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Supporting Information

Hydroxamate-Directed Access to β-Kdo Glycosides

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1. General Materials and Methods

Organic reactions were performed under an atmosphere of argon using anhydrous solvents unless otherwise noted. Thin layer chromatography (TLC) was carried out on Merck silica gel 60 F₂₅₄ coated aluminium sheets. TLC plates were detected with UV-absorption (254nm), and sprayed with 10% sulfuric acid in ethanol (1:9, v/v), followed by heating for visualization. Flash column chromatography was performed on silica gel (200-300 mesh). Proton nuclear magnetic resonance (1 H-NMR) spectra were recorded on a BrukerAvance III 400 spectrometer (at 400MHz). Multiplicities were given as singlet (s), broad signal (br), doublet (d), doublet of doublets (dd), triplet (t), quartet (q) or multiplet (m). Carbon nuclear magnetic resonance (13 C-NMR) spectra were recorded on a BrukerAvance III 400 Spectrometer (at 100 MHz). Non-decoupling 13 C NMR spectra were recorded on a BrukerAvance III 400 Spectrometer (at 100 MHz). The 1 H and 13 C NMR spectra were calibrated against the proton and carbon signals of the solvents as internal references (CDCl₃: $^{\delta}$ H = 7.26 ppm and $^{\delta}$ C = 77.2 ppm). The stereochemistry of the desired C-glycosides was assigned by the heteronuclear coupling constant between 13 C carbon at the C1 and the axial proton at C3 (3 JC1, H3ax) in the non-decoupling 13 C NMR spectra.

2. List of glycosyl acceptors (alcohols/thiol/hydroperoxide) used in the study

Figure S1

3. Preparation of Glycosyl Donors

Kdo-glycosyl acetate:3

Scheme 2 (from main text).

Transformation of compound 1 to 2 was carried out following the literature procedure. To a solution of compound 2 (0.5 g, 1.1 mmol, 1 equiv.) in dry CH₂Cl₂ (10.0 mL) and catalytic amount of DMF was added. The reaction mixture was cooled to 0° C and stirred for five minutes. Then, (COCI)₂ (0.2 mL, 1.5 equiv) was added dropwise to the reaction mixture and stirred at room temperature for 2h. To this solution was added O-benzylhydroxylamine hydrochloride (0.27 g, 1.7 mmol, 1.5 equiv) and Et₃N (0.4 mL, 3.0 equiv) in dry CH₂Cl₂ (10.0 mL) maintaining 0° C and resulting mixture was stirred at room temperature for 2h. Upon completion of the reaction (TLC monitored), it was quenched with water. Organic layer was separated, and aqueous layer was extracted two times with CH2Cl2 (5 mL x 2). The combined organic layer was again washed with saturated aqueous NaHCO₃ solution (5 mL), collected, dried over anhydrous Na₂SO₄ and finally concentrated in vacuo. The crude was purified by silica gel column chromatography (40% ethyl acetate in hexane as eluent) to afford the final product **3** (0.376 g, 61% yield); ¹**H NMR** (400 MHz, CDCl₃) δ 8.92 (s, 1H), 7.51 (d, J = 6.9 Hz, 2H), 7.42 - 7.39 (m, 3H), 5.35 - 5.29 (m, 2H), 5.03 - 4.97 (m, 2H), 4.86 (d, J = 11.3 Hz, 1H), 4.34 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (d, J = 9.9 Hz, 1H), 3.93 - 3.88 (m, J = 11.3 Hz, 1H), 4.34 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (d, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (d, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (d, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (d, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (d, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.10 (dd, J = 9.9 Hz, 1H), 4.94 (dd, J = 12.4, 1.5 Hz, 1H), 4.941H), 2.33 (dd, J = 13.3, 4.7 Hz, 1H), 2.15 (s, 3H), 2.05 (d, J = 2.8 Hz, 6H), 1.99 (s, 6H), 1.95 (m, 1H); ${}^{13}C\{{}^{1}H\}$ NMR (100 MHz, CDCl₃) δ 170.4, 170.3, 170.1, 169.7, 167.9, 164.3, 134.8, 130.1, 129.2, 128.8, 97.8, 78.5, 69.8, 67.1, 65.8, 63.9, 62.5, 32.0, 20.9(2), 20.8, 20.7(2); $^{3}J_{C1,H3ax}$:12 Hz; **HRMS(ESI-TOF)** m/z: [M + Na]⁺ C₂₅H₃₁NNaO₁₃ calcd. 576.1693, found, 576.1681.

^{1.} M. Mazur, B. Barycza, H. Andriamboavonjy, S. Lavoie, M. T. Kenfack, A. Laroussarie, Y. Bleriot and C. Gauthier, *J. Org. Chem.*, 2016,**81**,10585 - 10599.

Preparation of Kdo-glycosyl acetate 24

To a solution of compound 2 (0.060 g, 0.13 mmol, 1.0 equiv) in dry CH₂Cl₂ (3.0 mL) and catalytic amount of DMF was added. The reaction mixture was cooled to 0°C and stirred for five minutes. Then, (COCI)₂ (20 µL, 1.5 equiv) was added dropwise to the reaction mixture and stirred at room temperature for 2h. To this solution was added benzylamine (0.022 g. 0.20 mmol, 1.5 equiv) in dry CH₂Cl₂ (3.0 mL) maintaining 0° C and resulting mixture was stirred at room temperature for 2h. Upon completion of the reaction (TLC monitored), it was quenched with water. Organic layer was separated, and aqueous layer was extracted two times with CH₂Cl₂ (5 mL x 2). The combined organic layer was again washed with saturated aqueous NaHCO₃ solution (5 mL), collected, dried over anhydrous Na₂SO₄ and finally concentrated in vacuo. The crude was purified by silica gel column chromatography (30% ethyl acetate in hexane as eluent) to afford the final product 24 (0.040 g, 56% yield); The ${}^{3}J_{C1,H3ax}$: 6Hz indicated that α -anomer was formed predominantly in the transformation. ¹**H NMR** (400 MHz, CDCl₃) δ 7.37 - 7.29 (m, 5H), 6.85 (t, J = 5.6 Hz, 1H), 5.40 - 5.35 (m, 2H), 5.14 (d, J = 10.8 Hz, 1H), 4.62-4.56 (m, 1H), 4.48-4.43 (m, 2H), 4.15 (d, J = 9.9 Hz, 1H), 3.99 (dd, J = 12.4, 4.0 Hz, 1H), 2.37 (dd, J = 13.2, 4.8 Hz, 1H), 2.14 (s, 3H), 2.07 (s, 3H), 2.03 (s, 3H), 2.00 (s, 6H), 1.97 (m, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.4, 170.2, 170.0, 169.7, 167.8, 167.1, 137.6, 128.8, 127.6, 127.4, 97.6, 69.8, 67.1, 66.0, 64.0, 62.3, 43.3, 32.0, 20.8, 20.7(2), 20.6; **HRMS(ESI-TOF)** m/z: $[M + Na]^+ C_{25}H_{31}NNaO_{12}$ calcd. 560.1744, found, 560.1757.

4. General procedure for glycosylation

Conditions A: Kdo donor (1.0 equiv), acceptor (2.0 equiv) and freshly activated 4Å MS were taken in dry CH₂Cl₂ (0.1 M) and stirred under argon at room temperature for 30 min. The reaction mixture was then cooled to 0° C followed by addition of TMSOTf (1.0 equiv). Stirring was continued at room temperature until consumption of starting materials were observed (*ca.* 12h). The crude was then filtered through a pad of celite. The filtrate was washed with brine, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by silica gel column chromatography to give Kdo glycoside.

Conditions B: Kdo donor (1.0 equiv), acceptor (2.0 equiv) and freshly activated 4Å MS were taken in dry CH₂Cl₂ (0.1 M) and stirred under argon at room temperature for 30 min. The reaction mixture was then cooled to 0° C followed by addition of BF₃.OEt₂ (1.0 equiv). Stirring was continued at room temperature until consumption of starting materials were observed (*ca.* 12h). The crude was then filtered through a pad of celite. The filtrate was washed with brine, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure. The residue was purified by silica gel column chromatography to give Kdo glycoside.

5. Experimental details and characterization of glycosides

Synthesis of glycoside 5

Title compound was prepared according to the general procedure **A/B**. For procedure B, Kdo donor **3** (0.027 g, 0.05 mmol, 1.0 equiv), allyl alcohol (0.006 g, 0.10 mmol, 2.0 equiv) were used in presence of TMSOTf (8.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 64% (0.017 g) yield and isolated as mixture of anomer (β :α 9:1); R_f 0.4 (EtOAc: Hexane 4:6); $[\alpha]_D^{25} = -56.6$ (c = 0.120, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 9.55(s,0.1H), 8.94 (s, 0.91H), 7.43 -7.39 (m, 5H), 5.89 - 5.80 (m, 1H), 5.33 - 5.25 (m, 3H), 5.19 (d, J = 10.4 Hz, 1H), 5.05 (d, J = 10.1 Hz, 1H), 4.98 - 4.94 (m, 2H), 4.45 (dd, J = 12.3, 1.4 Hz, 1H), 4.08 (d, J = 9.8 Hz, 1H), 3.92 - 3.86 (m, 2H), 3.76 (dd, J = 12.0, 5.7 Hz, 1H), 2.28 (dd, J = 12.9, 4.7 Hz, 1H), 2.06 (s, 3H), 2.04 (s, 3H), 1.99 (s, 3H), 1.96 (s, 3H), 1.90 (t, J = 12.5 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.7, 170.3, 170.1, 169.9, 164.4, 134.7, 133.1, 129.7, 129.3, 128.8, 117.7, 99.9, 78.5, 68.7, 67.6, 66.2, 64.8, 64.5, 62.4, 32.3, 20.9(2), 20.8, 20.7; ³ $J_{C1,H3ax} = 11$ Hz; HRMS(ESI-TOF) m/z: [M + Na]+ C₂₆H₃₃NNaO₁₂ calcd. 574.1900, found, 574.1909; [The β :α ratio was determined by the NH protons at 9.55 -8.94 ppm].

Note: The anomeric configuration of glycosides was determined by the measurement of coupling constant ${}^3J_{C1,H3ax}$ of proton decoupled ${}^{13}C$ NMR experiment (see the figure below).

