Electronic Supporting Information

Enhanced Anion Recognition by Ammonium [2]Catenane Functionalisation of a Halogen Bonding Acyclic Receptor

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Table of Content		
1.	Synthetic Methods	2
2.	Pseudorotaxane Complexation of 5 with $6 \cdot PF_6$ in d_2 -dichloromethane	21
3.	Anion binding in organic and aqueous-organic mixture	21
4.	Synthesis and binding studies of 11XB	25
5.	References	28

1. Synthetic Methods

1.1 Synthesis of the bis-vinyldibenzyl ammonium thread 6·PF₆

Before embarking upon the azide catenane synthesis, it was necessary to prepare the appropriate bis-alkene thread, using a modified procedure reported by Stoddart and Iwamoto (Scheme S1).^{1, 2} Firstly, esterification of commercially available 4-aminomethyl benzoic acid **S1** with thionyl chloride and methanol afforded the methyl 4-aminomethyl benzoate **S2** in 88% yield. An equivalent of 4-formylmethylbenzoate was reacted with **S2** in dry methanol overnight to form imine **S3**, which was subsequently reduced using NaBH₄ to obtain the secondary amine **S4** without further purification. The reaction of **S4** with *tert*- di-*tert*-butyl dicarbonate in the presence of a catalytic amount DMAP gave the Boc-protected secondary amine **S5** in 82% yield. Reduction of the methyl ester group using LiAlH₄ was achieved in quantitative yield to obtain the bis-alcohol **S6**. A S_N2 reaction of **S6** and decenyl tosylate in the presence of NaH in DMF afforded the bis-alkene **S7** in 72% yield. Deprotection of the Boc group using acid gave the bis-alkene ammonium thread **6**·P**F**₆ in 90% yield after anion exchange.



Scheme S1 Synthesis of bis-vinyldibenzyl ammonium thread 6·PF₆.

1.2 Synthesis of the nitro-dibenzo[24]crown-8 macrocycle 3

Macrocycle precursors **S8**, **S9**, **S10**, and **2** were synthesised according to previously reported procedures (Scheme 2). Subsequent etherification of 2 with 4-nitrocatechol led to the formation of NO₂-Dibenzo[24]crown-8 **3**.^{3, 4}



Scheme S2 Synthesis of a novel nitro-dibenzo[24]crown-8 macrocycle 3.

(a) NO₂-Dibenzo[24]crown-8 (**3**)



Ditosylate **2** (2.87g, 4.2 mmol), 2-nitrocatechol (653 mg, 4.2 mmol), and KPF₆ (928 mg, 5.04 mmol) were mixed in MeCN (140 ml), and then degassed throughout a solution. K_2CO_3 (2.32g, 16.8 mmol) was added to a reaction mixture which was refluxed in the absence of light for 72 hours. After cool down to room temperature and removing solid residues, the reaction was dried and re-dissolved in DCM. The organic residue was washed with pyridine hydrochloride solution (3 times) and water (3 times), and subsequently dried over MgSO₄. Solvent was removed to obtain a yellow solid product of **2** in 90% yield.

¹**H-NMR** (400 MHz, CDCl₃) δ = 7.87 (H_a, dd, *J* = 8.9, 2.6 Hz, 1H), 7.72 (H_c, d, *J* = 2.6 Hz, 1H), 6.88 (H_b and H_{d-g}, q, *J* = 5.2, 4.6 Hz, 5H), 4.23 (H_{m,m'}, dt, *J* = 7.6, 4.0 Hz, 4H), 4.15 (H_{h,h'}, t, *J* = 4.3 Hz, 4H), 3.99 – 3.88 (H_{i,i} and H_{I,i}, m, 8H), 3.83 (H_{i,i} and H_{k,k'}d, *J* = 2.8 Hz, 8H).

¹³C-NMR (101 MHz, CDCl₃) δ = 154.46, 148.93, 148.85, 148.47, 141.57, 121.64, 121.57, 118.18, 114.17, 114.06, 111.47, 108.54, 71.47, 71.28, 69.95, 69.88, 69.73, 69.55, 69.49, 69.43, 69.30.

HRMS (ESI+ve) m/z: 511.22855 ([M+NH₄]⁺, C₂₄H₃₅N₂O₁₀ requires 511.22862)



Figure S2 ¹³C-NMR spectrum of 3 (101 MHz, CDCl₃)



Figure S3 HRMS spectrum of 3

1.3 Synthesis of the azido-dibenzo[24]crown-8 macrocycle 5



Scheme S3 Synthesis of a novel azide-dibenzo[24]crown-8 macrocycle 5.

