

## Helix-based screening with structure prediction using artificial intelligence has potential for the rapid development of peptide inhibitors targeting class I viral fusion

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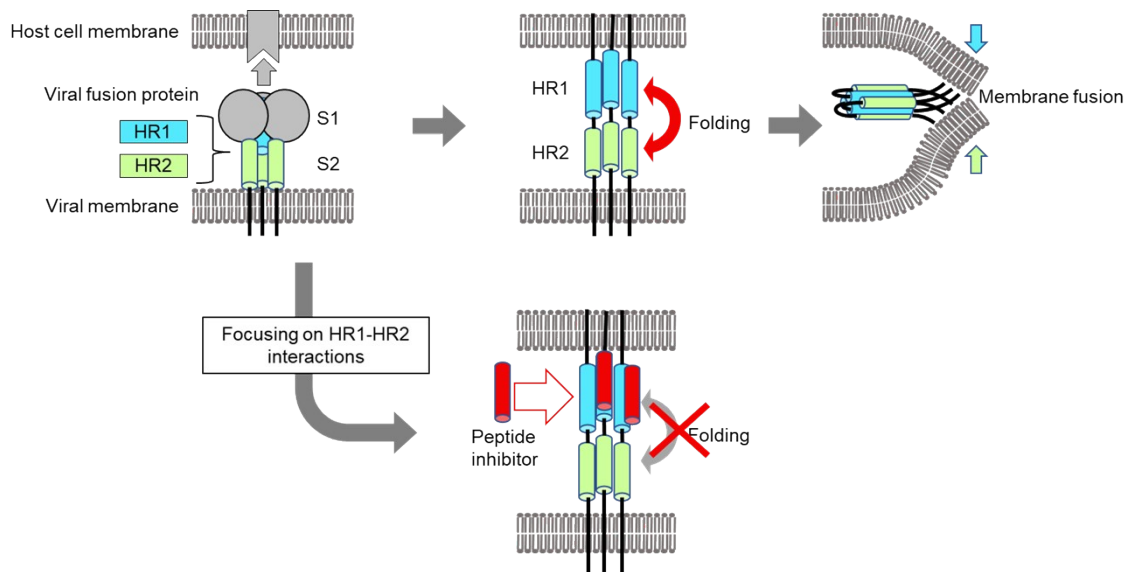
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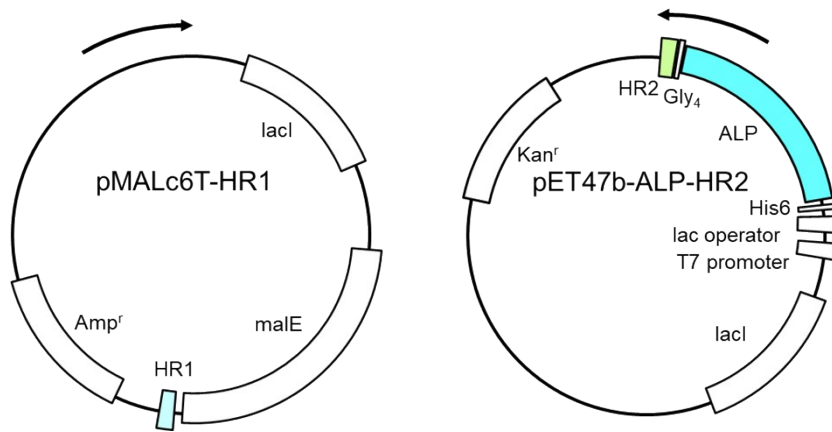
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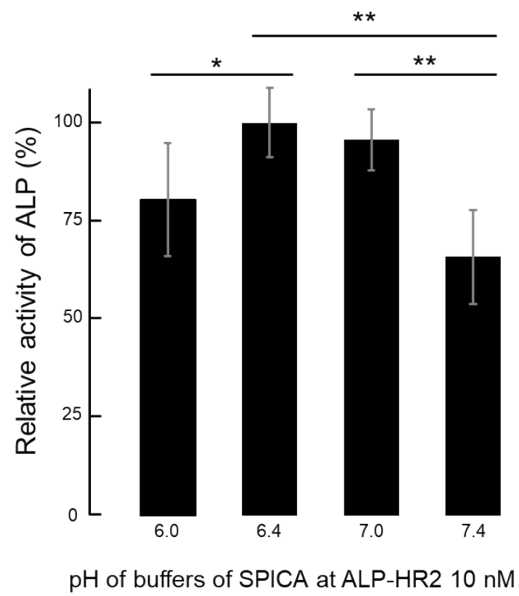
**Fig. S1. Mechanism of virus-cell fusion and the inhibition by peptides.**

The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) uses two spike (S) proteins to enter the host cells. The S1 protein binds to receptors of the host cells, such as angiotensin-converting enzyme 2. This binding activates the S2 protein and leads to penetration of the N-terminal fusion domain into the host cell membrane. After anchoring the cell and viral membranes with the S2 protein, the heptad repeat 1 (HR1) and heptad repeat 2 (HR2) interact with each other and form a six helix bundle (6HB) through intramolecular folding. The 6HB formation induces the fusion of the membranes. Agents preventing the intramolecular folding of the HR1-HR2 complex can inhibit viral entry.