Synthesis of glycoside7

Preparation of the title compound via general procedure **A** is provided here. It was prepared from Kdo donor **3** (0.025 g, 0.05 mmol, 1.0 equiv), 2-chloroethanol (0.008 g, 0.09 mmol, 2.0 equiv) in presence of BF₃.OEt₂ (5 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 50% (0.013 g) yield (β : α > 20:1); R_f 0.4 (EtOAc : Hexane 4:6); $[\alpha]_D^{25} = -5.0(c = 0.14, CHCl_3)$; ¹**H NMR** (400 MHz, CDCl₃) δ9.00 (s, 1H), 7.42 - 7.39 (m, 5H), 5.35 (s, 1H), 5.32 - 5.27 (m, 1H), 5.05 (dd, J = 10.1, 3.1 Hz, 1H), 4.98 (s, 2H), 4.48 (dd, J = 12.2, 1.3 Hz, 1H), 4.24 (d, J = 9.9 Hz, 1H), 3.89 (dd, J = 12.4, 4.5 Hz, 1H), 3.69 - 3.56 (m, 3H), 3.45 - 3.38 (m, 1H), 2.26 (dd, J = 13.1, 4.8 Hz, 1H), 2.10 (s, 3H), 2.05 (s, 3H), 2.01 (s, 3H), 1.97 (s, 3H), 1.89 (t, J = 12.7 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.9, 170.3, 170.0, 164.0, 134.6, 129.7, 129.3, 128.9, 99.6, 78.6, 68.8, 67.7, 66.0, 64.4, 63.9, 62.6, 42.5, 32.2, 21.0, 20.9, 20.8(2); ³ $J_{C1,H3ax} = 11Hz$; **HRMS (ESI-TOF)** m/z: [M + Na]⁺ C₂₅H₃₂CINNaO₁₂ calcd. 596.1511, found, 596.1525. [The β: α ratio was determined by the NH protons at ~ 9.0 ppm].

Synthesis of glycoside 8

Preparation of the title compound via general procedure **A** is provided here. It was prepared from Kdo donor **3** (0.025 g, 0.05 mmol, 1.0 equiv), propargyl alcohol (0.005 mg, 0.09 mmol, 2.0 equiv) in presence of BF₃.OEt₂ (5 µL) as promoter. Crude was purified by flash silica gel column chromatography (6:4 Hexane/EtOAc) to give the desired products as a colourless oil in 52% (0.013 g) yield (β : α > 20:1); R_f 0.4 (EtOAc : Hexane 4:6); [α] α = -77.0(α =0.1, CHCl₃); ¹**H NMR** (400 MHz, CDCl₃) α 9.01 (s, 1H), 7.43 - 7.39 (m, 5H), 5.35 - 5.28 (m, 2H), 5.08 (dd, α = 9.3, 4.8 Hz, 1H), 4.98 (s, 2H), 4.50 (d, α = 12.3 Hz, 1H), 4.20 (d, α = 9.8 Hz, 1H), 4.07 (dd, α = 14.9, 1.7 Hz, 1H), 3.92 - 3.84 (m, 2H), 2.48 (s, 1H), 2.26 (dd, α = 13.0, 4.7

Hz, 1H), 2.11 (s, 3H), 2.05 (s, 3H), 2.00 (s, 3H), 1.97 (s, 3H), 1.89 (t, J = 12.6 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ170.9, 170.3, 170.0(2), 163.7, 134.6, 129.7, 129.3, 128.9, 100.2, 78.8, 78.6, 75.4, 69.2, 67.8, 66.0, 64.4, 62.9, 52.2, 32.2, 20.9(2), 20.8(2); ${}^{3}J_{\text{C1,H3ax}} = 12$ Hz; HRMS(ESI-TOF) m/z: [M + Na]⁺ C₂₆H₃₁NNaO₁₂ calcd. 572.1744, found, 572.1761. [The β:α ratio was determined by the NH protons at ~ 9.0 ppm].

Synthesis of glycoside 9

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.025 g, 0.05 mmol, 1.0 equiv), 5-hexene-1-ol (0.01 g, 0.09 mmol, 2.0equiv) in presence of TMSOTf (8.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as colourless oil in 49% (0.013 g) yield (β : α > 20:1); R_f 0.25 (EtOAc : Hexane 2:8); $[\alpha]_D^{25} = -33.12$ (c = 0.16, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 8.93 (s, 1H), 7.41 - 7.37 (m, 5H), 5.84 - 5.73 (m, 1H), 5.35-5.33 (m, 1H), 5.28 - 5.24 (m, 1H), 5.05 - 4.95 (m, 5H), 4.46 (dd, J = 12.3, 1.3 Hz, 1H), 4.03 (d, J = 9.9 Hz, 1H), 3.88 (dd, J = 12.4, 4.1 Hz, 1H), 3.33 - 3.20 (m, 2H), 2.24 (dd, J = 12.9, 4.7 Hz, 1H), 2.07 (s, 3H), 2.04-2.02 (m, 4H), 1.99 (s, 3H), 1.97 (s, 3H), 1.91 - 1.82 (m, 2H), 1.55 (dd, J = 13.8, 6.8 Hz, 2H), 1.43 (dd, J = 14.5, 7.3 Hz, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.6, 170.3, 170.1, 169.9, 164.6, 138.4, 134.7, 129.6, 129.3, 128.8, 115.1, 99.9, 78.5, 68.5, 67.7, 66.3, 64.5, 63.9, 62.4, 33.6, 32.3, 29.0, 25.6, 20.9(2), 20.8(2); ³ $J_{C1,H3ax} = 11$ Hz; HRMS (ESI-TOF) m/z: [M + Na]⁺ C₂₉H₃₉NNaO₁₂ calcd. 616.2370, found, 616.2379. [The β :α ratio was determined by the NH protons at ~ 8.9 ppm].

Synthesis of glycoside 10

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.030 g, 0.05 mmol, 1.0 equiv), 11-bromo-1-undecanol (0.026g, 0.11 mmol, 2.0 equiv) in presence of TMSOTf (10.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 57% (0.023 g) yield (β:a>20:1); R_f 0.4 (EtOAc: Hexane 4:6); $[α]_0^{25} = -21.1(c = 0.26, CHCl_3)$; ¹H NMR (400 MHz, CDCl₃) δ 8.92 (s, 1H), 7.44 - 7.37 (m, 5H), 5.33 (s, 1H), 5.30 - 5.25 (m, 1H), 5.04 (ddd, J=9.8, 4.1, 2.2 Hz, 1H), 5.00 - 4.94 (m, 2H), 4.45 (dd, J=12.3, 2.0 Hz, 1H), 4.03 (dd, J=9.8, 0.7 Hz, 1H), 3.89 (dd, J=12.4, 4.2 Hz, 1H), 3.40 (t, J=6.9 Hz, 2H), 3.33 - 3.27 (m, 1H), 3.25 - 3.19 (m, 1H), 2.24 (dd, J=12.8, 4.6 Hz, 1H), 2.05 (d, J=10.2 Hz, 6H), 1.98 (d, J=11.3 Hz, 6H), 1.89 - 1.83 (m, 3H), 1.57 - 1.54 (m, 2H), 1.43-1.38 (dd, J=12.9, 3.6 Hz, 2H), 1.28 (s, 12H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.6, 170.3, 170.1, 169.9, 164.6, 134.8, 129.6, 129.2, 128.8, 99.9, 78.5, 68.5, 67.7, 66.3, 64.5, 64.1, 62.5, 34.2, 33.0, 32.3, 29.6, 29.5, 28.9, 28.3, 26.3, 20.9(2), 20.8, 20.7; ³ $J_{C1,H3ax}=12$ Hz; HRMS(ESI-TOF) m/z: [M + Na]* C₃₄H₅₀BrNNaO₁₂ calcd. 766.2414, found, 766.2437. [The β:α ratio was determined by the NH protons at ~ 8.9 ppm].

Synthesis of glycoside 11

Preparation of the title compound via general procedure **B** is provided here. It was prepared using Kdo donor **3** (0.030 g, 0.05 mmol, 1.0 equiv), benzyl alcohol (0.012g, 0.11 mmol, 2.0equiv) in presence of TMSOTf (10.0 µL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 52% (0.017 g) yield (β : α > 20:1); R_f 0.35 (EtOAc : Hexane 3:7); [α] $_D^{25}$ = -16.2(c =0.4, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 9.08 (s, 1H), 7.46 - 7.44 (m, 2H), 7.40 - 7.37 (m, 3H), 7.34 (d, J = 6.5 Hz, 2H), 7.31 - 7.29 (m, 3H), 5.35 (s, 1H), 5.33 - 5.28 (m, 1H),

5.06 (d, J = 9.7 Hz, 1H), 5.01 (s, 2H), 4.46 - 4.39 (m, 2H), 4.25 (m, 1H), 4.12 (d, J = 9.9 Hz, 1H), 3.87 (dd, J = 12.4, 4.0 Hz, 1H), 2.34 (dd, J = 13.0, 4.8 Hz, 1H), 2.09 (t, J = 9.3 Hz, 1H), 2.05 (s, 3H), 1.97 (d, J = 9.5 Hz, 6H), 1.91 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.8, 170.3, 170.1, 169.9, 164.4 136.4, 134.7, 129.7, 129.3, 128.8, 128.7, 128.2, 127.6, 100.1, 78.5, 68.6, 67.6, 66.2, 65.9, 64.4, 62.3, 32.3, 20.9, 20.8, 20.7(2); ³JC_{1,H3ax} = 10Hz; HRMS(ESI-TOF) m/z: [M + Na]⁺ C₃₀H₃₅NNaO₁₂ calcd. 624.2057, found, 624.2047. [The β:α ratio was determined by the NH protons at ~ 9.0 ppm].

Synthesis of glycoside 12

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.025 g, 0.05 mmol, 1.0 equiv), cyclohexanol (0.009 g, 0.09 mmol, 2.0 equiv) in presence of TMSOTf (8.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 53% (0.014 g) yield as mixture of anomer (β : α 10:1); R_f 0.5 (EtOAc: Hexane 4:6); $[\alpha]_D^{25}$ = -27.8 (c=0.140, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 9.50(s,0.09H), 9.03 (s, 0.91H), 7.45 - 7.38 (m, 5H), 5.33 (s, 1H), 5.29 - 5.24 (m, 1H), 5.04 - 5.01 (m, 1H), 4.99 - 4.93 (m, 2H), 4.45 (dd, J= 12.2, 1.7 Hz, 1H), 4.17 (d, J= 9.7 Hz, 1H), 3.85 (dd, J= 12.3, 4.3 Hz, 1H), 3.51 - 3.45 (m, 1H), 2.29 (dd, J= 12.8, 4.7 Hz, 1H), 2.06 (s, 3H), 2.04 (s, 3H), 1.99 (s, 3H), 1.97 (s, 3H), 1.87 (t, J= 12.6 Hz, 2H), 1.72 -1.70 (m, 3H), 1.50 -1.49 (m, 1H), 1.36 - 1.31 (m, 2H), 1.25 - 1.17 (m, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.7, 170.3, 170.2, 169.9, 165.4, 134.8, 129.5, 129.2, 128.8, 100.3, 78.5, 74.4, 68.9, 68.1, 66.4, 64.7, 62.3, 34.0, 33.3, 32.9, 25.4, 24.5, 24.3, 20.9(2), 20.8(2); ³ $J_{C1,H3ax}$ = 11Hz; HRMS(ESI-TOF) m/z: [M + Na]+ C₂₉H₃₉NNaO₁₂ calcd. 616.2370, found, 616.2397; [The β :α ratio was determined by the NH protons at ~ 9.5-9.0 ppm].

