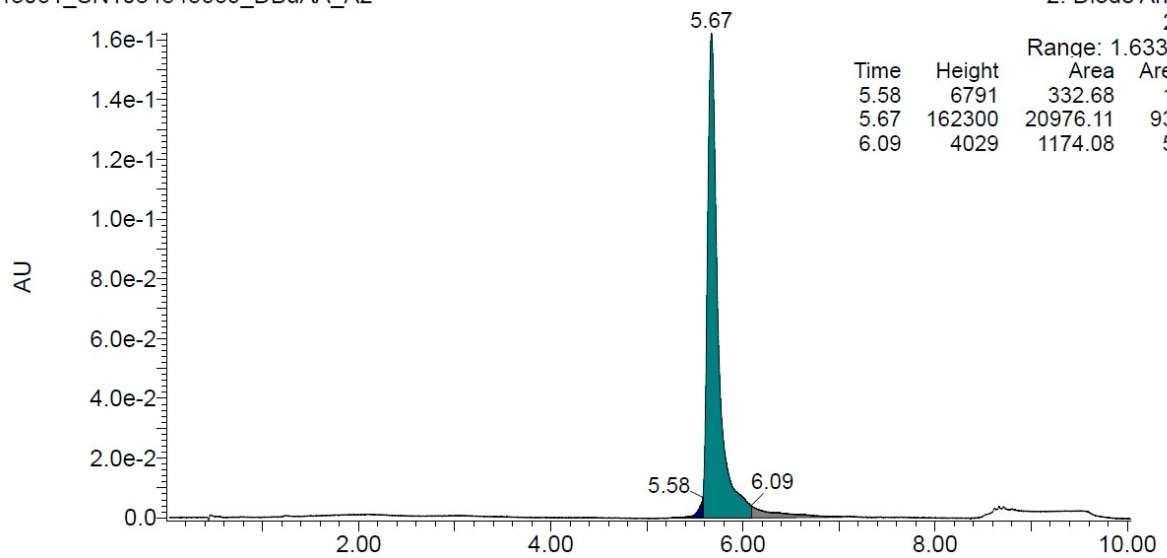
43061\_SN1084545669\_DBuAA\_A2



### ASO2

SN1084545669\_DBuAA\_5\_50

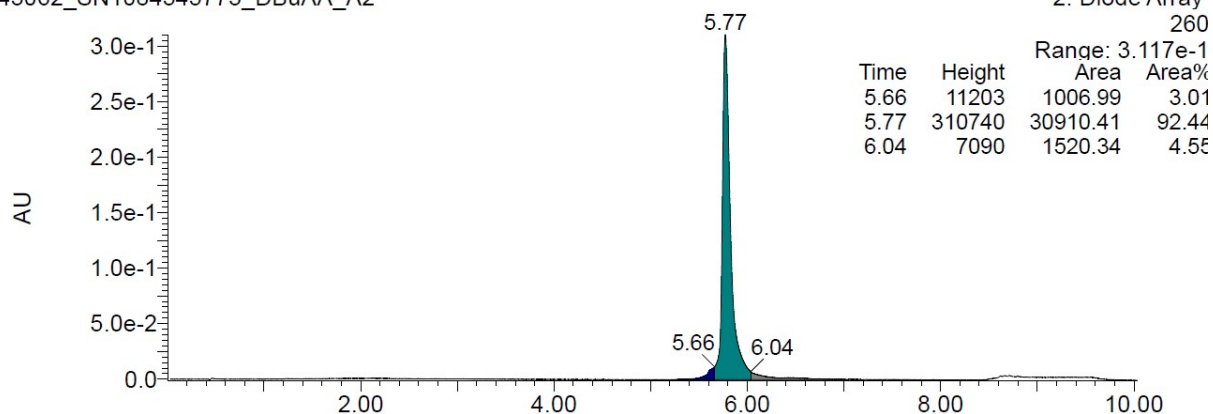
43061\_SN1084545669\_DBuAA\_A2



### ASO3

SN1084545773\_DBuAA\_5\_50

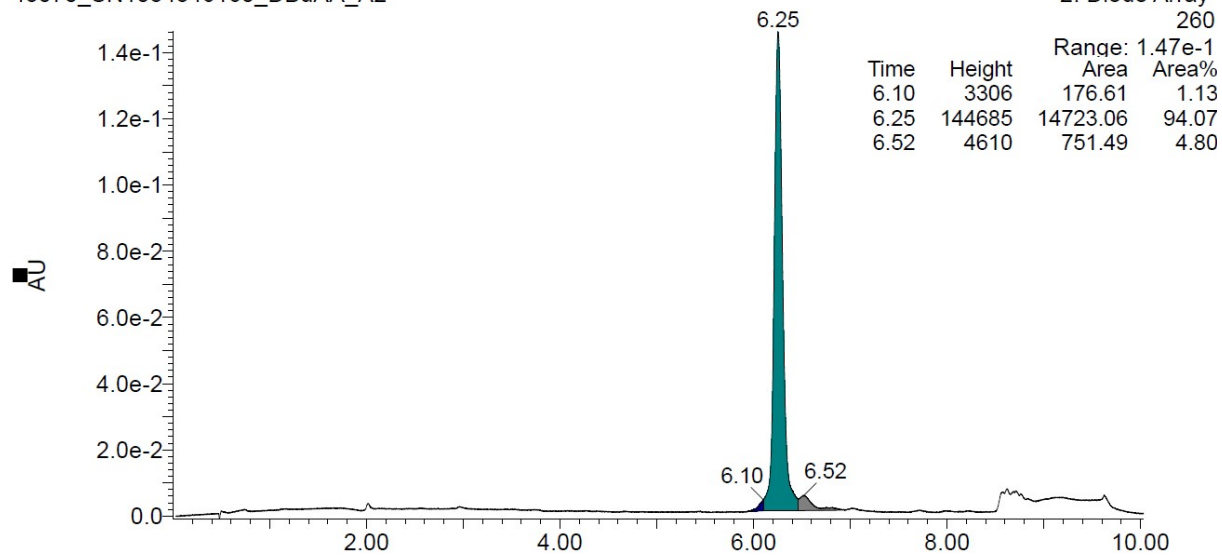
43062\_SN1084545773\_DBuAA\_A2



### ASO4

SN1084546160\_DBuAA\_5\_50

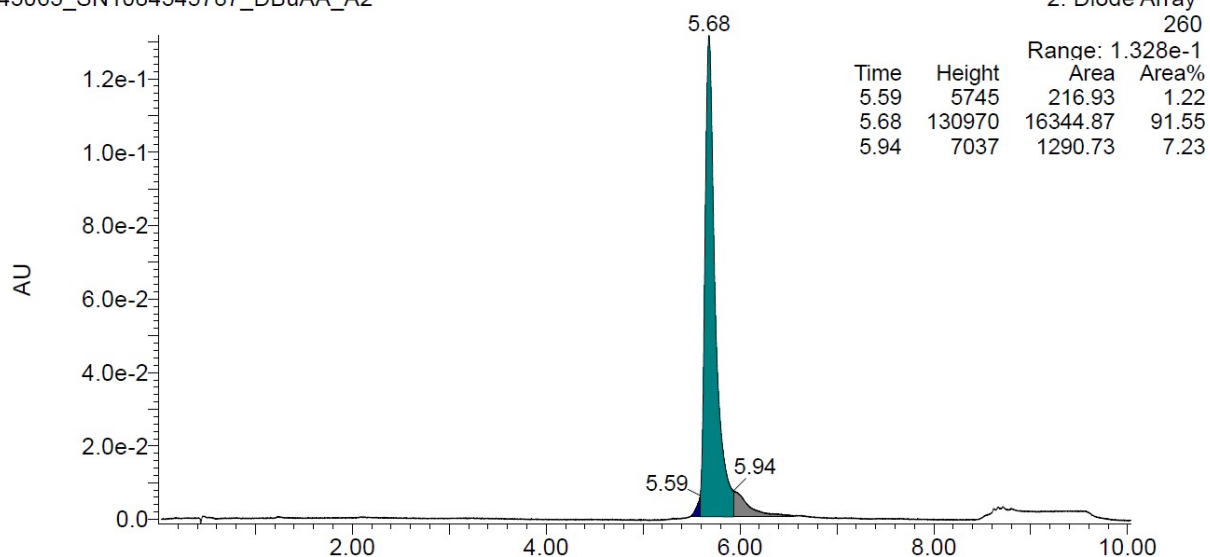