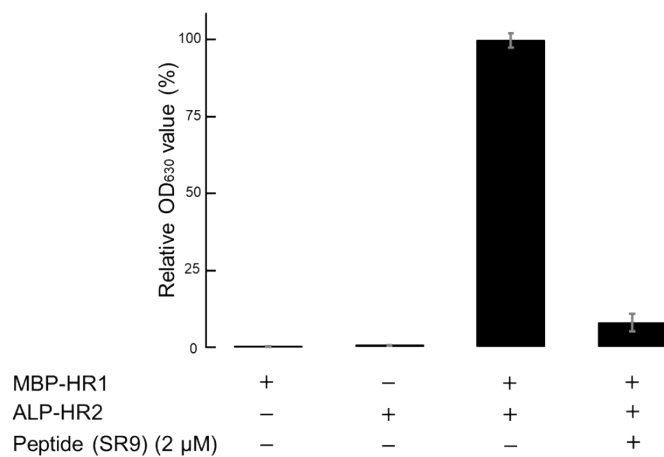
**Fig. S2. Construction of plasmids.**

The amplified DNA fragment coding HR1 was inserted into the pMAL-c6T vector to generate maltose binding protein (MBP)-fused HR1 (MBP-HR1). The DNA fragment of the alkaline phosphatase (ALP) coding region (amino acids 22–471) from *Escherichia coli*, JM109 genome cDNA for a tetra-glycine (Gly<sub>4</sub>) linker, and cDNA for HR2 were sequentially inserted downstream of the original a hexa-histidine tag (His6) coding frame of a pET47b vector. The different lengths of DNA fragments derived from the HR2 of SARS-CoV-2 were inserted into the pET47b-ALP vector. ALP-HR2 was designed to be expressed with a His6 fused to the N-terminal amino acids of ALP.



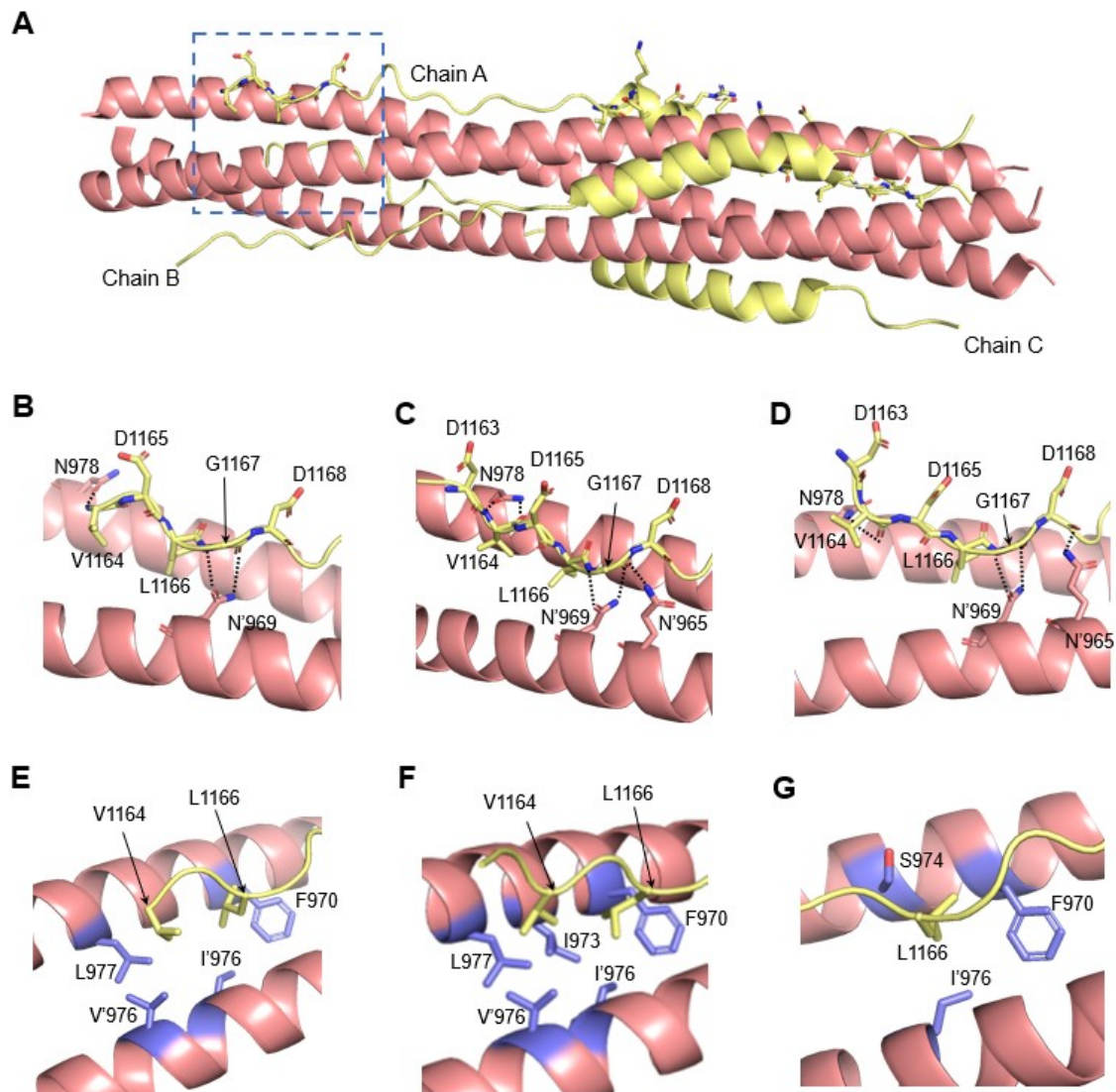
**Fig. S3. Optimization of the pH value for the interaction of ALP-HR2 with MBP-HR1 in SPICA.**