Synthesis of glycoside 13

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.025 g, 0.05 mmol, 1.0 equiv), (—)-menthol (0.015 g, 0.09 mmol, 2.0 equiv) in presence of TMSOTf (9.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 48% (0.014 g) yield (β : α > 20:1); R_f 0.4 (EtOAc : Hexane 3:7); [α] $_0$ ²⁵ = -21.1(c=0.18, CHCl $_3$); ¹**H NMR** (400 MHz, CDCl $_3$) δ 9.11 (s, 1H), 7.44 - 7.37 (m, 5H), 5.33 - 5.27 (m, 2H), 5.07 - 4.94 (m, 3H), 4.54 (dd, J= 12.1, 2.1 Hz, 1H), 4.23 (d, J= 9.6 Hz, 1H), 3.72 (dd, J= 12.1, 5.7 Hz, 1H), 3.55 (td, J= 10.5, 4.3 Hz, 1H), 2.26 (dd, J= 12.5, 4.5 Hz, 1H), 2.21 - 2.14 (m, 1H), 2.05 (d, J= 5.3 Hz, 6H), 1.97 (d, J= 2.2 Hz, 6H), 1.88 (t, J= 12.5 Hz, 1H), 1.60 (s, 4H), 1.27 (m, 2H), 1.19 -1.14 (m, 1H), 0.97 - 0.93 (m, 1H), 0.88 (d, J= 7.1 Hz, 3H), 0.84 (d, J= 6.5 Hz, 3H), 0.80 (d, J= 6.8 Hz, 3H); ¹³C{¹H} NMR (100 MHz, CDCl $_3$) δ 170.7, 170.4, 170.3, 169.9, 164.4, 134.9, 129.4, 129.2, 128.9, 101.0, 78.5, 69.1, 68.3, 66.4, 64.8, 62.8, 48.6, 43.0, 34.3, 33.7, 31.4, 25.8, 23.2, 22.4, 21.2, 21.0, 20.8, 16.4; ³ $J_{\text{C1,H3ax}}$ = 11Hz; HRMS(ESI-TOF) m/z: [M + Na]+ C₃₃H₄₇NNaO₁₂ calcd. 672.2996, found, 672.2992. [The β :α ratio was determined by the NH protons at ~ 9.0 ppm].

Synthesis of glycoside 14

Title compound was prepared according to the general procedure B, using Kdo donor 3 (0.020 g, 0.04 mmol, 1.0 equiv), cholesterol (0.028 g, 0.07 mmol, 2.0 equiv) in presence of TMSOTf (6.0 µL) as promoter. Crude was purified by flash silica gel column chromatography (8:2 Hexane/EtOAc) to give the desired products as a colourless oil in 32% (0.01 g) yield $(\beta:\alpha>20:1)$; R_f 0.4 (EtOAc: Hexane 2:8); $[\alpha]_D^{25} = -66.1(c=0.18, CHCl_3)$; ¹H NMR (400 MHz, CDCl₃) δ 9.04 (s, 1H), 7.41 - 7.38 (m, 5H), 5.34 (s, 1H), 5.29 - 5.27 (m, 2H), 5.06 - 5.03 (m, 1H), 4.96 (dd, J = 24.0, 11.1 Hz, 2H), 4.52 (dd, J = 12.2, 1.7 Hz, 1H), 4.17 (d, J = 9.8 Hz, 1H), 3.84 (dd, J = 12.6, 5.0 Hz, 1H), 3.43 - 3.35 (m, 1H), 2.64 - 2.60 (m, 1H), 2.43 - 2.33 (m, 2H), 2.27 (dd, J = 13.2, 5.0 Hz, 1H), 2.16 - 2.11 (m, 2H), 2.04 (d, J = 7.3 Hz, 6H), 1.97 (d, J = 7.3 Hz, 6 = 7.3 Hz, 6H), 1.88 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 Hz, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 \text{ Hz}, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 \text{ Hz}, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 \text{ Hz}, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 \text{ Hz}, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 \text{ Hz}, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 \text{ Hz}, 2H), 1.83-1.79 (m, 3H), 1.43-1.42 (m, 3H), 1.33 (d, J = 12.8 \text{ Hz} 4.3 Hz, 2H), 1.28 -1.25 (m, 4H), 1.15 -1.09 (m, 6H), 0.98 (s, 3H), 0.91 (d, J = 6.3 Hz, 4H), 0.86 (d, J = 6.5 Hz, 9H), 0.67 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.8, 170.4, 170.2, 169.9, 165.3, 140.6, 134.8, 129.6, 129.2, 128.9, 122.3, 100.5, 78.5, 76.3, 69.2, 68.2, 66.4, 64.7, 62.6, 56.9, 56.3, 50.3, 42.5, 40.6, 39.9, 39.7, 37.4, 36.7, 36.3, 35.9, 32.1, 32.0, 29.3, 28.4, 28.2, 24.4, 24.0, 23.0, 22.7, 21.0, 20.9, 20.8(2), 19.4, 18.9, 12.0; HRMS(ESI-TOF) m/z: [M + Na]⁺ C₅₀H₇₃NNaO₁₂ calcd. 902.5030, found, 902.5030. [The β:α ratio was determined by the NH protons at ~ 9.0 ppm].

Synthesis of glycoside 15

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.020 g, 0.04 mmol, 1.0 equiv), 4-bromophenol (0.013 g, 0.07 mmol, 2.0 equiv) in presence of TMSOTf (7.0 µL) as promoter. Crude was purified by flash silica gel column

chromatography (6:4 Hexane/EtOAc) to give the desired products as a colourless oil in 63% (0.015 g) yield (β : α > 20:1); R_f 0.35 (EtOAc : Hexane 4:6); [α] $_0$ 25 = -5.41(c =0.2, CHCl $_3$); ¹H NMR (400 MHz, CDCl $_3$) δ 8.98 (s, 1H), 7.41 - 7.39 (m, 3H), 7.35-7.32 (m, 4H), 6.84 (d, J = 8.9 Hz, 2H), 5.47 - 5.42 (m, 1H), 5.40 (s, 1H), 5.07 (ddd, J = 9.8, 3.4, 2.2 Hz, 1H), 4.93 (d, J = 11.3 Hz, 1H), 4.82 (d, J = 11.3 Hz, 1H), 4.27 - 4.20 (m, 2H), 3.79 (dd, J = 12.5, 3.7 Hz, 1H), 2.45 (dd, J = 13.0, 4.6 Hz, 1H), 2.07 (s, 3H), 2.03 (m, 1H), 2.00 (s, 3H), 1.98 (s, 3H), 1.78 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl $_3$) δ 170.6, 170.2, 170.1, 169.8, 163.6, 152.7, 134.6, 132.6, 129.4, 129.3, 128.9, 120.0, 116.4, 100.4, 78.6, 69.5, 67.3, 65.9, 64.3, 62.1, 33.3, 20.9, 20.7, 20.5; ³ $J_{C1,H3ax}$ =12Hz; HRMS(ESI-TOF) m/z: [M + Na]+ C₂₉H₃₂BrNNaO₁₂ calcd. 688.1006, found, 681.1024. [The β : α ratio was determined by the NH protons at ~ 8.9 ppm].

Synthesis of glycoside 16

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.022 g, 0.04 mmol, 1.0 equiv), thiophenol (0.009 g, 0.08 mmol, 2.0equiv) in presence of TMSOTf (7.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to obtain α- andβ-anomer separately. α-anomer. The α-anomer was obtained as a colourless oil in 55% (0.013 g) yield; R_f 0.4 (EtOAc: Hexane 4:6); $[\alpha]_D^{25}$ = 17.2(c =0.18, CHCl₃); ¹**H NMR** (400 MHz, CDCl₃) δ 8.64 (s, 1H), 7.53 (d, J = 7.5 Hz, 2H), 7.43 - 7.39 (m, 2H), 7.36 (s, 1H), 7.34 - 7.33 (m, 3H), 7.21 - 7.19 (m, 2H), 5.41 - 5.37 (m, 2H), 5.05 - 5.03 (m, 1H), 4.78 (d, J = 10.9 Hz, 1H), 4.67 (d, J = 9.4 Hz, 1H), 4.45 (d, J = 11.0 Hz, 1H), 4.24 (dd, J = 12.5, 1.1 Hz, 1H), 3.74 (dd, J = 12.5, 5.0 Hz, 1H), 2.44 (dd, J = 13.8, 4.7 Hz, 1H), 2.24 (t, J = 13.0 Hz, 1H), 2.08 (s, 3H), 2.04 (s, 3H), 2.01 (s, 3H), 1.99 (s, 3H); 13 C{¹H} NMR (100 MHz, CDCl₃) δ 170.8, 170.3, 170.0, 169.8, 165.5, 136.6, 134.7, 130.2, 129.5, 129.3, 129.1, 129.0, 128.7, 91.1, 78.5, 70.0, 67.9, 66.7, 64.5, 62.8, 32.2, 21.0, 20.9, 20.8, 20.7.