43070\_SN1084546160\_DBuAA\_A2



### ASO5

SN1084545787\_DBuAA\_5\_50

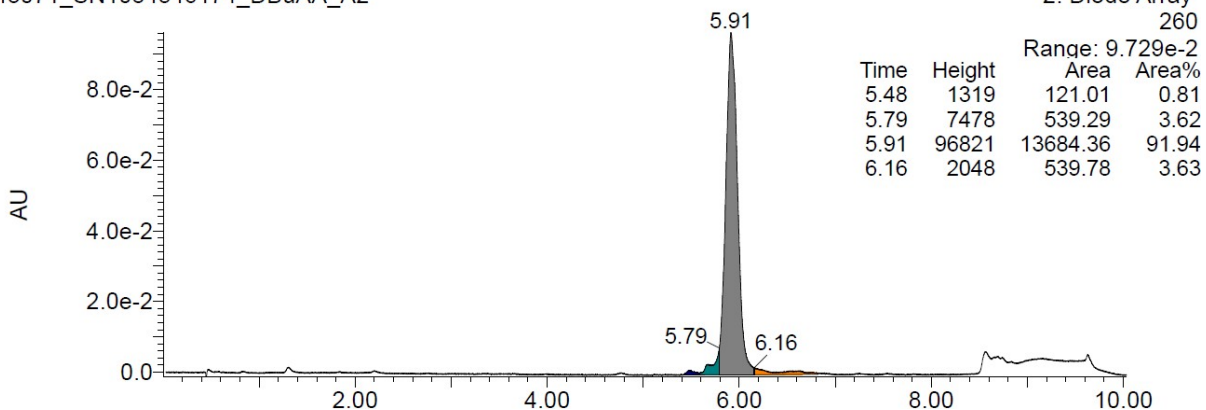
43063\_SN1084545787\_DBuAA\_A2



### ASO6

SN1084546174\_DBuAA\_5\_50

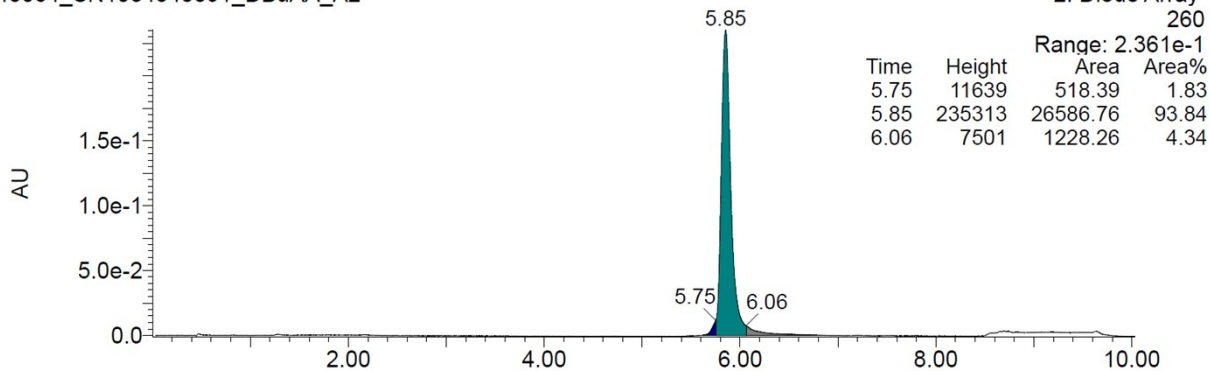
43071\_SN1084546174\_DBuAA\_A2



### ASO 7

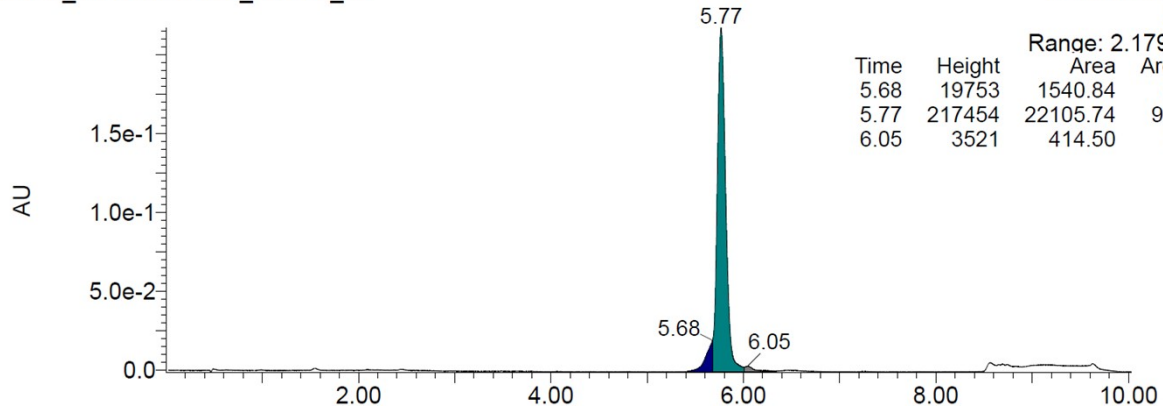
SN1084545891\_DBuAA\_5\_50