The pH value of the SPICA buffer (to match the optimal buffer range for each buffer, 100 mM MES at pH 6.0 and 6.4, 100 mM HEPES at pH 7.0, PBS at pH 7.4, with 100 mM NaCl and 2 mM MgCl<sub>2</sub>, were used) influenced the activity of ALP. The activity of ALP at different pH values was calculated relative to the ALP activity at pH 6.4. Single asterisk means p value < 0.05, double asterisks mean p value < 0.01.



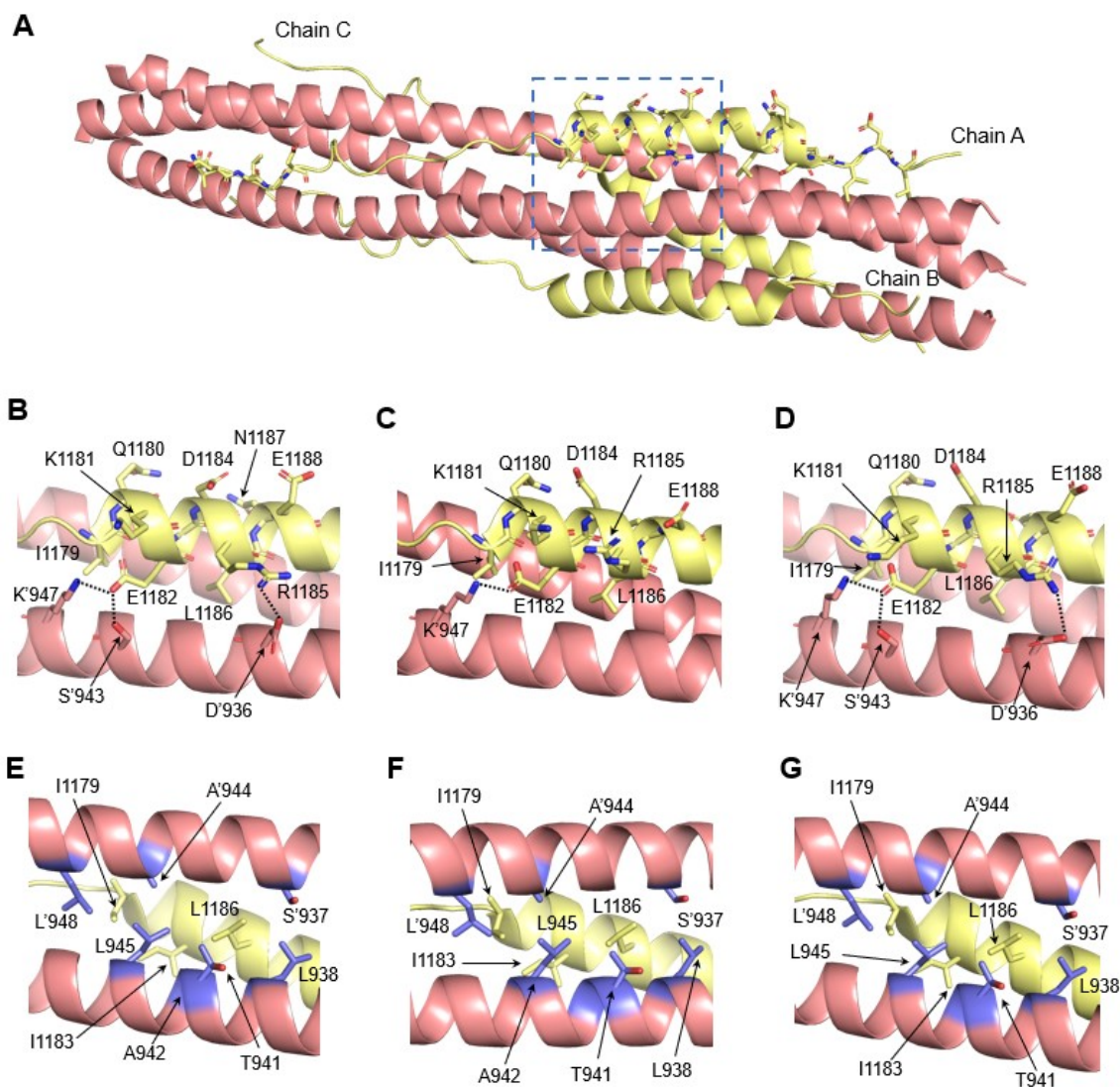
**Fig. S4. SPICA with control well.**

Little ALP activity was observed as indicated by the absorbance at 630 nm (OD<sub>630</sub>) with MBP-HR1 or ALP-HR2 alone, but ALP activity was detected with MBP-HR1 and ALP-HR2. The activity of ALP was suppressed with SR9.