β-anomer: It was obtained as a colourless oil in 17% (0.004 g) yield; R_f 0.35 (EtOAc : Hexane 4:6); [α] $D^{25} = -67.9(c = 0.24, CHCl_3)$; ¹H NMR (400 MHz, CDCl₃) δ 8.89 (s, 1H), 7.56 (d, J = 0.000)

6.9 Hz, 2H), 7.40 (d, J = 7.1 Hz, 1H), 7.36 (d, J = 7.5 Hz, 2H), 7.34 - 7.31 (m, 3H), 7.18 - 7.16 (m, 2H), 5.32 (dd, J = 6.6, 3.0 Hz, 1H), 5.08 (td, J = 6.2, 2.5 Hz, 1H), 4.85 - 4.80 (m, 1H), 4.68 (d, J = 11.0 Hz, 1H), 4.51 (dd, J = 5.7, 3.0 Hz, 1H), 4.29 (dd, J = 12.5, 2.6 Hz, 1H), 4.24 (d, J = 11.0 Hz, 1H), 3.97 (dd, J = 12.5, 5.8 Hz, 1H), 2.98 (dd, J = 14.8, 7.9 Hz, 1H), 2.32 (dd, J = 14.8, 5.0 Hz, 1H), 2.10 (s, 6H), 2.06 - 2.01 (s, 3H), 1.95 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.6, 170.3, 170.1, 169.6, 165.7, 136.6, 134.9, 130.4, 129.9, 129.4, 129.0, 128.6, 127.7, 95.0, 82.1, 78.4, 72.0, 69.9, 69.6, 61.9, 42.8, 21.1, 20.9(2), 20.6; HRMS(ESI-TOF) m/z: [M + Na]⁺ C₂₉H₃₃NNaO₁₁Scalcd. 626.1672, found, 626.1689.

Note: Experiment performed to determine the heteronuclear coupling constant between the carbonyl carbon at C1 and axial proton at C3 for one anomer did not give any splitting of the target carbonyl, which we characterized as the α -anomer. The other one provided $^3J_{\text{C1,H3ax}}=12$ Hz and characterized as β -anomer.

Synthesis of glycoside 17

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.027 g, 0.05 mmol, 1.0 equiv), *tert*-butyl hydroperoxide (0.010 g, 0.10 mmol, 2.0equiv) in presence of TMSOTf (8.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 49% (0.014 g) yield (β : α > 20:1); R_f 0.35 (EtOAc : Hexane 3:7); [α] $_0$ ²⁵= -25.6 (c =0.18, CHCl₃); 1 **H NMR** (400 MHz, CDCl₃) δ 9.03 (s, 1H), 7.45 - 7.44 (m, 2H), 7.41 - 7.36 (m, 3H), 5.34 (d, J = 2.0 Hz, 1H), 5.17 - 5.06 (m, 2H), 4.99 (d, J = 11.1 Hz, 1H), 4.90 (d, J = 11.1 Hz, 1H), 4.41 (dd, J = 12.2, 1.6 Hz, 1H), 4.19 (d, J = 10.0 Hz, 1H), 4.02 (dd, J = 12.3, 4.7 Hz, 1H), 2.33 (dd, J = 13.7, 5.1 Hz, 1H), 2.07 (s, 3H), 2.05 (s, 3H), 2.01 (s, 3H), 2.00 - 1.93 (m, 4H), 1.24 (s, 9H); 13 **C**{ 1 **H**} **NMR** (100 MHz, CDCl₃) δ 170.7, 170.5, 170.1, 169.9, 163.7, 134.9, 129.7, 129.1, 128.8, 101.6, 81.6, 78.5, 68.7, 67.9, 66.3, 64.3, 63.0, 29.7, 26.6, 21.0, 20.9, 20.8, 20.7; 3 J_{C1,H3ax} =12Hz; **HRMS(ESI-TOF)** m/z: [M + Na]+ C₂₇H₃₇NNaO₁₃calcd. 606.2163, found, 606.2174. [The β : α ratio was determined by the NH protons at ~ 9.0 ppm].

Synthesis of glycoside 18

Title compound was prepared according to the general procedure B, using Kdo donor 3 (0.050 g, 0.09 mmol, 1.0 equiv), 3,4,5-tris(benzyloxy)-6-methoxytetrahydro-2H-pyran-2-yl) methanol (0.084 g, 0.18 mmol, 2.0equiv) in presence of TMSOTf (17.0 µL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 60% (0.052 g) yield ($\beta:\alpha>20:1$); R_f 0.35 (EtOAc: Hexane 3:7); $[\alpha]_D^{25} = -4.17$ (c=0.12, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 9.26 (s, 1H), 7.34 - 7.32 (m, 9H), 7.30 - 7.29 (m, 6H), 7.26 - 7.24 (m, 5H), 5.25 (s, 1H), 5.23 - 5.19 (m, 1H), 5.06 - 5.01 (m, 1H), 4.97 - 4.88 (m, 3H), 4.82 - 4.74 (m, 3H), 4.64 (d, J = 12.1 Hz, 1H), 4.51(dd, J = 16.0, 7.3 Hz, 2H), 4.34 (d, J = 11.8 Hz, 1H), 3.96 (t, J = 10.1 Hz, 2H), 3.79 (dd, J = 10.1 Hz, 2H), 3.70 (dd, J = 10.1 Hz, 2H), 3.70 (dd, J = 10.1 Hz, 2H), 3.70 (dd, J = 10.1 Hz, 2H), 3.12.3, 4.4 Hz, 2H), 3.62 (d, J = 9.9 Hz, 1H), 3.43 (dd, J = 9.6, 3.3 Hz, 1H), 3.36 - 3.30 (m, 5H), 2.12 (dd, J = 11.4, 6.6 Hz, 1H), 2.03 (s, 3H), 1.99 (brs, 1H), 1.97 (s, 3H), 1.95 (s, 3H), 1.82 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.6, 170.4, 170.0, 169.8, 164.4, 138.6, 138.1(2), 134.9, 129.5, 129.1, 128.7, 128.6(2), 128.5, 128.2, 128.1(2), 127.9, 127.8, 99.7, 97.9, 82.1, 80.0, 78.4, 77.8, 75.9, 75.0, 73.4, 69.3, 68.9, 67.7, 66.2, 64.4, 63.3, 62.6, 55.4, 32.3, 20.9, 20.8, 20.7(2); ${}^{3}J_{C1,H3ax} = 11Hz$; **HRMS(ESI-TOF)** m/z: [M + Na]⁺ C₅₁H₅₉NNaO₁₇ calcd. 980.3681, found, 980.3678. [The β : α ratio was determined by the NH protons at \sim 9.0 ppm].

Synthesis of glycoside 19

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.038 g, 0.07 mmol, 2.0 equiv), 6-(allyloxy)-3,4,5-tris(benzyloxy)tetrahydro-2H-pyran-2-yl) methanol (0.017 g, 0.03 mmol, 1.0equiv) in presence of TMSOTf (7.0 μ L) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 35% (0.012 g) yield which was isolated as mixture of anomer (β : α > 20:1); R_f 0.3 (EtOAc : Hexane

3:7); $[\alpha]_D^{25} = -49.28$ (c=0.140, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 9.04 (s, 1H), 7.47- 7.44 (3H), 7.44 - 7.37 (m, 6H), 7.35 (s, 2H), 7.34 (s, 2H), 7.32 - 7.31 (m, 4H), 7.29 - 7.28 (m, 3H), 5.91 - 5.78 (m, 1H), 5.35 - 5.28 (m, 4H), 5.24 - 5.18 (m, 1H), 5.06 (ddd, J = 9.9, 3.9, 2.1 Hz, 2H), 5.02 (s, 2H), 4.98 - 4.81 (m, 2H), 4.74 - 4.66 (m, 1H), 4.62 - 4.51 (m, 2H), 4.46 - 4.40 (m, 3H), 4.26 (d, J = 11.0 Hz, 1H), 4.19 - 4.10 (m, 2H), 3.95 - 3.73(m, 5H), 2.35 (dd, J = 12.8, 4.7 Hz, 1H), 2.05 (s, 3H), 1.99 (s, 3H), 1.96 (s, 3H), 1.92 (s, 3H), 1.87 (s, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.7, 170.3, 170.0, 169.9, 164.4, 138.4, 136.5, 134.8, 133.6, 129.7, 129.5, 129.3, 128.9, 128.7, 128.6, 128.5(2), 128.2, 128.0(2), 127.8, 127.7, 117.7, 100.2, 99.7, 78.6, 78.5, 75.1, 75.0, 73.1, 72.3, 71.0, 68.8, 68.0, 67.7, 66.3, 66.0, 64.6, 64.5, 62.4, 32.3, 20.9, 20.8(2), 20.7; ${}^3J_{\text{C1,H3ax}}$ = 10 Hz; HRMS(ESI-TOF) m/z: [M + Na]+ C₅₃H₆₁NNaO₁₇ calcd. 1006.3837, found, 1006.3829. [The β : α ratio was determined by the NH protons at ~ 9.0 ppm].

Synthesis of glycoside 20

Title compound was prepared according to the general procedure **B**, using Kdo donor **3** (0.058 g, 0.10 mmol, 1.2equiv), 3,4-bis(benzyloxy)-5-methoxytetrahydrofuran-2-yl) methanol (0.030 g, 0.09 mmol, 1.0 equiv) in presence of TMSOTf (10.0 μL) as promoter. Crude was purified by flash silica gel column chromatography (6:4 Hexane/EtOAc) to give the desired products as a colourless oil in 42% (0.031 g) yield which was isolated as anomeric mixture (β : α > 20:1); R_f 0.35 (EtOAc: Hexane 4:6); $[\alpha]_D^{25} = -19.4(c=0.180, CHCl_3)$; ¹**H NMR** (400 MHz, CDCl₃) δ 9.13 (s, 1H), 7.30 - 7.26 (m, 9H), 7.24 - 7.22(m, 4H), 7.20 (s, 2H), 5.25 (s, 1H), 5.18 - 5.15 (m, 1H), 5.04 - 5.01 (m, 1H), 4.89 - 4.80 (m, 3H), 4.56 - 4.34 (m, 6H), 4.11 (d, J = 4.3 Hz, 1H), 4.04 (d, J = 9.7 Hz, 1H), 3.90 (brs, 1H), 3.81 - 3.74 (m, 2H), 3.45 (dd, J = 9.9, 3.5 Hz, 1H), 3.31 (s, 3H), 2.09 (dd, J = 12.9, 4.9 Hz, 1H), 1.99 (s, 3H), 1.94 (d, J = 3.8 Hz, 6H), 1.91 (s, 3H), 1.85(m, 1H); ¹³**C**{¹**H**} **NMR**(100 MHz, CDCl₃) δ 170.8, 170.4, 170.0, 169.9, 164.3, 137.8, 137.5, 134.8, 129.6, 129.1, 128.8, 128.6(2), 128.1(2), 128.0, 107.5, 99.8, 87.6, 83.4, 80.5, 78.4, 72.3, 72.0, 68.9, 67.8, 66.2, 64.6, 64.4, 63.1, 55.1, 32.3, 20.9, 20.8(3); ³J_{C1,H3ax} =12Hz; **HRMS(ESI-TOF)** m/z: [M + Na]⁺ C₄₃H₅₁NNaO₁₆ calcd. 860.3106, found, 860.3102. [The β :α ratio was determined by the NH protons at ~ 9.0 ppm].