43064\_SN1084545891\_DBuAA\_A2



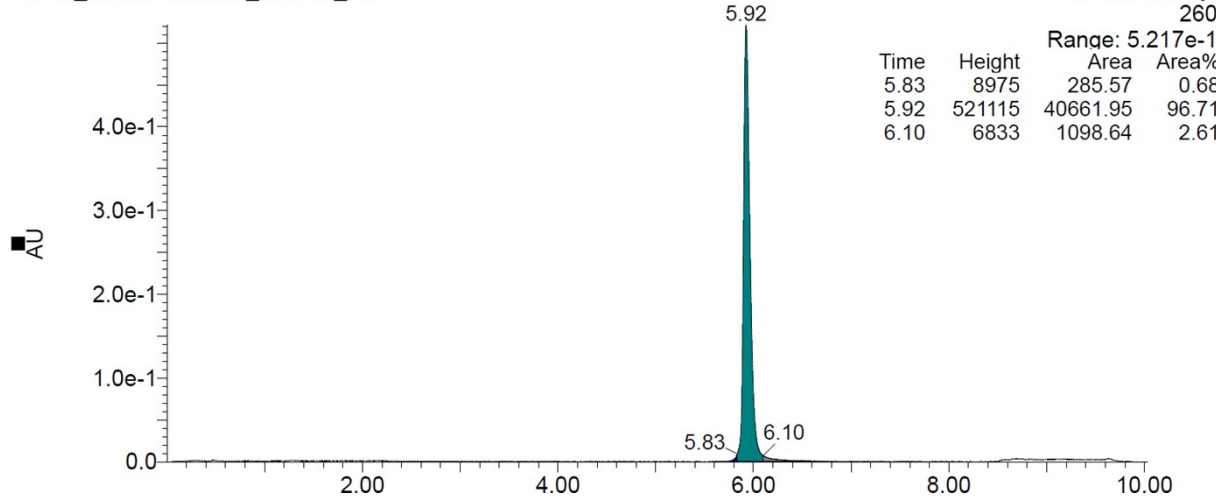
### ASO8

SN1084546188\_DBuAA\_5\_50  
43072\_SN1084546188\_DBuAA\_A2



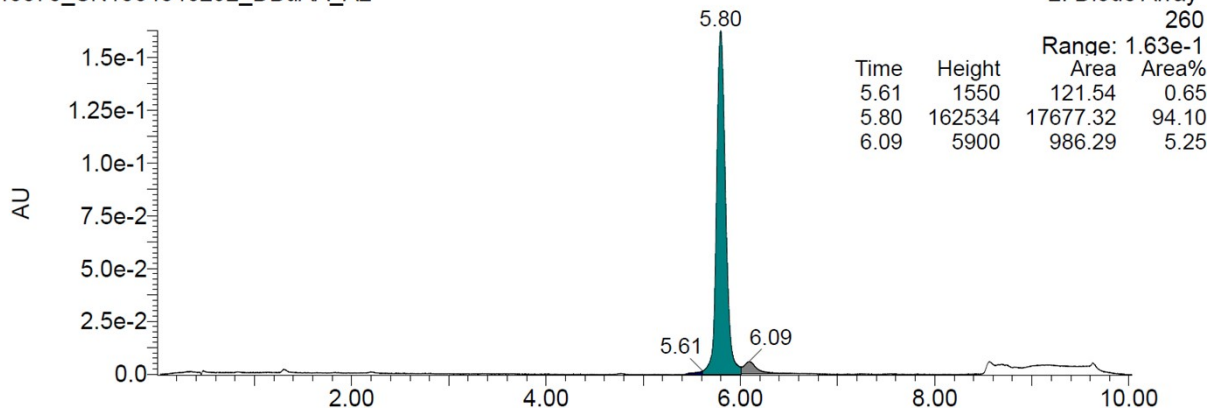
### ASO9

SN1084545806\_DBuAA\_5\_50  
43065\_SN1084545806\_DBuAA\_A2



### ASO10

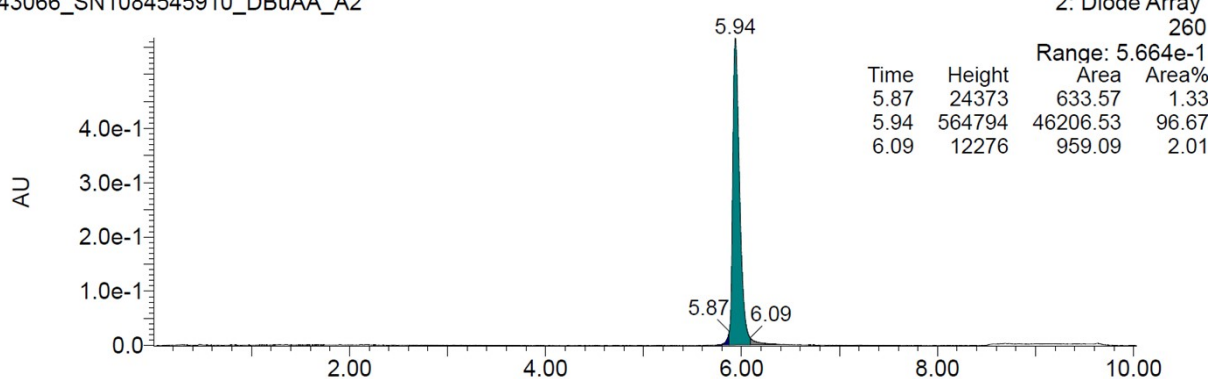
SN1084546292\_DBuAA\_5\_50  
43073\_SN1084546292\_DBuAA\_A2



