

Supporting Information

## Electrochemical Decarboxylative Alkylation of $\beta$ -Keto Acids with Phenol Derivatives

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### 1. General information

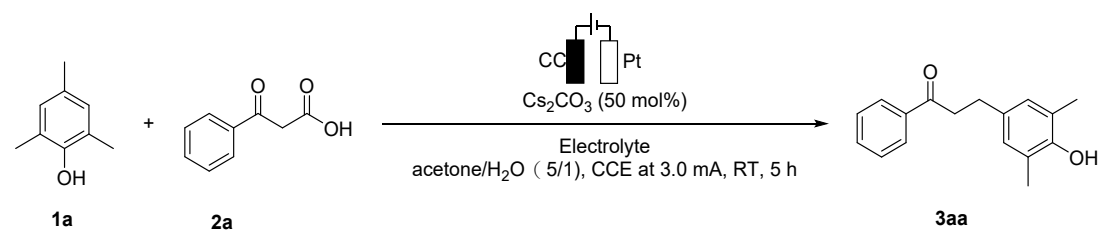
$^1\text{H}$ -NMR,  $^{13}\text{C}$ -NMR and  $^{19}\text{F}$ -NMR spectra were recorded with solvent  $\text{CDCl}_3$  on Bruker avance DPX 400 spectrometer (400 MHz for  $^1\text{H}$ , 101 MHz for  $^{13}\text{C}$ ). Chemical shifts were reported in parts per million (ppm), and the residual solvent peak was used as an internal reference: proton (chloroform  $\delta$  7.26), carbon (chloroform  $\delta$  77.16). Multiplicity was indicated as follows: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), dd (doublet of doublet), td (doublet of triplet).

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Exact ESI mass spectra were recorded on a Bruker Daltonics MicroTOF-Q. ESI-MS were obtained on a Thermo-ITQ. Mass spectral data (MS) was recorded using an Agilent-6110 mass spectrometer. For thin layer chromatography (TLC), pre-coated Qingdao Haiyang TLC plates (GF254) were used, and compounds were visualized with a UV light at 254 nm. Flash chromatographic separations were performed on 200-300 mesh silica gel (from Qingdao Haiyang Chem. Company, Ltd.). Unless otherwise noted, all reagents were purchased from commercial sources (Adamas, Energy, Aldrich) and used as received without further purification. Solvents were dried and purified according to the procedure from “Purification of Laboratory Chemicals book”. The final cross-coupling reaction was carried out in an open vial under ambient condition.

## 2. Optimization of The Reaction Conditions

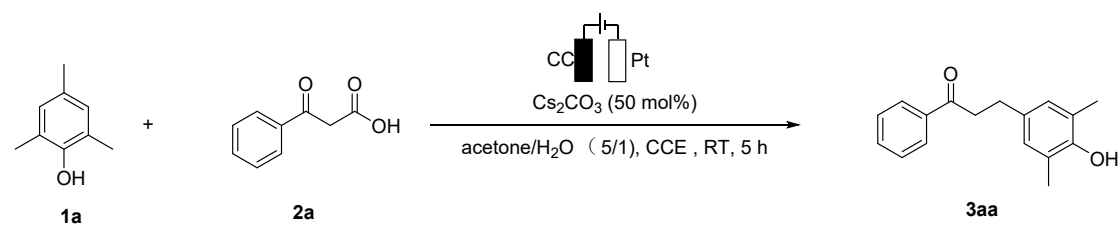
**Table S1: Evaluation of electrolyte** <sup>[a]</sup>



Entry	Electrolyte	Yield(%) <sup>[b]</sup>
1	<i>n</i> -Bu <sub>4</sub> NPF <sub>6</sub>	52%
2	<i>n</i> -Bu <sub>4</sub> NBF <sub>4</sub>	50%
3	none	<b>53%</b>

<sup>a</sup>Reaction conditions: undivided cell, Graphite anode, Pt cathode, **1a** (0.3 mmol), **2a** (0.6 mmol),  $\text{Cs}_2\text{CO}_3$  (0.5 equiv.), electrolyte (0.1 M), acetone/ $\text{H}_2\text{O}$  (5/1, 6 mL), CCE = 3.0 mA, 5 h, RT, under air. <sup>b</sup>Yield is that of the isolated product. CCE = constant current electrolysis.

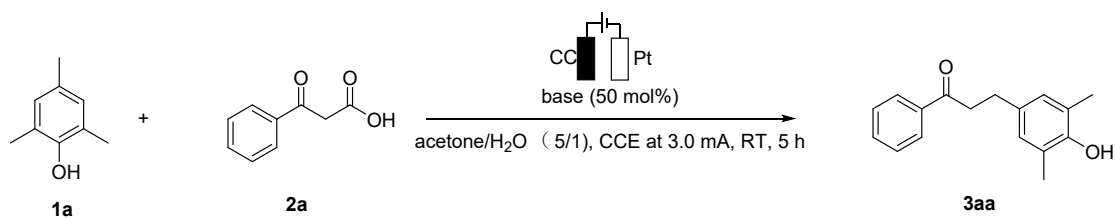
**Table S2: Evaluation of electric current** <sup>[a]</sup>



Entry	Electric current (mA)	Yield (%) <sup>[b]</sup>
1	0	N.R
2	1.0	38
3	3.0	<b>53</b>
4	5.0	39
7	7.0	41

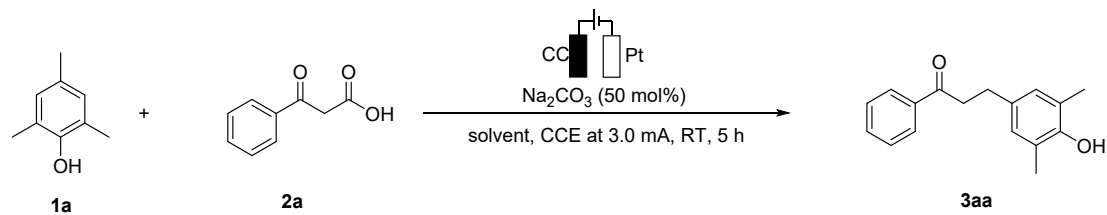
<sup>a</sup>Reaction conditions: undivided cell, Graphite anode, Pt cathode, **1a** (0.3 mmol), **2a** (0.6 mmol),  $\text{Cs}_2\text{CO}_3$  (0.5 equiv.), acetone/ $\text{H}_2\text{O}$  (5/1, 6 mL), CCE, 5 h, RT, under air. <sup>b</sup>Yield is that of the isolated product. CCE = constant current electrolysis.

**Table S3: Evaluation of base<sup>[a]</sup>**



Entry	Base	Yield(%) <sup>[b]</sup>
1	Cs <sub>2</sub> CO <sub>3</sub> as base	55
2	K <sub>2</sub> CO <sub>3</sub> as base	46
3	Na <sub>2</sub> CO <sub>3</sub> as base	<b>64</b>
4	NaOAc as base	35
5	NaHCO <sub>3</sub> as base	57
6	NaH <sub>2</sub> PO <sub>4</sub> as base	33
7	DABCO as base	33
8	DBU as base	28
9	without base	45

<sup>a</sup>Reaction conditions: undivided cell, Graphite anode, Pt cathode, **1a** (0.3 mmol), **2a** (0.6 mmol), Base (0.5 equiv.), acetone/H<sub>2</sub>O (5/1, 6 mL), CCE = 3.0 mA, 5 h, RT, under air. <sup>b</sup>Yield is that of the isolated product. CCE = constant current electrolysis.