6. Follow-up transformations

Scheme 3 (from main text).

Step-(i): To a solution of **18** (0.030g, 0.03mmol, 1.0equiv) in MeOH (0.05M) was added 20 wt% MeONa in MeOH (freshly prepared). The reaction mixture was stirred for 30min at room temperature and then neutralized with Amberlite® IR-120-H resin and filtered. The filtered was concentrated in *vacuo* to give a crude of product **21** which was used in the next reaction. Step-(ii): Crude product **21** and Pd(OH)₂ (0.2 equiv.) were taken in absolute ethanol (0.05M), purged two times consecutively with N₂ and H₂ and stirring was continued with H₂balloon at room temperature for 12h. Upon completion (TLC monitored) reaction was filtered through celite and concentrated in *vacuo* to obtain crude product **22** and was analysed with LC-MS. The crude was directly used in the next step.

Step-(iii): Crude **22** was taken in a mixture of pyridine (30μL) in acetic anhydride (30μL)and stirred under argon at room temperature for 12h. Solvents were removed in vacuo and the crude was purified by silica gel column chromatography (using 20% EtOAc/ hexane as eluent) to afford compound **23** (0.010g; 45% overall yield); R_f 0.3 (EtOAc : Hexane 2:8 ; [α] p^{25} = -20.8(c =0.12, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 6.67 (d, J = 2.6 Hz, 1H), 5.63 (d, J = 2.9 Hz, 1H), 5.47 (t, J = 9.7 Hz, 1H), 5.36 - 5.33 (d, J = 10.7 Hz, 2H), 5.22 - 5.18 (m, 1H), 4.97 - 4.95 (m, 1H), 4.89 - 4.86 (m, 1H), 4.84 - 4.80 (m, J = 10.3, 3.8 Hz, 1H), 4.63 (dd, J = 12.2, 2.2 Hz, 1H), 4.24 (d, J = 9.2 Hz, 1H), 4.06-4.02 (m, 1H), 4.01 - 3.98 (m, 1H), 3.45 (s, 3H), 3.43-3.41 (m, 2H), 2.23 (dd, J = 12.8, 5.4 Hz, 1H), 2.10 (s, 3H), 2.08 (s, 6H), 2.05 (s, 1H), 2.04 (s, 3H), 2.03 (s, 3H), 2.00 (s, 3H), 1.96 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.9, 170.4, 170.1, 170.0(3), 169.3, 99.2, 96.5, 71.1, 70.2, 69.4, 69.3, 68.1(2), 66.2, 64.7,

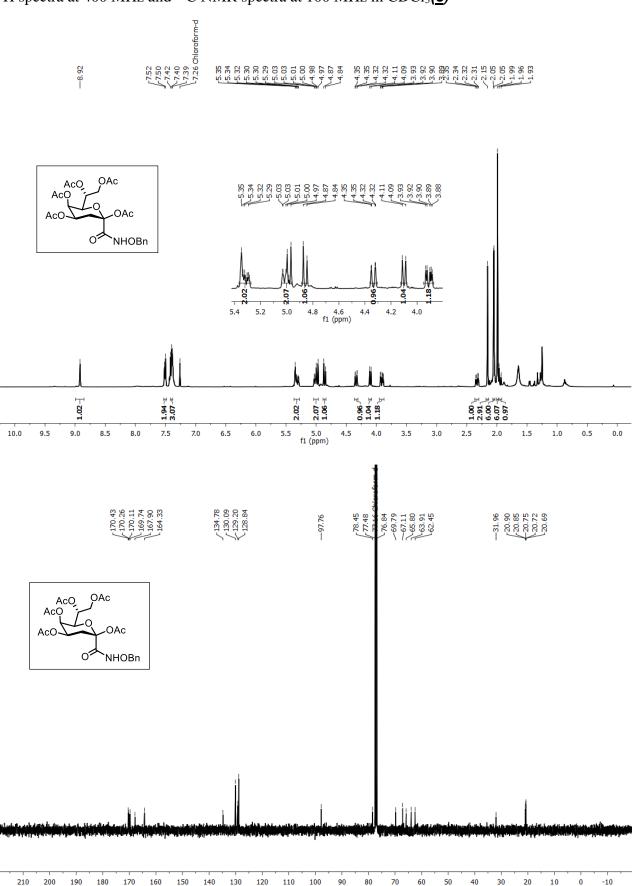
63.1, 62.8, 55.6, 32.1, 21.0, 20.9, 20.8(2); **HRMS(ESI-TOF)** m/z: [M + Na]⁺ C₂₉H₄₁NNaO₁₉ calcd. 730.2170, found, 730.2168

Synthesis of glycoside 25

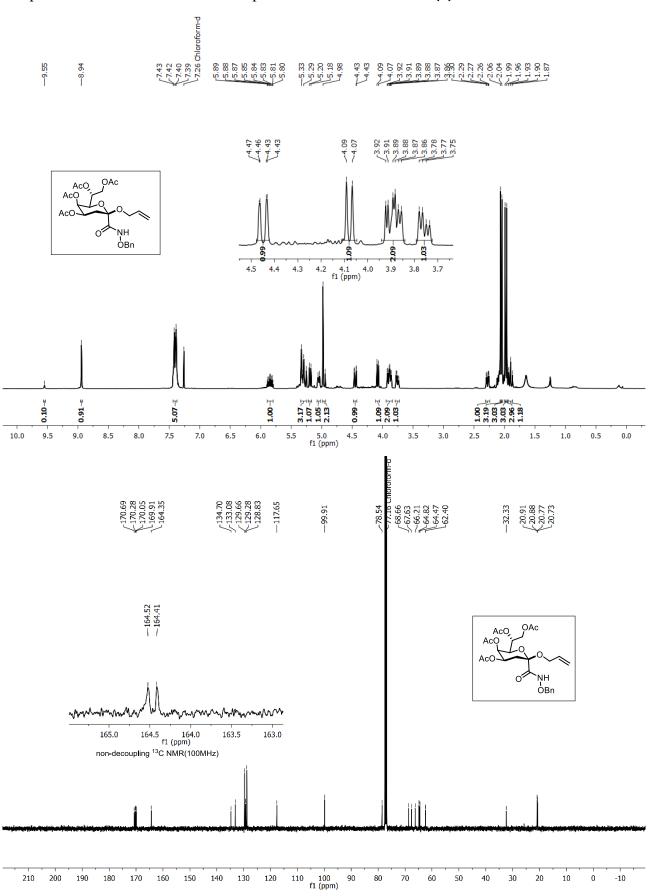
Title compound was prepared according to the general procedure **B**, using Kdo donor **24** (0.05 g, 0.09 mmol, 1.0 equiv), allyl alcohol (0.01 g, 0.19 mmol, 2.0 equiv) in presence of TMSOTf (17.0 µL) as promoter. Crude was purified by flash silica gel column chromatography (7:3 Hexane/EtOAc) to give the desired products as a colourless oil in 22% (0.011 g) yield as single anomer; R_f 0.5 (EtOAc : Hexane 4:6); $[\alpha]_D^{25}$ = -33.5(c=0.140, CHCl₃); ¹H NMR (400 MHz, CDCl₃) δ 7.37 (m, 1H), 7.35 (t, J = 1.7 Hz, 1H), 7.30 (s, 2H), 7.28 (s, 1H), 6.94 (t, J = 5.9 Hz, 1H), 5.89 (m, 1H), 5.41-5.36 (m, 2H), 5.31 - 5.26 (m, 1H), 5.21-5.15 (m, 2H), 4.54 - 4.50 (m, 4H), 4.18 (dd, J = 9.9, 1.0 Hz, 1H), 4.05 (dd, J = 12.4, 4.1 Hz, 1H), 3.95 (ddt, J = 12.1, 5.3, 1.4 Hz, 1H), 3.86 (ddt, J = 12.1, 5.6, 1.4 Hz, 1H), 2.34 (ddd, J = 13.0, 4.5, 1.4 Hz, 1H), 2.06 (s, 6H), 2.00 (s, 3H), 1.97 (s, 3H); ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 170.8, 170.4, 170.1, 170.0, 167.5, 137.8, 133.3, 129.0, 127.9, 127.7, 117.5, 99.7, 68.8, 67.7, 66.5, 64.8, 64.7, 62.5, 43.4, 32.5, 20.9, 20.8; ³J_{C1,H3ax} = 3 Hz; HRMS(ESI-TOF) m/z: [M + Na]+C₂eH₃₃NNaO₁₁ calcd. 558.1951, found, 558.1951.

7. NMR Spectra of new compounds

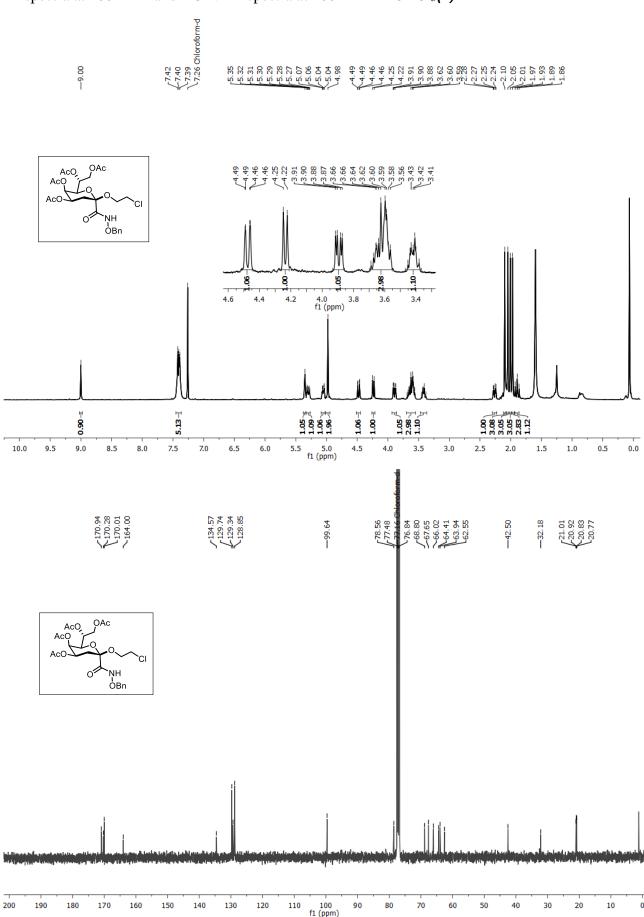
¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(3)