### ASO11

SN1084545910\_DBuAA\_5\_50

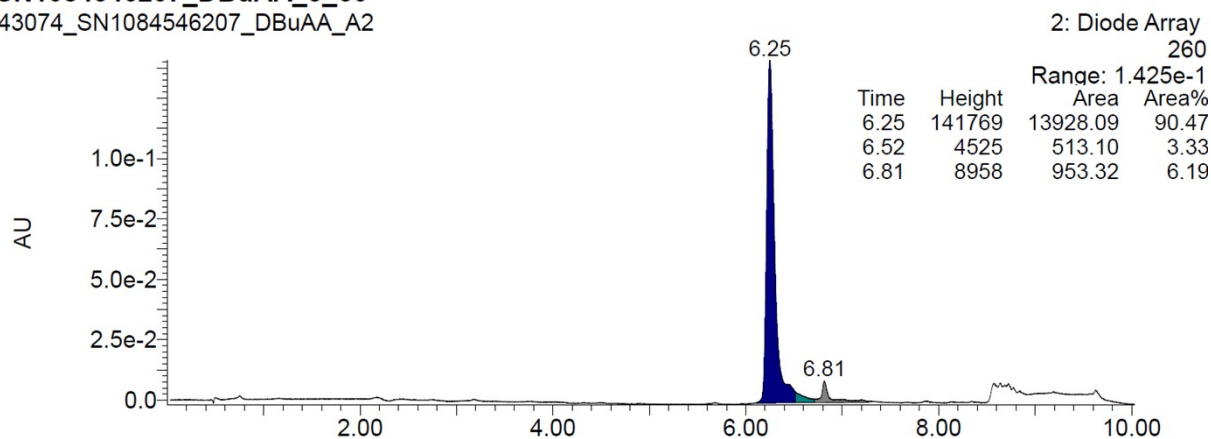
43066\_SN1084545910\_DBuAA\_A2



### ASO12

SN1084546207\_DBuAA\_5\_50

43074\_SN1084546207\_DBuAA\_A2



### ASO13

SN1084545924\_DBuAA\_5\_50

43067\_SN1084545924\_DBuAA\_A2

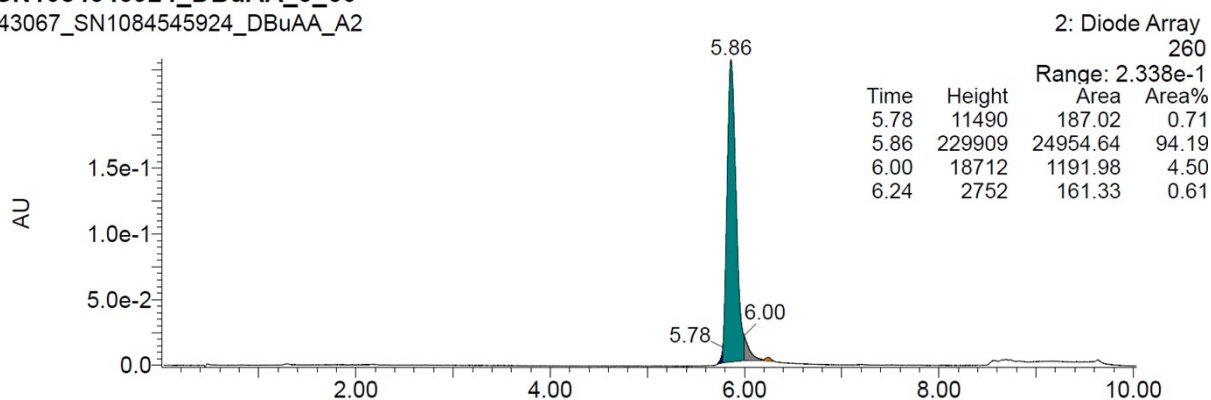


Figure S1. Oligonucleotide analytics for the sequences used in this study. Oligonucleotides were purified by preparative HPLC on aXBridge C18, 5µm 19x150mm column. A gradient from 15-50% solvent A (60mM dibutylammonium-acetate, pH 7, in acetonitrile:water=5:95) to solvent B (60mM dibutylammonium-acetate, pH 7 in acetonitrile) in 12 minutes using a flowrate of 20 mL/min. Analytical HPLC was performed on a Waters Acquity BEH C18, 1.7µm 2.1x100mm column using a gradient from 5-50% solvent A (60mM dibutylammonium-acetate, pH 7, in acetonitrile:water=5:95) to solvent B (60mM dibutylammonium-acetate, pH 7 in acetonitrile) at 60 °C and chromatograms were collected by UV detection at a wavelength of 260nm.

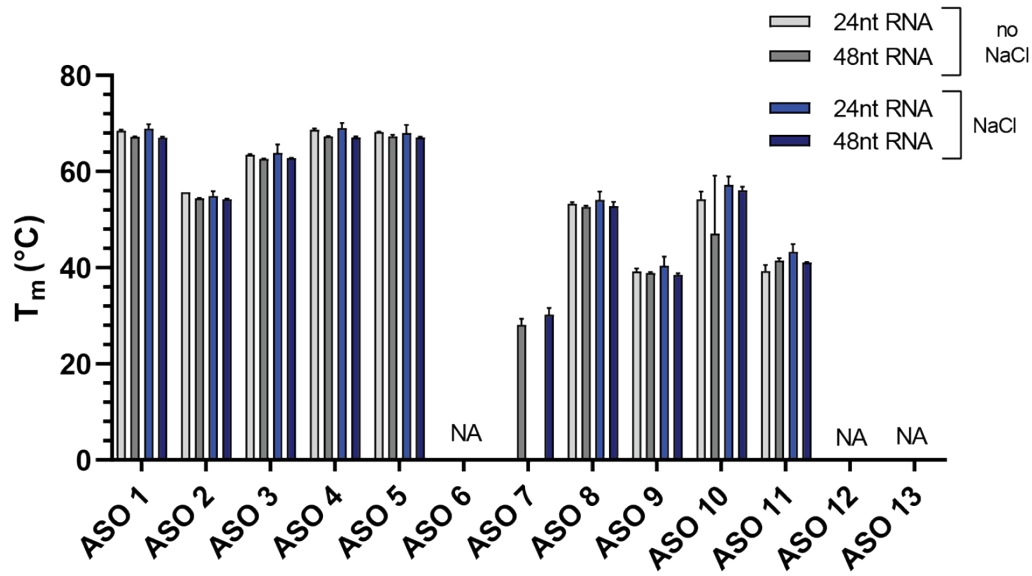


Figure S2. Differential scanning fluorimetry data for all ASOs and RNAs in two different buffer conditions, with salts and without salts.