**Table S4: Evaluation of solvents<sup>[a]</sup>**

Entry	Solvent (mL)	Yield(%) <sup>[b]</sup>
1	acetone /H <sub>2</sub> O (5/1)	65
2	acetone /H <sub>2</sub> O (4/2)	60
3	acetone (6)	35
4	HFIP/H <sub>2</sub> O (5/1)	45
5	THF/H <sub>2</sub> O (5/1)	46
6	1,4-Dioxane/H <sub>2</sub> O(5/1)	50
7	MeCN/H <sub>2</sub> O (5/1)	65
8	MeCN/H <sub>2</sub> O (4/1)	60
10	MeCN/H <sub>2</sub> O (3/1)	<b>78</b>
12	MeCN/H <sub>2</sub> O (2/1)	60
13	MeCN/H <sub>2</sub> O (1/1)	60
14	MeCN (4)	42
15	H <sub>2</sub> O (4)	N.R

<sup>a</sup>Reaction conditions: undivided cell, Graphite anode, Pt cathode, **1a** (0.3 mmol), **2a** (0.6 mmol),  $\text{Na}_2\text{CO}_3$  (0.5 equiv.), Solvent (xx mL), CCE = 3.0 mA, 5 h, RT, under air. <sup>b</sup>Yield is that of the isolated product. HFIP = 1,1,1,3,3,3-hexafluoro-2-propanol, CCE = constant current electrolysis.

**Table S5: Evaluation of electrode<sup>[a]</sup>**

Entry	Electrode	Yield(%) <sup>[b]</sup>
1	Graphite(+), Pt(-)	78
2	Pt(+), Pt(-)	65
3	Graphite (+), Graphite (-)	62

<sup>a</sup>Reaction conditions: undivided cell, xx anode, xx cathode, **1a** (0.3 mmol), **2a** (0.6 mmol), Na<sub>2</sub>CO<sub>3</sub> (0.5 equiv.), MeCN/H<sub>2</sub>O (3/1, 4 mL), CCE = 3.0 mA, 5 h, RT, under air. <sup>b</sup>Yield is that of the isolated product. CCE = constant current electrolysis.

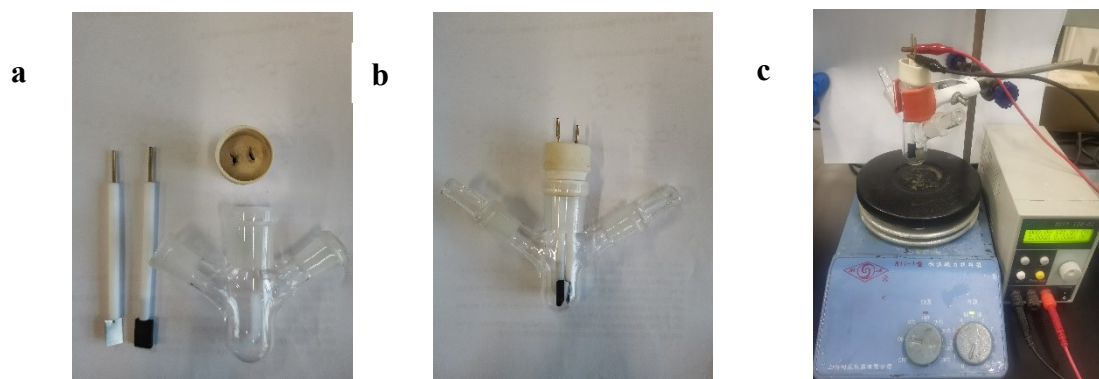
**Table S6: Evaluation of the conductivity of different kind of solutions<sup>[a]</sup>**

Entry	Variation of mixture	Description	Conductivity <sup>[b]</sup>
1	MeCN/H <sub>2</sub> O	solvents	13.87 μS/cm
2	MeCN/H <sub>2</sub> O + <i>n</i> -Bu <sub>4</sub> NPF <sub>6</sub>	with electrolyte	6250 μS/cm
3	MeCN/H <sub>2</sub> O + Na <sub>2</sub> CO <sub>3</sub>	with Na <sub>2</sub> CO <sub>3</sub>	516 μS/cm
4	MeCN/H <sub>2</sub> O + <i>n</i> -Bu <sub>4</sub> NPF <sub>6</sub> + Na <sub>2</sub> CO <sub>3</sub>	with electrolyte and Na <sub>2</sub> CO <sub>3</sub>	9300 μS/cm
5	MeCN/H <sub>2</sub> O + <b>1a</b> + <b>2a</b>	without Na <sub>2</sub> CO <sub>3</sub>	225 μS/cm
6	MeCN/H <sub>2</sub> O + <b>1a</b> + <b>2a</b> + Na <sub>2</sub> CO <sub>3</sub>	without electrolyte	1855 μS/cm
7	MeCN/H <sub>2</sub> O + <b>1a</b> + <b>2a</b> + <i>n</i> -Bu <sub>4</sub> NPF <sub>6</sub> + Na <sub>2</sub> CO <sub>3</sub>	with electrolyte and Na <sub>2</sub> CO <sub>3</sub>	10110 μS/cm

[a] Measure conditions: **1a** (0.8 mmol), **2a** (1.6 mmol), Na<sub>2</sub>CO<sub>3</sub> (0.5 equiv.), MeCN /H<sub>2</sub>O (3/1, 4 mL) · *n*-Bu<sub>4</sub>NPF<sub>6</sub> (0.1 M). Conductivity was measured by DDSJ-308F Conductivity meter. [b] The data of each entry was measured three times and averaged.

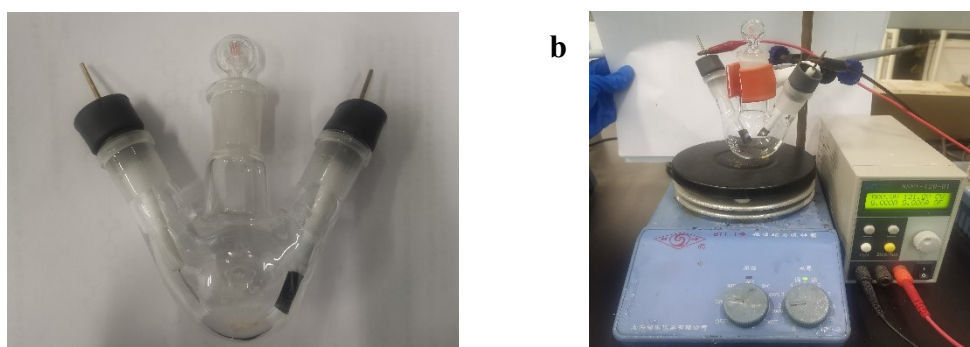
### 3. General procedure

#### 3.1 Graphical guide for the set-up



As experimental set-up, a graphite electrode (10 mm×15 mm×3 mm), a platinum plate electrode (10 mm×15mm×0.1 mm), rubber plugs, an undivided three-necked bottle and Hansheng Puyuan Programmable DC Power Supplies (HPYS-120-01) were used.

**Figure S1. Setup for milligram scale photoelectrocatalytic reactions. a) Electrodes; b) Assembled reactor; c) Reaction in progress.**



**Figure S2. Setup for the gram scale synthesis of 3. a) Assembled reaction setup. b) Reaction in progress.**

#### 3.2 Substrate preparation

All the  $\beta$ -ketoacids derivatives<sup>[1-2]</sup> and phenol derivatives<sup>[3-4]</sup> are known compounds and prepared according to the reported procedure.