(a) NH₂-Dibenzo[24]crown-8 (4)



To a solution of NO₂-dibenzo[24]crown-8 **3** (1.07g, 2.03 mmol) in EtOH (10 ml) was added a suspension of 10%(w/w) Pd/C (100 mg) in water (0.5ml) and hydrazine monohydrate (5.5ml). The mixture was refluxed at 80°C for 5 hrs, followed by filtered through Celite® and removed solvent to afford a white solid in quantitative yield of **4**. Product can be used in the next step without further purification.

¹**H-NMR** (400 MHz, CDCl₃) δ = 6.88 (H_{d-g}, d, *J* = 2.4 Hz, 4H), 6.70 (H_b, d, *J* = 8.4 Hz, 1H), 6.25 (H_c, d, *J* = 2.6 Hz, 1H), 6.18 (H_a, dd, *J* = 8.4, 2.6 Hz, 1H), 4.14 (H_{h,h'}, t, *J* = 4.3 Hz, 4H), 4.06 (H_{m,m'}, q, *J* = 5.0 Hz, 4H), 3.96 - 3.71 (H_{i-1} and H_{i-1}, m, 16H).

¹³**C-NMR** (101 MHz, CDCl₃) δ = 150.22, 148.98, 148.95, 141.70, 141.55, 121.49, 121.46, 117.32, 114.25, 114.18, 107.20, 102.55, 71.19, 70.98, 70.65, 70.18, 69.91, 69.89, 69.86, 69.44, 69.33, 69.03.

HRMS (ESI+ve) m/z: 464.22760 ([M+H]⁺, C₂₄H₃₄NO₈ requires 464.22789)



Figure S5 ¹³C-NMR spectrum of 4 (101 MHz, CDCl₃)



Figure S6 HRMS spectrum of 4

(b) N₃-Dibenzo[24]crown-8 (5)



A solution of NH₂-Dibenzo[24]crown-8 **4** (139 mg, 0.3 mmol) in EtOH (5ml) was cooled down and added 1:1 HCl (37%)/H₂O. NaNO₂ (62 mg, 0.9 mmol) was subsequently added to a reaction mixture, then stirring at 0°C for 1 hour. After addition of NaN₃ (59 mg, 0.9 mmol), the reaction was left stirring overnight, followed by dilute with water. An aqueous solution was extracted using Et₂O, the organic layer was dry over MgSO₄ and removed solvent to obtain the target product **5** as a brown solid in 90% yield.

¹**H-NMR** (400 MHz, CDCl₃) δ = 6.94 – 6.80 (H_{d-g} and H_a m, 5H), 6.58 (H_b, dd, *J* = 8.5, 2.6 Hz, 1H), 6.52 (H_c, d, *J* = 2.6 Hz, 1H), 4.14 (H_{m,m'} and H_{h,h'}, dt, *J* = 8.6, 4.0 Hz, 8H), 3.90 (H_{i,i'} and H_{l,i'}, q, *J* = 6.2, 5.3 Hz, 8H), 3.82 (H_{j,j'} and H_{k,k'}, s, 8H).

¹³**C-NMR** (101 MHz, CDCl₃) δ = 150.09, 148.99, 148.96, 146.34, 133.45, 121.61, 115.45, 114.32, 114.26, 111.27, 105.57, 71.19, 70.00, 69.90, 69.70, 69.47, 69.45, 69.39.

HRMS (ESI+ve) m/z: 512.20007 ([M+Na]⁺, C₂₄H₃₁N₃O₈ requires 512.20034)



Figure S8 ¹³C-NMR spectrum of 5 (101 MHz, CDCl₃)



Figure S9 HRMS spectrum of 5

1.4 Synthesis of Azido-catenane (8·PF₆)



A solution of N₃-dibenzo[24]crown-8 (100 mg, 0.204 mmol) in dry and degassed DCM (100 ml) was added the bis-vinyl thread (139 mg, 0.204 mmol). After stirring at room temperature for 30 minutes, the first-generation Grubbs' catalyst (35 mg, 0.043 mmol) was added to a reaction mixture and refluxed at 40°C for 48 hours. The crude mixture was stirred with activated charcoal (1 g) then filtered through Celite® and the organic residue was collected. The mixture was purified by silica gel column chromatography (5% MeOH/DCM) followed by passing through the size exclusion chromatography to obtain the desire product in 16% yield.