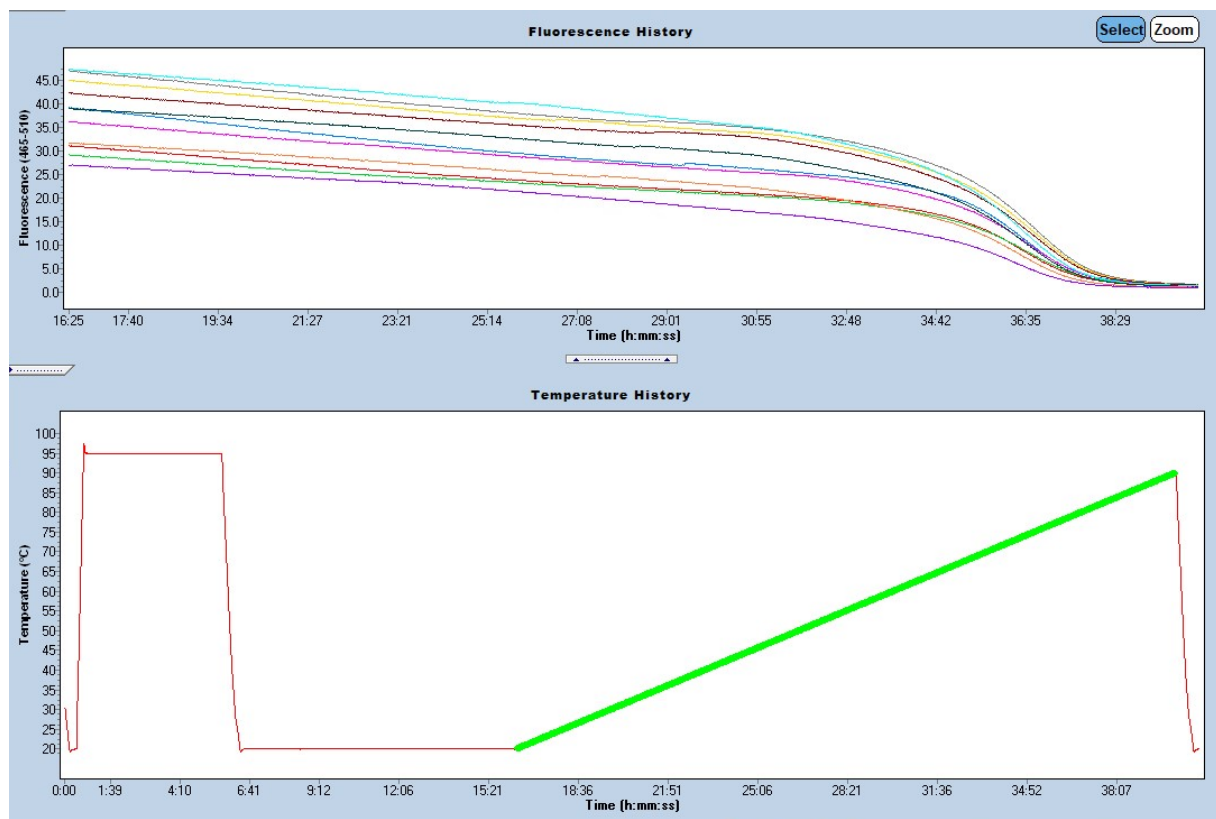


Figure S3. Fluorescence curves produced from the Roche 480 Lightcycler using RIBOGreen reporter dye. The fluorescence intensity curves for of ASOs/RNA hybridized duplexes and RNA used in this study where measures at 4  $\mu$ M in 10  $\mu$ l total volume and 50 nl of Quant-iT<sup>TM</sup> RiboGreen<sup>®</sup> (RIBO, Life Technologies, Ex max= $\sim$ 500 nm, Em max= $\sim$ 525 nm) dye stock solution in DMSO. The fluorescence intensity was monitored using the SYBR Green I/HRM dye filter combination (465–510 nm) from 20 to 95  $^{\circ}$ C at a ramp rate of 0.03  $^{\circ}$ C/s with 36 acquisitions per  $^{\circ}$ C.