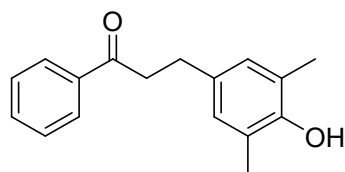
##### Electrochemical Decarboxylative Alkylation





## 5. Characterization data of products

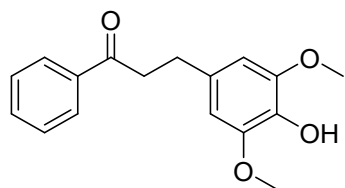
### 3-(4-hydroxy-3,5-dimethylphenyl)-1-phenylpropan-1-one(3aa)



**3aa**

Following the general procedure A. A yellow oily liquid. Isolated yield (PE/EA = 10:1), 79.3 mg, 78%. <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.97 (dd, J = 8.4, 1.3 Hz, 2H), 7.61 – 7.52 (m, 1H), 7.46 (t, J = 7.5 Hz, 2H), 6.87 (s, 2H), 4.62 (s, 1H), 3.30 – 3.22 (m, 2H), 2.98 – 2.90 (m, 2H), 2.23 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 199.72, 150.64, 137.03, 133.16, 132.96, 128.72, 128.64, 128.20, 123.19, 41.08, 29.43, 16.05. HR-MS (ESI-TOF) m/z: [M+Na]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>NaO<sub>2</sub> 277.1204, found 277.1216.

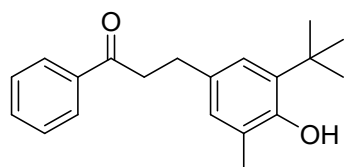
### 3-(4-hydroxy-3,5-dimethoxyphenyl)-1-phenylpropan-1-one (3ba)



**3ba**

Following the general procedure A. A yellow solid. Isolated yield (PE/EA = 10:1), 63 mg, 55%. <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.95 (dd, J = 8.5, 1.4 Hz, 2H), 7.58 – 7.53 (m, 1H), 7.45 (t, J = 7.6 Hz, 2H), 6.46 (s, 2H), 5.43 (s, 1H), 3.86 (s, 6H), 3.31 – 3.24 (m, 2H), 3.00 (d, J = 8.0 Hz, 2H). <sup>13</sup>C NMR (101 MHz, Chloroform-d) δ 199.56, 147.08, 136.97, 133.21, 133.08, 132.50, 128.72, 128.13, 105.10, 56.35, 40.95, 30.54. HR-MS (ESI-TOF) m/z: [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>19</sub>O<sub>4</sub> 287.1283, found 287.1281.

### 3-(3-(tert-butyl)-4-hydroxy-5-methylphenyl)-1-phenylpropan-1-one (3ca)

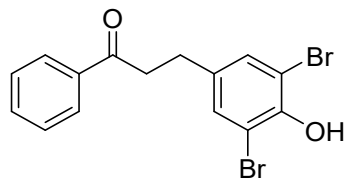


**3ca**

Following the general procedure A. A brownish yellow oily liquid. Isolated yield (PE/EA = 10:1), 81.7 mg, 69%. <sup>1</sup>H NMR (400 MHz, Chloroform-d) δ 7.98 (dd, J = 8.4, 1.4 Hz, 2H), 7.59 – 7.54 (m, 1H), 7.47 (tt, J = 6.7, 1.4 Hz, 2H), 7.02 (d, J = 2.3 Hz, 1H), 6.91 (d, J = 2.1 Hz, 1H), 4.79 (s, 1H),

3.31 – 3.26 (m, 2H), 3.01 – 2.96 (m, 2H), 2.25 (s, 3H), 1.42 (s, 9H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 199.93, 151.13, 137.03, 135.85, 133.13, 132.42, 128.69, 128.57 – 128.31 (m), 128.20, 125.41 – 124.93 (m), 123.26, 41.16, 34.62, 29.90, 29.87, 16.15. HR-MS (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>20</sub>H<sub>25</sub>O<sub>2</sub> 297.1855, found 297.1856.

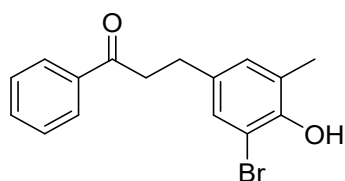
### 3-(3,5-dibromo-4-hydroxyphenyl)-1-phenylpropan-1-one (3da)



**3da**

Following the general procedure A. A light yellow solid. Isolated yield (PE/EA = 10:1), 62.6 mg, 41%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.98 – 7.91 (m, 2H), 7.59 – 7.54 (m, 1H), 7.46 (t, *J* = 7.6 Hz, 2H), 7.34 (s, 2H), 5.84 (s, 1H), 3.26 (t, *J* = 7.5 Hz, 2H), 2.96 (t, *J* = 7.4 Hz, 2H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 198.60, 147.82, 136.72, 136.11, 133.43, 132.10, 128.82, 128.15, 109.81, 40.16, 28.60. HR-MS (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>15</sub>H<sub>13</sub>Br<sub>2</sub>O<sub>4</sub> 382.9282, found 382.9279.

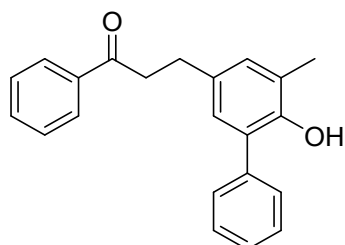
### 3-(3-bromo-4-hydroxy-5-methylphenyl)-1-phenylpropan-1-one (3ea)



**3ea**

Following the general procedure A. A yellow oily liquid. Isolated yield (PE/EA = 10:1), 67.4 mg, 53%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.96 (dd, *J* = 8.4, 1.4 Hz, 1H), 7.61 – 7.52 (m, 1H), 7.50 – 7.39 (m, 2H), 7.19 (d, *J* = 1.3 Hz, 1H), 6.95 (d, *J* = 2.2 Hz, 1H), 5.46 (s, 1H), 3.29 – 3.19 (m, 2H), 2.99 – 2.90 (m, 2H), 2.27 (s, 3H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 199.20, 148.83, 136.86, 134.37, 133.29, 130.72, 129.05, 128.76, 128.16, 125.87, 110.04, 40.61, 29.01, 16.82. HR-MS (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>19</sub>O<sub>4</sub> 287.1283, found 287.1281.

### 3-(6-hydroxy-5-methyl-[1,1'-biphenyl]-3-yl)-1-phenylpropan-1-one (3fa)

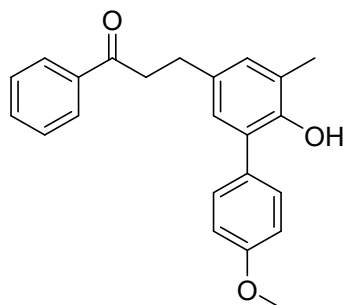


**3fa**

Following the general procedure A. A white solid. Isolated yield (PE/EA = 10:1 to 5:1), 68.7 mg, 54%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.98 (dd, *J* = 8.5, 1.3 Hz, 2H), 7.59 – 7.53 (m, 1H), 7.52 – 7.39 (m, 7H), 7.06 (d, *J* = 2.3 Hz, 1H), 6.98 (d, *J* = 3.0 Hz, 1H), 5.23 (s, 1H), 3.36 – 3.27 (m, 2H),

3.06 – 2.98 (m, 2H), 2.32 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  199.57, 149.03, 137.49, 136.93, 133.16, 132.93, 130.62, 129.42, 129.21, 128.69, 128.17, 127.93, 127.74, 127.64, 124.83, 40.91, 29.43, 16.37. **HR-MS** (ESI-TOF)  $m/z$ :  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{22}\text{H}_{21}\text{O}_2$  317.1542, found 317.1544.