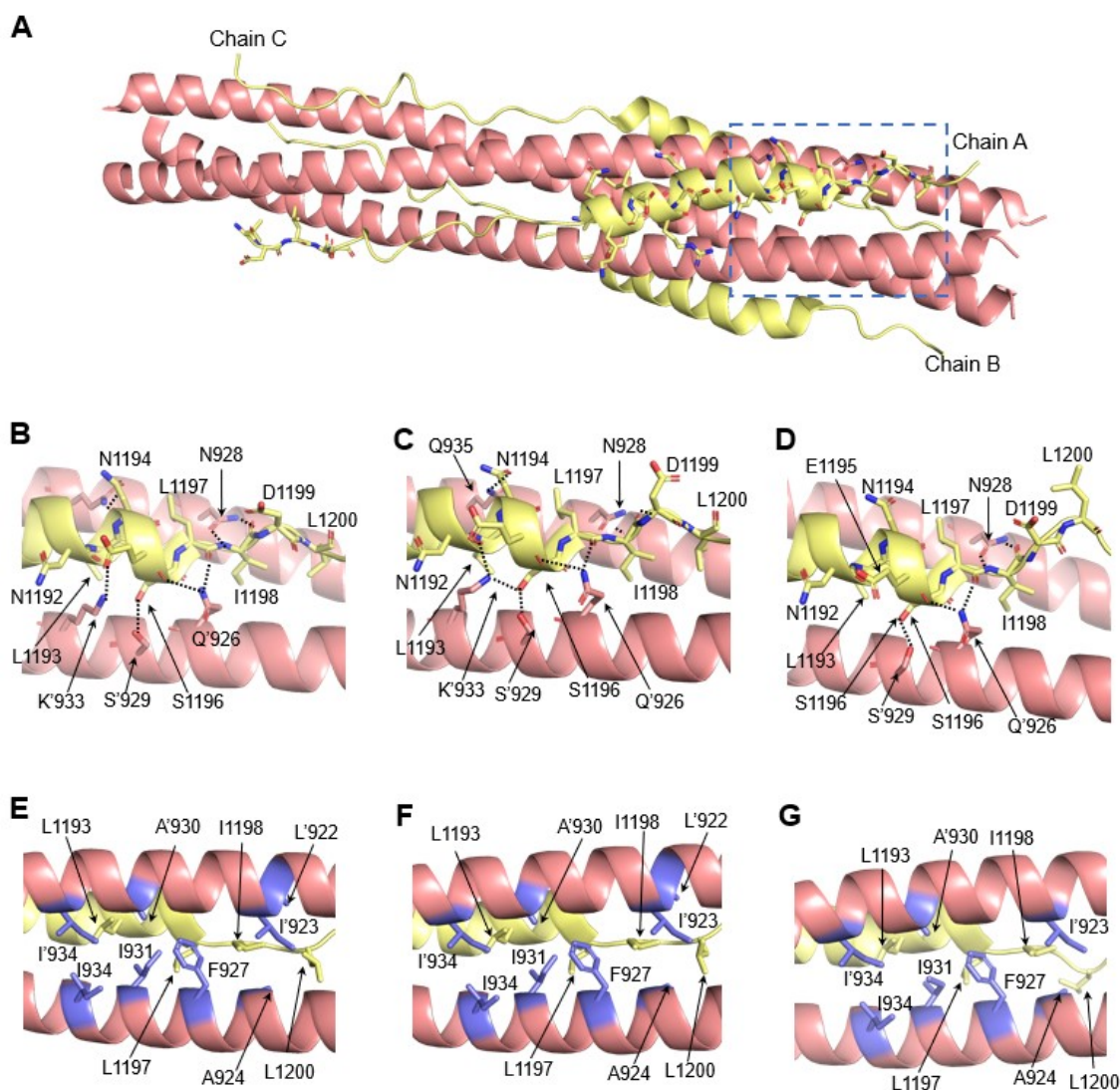
**Fig. S5. Binding mode between DVDLGD and a HR1 trimer in previous reported crystal structure.**

(A) showed a crystal structure of the six-helix bundle formed by a HR1 trimer and three HR2s (Chain A, B and C). HR1 and HR2 in 6HB (6LXT) are shown in ribbon model, and colored pink and light yellow, respectively. DVDLGD was surrounded by a dotted line. (B) to (D) showed hydrogen bonds (H-Bonds) formed between DVDLGD and HR1 trimer. DVDLGD was displayed in stick model. Amino acids of HR1 forming H-bonds with DVDLGD are shown in stick model. Black dotted lines mean H-bonds. (E) to (G) exhibited amino acids forming hydrophobic interactions. Amino acids forming hydrophobic interactions with V1164 and/or L1166 of HR2 were displayed in stick model and colored light blue. Additionally, (B) and (E) show interactions around chain A in the crystal structure. (C) and (F) are chain B. (D) and (G) are chain C.