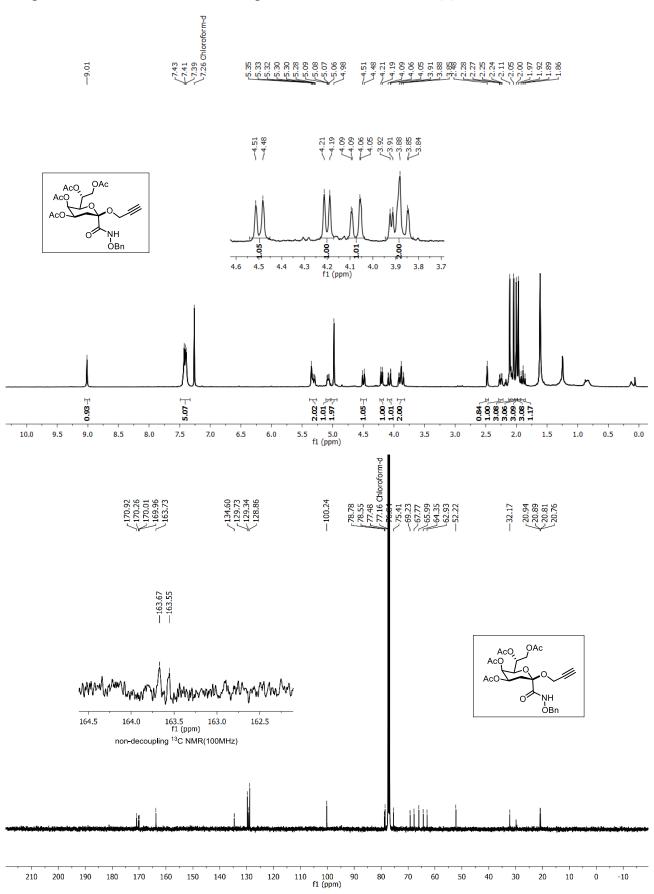
 ^{1}H spectra at 400 MHz and ^{13}C NMR spectra at 100 MHz in CDCl₃(5)



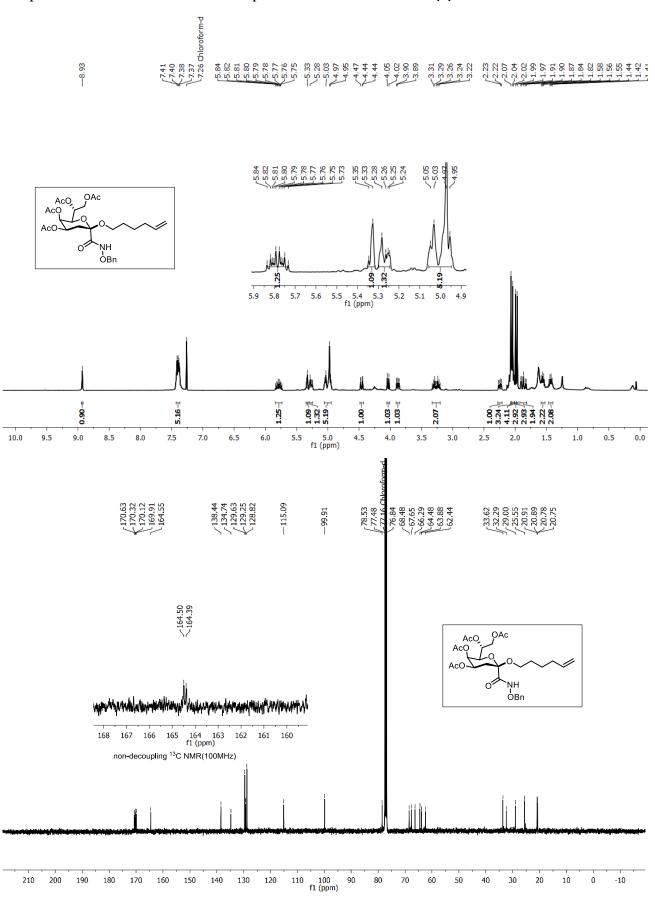




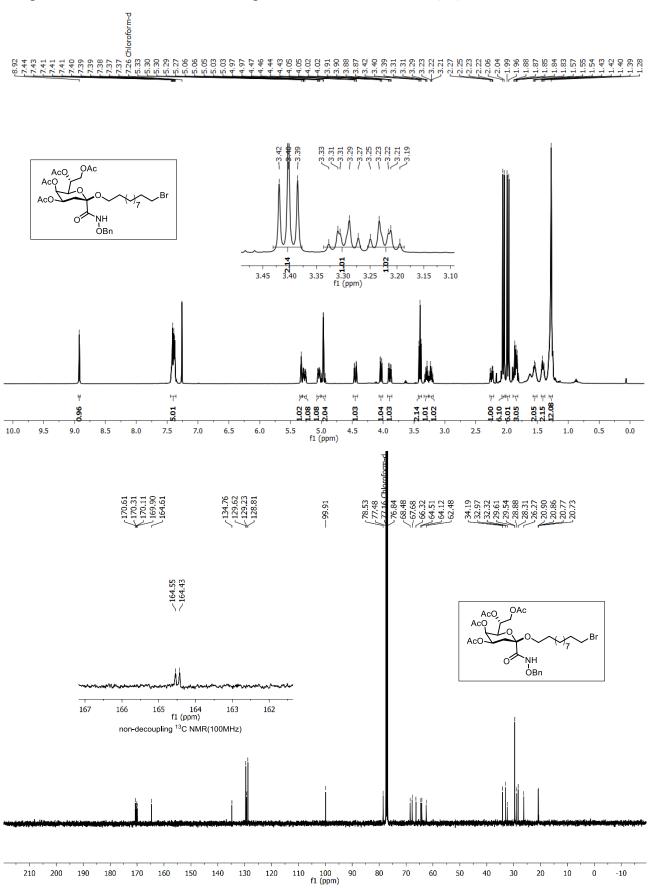




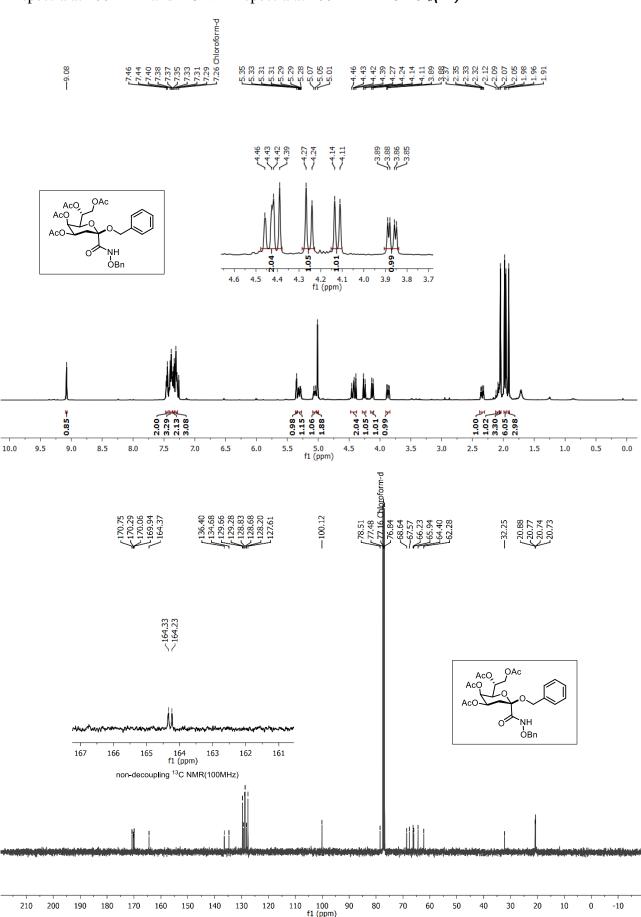
 ^{1}H spectra at 400 MHz and ^{13}C NMR spectra at 100 MHz in CDCl₃(9)



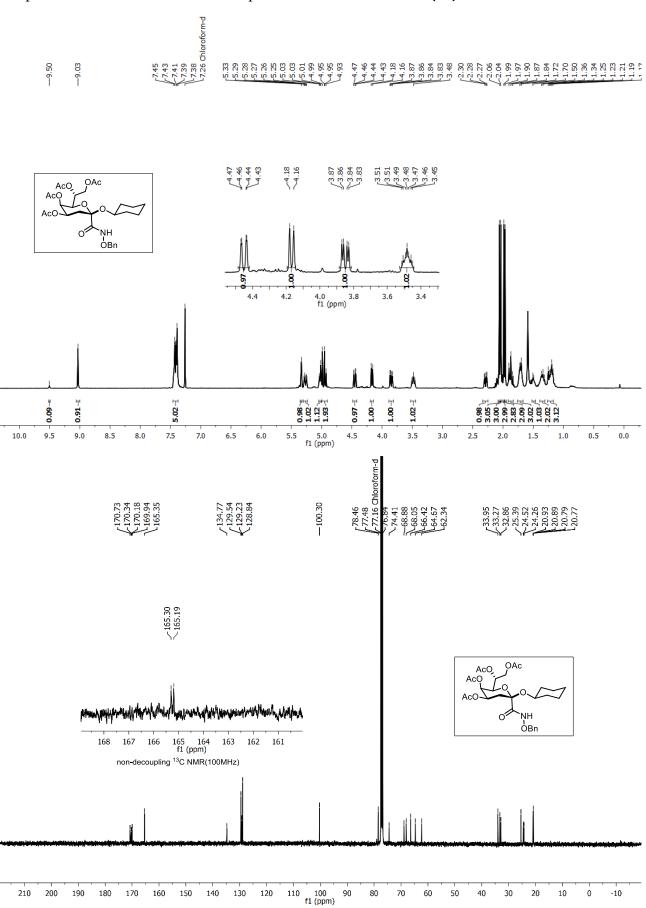




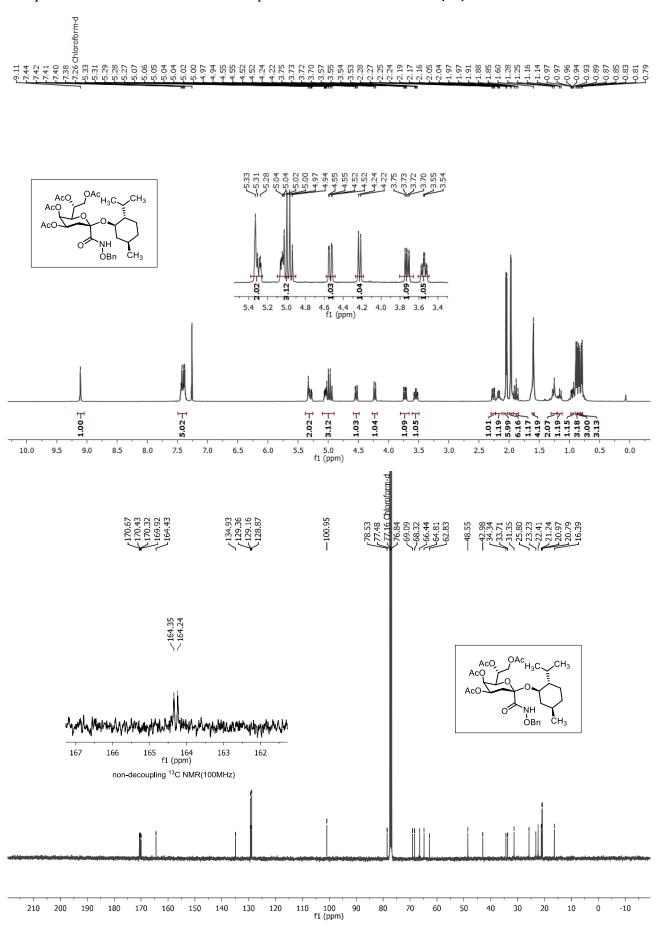
¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(11)