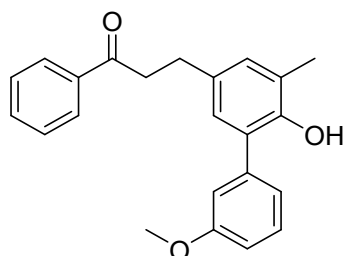
**3-(6-hydroxy-4'-methoxy-5-methyl-[1,1'-biphenyl]-3-yl)-1-phenylpropan-1-one (3ga)**



**3ga**

Following the general procedure A. A brownish yellow oily liquid. Isolated yield (PE/EA = 10:1), 76.2 mg, 55%.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.97 (dd,  $J$  = 8.4, 1.4 Hz, 2H), 7.60 – 7.51 (m, 1H), 7.50 – 7.41 (m, 2H), 7.41 – 7.31 (m, 2H), 7.05 – 6.97 (m, 3H), 6.95 – 6.90 (m, 1H), 5.15 (s, 1H), 3.86 (s, 4H), 3.34 – 3.25 (m, 2H), 3.03 – 2.95 (m, 2H), 2.28 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  199.65, 159.38, 149.13, 136.95, 133.16, 132.84, 130.40, 130.28, 129.53, 128.70, 128.17, 127.66, 127.43, 124.67, 114.83, 55.47, 40.95, 29.45, 16.35. **HR-MS** (ESI-TOF)  $m/z$ :  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{23}\text{H}_{23}\text{O}_3$  347.1647, found 347.1645.

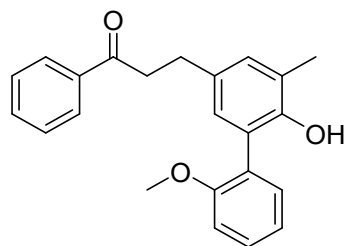
**3-(6-hydroxy-3'-methoxy-5-methyl-[1,1'-biphenyl]-3-yl)-1-phenylpropan-1-one (3ha)**



**3ha**

Following the general procedure A. A light yellow oily liquid. Isolated yield (PE/EA = 10:1), 69.2 mg, 50%.  $^1\text{H}$  NMR (400 MHz, Chloroform-*d*)  $\delta$  7.99 – 7.95 (m, 2H), 7.55 (d,  $J$  = 7.4 Hz, 1H), 7.48 – 7.43 (m, 2H), 7.40 (t,  $J$  = 7.7 Hz, 1H), 7.05 – 7.01 (m, 2H), 6.97 – 6.92 (m, 3H), 5.28 (s, 1H), 3.84 (s, 3H), 3.34 – 3.26 (m, 2H), 3.04 – 2.96 (m, 2H), 2.29 (s, 3H).  $^{13}\text{C}$  NMR (101 MHz, Chloroform-*d*)  $\delta$  199.60, 160.46, 149.03, 138.86, 136.97, 133.20, 132.90, 130.73, 130.60, 128.74, 128.20, 127.56, 127.44, 124.89, 121.28, 114.55, 113.80, 55.47, 40.94, 29.84, 16.39. **HR-MS** (ESI-TOF)  $m/z$ :  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{23}\text{H}_{23}\text{O}_3$  347.1647, found 347.1651.

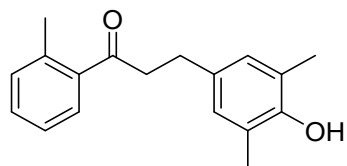
**3-(6-hydroxy-2'-methoxy-5-methyl-[1,1'-biphenyl]-3-yl)-1-phenylpropan-1-one (3ia)**



**3ia**

Following the general procedure A. A yellow oily liquid. Isolated yield (PE/EA = 10:1), 90 mg, 65%.  $^1\text{H NMR}$  (400 MHz, Chloroform-*d*)  $\delta$  7.97 – 7.94 (m, 2H), 7.57 – 7.52 (m, 1H), 7.48 – 7.42 (m, 2H), 7.38 (ddd,  $J = 8.3, 7.4, 1.8$  Hz, 1H), 7.31 (dd,  $J = 7.6, 1.8$  Hz, 1H), 7.09 (td,  $J = 7.5, 1.1$  Hz, 1H), 7.05 (td,  $J = 8.3, 7.9, 1.1$  Hz, 2H), 6.99 – 6.94 (m, 1H), 6.11 (s, 1H), 3.90 (s, 3H), 3.34 – 3.26 (m, 2H), 3.03 – 2.98 (m, 2H), 2.31 (s, 3H).  $^{13}\text{C NMR}$  (101 MHz, Chloroform-*d*)  $\delta$  199.70, 155.58, 150.31, 137.01, 133.16, 132.70, 130.75, 129.37, 128.86, 128.73, 128.20, 127.34, 126.38, 125.89, 122.19, 111.40, 56.17, 40.99, 29.54, 16.70. **HR-MS** (ESI-TOF)  $m/z$ :  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{23}\text{H}_{23}\text{O}_3$  347.1647, found 347.1645.

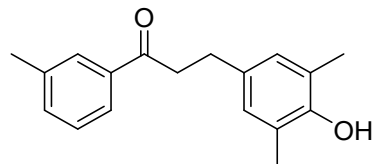
### 3-(4-hydroxy-3,5-dimethylphenyl)-1-(*o*-tolyl)propan-1-one (3ab)



**3ab**

Following the general procedure A. A brown oily liquid. Isolated yield (PE/EA = 10:1), 54.7 mg, 51%.  $^1\text{H NMR}$  (400 MHz, Chloroform-*d*)  $\delta$  7.61 (dd,  $J = 8.0, 1.4$  Hz, 1H), 7.36 (td,  $J = 7.5, 1.4$  Hz, 1H), 7.24 (d,  $J = 5.0$  Hz, 2H), 6.83 (s, 2H), 4.55 (s, 1H), 3.23 – 3.13 (t, 2H), 2.94 – 2.86 (t, 2H), 2.47 (s, 3H), 2.22 (s, 6H).  $^{13}\text{C NMR}$  (101 MHz, Chloroform-*d*)  $\delta$  203.98, 150.61, 138.18, 138.09, 132.81, 132.06, 131.33, 128.62, 128.53, 125.77, 123.16, 43.87, 29.64, 21.37, 16.04. **HR-MS** (ESI-TOF)  $m/z$ :  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{18}\text{H}_{21}\text{O}_2$  269.1542, found 269.1535.

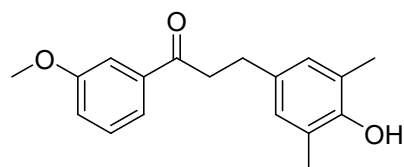
### 3-(4-hydroxy-3,5-dimethylphenyl)-1-(*m*-tolyl)propan-1-one (3ac)



**3ac**

Following the general procedure A. A brown solid. Isolated yield (PE/EA = 10:1), 72.9 mg, 68%.  $^1\text{H NMR}$  (400 MHz, Chloroform-*d*)  $\delta$  7.82 – 7.75 (m, 2H), 7.43 – 7.31 (m, 2H), 6.89 (s, 2H), 4.89 (s, 1H), 3.31 – 3.22 (m, 2H), 2.99 – 2.91 (m, 2H), 2.42 (s, 3H), 2.26 (s, 6H).  $^{13}\text{C NMR}$  (101 MHz, Chloroform-*d*)  $\delta$  200.14, 150.73, 138.49, 137.01, 133.95, 132.92, 128.77, 128.62, 128.59, 125.44, 123.35, 41.17, 29.50, 21.48, 16.12. **HR-MS** (ESI-TOF)  $m/z$ :  $[\text{M}+\text{H}]^+$  calcd for  $\text{C}_{18}\text{H}_{21}\text{O}_2$  269.1542, found 269.1541.