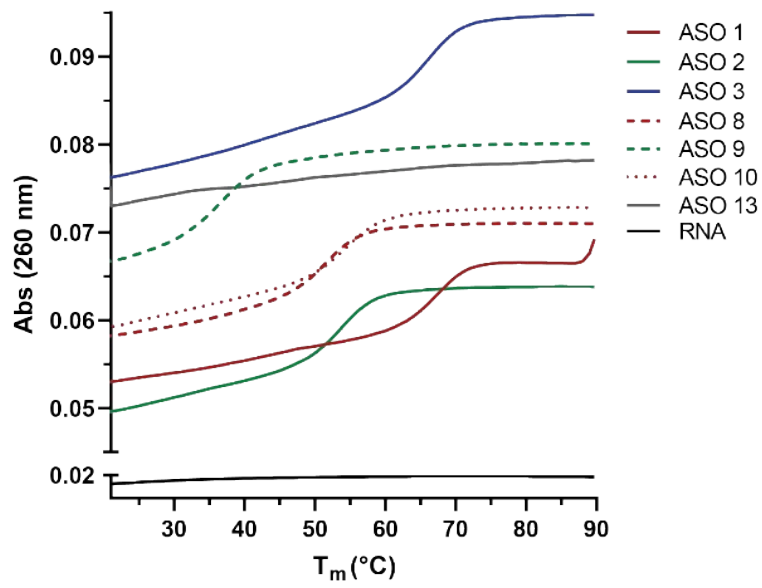
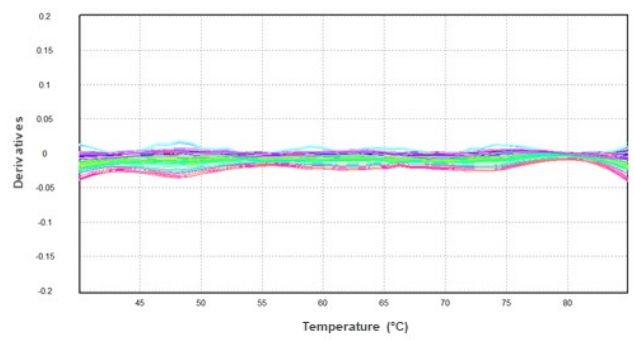
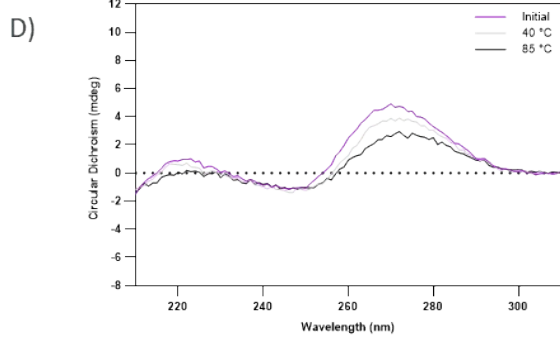
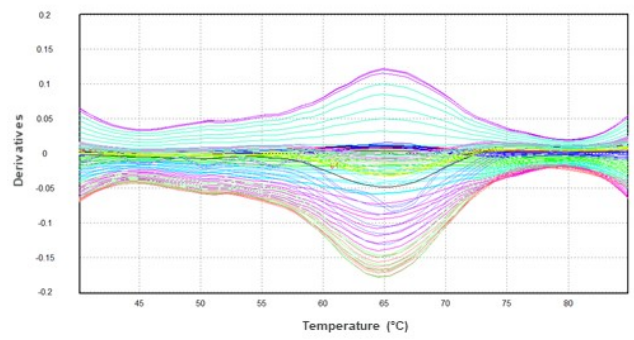
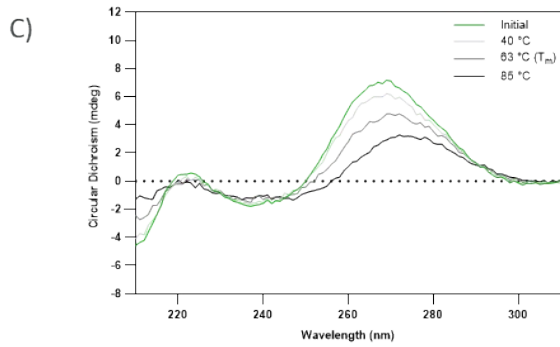
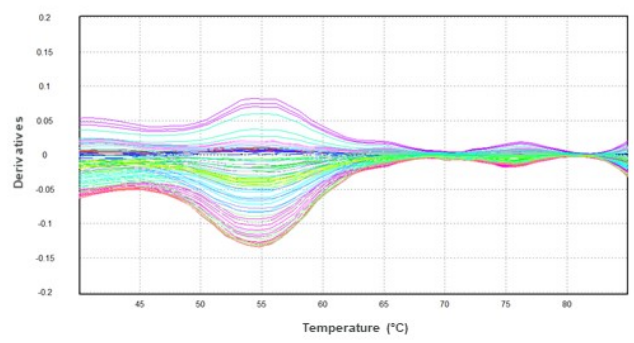
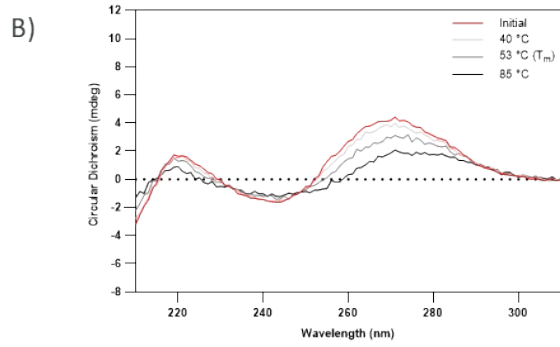
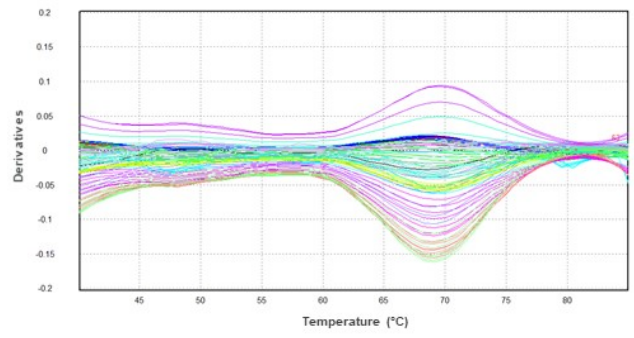
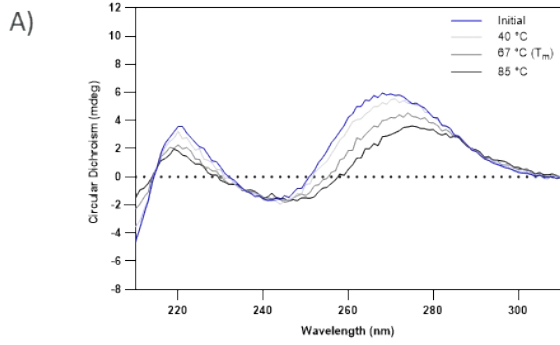
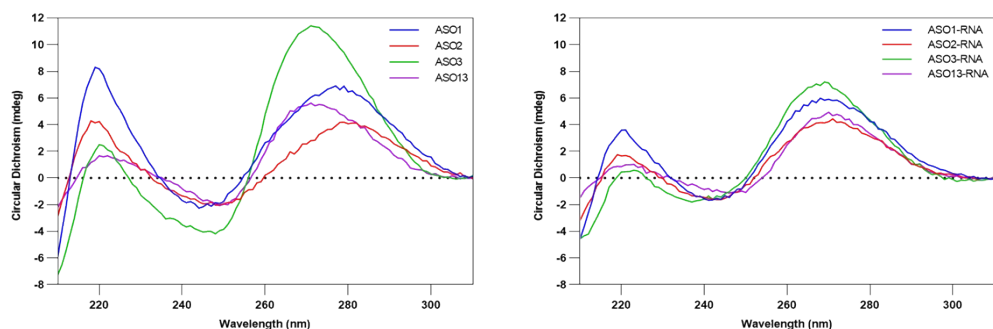


Figure S4. Raw denaturation melting temperature curves measured by absorbance at 260nm through UV spectroscopy.

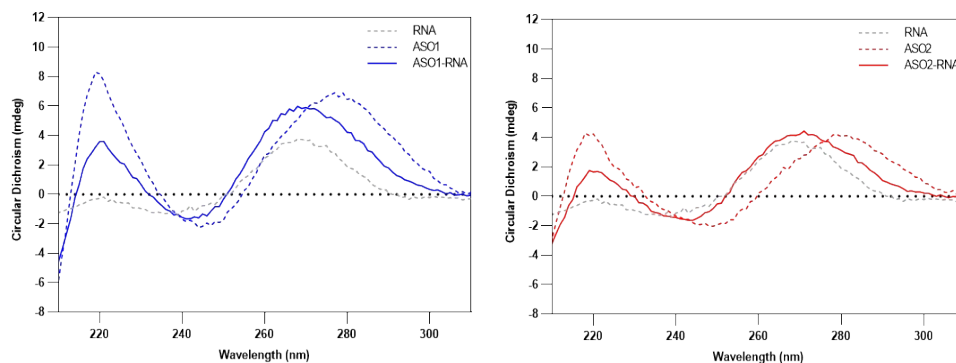




E)



F)



G)