**Fig. S6. Binding mode between IQKEIDRLNE and a HR1 trimer in previous reported crystal structure.**

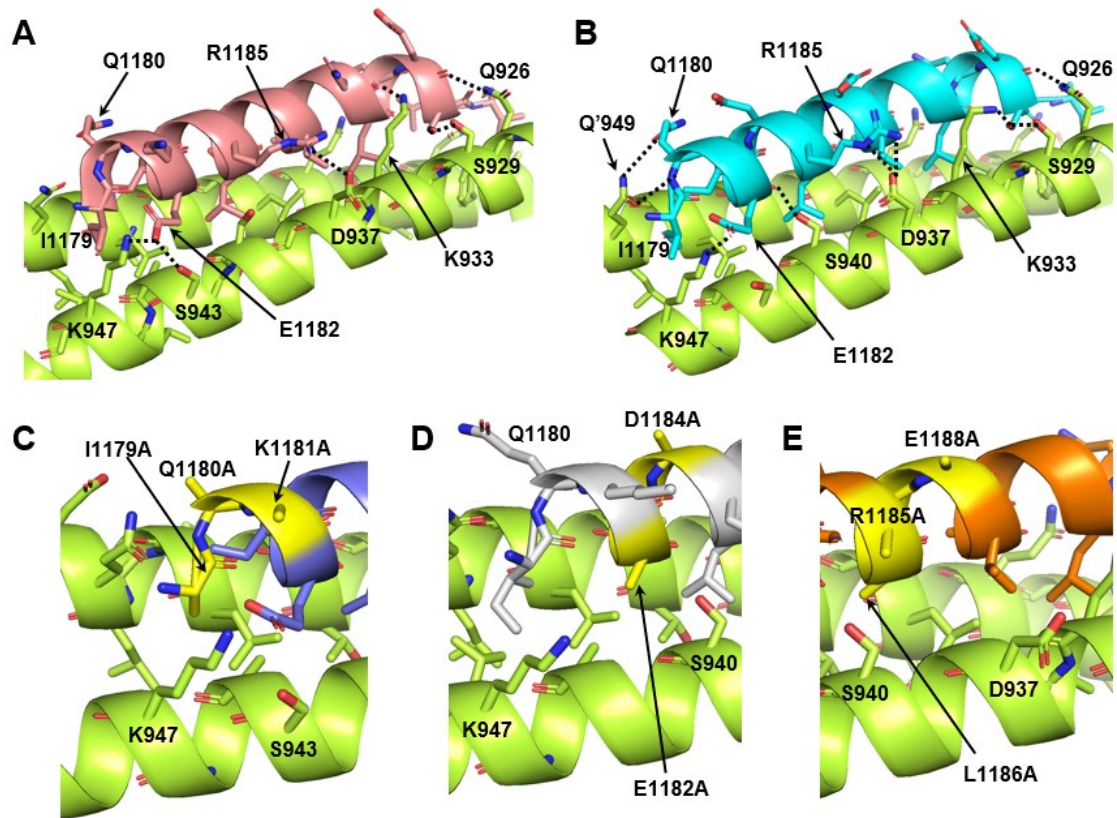
(A) showed a crystal structure of the six-helix bundle formed by a HR1 trimer and three HR2s (Chain A, B and C). HR1 and HR2 in 6HB (6LXT) are shown in ribbon model, and colored pink and light yellow, respectively. IQKEIDRLNE was surrounded by a dotted line. (B) to (D) showed hydrogen bonds (H-Bonds) formed between IQKEIDRLNE and HR1 trimer. IQKEIDRLNE is displayed in stick model. Amino acids of HR1 forming H-bonds with IQKEIDRLNE are shown in stick model. Black dotted lines mean H-bonds. (E) to (G) exhibited amino acids forming hydrophobic interactions. Amino acids forming hydrophobic interactions with I1179, I1183 and/or L1186 of HR2 were displayed in stick model and colored light blue. Additionally, (B) and (E) show interactions around chain A in the crystal structure. (C) and (F) are chain B. (D) and (G) are chain C.