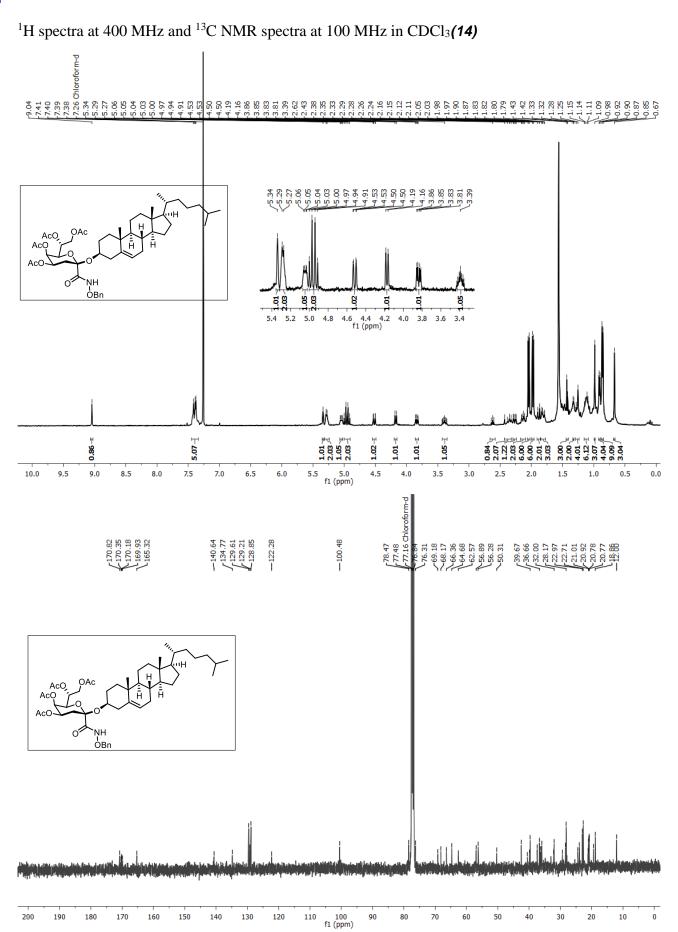


¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(12)

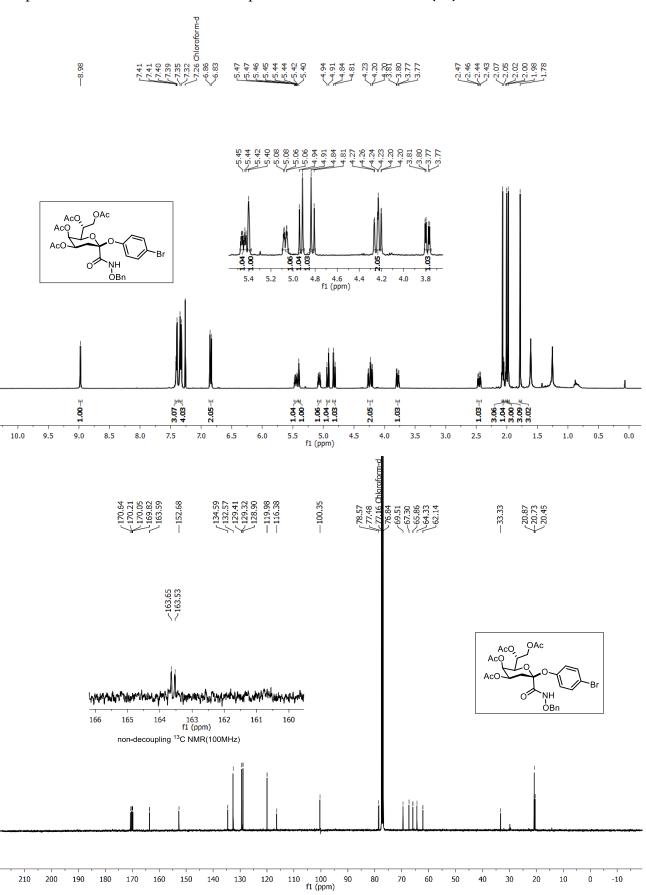


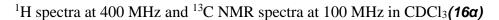
¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(13)

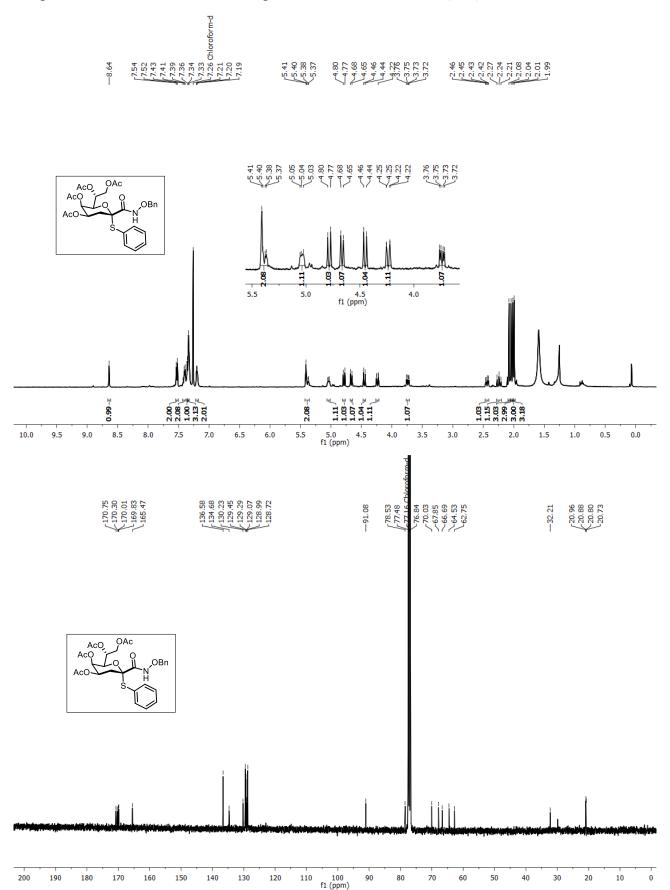




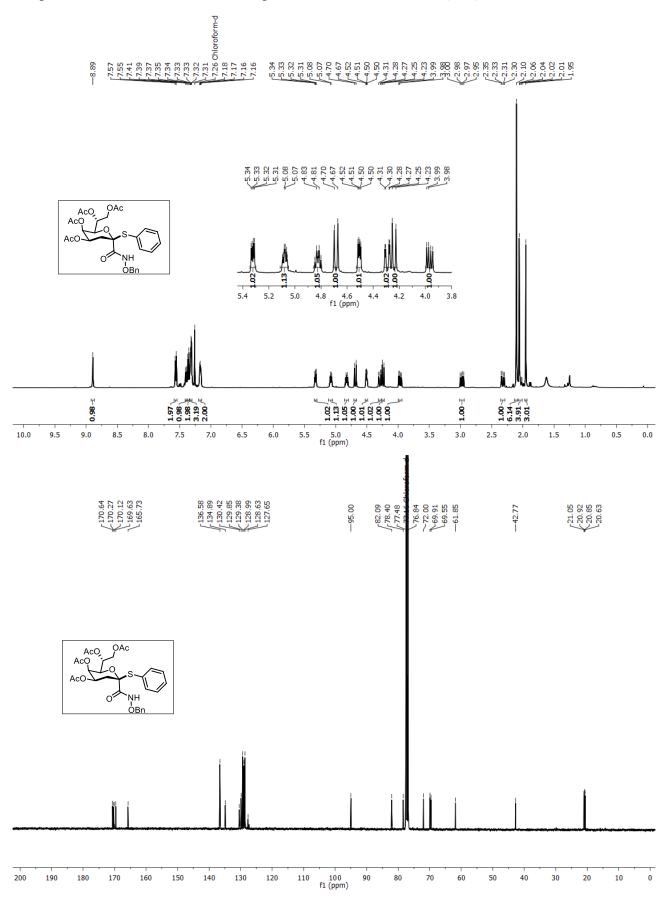
¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(15)