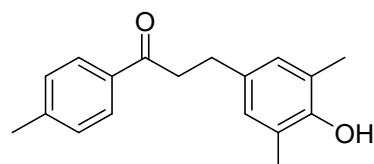
### 3-(4-hydroxy-3,5-dimethylphenyl)-1-(3-methoxyphenyl)propan-1-one (3ad)



**3ad**

Following the general procedure A. A brown oily liquid. Isolated yield (PE/EA = 10:1), 68.2 mg, 60%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.55 – 7.52 (m, 1H), 7.48 (dd, *J* = 2.6, 1.6 Hz, 1H), 7.35 (t, *J* = 7.9 Hz, 1H), 7.11 – 7.06 (m, 1H), 6.85 (s, 2H), 4.63 (s, 1H), 3.84 (s, 3H), 3.27 – 3.19 (m, 2H), 2.96 – 2.88 (m, 2H), 2.22 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 199.62, 159.91, 150.67, 138.35, 132.85, 129.69, 128.61, 123.24, 120.84, 119.69, 112.33 (d, *J* = 4.7 Hz), 55.55 (t, *J* = 4.1 Hz), 41.18, 29.47, 16.07. **HR-MS** (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>19</sub>O<sub>4</sub> 285.1491, found 285.1493.

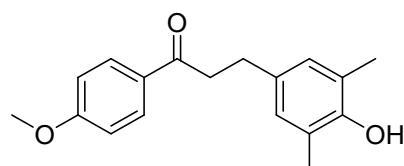
### 3-(4-hydroxy-3,5-dimethylphenyl)-1-(*p*-tolyl)propan-1-one (3ae)



**3ae**

Following the general procedure A. A white solid. Isolated yield (PE/EA = 10:1), 80.4 mg, 75%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.88 (d, *J* = 8.3 Hz, 2H), 7.27 (s, 1H), 7.25 (s, 1H), 6.87 (s, 2H), 4.75 (s, 1H), 3.28 – 3.19 (m, 2H), 2.97 – 2.89 (m, 2H), 2.42 (s, 3H), 2.24 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 199.47, 150.63, 143.92, 134.50, 132.99, 129.38, 128.60, 128.31, 123.22, 40.96, 29.51, 21.74, 16.06. **HR-MS** (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>21</sub>O<sub>2</sub> 269.1542, found 269.1542.

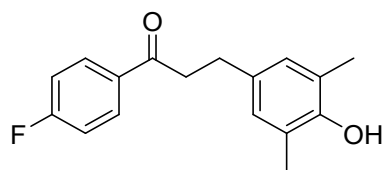
### 3-(4-hydroxy-3,5-dimethylphenyl)-1-(4-methoxyphenyl)propan-1-one (3af)



**3af**

Following the general procedure A. A yellow oily liquid. Isolated yield (PE/EA = 10:1), 87.51 mg, 77%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, *J* = 9.0 Hz, 1H), 6.93 (d, *J* = 8.9 Hz, 2H), 6.86 (s, 2H), 4.62 (s, 1H), 3.87 (s, 3H), 3.24 – 3.16 (m, 2H), 2.96 – 2.88 (m, 2H), 2.23 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 198.33, 163.54, 150.61, 133.12, 130.47, 130.13, 128.63, 123.18, 113.83, 55.60, 40.76, 29.64, 16.06. **HR-MS** (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>18</sub>H<sub>21</sub>O<sub>3</sub> 285.1491, found 285.1491.

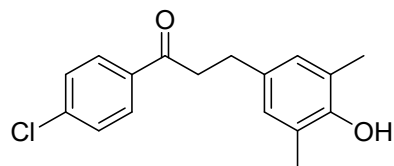
**1-(4-fluorophenyl)-3-(4-hydroxy-3,5-dimethylphenyl)propan-1-one (3ag)**



**3ag**

Following the general procedure A. A light yellow oily liquid. Isolated yield (PE/EA = 10:1), 59.9 mg, 55%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.99 (dd, *J* = 8.9, 5.4 Hz, 2H), 7.12 (t, *J* = 8.6 Hz, 2H), 6.86 (s, 2H), 4.63 (s, 1H), 3.25 – 3.20 (m, 2H), 2.96 – 2.90 (m, 2H), 2.23 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 197.98, 165.65 (d, *J* = 254.5 Hz) 150.52, 133.25 (d, *J* = 3.0 Hz), 132.58, 130.64 (d, *J* = 9.3 Hz), 128.44, 123.08, 115.63 (d, *J* = 21.8 Hz), 40.82, 29.22, 15.90. <sup>19</sup>F-NMR (376 MHz, Chloroform-*d*) δ -105.25 (t, *J* = 6.6 Hz). HR-MS (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>FO<sub>2</sub> 273.1291, found 273.1283.

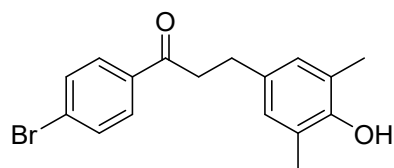
**1-(4-chlorophenyl)-3-(4-hydroxy-3,5-dimethylphenyl)propan-1-one (3ah)**



**3ah**

Following the general procedure A. A light yellow solid. Isolated yield (PE/EA = 10:1), 66.8 mg, 58%. <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.94 – 7.86 (m, 2H), 7.46 – 7.38 (m, 2H), 6.85 (s, 2H), 4.65 (s, 1H), 3.25 – 3.19 (m, 2H), 2.95 – 2.89 (m, 2H), 2.23 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 198.50, 150.69, 139.56, 135.27, 132.65, 129.59, 129.00, 128.59, 123.24, 41.02, 29.31, 16.05. HR-MS (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>ClO<sub>2</sub> 289.0995, found 289.1004.

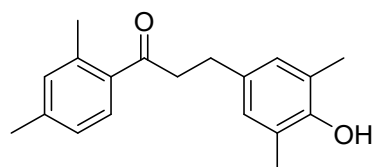
**1-(4-bromophenyl)-3-(4-hydroxy-3,5-dimethylphenyl)propan-1-one (3ai)**



**3ai**

Following the general procedure A. A brown solid. Isolated yield 69.1 mg, 52%. Purification by column chromatography on silica gel (petroleum ether/EtOAc: 15/1) <sup>1</sup>H NMR (400 MHz, Chloroform-*d*) δ 7.86 – 7.78 (m, 2H), 7.63 – 7.56 (m, 2H), 6.85 (s, 2H), 4.58 (s, 1H), 3.25 – 3.17 (m, 2H), 2.96 – 2.88 (m, 2H), 2.23 (s, 6H). <sup>13</sup>C NMR (101 MHz, Chloroform-*d*) δ 198.65, 150.70, 135.72, 132.68, 132.03, 129.73, 128.61, 128.31, 123.23, 41.03, 29.32, 16.06. HR-MS (ESI-TOF) *m/z*: [M+H]<sup>+</sup> calcd for C<sub>17</sub>H<sub>18</sub>BrO<sub>2</sub> 333.0491, found 333.0496.

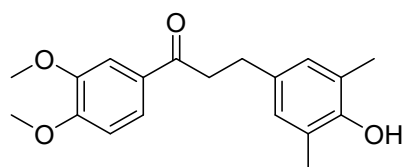
**1-(2,4-dimethylphenyl)-3-(4-hydroxy-3,5-dimethylphenyl)propan-1-one (3aj)**



**3aj**

Following the general procedure A. A brown solid. Isolated yield 79.0 mg, 70%. Purification by column chromatography on silica gel (petroleum ether/EtOAc: 10/1). **<sup>1</sup>H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.58 (d,  $J = 7.7$  Hz, 1H), 7.06 (d,  $J = 8.1$  Hz, 2H), 6.84 (s, 2H), 4.67 (s, 1H), 3.23 – 3.13 (m, 2H), 2.96 – 2.86 (m, 2H), 2.49 (s, 3H), 2.36 (s, 3H), 2.23 (s, 6H). **<sup>13</sup>C NMR** (101 MHz, Chloroform-*d*)  $\delta$  203.21, 150.59, 141.96, 138.78, 134.95, 132.98, 132.92, 129.16, 128.60, 126.39, 123.16, 43.56, 29.80 (d,  $J = 5.7$  Hz), 21.64, 21.46, 16.03. **HR-MS** (ESI-TOF)  $m/z$ :  $[M+H]^+$  calcd for C<sub>17</sub>H<sub>19</sub>O<sub>4</sub> 283.1698, found 283.1701.