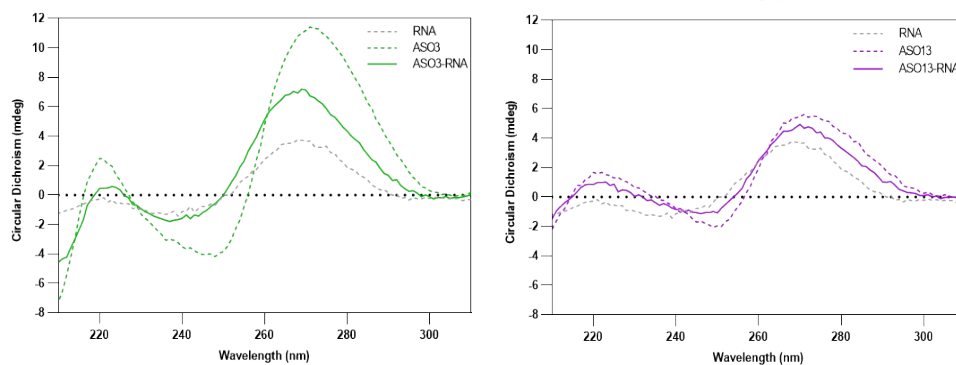


Figure S5. Circular Dichroism Spectroscopy. The CD spectra at 20°C (initial), 40°C, melting temperature ( $T_m$ ) and 85 °C (left) and the derivative plots including all wavelengths (right) for A) ASO 1/RNA B) ASO 2/RNA C) ASO 3-RNA D) ASO 13/RNA. The CD derivative plots for E) ASO 1-RNA F) ASO 2-RNA G) ASO 3-RNA D) ASO 13-RNA at 85°C

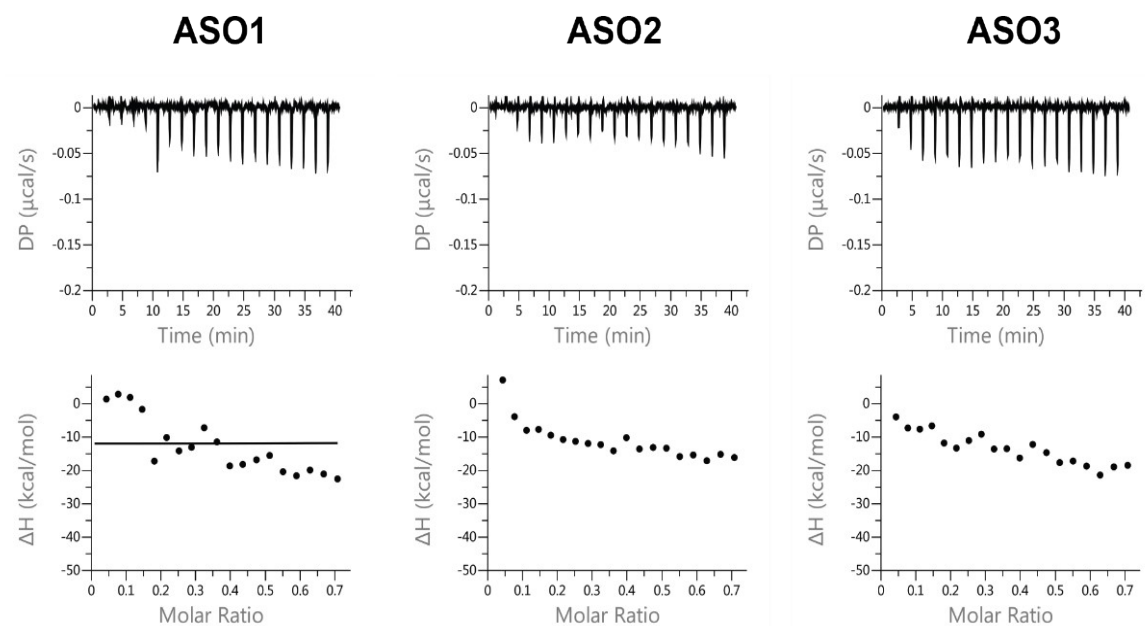


Figure S6. Isothermal calorimetry. Controls binding isotherms for ASO 1, 2 and 3 AT 37°C.

Table S2. SPR kinetics values measured at 25°C through a 1:1 binding model

ASO	AZ Sequence	Sugar Composition	Sugar Pattern	ka (1/Ms)	kd (1/s)	KD (M)	St dev KD
1	TGmCTACAAAACmCmCA	LNA-DNA-LNA	3-8-3	5.99E+06	2.59E-03	4.28E-10	1.71E-10
2	TGmCTACAAAACmCmCA	MOE-DNA-MOE	3-8-3	2.33E+05	5.38E-05	2.34E-10	9.06E-11
3	UGCUACAAAACCCA	OMe	14	3.02E+05	2.81E-05	7.81E-11	7.46E-11
8	TGATACAAAACmCmCA	LNA-DNA-LNA	3-8-3	4.77E+05	2.75E-04	3.87E-10	9.06E-11
9	TGATACAAAACmCmCA	MOE-DNA-MOE	3-8-3	4.61E+08	1.32E+01	2.83E-08	3.97E-10
10	TGmCTAAAAAACmCmCA	LNA-DNA-LNA	3-8-3	3.52E+08	9.73E+00	1.81E-08	7.79E-11
11	TGmCTAAAAAACmCmCA	MOE-DNA-MOE	3-8-3	9.95E+05	7.79E-03	7.92E-09	1.12E-09
13	mCGTCAGTATGCGAATC	LNA-DNA-LNA	3-8-3	NA	NA	NA	NA

Table S3. . SPR kinetics values measured at 37°C through a 1:1 binding model.

ASO	AZ Sequence	Sugar Composition	Sugar Pattern	ka (1/Ms)	kd (1/s)	KD (M)	St dev KD
1	TGmCTACAAAACmCmCA	LNA-DNA-LNA	3-8-3	1.50E+06	2.15E-04	1.41E-10	3.11E-11
2	TGmCTACAAAACmCmCA	MOE-DNA-MOE	3-8-3	1.11E+06	1.32E-03	2.79E-09	2.31E-09
3	UGCUACAAAACCCA	OMe	14	6.73E+05	1.11E-03	1.88E-09	2.15E-09
8	TGATACAAAACmCmCA	LNA-DNA-LNA	3-8-3	8.86E+05	4.23E-03	5.50E-09	2.31E-09
9	TGATACAAAACmCmCA	MOE-DNA-MOE	3-8-3	NA	NA	NA	NA
10	TGmCTAAAAAACmCmCA	LNA-DNA-LNA	3-8-3	5.80E+05	5.31E-03	1.89E-08	2.47E-08
11	TGmCTAAAAAACmCmCA	MOE-DNA-MOE	3-8-3	NA	NA	NA	NA
13	mCGTCAGTATGCGAATC	LNA-DNA-LNA	3-8-3	NA	NA	NA	NA

Table S4. ITC values measured at 37°C and 45°C for ASO 1, 2, 3 and 24nt RNA duplexes.