¹**H-NMR** (500 MHz, 1:1 CDCl₃/MeOD) δ = 7.30 (H_y, dd, *J* = 8.2, 2.0 Hz, 4H), 7.19 (H_x, dd, *J* = 8.0, 3.1 Hz, 4H), 6.91 – 6.87 (H_d and H_g,m, 2H), 6.86 – 6.75 (H_e,H_f, and H_a m, 3H), 6.60 (H_b, ddd, *J* = 8.7, 4.0, 2.5 Hz, 1H), 6.44 (H_c, dd, *J* = 4.1, 2.5 Hz, 1H), 5.33 – 5.22 (H_u, m, 2H), 4.41 (H_w, d, *J* = 3.6 Hz, 4H), 4.18 – 4.03 (H_{crown}, m, 8H), 3.84 – 3.70 (H_{crown}, m, 8H), 3.54 – 3.35 (H_{crown} and H_v, m, 12H), 1.90 (H_{methylene}, dq, *J* = 16.8, 6.3 Hz, 4H), 1.58 (H_{methylene}, hept, *J* = 7.4 Hz, 5H), 1.38 – 1.32 (H_{methylene}, m, 4H), 1.31 – 1.18 (H_{methylene}, m, 18H).

Note

- 1. H_z proton signals are hidden under the peak of water in MeOD at the chemical shift around 4.6 ppm.
- H_{crown} represents proton signals of methylene protons derived from the dibenzo[24]crown-8 macrocycle.

¹³**C-NMR** (126 MHz, 1:1 CDCl₃/MeOD) δ = 149.20, 149.19, 147.98, 147.95, 147.93, 145.68, 145.66, 140.78, 140.75, 134.39, 134.35, 131.52, 130.92, 130.31, 130.03, 129.94, 128.47,

128.39, 122.45, 122.42, 122.40, 114.30, 113.36, 113.34, 113.31, 113.29, 111.83, 111.80, 105.10, 105.07, 72.64, 72.59, 71.29, 71.23, 71.19, 70.92, 70.89, 70.83, 70.77, 70.71, 70.68, 69.30, 69.24, 68.97, 68.93, 68.91, 68.86, 68.81, 52.79, 32.97, 30.39, 30.24, 30.20, 30.16, 30.06, 29.88, 29.71, 29.67, 29.25, 27.71, 26.97, 26.88.

HRMS (ESI+ve) m/z: 995.60962 ([M-PF₆]⁺, C₅₈H₈₃N₄O₁₀ requires 995.61037)



Figure S10 ¹H-NMR spectrum of (8·PF₆) (400 MHz, CDCl₃)



Figure S11 ¹³C-NMR spectrum of (8·PF₆) (126 MHz, CDCl₃)

Further evidence of the interlocked nature of $8 \cdot PF_6$ was observed by ¹H-¹H ROESY NMR spectroscopy (Figure S12), in which through space interactions between proton resonances of the two macrocycles (H_a and H_c from the dibenzyl ammonium macrocycle with H_{crown ether}) are observed. In addition, strong through space coupling between H_c and methylene protons from the crown ether group indicates that the secondary ammonium group is stabilised in the cavity of dibenzo[24]crown-8.



Figure S12 ROESY ¹H-NMR spectrum of the azido-[2]catenane **8**·**PF**₆ in 1:1 CD₃OD/CDCl₃ (298K, 500 MHz).



Figure S13 HRMS spectrum of 8.PF₆

1.5 Synthesis of Ammonium catenane functionalised XB Receptor (1·(PF₆)₂)



A mixture of Cu(MeCN)₄PF₆ (2.3 mg, 0.006 mmol) and TBTA (2.4 mg, 0.005 mmol) was stirred in dry and degassed DCM (0.5 ml) at room temperature for 15 minutes. 3,5-Diiodoethynylpyridine (5.7 mg, 0.015 mmol) was added to a solution mixture, followed by azidocatenane **8·PF₆** (36 mg, 0.036 mmol). The reaction was stirred at room temperature for 72 hours prior to purify by preparative plate chromatography in 3% MeOH/DCM to afford a pure product in 85% yield after anion exchange to PF_6^- salt.