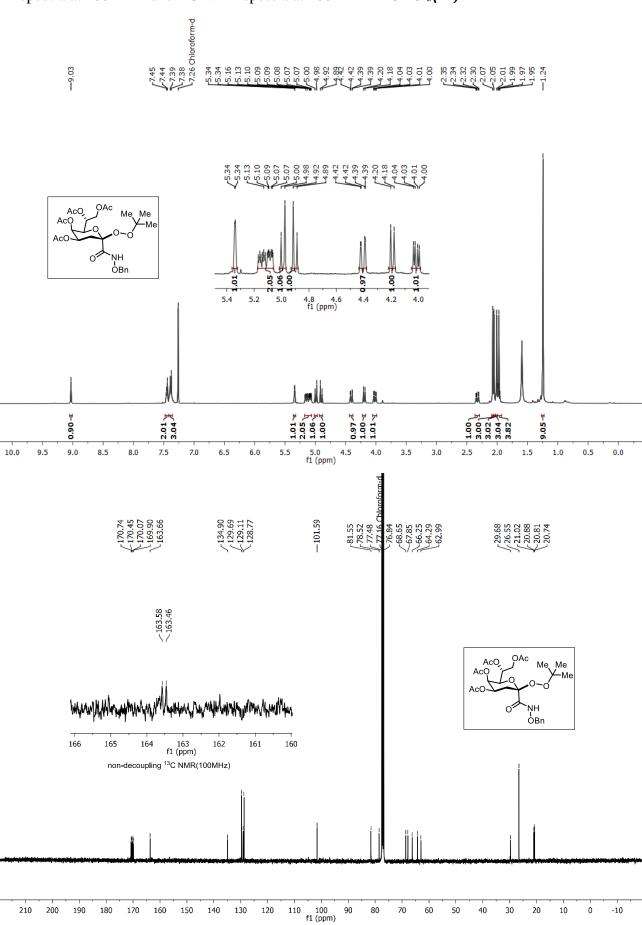




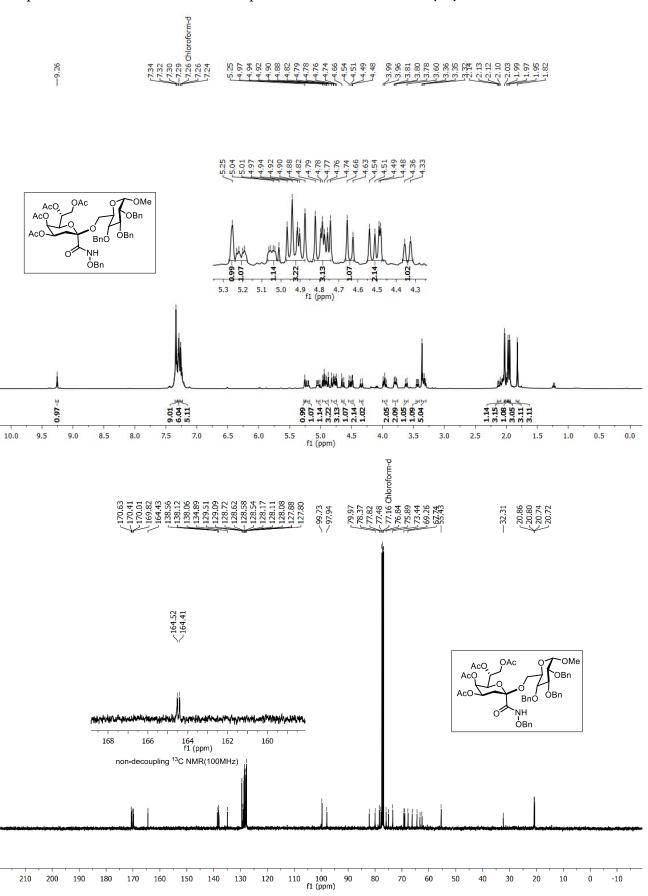
¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(16β)



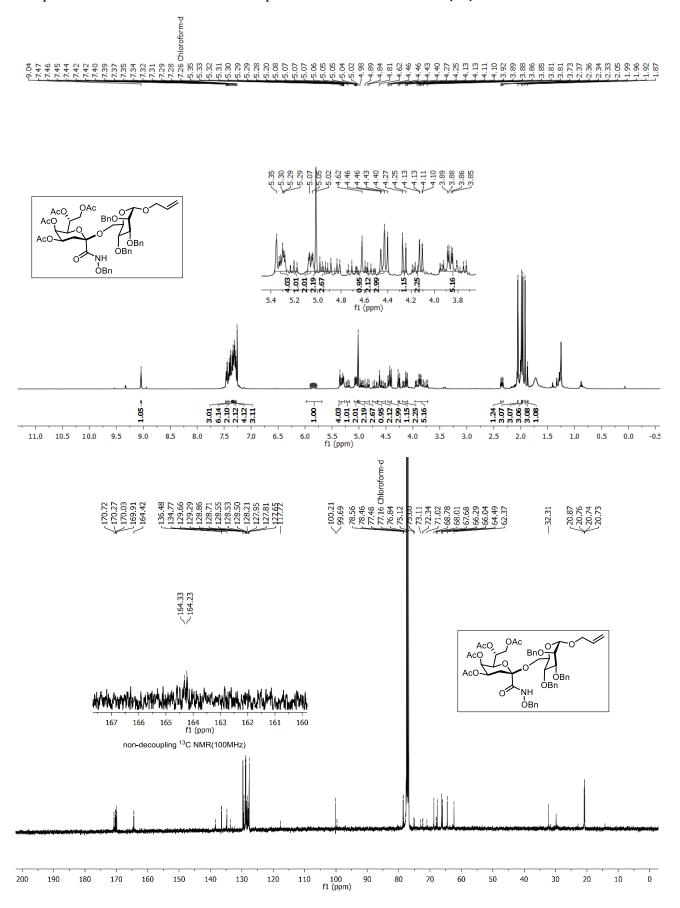
¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(17)



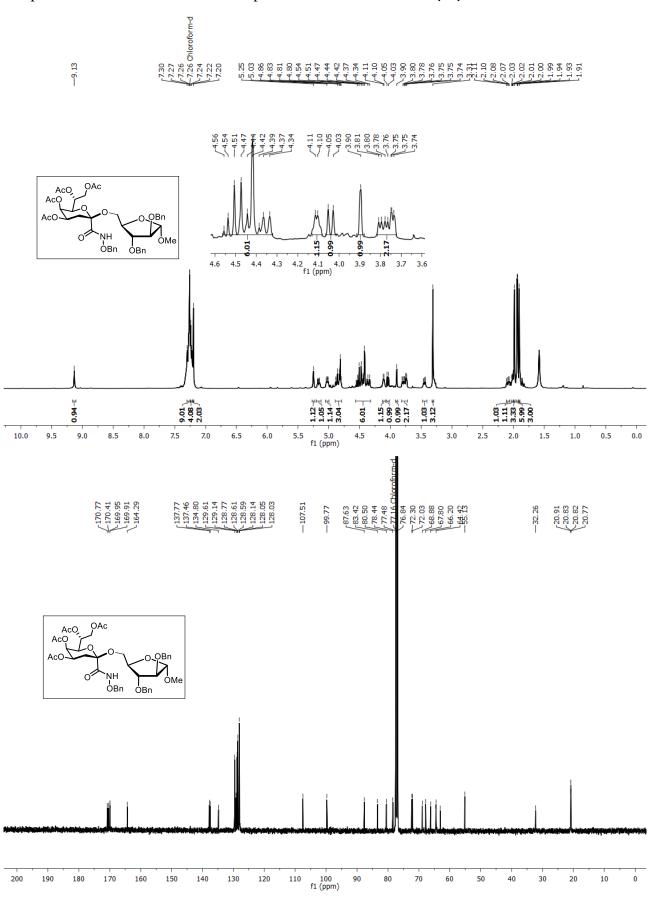
¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(18)



¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(19)

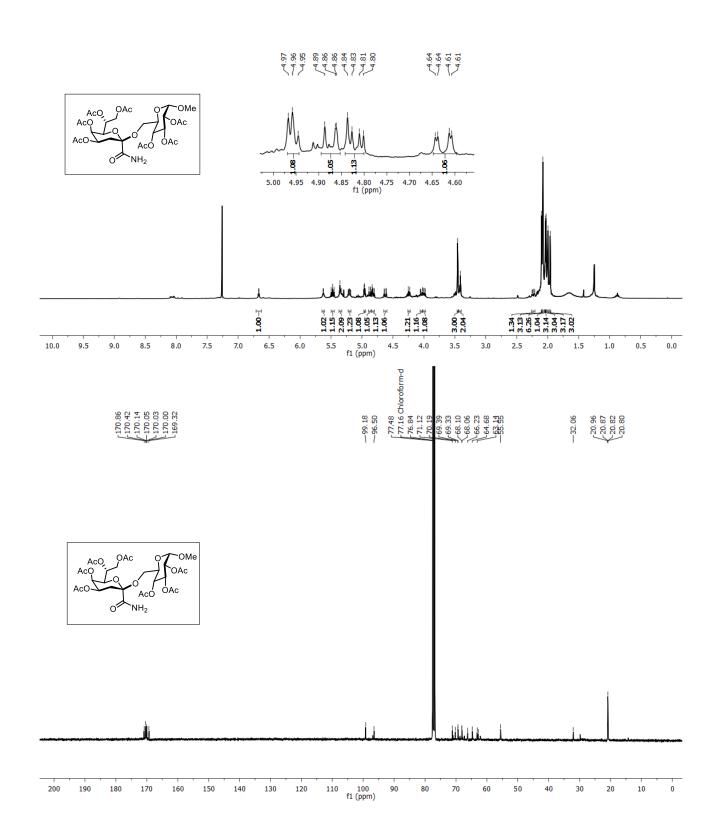


¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(20)

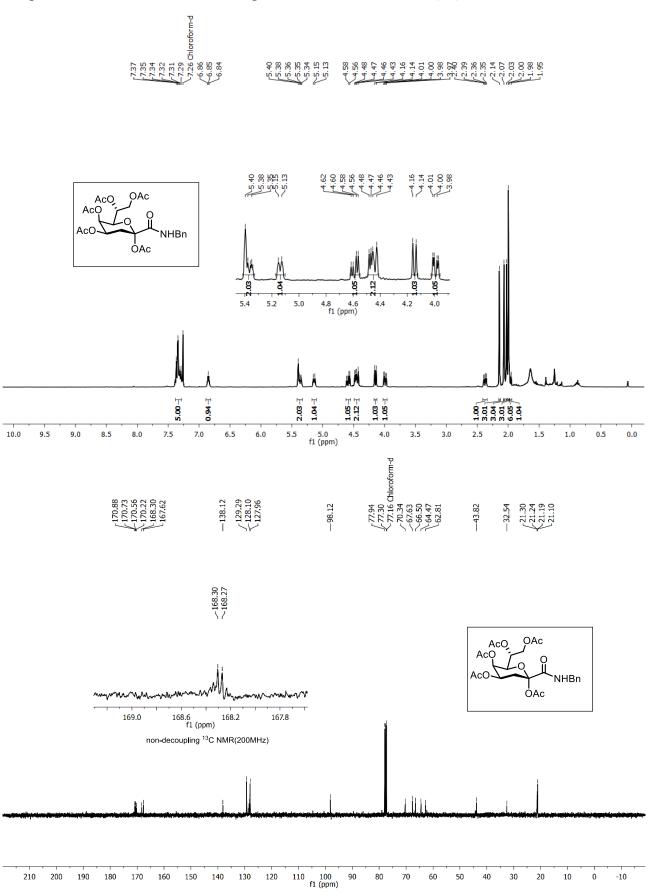


¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(23)









¹H spectra at 400 MHz and ¹³C NMR spectra at 100 MHz in CDCl₃(25)