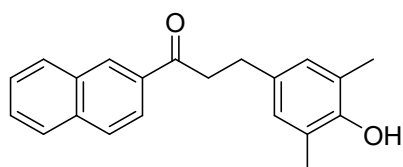
**1-(3,4-dimethoxyphenyl)-3-(4-hydroxy-3,5-dimethylphenyl)propan-1-one (3ak)**



**3ak**

Following the general procedure A. A white solid. Isolated yield 81.7 mg, 65%. Purification by column chromatography on silica gel (petroleum ether/EtOAc: 15/1). **<sup>1</sup>H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.58 (dd,  $J = 8.4, 2.0$  Hz, 1H), 7.52 (d,  $J = 2.0$  Hz, 1H), 6.87 (s, 1H), 6.86 (s, 2H), 4.57 (s, 1H), 3.93 (s, 3H), 3.92 (s, 3H), 3.23 – 3.16 (m, 2H), 2.95 – 2.87 (m, 2H), 2.22 (s, 6H). **<sup>13</sup>C NMR** (101 MHz, Chloroform-*d*)  $\delta$  198.36, 153.29, 150.63, 149.09, 133.08, 130.26, 128.63, 123.18, 122.79, 110.22 (d,  $J = 5.6$  Hz), 110.08, 56.14, 40.66, 29.75, 16.06. **HR-MS** (ESI-TOF)  $m/z$ :  $[M+H]^+$  calcd for C<sub>19</sub>H<sub>23</sub>O<sub>4</sub> 315.1696, found 315.1600.

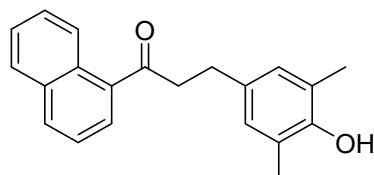
**3-(4-hydroxy-3,5-dimethylphenyl)-1-(naphthalen-2-yl)propan-1-one (3al)**



**3al**

Following the general procedure A. A yellow brown oily liquid. Isolated yield 86.4 mg, 71%. Purification by column chromatography on silica gel (petroleum ether/EtOAc: 15/1). **<sup>1</sup>H NMR** (400 MHz, Chloroform-*d*)  $\delta$  8.47 (s, 1H), 8.05 (dd,  $J = 8.6, 1.8$  Hz, 1H), 7.94 (d,  $J = 8.2$  Hz, 1H), 7.91 – 7.85 (m, 2H), 7.64 – 7.51 (m, 2H), 6.91 (s, 2H), 4.75 (s, 1H), 3.44 – 3.35 (m, 2H), 3.05 – 2.96 (m, 2H), 2.25 (s, 6H). **<sup>13</sup>C NMR** (101 MHz, Chloroform-*d*)  $\delta$  199.70, 150.69, 135.66, 134.29, 132.94, 132.62, 129.86, 129.65, 128.64, 128.54, 127.88, 126.86, 123.98, 123.27, 41.18, 29.56, 16.08. **HR-MS** (ESI-TOF)  $m/z$ :  $[M+H]^+$  calcd for C<sub>21</sub>H<sub>21</sub>O<sub>2</sub> 305.1542, found 305.1546.

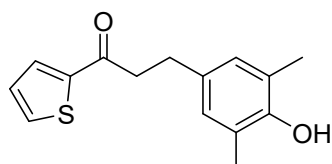
**3-(4-hydroxy-3,5-dimethylphenyl)-1-(naphthalen-2-yl)propan-1-one (3am)**



**3am**

Following the general procedure A. A light brown oily liquid. Isolated yield 76.6 mg, 63%. Purification by column chromatography on silica gel (petroleum ether/EtOAc: 15/1). **<sup>1</sup>H NMR** (400 MHz, Chloroform-*d*)  $\delta$  8.54 (d,  $J = 8.6$  Hz, 1H), 7.97 (d,  $J = 7.8$  Hz, 1H), 7.87 (d,  $J = 6.3$  Hz, 1H), 7.82 (d,  $J = 7.2$  Hz, 1H), 7.62 – 7.55 (m, 1H), 7.55 – 7.51 (m, 1H), 7.47 (t,  $J = 7.7$  Hz, 1H), 6.85 (s, 2H), 4.65 (s, 1H), 3.33 (t,  $J = 7.7$  Hz, 2H), 3.00 (t,  $J = 7.7$  Hz, 2H), 2.21 (s, 6H). **<sup>13</sup>C NMR** (101 MHz, Chloroform-*d*)  $\delta$  204.25, 150.64, 136.16, 134.04, 132.97 – 132.26 (m), 130.20, 128.80 (d,  $J = 11.6$  Hz), 128.69 – 128.23 (m), 128.23 – 127.70 (m), 127.73 – 127.15 (m), 126.52, 126.02, 125.77, 124.82 – 123.99 (m), 123.19, 44.46, 29.96, 16.02 (d,  $J = 6.1$  Hz). **HR-MS** (ESI-TOF)  $m/z$ :  $[M+H]^+$  calcd for C<sub>21</sub>H<sub>21</sub>O<sub>2</sub> 305.1542, found 305.1548.

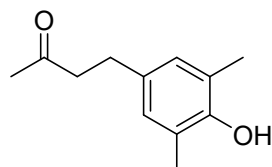
### 3-(4-hydroxy-3,5-dimethylphenyl)-1-(thiophen-2-yl)propan-1-one (3an)



**3an**

Following the general procedure A. A yellow oily liquid. Isolated yield 81.1 mg, 78%. Purification by column chromatography on silica gel (petroleum ether/EtOAc: 15/1). **<sup>1</sup>H NMR** (400 MHz, Chloroform-*d*)  $\delta$  7.70 (dt,  $J = 3.8, 1.0$  Hz, 1H), 7.63 (dt,  $J = 5.0, 1.0$  Hz, 1H), 7.12 (ddd,  $J = 4.8, 3.8, 0.8$  Hz, 1H), 6.86 (s, 2H), 4.73 (s, 1H), 3.22 – 3.15 (m, 2H), 2.97 – 2.91 (m, 2H), 2.23 (s, 6H). **<sup>13</sup>C NMR** (101 MHz, Chloroform-*d*)  $\delta$  192.73, 150.72, 144.31, 133.69, 132.58, 132.01, 128.61, 128.22, 123.27, 41.77, 29.72, 16.07. **HR-MS** (ESI-TOF)  $m/z$ :  $[M+H]^+$  calcd for C<sub>17</sub>H<sub>19</sub>O<sub>4</sub> 261.0949, found 261.0948.

### 4-(4-hydroxy-3,5-dimethylphenyl)butan-2-one (3ao)



**3ao**

Following the general procedure A. A light-yellow oily liquid. Isolated yield 23 mg, 31%. Purification by column chromatography on silica gel (petroleum ether/EtOAc: 15/1). **<sup>1</sup>H NMR** (400 MHz, Chloroform-*d*)  $\delta$  6.79 (s, 2H), 4.86 (s, 1H), 2.81 – 2.67 (m, 4H), 2.22 (s, 6H), 2.14 (s, 3H). **<sup>13</sup>C NMR** (101 MHz, Chloroform-*d*)  $\delta$  208.89, 150.66, 132.48, 128.44, 123.26, 45.75, 30.18, 29.00, 16.05. **HR-MS** (ESI-TOF)  $m/z$ :  $[M+Na]^+$  calcd for C<sub>12</sub>H<sub>16</sub>O<sub>2</sub>Na 215.1048, found 215.1056.