¹**H-NMR** (500 MHz, 1:1 CDCl₃/MeOD) δ = 7.37(m, 1H), 7.31 (d, *J* = 7.8 Hz, 8H), 7.24 – 7.14 (m, 8H), 7.10 (s, 3H), 6.99 (d, *J* = 7.2 Hz, 4H), 6.87 (q, *J* = 4.5 Hz, 5H), 6.74 (dd, *J* = 6.5, 3.7 Hz, 4H), 5.26 (dt, *J* = 9.7, 4.3 Hz, 4H), 4.68 (d, *J* = 3.0 Hz, 8H), 4.39 (d, *J* = 3.5 Hz, 8H), 4.21 (d, *J* = 22.7 Hz, 8H), 4.05 (q, *J* = 6.2, 5.7 Hz, 8H), 3.91 (s, 8H), 3.73 (t, *J* = 7.5 Hz, 8H), 3.61 – 3.51 (m, 8H), 3.51 – 3.43 (m, 8H), 3.40 (dt, *J* = 11.4, 6.1 Hz, 8H), 1.89 (dq, *J* = 16.8, 6.2 Hz, 8H), 1.55 (p, *J* = 6.5 Hz, 8H), 1.37 – 1.10 (m, 40H).

¹³**C-NMR** (126 MHz, 1:1 CDCl₃/MeOD) δ 149.76, 148.46, 147.62, 140.74, 131.36, 130.84, 130.67, 130.24, 129.84, 129.75, 128.46, 128.39, 122.29, 120.30, 113.01, 111.75, 72.55, 72.49, 71.32, 71.09, 70.86, 70.81, 70.67, 69.21, 68.62, 52.73, 32.90, 30.32, 30.16, 30.13, 30.09, 29.99, 29.87, 29.81, 29.64, 29.59, 29.19, 27.65, 26.90, 26.81.

HRMS (ESI+ve) m/z: 1185.02466 ([M-2PF₆]²⁺, C₁₂₅H₁₆₉N₉O₂₀I₂ requires 1185.02811)





Figure S15 ¹³C-NMR spectrum of 1·(PF₆)₂ (101 MHz, 1:1 CDCl₃/MeOD)



Figure S16 HRMS spectrum of 1 · (PF₆)₂



2. Pseudorotaxane Complexation of 5 with $6 \cdot PF_6$ in d_2 -dichloromethane

Figure S17 Comparative truncated ¹H-NMR spectra of (a) a thread $\mathbf{6} \cdot \mathsf{PF}_6$, (b) an equimolar mixture of $\mathbf{6} \cdot \mathsf{PF}_6$ and $\mathbf{5}$, and (c) a macrocycle $\mathbf{5}$ in d_2 -dichloromethane at room temperature.

3. Anion binding in organic and aqueous-organic mixture

Spectra for ¹H NMR titrations were recorded at 293 K on a Varian Unity Plus 500 spectrometer with ¹H operating at 500 MHz. Initial sample volumes were 0.50 mL and concentrations were 2.0 mM of host in all cases. Anion solution (100 mM) as the tetrabutylammonium salts were added in aliquots, the samples thoroughly shaken and spectra recorded. Spectra were recorded at 0, 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.5, 3.0, 4.0, 5.0, 7.0 and 10 equivalents of anion. In all cases where association constants were calculated, bound and unbound species were found to be in fast exchange on the NMR timescale. In all cases where association constants were found to be in fast exchange on the NMR timescale. In all cases where association constants were found to be in fast exchange and unbound species were found to be in fast exchange on the NMR timescale. In all cases where association constants were calculated, bound and unbound species were found to be in fast exchange on the NMR timescale. In all cases where association constants were obtained by analysis of the resulting binding isotherm data monitoring aryl proton a using WinEQNMR2 and Bindfit v.05.⁵⁻⁷



Figure S18 Truncated ¹H-NMR spectra of $1 \cdot (PF_6)_2$ in 5%D₂O/d₆-acetone upon addition of TBAI (T=298K, 500 MHz).



Figure S19 Anion binding isotherm of $1 \cdot (PF_6)_2$, [host] = 2mM and [guest] = 100 mM (5% D₂O/ d₆-Acetone, 500MHz, 298K)



Figure S20 Truncated ¹H-NMR spectra of **XB9**·2**NaPF**₆ in $5\%D_2O/d_6$ -acetone upon addition of TBAI (T=298K, 500 MHz), counter-anions in the structure are omitted for clarity.



Figure S21 Anion binding isotherm of $XB9 \cdot 2NaPF_6$, [host] = 2mM and [guest] = 100 mM (5% D₂O/d₆-Acetone, 500MHz, 298K)



Figure S22 Truncated ¹H-NMR spectra of **HB10·2NaPF**₆ in $5\%D_2O/d_6$ -acetone upon addition of TBAI (T=298K, 500 MHz), counter-anions in the structure are omitted for clarity.