**Fig. S7. Binding mode between NLNESLIDL and a HR1 trimer in previous reported crystal structure.**

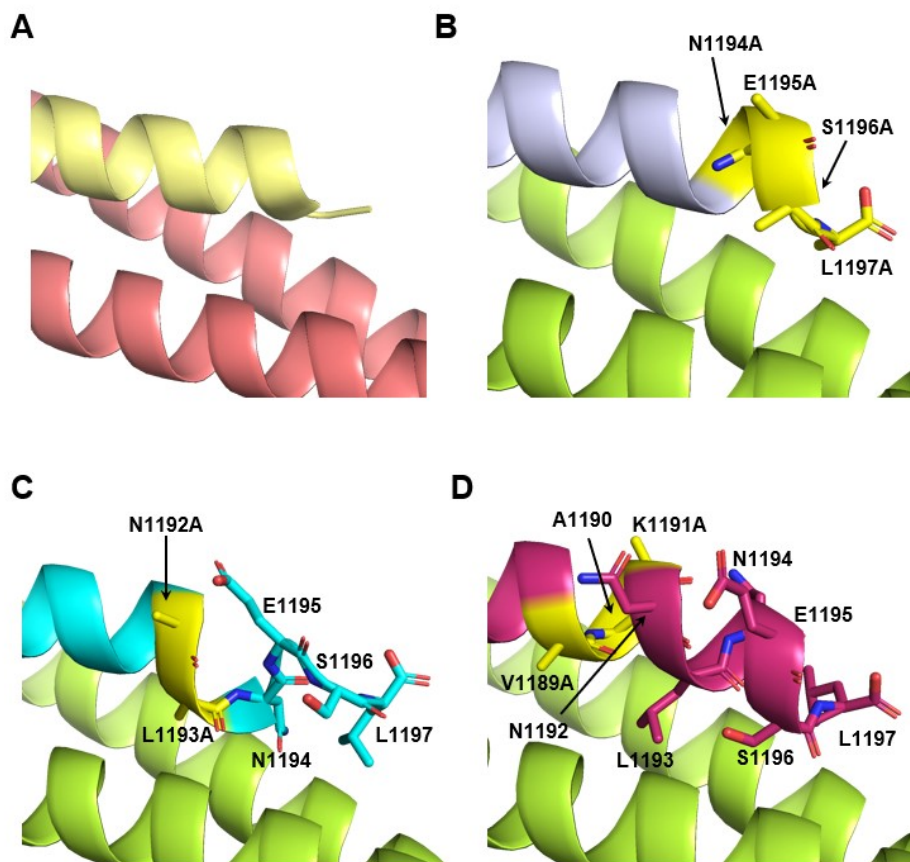
(A) showed a crystal structure of the six-helix bundle formed by a HR1 trimer and three HR2s (Chain A, B and C). HR1 and HR2 in 6HB (6LXT) are shown in ribbon model, and colored pink and light yellow, respectively. NLNESLIDL was surrounded by a dotted line. (B) to (D) showed hydrogen bonds (H-Bonds) formed between NLNESLIDL and HR1 trimer. NLNESLIDL was displayed in stick model. Amino acids of HR1 forming H-bonds with NLNRSLIDL are shown in stick model. Black dotted lines mean H-bonds. (E) to (G) exhibited amino acids forming hydrophobic interactions. Amino acids of HR1 forming hydrophobic interactions with L1193, L1197, I1198 and/or L1200 of HR2 were displayed in stick model and colored light blue. Additionally, (B) and (E) show interactions around chain A in the crystal structure. (C) and (F) are chain B. (D) and (G) are chain C.





**Fig. S8. Predicted 3D structures of Vir<sup>SC2</sup>4 and derivatives.**

**(A)** Crystal structure (PDB code: 6LXT) of the 6HB from SARS-CoV-2. Light yellow and light pink indicate the  $\alpha$ -helices in the HR2 and HR1 regions, respectively. The dotted lines indicate hydrogen bonds (H-bonds). **(B)** Predicted 3D structure of the 6HB formed by Vir<sup>SC2</sup>4 and HR1. Cyan represents the  $\alpha$ -helix in Vir<sup>SC2</sup>4. Light green represents HR1. The dotted lines indicate predicted H-bonds. **(C)** to **(E)** N-terminal region of the predicted structures formed by the Vir<sup>SC2</sup>4 derivatives and HR1. Blue, white, and orange represents Vir<sup>SC2</sup>4<sub>I<sub>Q</sub>K</sub>, Vir<sup>SC2</sup>4<sub>E<sub>ID</sub></sub>, and Vir<sup>SC2</sup>4<sub>R<sub>LNE</sub></sub>, respectively. Light green represents HR1. Substitutions with alanine in Vir<sup>SC2</sup>4 are indicated in yellow. The 3D structures suggested that alanine substitutions in Vir<sup>SC2</sup>4 resulted in the loss of H-bonds and/or hydrophobic interactions.



**Fig. S9. Predicted 3D structures of Vir<sup>SC28</sup> and derivatives.**

**(A)** Light yellow and light pink represents C-terminal HR2 region of the  $\alpha$ -helix and HR1 derived from a previously reported crystal structure, respectively (PDB code: 6LXT). **(B)** to **(D)** C-terminal region in the predicted structures of the complexes of Vir<sup>SC28</sup> derivatives with HR1. **(B)** Vir<sup>SC28</sup><sub>NESL</sub> (light blue). **(C)** Vir<sup>SC28</sup><sub>NL</sub> (cyan). **(D)** Vir<sup>SC28</sup><sub>VAK</sub> (wine red). HR1 is shown in light green. Alanine substitutions in Vir<sup>SC28</sup> are indicated in yellow.