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## 6. Cyclic voltammetry studies and control experiments

### 6.1 Cyclic voltammetry studies

Unless otherwise noted, the cyclic voltammograms were recorded on a CHI 760E instrument using a graphite sheet as the working electrode (10\*15 \*3 mm), a platinum sheet auxiliary electrode (10\*15\*0.1 mm) and a SCE as reference electrode,.Scan rate is 100 mV/s.

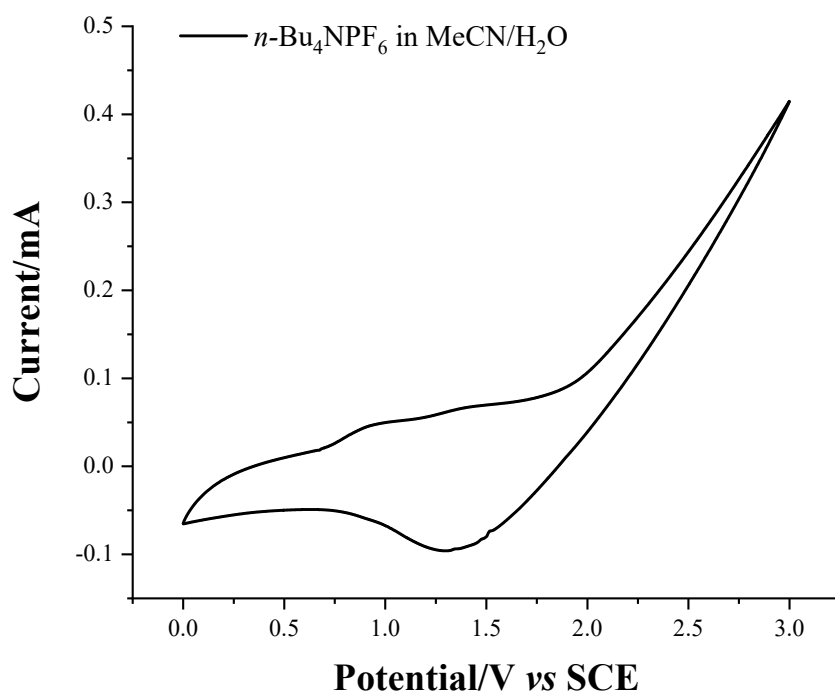


Figure S3. Cyclic voltammogram of  $n\text{-Bu}_4\text{NPF}_6$  (0.1 M) in MeCN/H<sub>2</sub>O (3/1).

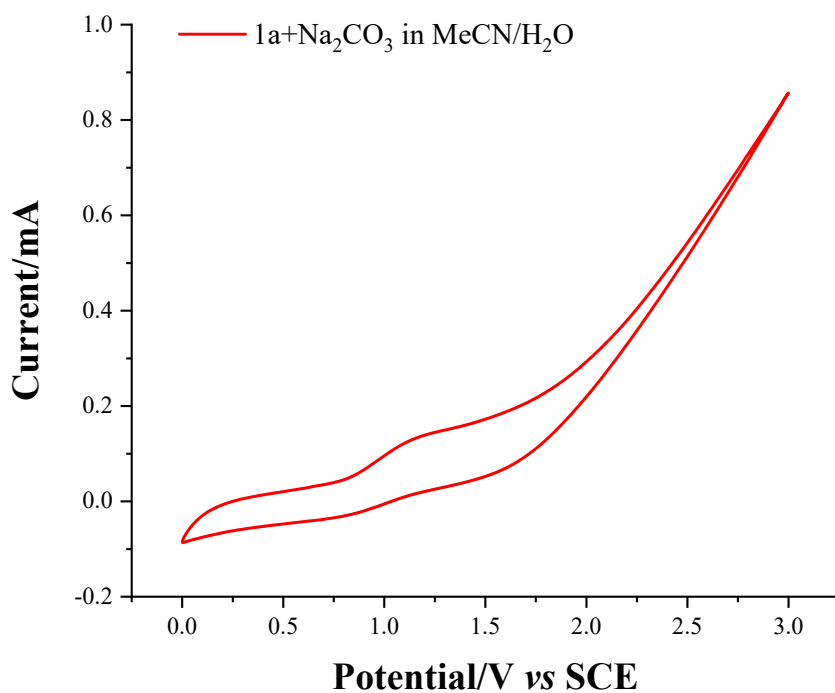


Figure S4. Cyclic voltammogram of **1a** (0.8 mmol) in an electrolyte of *n*-Bu<sub>4</sub>NPF<sub>6</sub> (0.1 M) and Na<sub>2</sub>CO<sub>3</sub> (0.4 mmol) in MeCN/H<sub>2</sub>O (3/1).  $E_{\text{oxi}} = 1.18$  V vs. SCE

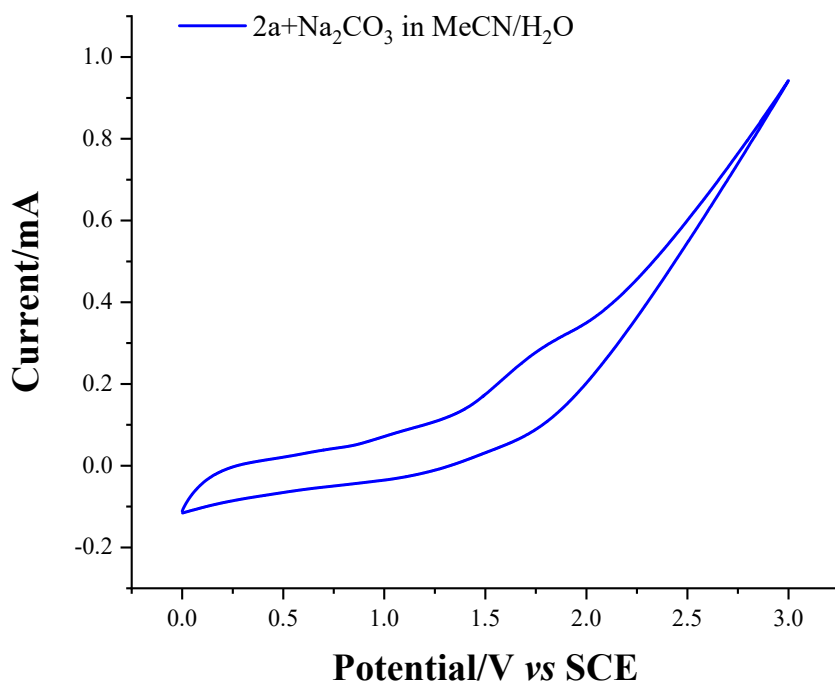


Figure S5. Cyclic voltammogram of **2a** (0.8 mmol) in an electrolyte of *n*-Bu<sub>4</sub>NPF<sub>6</sub> (0.1 M) and Na<sub>2</sub>CO<sub>3</sub> (0.4 mmol) in MeCN/H<sub>2</sub>O (3/1).  $E_{\text{oxi}} = 1.76$  V vs. SCE