Figure S23 Anion binding isotherm of $HB10 \cdot 2NaPF_6$, [host] = 2mM and [guest] = 100 mM (5% D₂O/ d₆-Acetone, 500MHz, 298K)

- 4. Comparative anion binding studies of XB11
 - 4.1 Synthesis of XB11



A mixture of $Cu(MeCN)_4PF_6$ (15 mg, 0.04 mmol) and TBTA (7 mg, 0.013 mmol) was stirred in dry and degassed THF (1.0 ml) at room temperature for 15 minutes. 3,5-Diiodoethynylpyridine (50.3 mg, 0.013 mmol) was added to a solution mixture, followed by an azide **5** (143 mg, 0.29 mmol). The reaction was stirred at room temperature for 48 hours prior to purify by diluting with DCM, stirring with EDTA solution to remove copper residues, and washing the organic solution with water twice. After concentrating the solution, MeOH was added to crash out the product. The solid residue was collected by a vacuum filtration and washed with cold MeOH. The product was obtained as an off-white solid in 44 %yield.

¹**H-NMR** (500 MHz, CDCl₃) δ = 9.34 (H_b, d, J = 2.1 Hz, 2H), 8.98 (H_a, t, J = 2.1 Hz, 1H), 7.17 – 6.96 (H_c, H_d, and H_e, 6H), 6.96 – 6.85 (H_f, H_g, H_h, and H_i, 8H), 4.31 – 4.13 (m, 8H), 4.04 – 3.89 (m, 8H), 3.85 (m, 16H).

¹³**C-NMR** (126 MHz, CDCl₃) δ = 150.52, 149.19, 149.00, 148.50, 147.42, 126.39, 121.67, 119.67, 114.23, 71.44, 70.03, 69.79, 69.48 ppm.

HRMS (ESI +ve) m/z: 1380.2457 ([M+Na]⁺, C₅₇H₆₅N₇O₁₆I₂Na⁺ requires 1380.2469).



Figure S25 ¹³C-NMR spectrum of XB11 (126 MHz, CDCl₃)





Figure S26 Stacking of truncated ¹H-NMR spectra of (a) **XB11**, (b) **XB11**[.]**2DBAPF**₆ and (c) **2DBAPF**₆ · in 1:1 CDCl₃:CD₃CN, (400 MHz, 298 K)

4.3 Anion Binding Studies of XB11





Figure S27 Truncated ¹H-NMR titration spectra of **XB11** upon addition of 0-10 equivalents of TBAI (500 MHz, 298 K, 1:1 CDCI₃:CD₃CN, [XB11] = 1.0 mM).



Figure S28 Changes in chemical shift of the internal pyridine proton b of **XB11** upon addition of halide anions as their TBA salts (1:1 CDCI₃:CD₃CN, 298K). Experimental titration data (squares, circles), and fitted binding isotherms (solid lines) by *Bindfit*.

4.4 Anion Binding Studies of XB11 complexed with DBAPF₆



Figure S29 Truncated ¹H-NMR titration spectra of **XB11** \cdot **2DBAPF**₆ upon addition of 0-10 equivalents of TBABr (500 MHz, 298 K, 1:1 CDCl₃:CD₃CN, [XB11 + 2DBA \cdot PF6] = 1.0 mM).



Figure S30 Truncated ¹H-NMR titration spectra of **XB11** \cdot **2DBAPF**₆ upon addition of 0-10 equivalents of TBAI (500 MHz, 298 K, 1:1 CDCI₃:CD₃CN, XB11 + 2DBA \cdot PF6] = 1.0 mM).

Table S1 Anion association constants of **XB11** and **XB11** complexed with 2 equivalents of **DBA·PF**₆ (**XB11·2DBAPF**₆) in 1:1 CDCl₃/CD₃CN, 298 K.^{a,b}

Anion	Association Constant (K _a , M ⁻¹) ^a		
Anion	XB11	XB11·2DBAPF ₆	
Br⁻	3640 ^b	779°	
I-	853	1752 ^d	

 a K_a values obtained by fitting the binding isotherm (monitored chemical shift changes of H_a) to a host-guest 1:1 stoichiometric binding model using Bindfit v.05 error (± 10%); each anion added as its tetrabutylammonium (TBA) salt. b Bindfit v.05 error (± 15%); Quantitative binding data could not be determined from Bindfit analysis of the TBACI titration isotherms.

^c Binding data can be access via http://app.supramolecular.org/bindfit/view/d5be64e8-5869-422f-9190-dac8e466bd86

^d Binding data can be access via <u>http://app.supramolecular.org/bindfit/view/0e0e364b-4641-4632-b7f0-08d640115bea</u>

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