**Table S1. Peptide inhibitors reported in previous studies.**

Peptide	Amino-acid sequence	Reference
EK1	SLDQINVTFDLEYEMKKLEEAIKKLEESYIDLKEL	1
IPB1 (SR9)	ISGINASVVNIQKEIDRLNEVAKNLNESLIDLQEL	2
#2	DISGINASVVNIQKEIDRLNEVAKNLNESLIDLQEL	3
IPB19	SVVNIQKEIDRLNEVAKNLNESLIDLQELGKYEQYIK	4
EKL1	NVTFLDLEYEMKKLEEAIKKLEESYIDLKELGTYEY	5
AntiSCV2P1	AAVEQIQKEIERLQEAAKNLLESK	6
AntiSCV2P7	ASVRNIQEEIKRLNEVAKKLAESLI	6
longHR2_42	PDVDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQEL	7
42G	PDVDLGGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQEL	7
HR2-P40	VDLGDISGINASVVNIQKEIDRLNEVAKNLNESLIDLQEL	8
3a	ISGINASVVNIQEEIKRLNEVAKKLNESLIDLQE	9

**Table S2. Interactions between amino acids in HR1 and 2 measured by protein crystallography.**

Structure of HR2	Amino acids of HR2 (chain A)	Amino acids of HR1 (chain A)	Amino acids of HR1 (chain B)	Binding	Consistency with the results of SPICA	
Disordered	Asp1163	Asn978		Hydrogen	+	
	Val1164	Asn978		Hydrogen**	+	
	Leu1166		Leu977		Hydrophobic	+
				Val1976		Hydrophobic
	Gly1167		Phe970	Ile973	Hydrophobic	+
						Hydrophobic
	Ile1169		Leu966	Asn969	Hydrogen**	+
					Leu962*	
	Ile1172		Val963		Hydrophobic	-
						Hydrophobic
	Asn1173		Asn960	Leu962	Hydrophobic	-
					Ala958	
	Ala1174		Asn960		Hydrogen	-
						Hydrogen**
	Ser1175		Leu959		Hydrophobic	-
						Hydrophobic
	Val1176		Ala956	Asn955	Hydrogen**	-
					Gln954	
	Val1177		Asn953		Hydrophobic	-
						Hydrogen**
		Val952		Hydrophobic	-	
				Val951		Hydrophobic
		Gln949		Hydrogen**	-	
			Leu948	Hydrophobic	-	
Helix	Ile1179	Leu945	Leu948	Hydrophobic	+	
				Ala944	Hydrophobic	+
	Glu1182			Lys947	Hydrogen	+
					Ser943	
	Ile1183	Leu945	Ala942		Hydrophobic	+
						Hydrophobic
	Arg1185			Asp936	Hydrogen	+
						Hydrophobic
	Leu1186	Leu938*	Ala942		Hydrophobic	+
					Thr941	
	Val1189	Leu938		Ser937	Hydrophobic	+
						Hydrophobic
	Ala1190	Leu938			Hydrophobic	-
					Gln935	
	Asn1192			Lys933	Hydrogen**	+
						Hydrophobic
	Leu1193	Leu938	Ile934		Hydrophobic	+
				Ile931		Hydrophobic
Asn1194	Gln935		Ala930	Hydrophobic	+	
					Hydrogen	+
Glu1195			Gln926	Hydrogen**	+	
					Ser929	Hydrogen
Ser1196				Hydrogen	+	
					Hydrophobic	+
Leu1197	Ile931			Hydrophobic	+	
					Gln926	Hydrogen**
Disordered	Ile1198	Asn928		Hydrogen**	+	
				Phe927	Hydrophobic	+
	Leu1200	Ala924			Hydrophobic	+
						Ile923
	Gln1201			Leu922	Hydrophobic	+
						Ile923
			Asn919	Hydrogen**	-	
		Tyr917*		Hydrogen**	-	

The crystal structure from the Protein Data Bank data, 6LXT<sup>10</sup> (Asp1163 from 6XRA<sup>11</sup>), was analyzed and compared with the results of SPICA. \* indicates hydrogen bonds or hydrophobic interactions observed only on some chains; \*\* indicates hydrogen bonds to peptide bonds. The gray columns show hydrophobic interactions. The structure of Leu1203 to Pro1213 was not able to be observed in the crystal structure and cryo-electron microscopy (cryo-EM) structure.<sup>11</sup>