Samples	T(°C)	N (sites)	KD (M)	$\Delta H$ (kcal/mol)	$\Delta G$ (kcal/mol)	$-T\Delta S$ (kcal/mol)	Offset (kcal/mol)	Red. Chi-Sqr. (kcal/mol) <sup>2</sup>
24nt RNA + ASO1	37	0.927	8.09E-09	-35.8	-11.6	24.2	-5.6	7.2
24nt RNA + ASO2	37	0.972	3.91E-09	-21.9	-11.9	9.9	-7.0	1.4
24nt RNA + ASO3	37	1.1	3.32E-09	-24.5	-12.4	12.1	-13.4	3.5
24nt RNA + ASO1	45	0.927	1.22E-08	-46.0	-11.5	34.5	-2.1	18.0
24nt RNA + ASO2	45	1	4.87E-08	-38.3	-10.6	27.6	0.1	5.2
24nt RNA + ASO3	45	1.02	8.52E-09	-43.8	-11.7	32.0	-1.8	3.7

Table S5. Invariant parameters of ASO 2 in the presence and absence of RNA

Sample	Maximum Dimension (Å)	Radius of gyration (Å)	Molecular weight estimate (Da)
ASO	43	14.2	4050-5450
ASO + 24ntRNA	63	20.5	9950-11650

Table S6. Replicates SPR assay at 25°C.

Sample	ka (1/Ms)	kd (1/s)	KD (M)	Rmax (RU)	Chi <sup>2</sup> (RU <sup>2</sup> )	Model
ASO 1	5.00E+06	1.74E-03	3.48E-10	65.8	1.57	1:1 w Drift
	6.64E+06	2.07E-03	3.12E-10	54.7	1.7	1:1 w Drift
	6.33E+06	3.96E-03	6.25E-10	75.6	1.8	1:1 w Drift
ASO 2	2.30E+05	4.20E-05	1.83E-10	110.2	0.526	1:1 w Drift
	2.52E+05	4.56E-05	1.81E-10	93	1.03	1:1 w Drift
	2.18E+05	7.37E-05	3.39E-10	116.4	1.21	1:1 w Drift
ASO 3	2.98E+05	2.53E-05	8.49E-11	136.4	0.848	1:1 w Drift
	3.94E+05	5.88E-05	1.49E-10	108.6	1.23	1:1 w Drift
	2.14E+05	5.84E-08	2.73E-13	168.1	1.41	1:1 w Drift
ASO 8	2.74E+05	5.02E-05	1.83E-10	141.1	2.24	1:1 w Drift
	2.86E+05	3.85E-05	1.34E-10	123.5	2.31	1:1 w Drift
	8.71E+05	7.35E-04	8.44E-10	102.4	2.22	1:1 w Drift
ASO 9	8.78E+08	2.47E+01	2.81E-08	89.3	1.05	1:1 w Drift
	1.76E+08	4.49E+00	2.55E-08	77.9	1.43	1:1 w Drift
	3.30E+08	1.03E+01	3.12E-08	94.1	1.53	1:1 w Drift
ASO 10	1.31E+06	2.62E-04	2.00E-10	122.3	2.6	1:1 w Drift
	1.38E+06	2.31E-04	1.68E-10	108	2.52	1:1 w Drift
	3.04E+06	9.63E-04	3.16E-10	129.9	3.39	1:1 w Drift
ASO 11	1.01E+06	7.85E-03	7.75E-09	80.1	0.548	1:1 w Drift
	1.11E+06	7.64E-03	6.89E-09	69.3	0.983	1:1 w Drift
	8.66E+05	7.89E-03	9.11E-09	82.2	1.11	1:1 w Drift
ASO 13	2.37E+10	2.22E+03	9.36E-08	7.2	0.0688	1:1 w Drift
	4.23E+07	5.06E+00	1.20E-07	8.6	0.596	1:1 w Drift
	4.67E+07	2.42E+00	5.18E-08	5.4	0.579	1:1 w Drift

Table S7. Replicates SPR assay at 37°C.

Sample	ka (1/Ms)	kd (1/s)	KD (M)	Rmax (RU)	Chi <sup>2</sup> (RU <sup>2</sup> )	Model
ASO 1	1.34E+06	1.60E-04	1.19E-10	187.7	2.31	1:1 w drift
	1.65E+06	2.69E-04	1.63E-10	189	4.16	1:1 w drift
	2.43E+06	3.25E-04	1.34E-10	164.4	2.93	1:1 w drift
ASO 2	4.34E+05	1.68E-03	3.88E-09	74.6	0.283	1:1 w drift
	4.51E+05	1.96E-03	4.35E-09	72.7	0.923	1:1 w drift
	5.47E+05	2.38E-03	4.36E-09	52.2	0.782	1:1 w drift
ASO 3	6.45E+05	3.53E-04	5.48E-10	109.8	0.762	1:1 w drift
	8.28E+05	6.03E-04	7.28E-10	102.7	1.59	1:1 w drift
	1.16E+06	8.02E-04	6.93E-10	80.1	1.25	1:1 w drift
ASO 8	7.02E+05	5.37E-03	7.65E-09	74	0.369	1:1 w drift
	7.97E+05	6.51E-03	8.16E-09	75	1.02	1:1 w drift
	8.91E+05	7.29E-03	8.18E-09	56.6	0.803	1:1 w drift
ASO 9	NA	NA	NA	NA	NA	1:1 w drift
	NA	NA	NA	NA	NA	1:1 w drift
	NA	NA	NA	NA	NA	1:1 w drift
ASO 10	2.16E+06	5.27E-03	2.44E-09	95.4	0.604	1:1 w drift
	2.23E+06	5.71E-03	2.57E-09	96.3	1.05	1:1 w drift
	2.53E+06	6.29E-03	2.48E-09	83.5	0.779	1:1 w drift
ASO 11	NA	NA	NA	NA	NA	1:1 w drift
	NA	NA	NA	NA	NA	1:1 w drift
	NA	NA	NA	NA	NA	1:1 w drift
ASO 13	NA	NA	NA	NA	NA	1:1 w drift
	NA	NA	NA	NA	NA	1:1 w drift
	NA	NA	NA	NA	NA	1:1 w drift