**Table S3.** Nucleotide sequence of the primers for RT-PCR, the sequence reaction, and cloning.

	Primer name	Sequences	
RT-PCR	RT-F	5'- CTTGGTGATATTGCTGCTAGAGA -3'	
	RT-R	5'- CGTCTTCATCAAATTTGCAGCAGGA -3'	
For Sequencing	HR2-F1	5'- CACGATGCGTCCGGCGTAGAGGATCGA -3'	
	HR2-F2	5'- GGA CTACGTCACCGACTCGGCTGCA -3'	
	HR2-F3	5'- AGAGGATTCACAAGAACATACCGGCA -3'	
	HR2-R1	5'- GCGCGTCCCATTGCGCAATCCGGATA -3'	
	HR2-R2	5'- CTTTGGTATTTAGCGCCTGGGTGA -3'	
	T7 pro	5'- TAATACGACTCACTATAGGG -3'	
	T7 term	5'- ATGCTAGTTATTGCTCAGCGG-3'	
	HR1-F1	5'- GACAATTAATCATCGGCTCGTATA -3'	
	HR1-F2	5'- CCCGCCAAAAACCTGGGAAGAGA -3'	
	HR1-R1	5'- TCCGCTCCCGCGGATTTGTCCTA -3'	
	HR1-R2	5'- TCGTAAGACTTCAGCGCTACGGCA -3'	
	For Cloning	ALP-F	5'- GACGGATCCCGGACACCAAGAAATGCCTGTTCTGGAA -3'
		ALP-R	5'- CCCAAGCTTACCACCACCACCTTTTCAGCCCCAGAGCGGCTTCATGGTG -3'
HR1-F		5'- CCCGTCGACACACAGAATGTTCTCTATGAGAACCA -3'	
HR1-R		5'- CCCGGATCCTTAAAGACGTGAAAGGATATCATTAAAA -3'	
HR2-F		5'- CCCAAGCTTIGATGTTGATTAGGTGACATCTCTGGCA -3'	
HR2-F-Vir <sup>SC21</sup>		5'- CCCAAGCTTATCTCTGGCATTAAATGCTTCAGTTGTAAA -3'	
HR2-F-Vir <sup>SC22</sup>		5'- CCCAAGCTTAATGCTTCAGTTGTAAACATTCAAA -3'	
HR2-F-Vir <sup>SC23</sup>		5'- CCCAAGCTTIGTTGTAACATTCAAAAAGAAA -3'	
HR2-F-Vir <sup>SC24</sup>		5'- CCCAAGCTTATTCAAAAAGAAATTGACCGCCTCA -3'	
HR2-F-Vir <sup>SC24</sup> <sub>TOK</sub>		5'- CCCAAGCTTIGCAGCAGCAGAAATGACCGCCTCAATGAGGTTGCCA -3'	
HR2-F-Vir <sup>SC24</sup> <sub>EID-1</sub>		5'- GCAGCAGCAGCCTCAATGAGGTTGCCAAGAATTA -3'	
HR2-F-Vir <sup>SC24</sup> <sub>EID-2</sub>		5'- CCCAAGCTTATTCAAAAAGCAGCAGCAGCCTCAA -3'	
HR2-F-Vir <sup>SC24</sup> <sub>RLNE-1</sub>		5'- GCAGCAGCAGCAGTTGCCAAGAATTTAAATGAATCTCTCA -3'	
HR2-F-Vir <sup>SC24</sup> <sub>RLNE-2</sub>		5'- CCCAAGCTTATTCAAAAAGAAATTGACGCGCAGCAGCAGTTG -3'	
HR2-R		5'- CCCCTCGAGTTATGGCCATTTTATATACTGCTCATA -3'	
HR2-R-Vir <sup>SC25</sup>		5'- CCCCTCGAGTTACTGCTCATACTTTCCAAGTTCTTGGAGA -3'	
HR2-R-Vir <sup>SC26</sup>		5'- CCCCTCGAGTTAAAGTTCTTGGAGATCGATGAGAGATTCA -3'	
HR2-R-Vir <sup>SC27</sup>		5'- CCCCTCGAGTTAGAGATCGATGAGAGATTCATTTA -3'	
HR2-R-Vir <sup>SC28</sup>		5'- CCCCTCGAGTTAGAGAGATTCATTTAAATCTTGGCA -3'	
HR2-R-Vir <sup>SC29</sup>		5'- CCCCTCGAGTTATAAATCTTGGCAACCTCATTGA -3'	
HR2-R-Vir <sup>SC28</sup> <sub>BESL-1</sub>		5'- GCGCGCGCGGCTAAATCTTGGCAACCTCATTGA -3'	

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HR2-R-Vir <sup>SC2</sup> <sub>8_NESL</sub> -2	5' - <u>CCCCTCGAGT</u> TAGGCGGCGGGCTAAATTCCTGGCA -3'
HR2-R-Vir <sup>SC2</sup> <sub>8_NL</sub> -1	5' - GAGAGATTCATTTGCTGCCTTGGCAACCTCATTGAGGCGGTCA -3'
HR2-R-Vir <sup>SC2</sup> <sub>8_NL</sub> -2	5' - <u>CCCCTCGAGT</u> TAGAGAGATTCATTTGCTGCCTGG -3'
HR2-R-Vir <sup>SC2</sup> <sub>8_VAK</sub> -1	5' - TAAATTTGCTGCTGCCTCATTGAGGCGGTCAATTTCTTTTGGAA -3'
HR2-R-Vir <sup>SC2</sup> <sub>8_VAK</sub> -2	5' - <u>CCCCTCGAGT</u> TAGAGAGATTCATTTAAATTTGCTGCTGCCTCA -3'

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Underlining indicates restriction enzyme sites. Orange letters indicate a tetra-glycine linker (Gly<sub>4</sub>).

## References

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