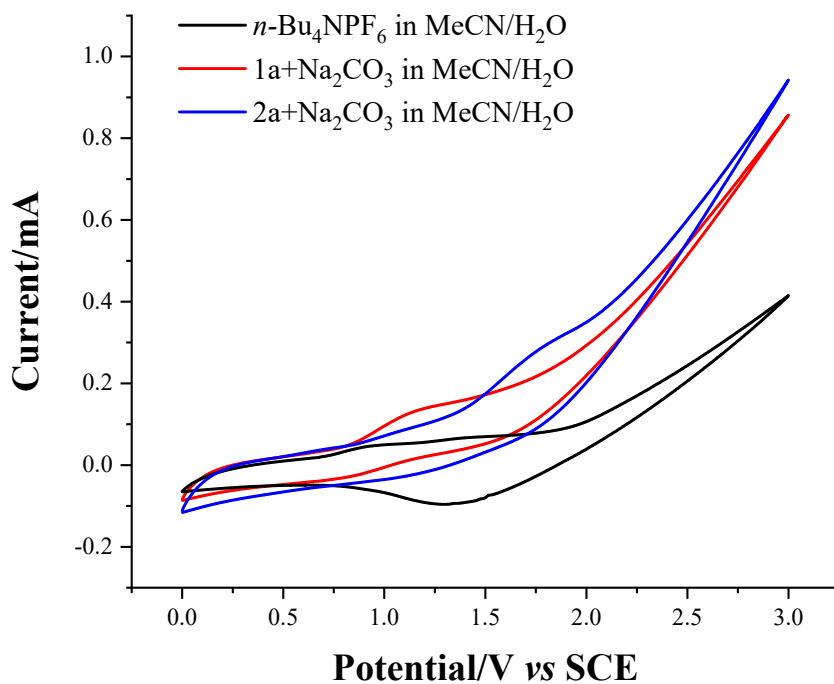
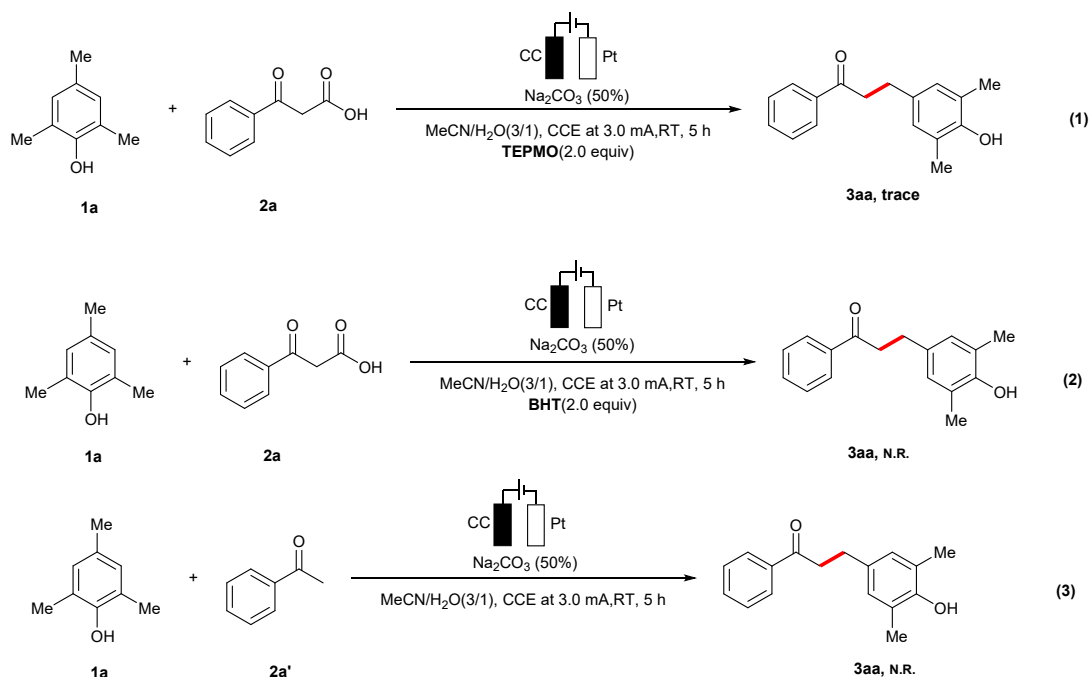


Figure S6. Cyclic voltammograms of *n*-Bu<sub>4</sub>NPF<sub>6</sub> (0.1 M) in MeCN/H<sub>2</sub>O (3/1) in (blank, black line), substrate **1a**+ Na<sub>2</sub>CO<sub>3</sub> (red line), **2a**+ Na<sub>2</sub>CO<sub>3</sub> (purple line),. Reference electrode: SCE, Scan rate = 100 mV/s.

Cyclic voltammetry experiments showed that in the presence of sodium carbonate, trimethylphenol **1a** exhibited a significant irreversible oxidation potential at 1.18 V (*vs.* SCE), whereas the oxidation potential of  $\beta$ -ketoacids **2a** was observed at a higher potential ( $E_{\text{oxi}} = 1.76$  V *vs.* SCE). This result indicated that trimethylphenol **1a** was initially oxidized in a mixed solution of acetonitrile and H<sub>2</sub>O. The mechanism of the reaction is also being further explored.



## 6.2 Control experiments

<sup>a</sup>Reaction conditions: undivided cell, Graphite anode, Pt cathode, **1a** (0.3 mmol), **2a** (0.6 mmol),  $\text{Na}_2\text{CO}_3$  (0.5 equiv.), MeCN/ $\text{H}_2\text{O}$  (3/1, 4 mL), CCE = 3.0 mA, 5 h, RT, under air. CCE = constant current electrolysis.

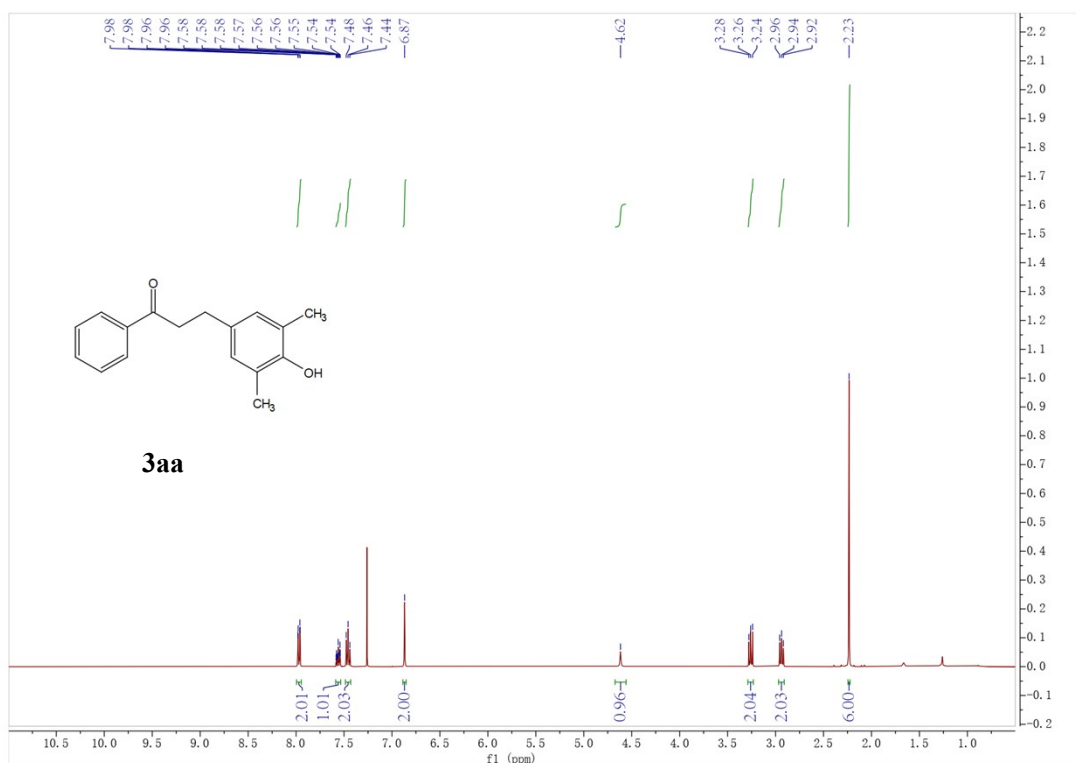
In free radical validation experiments, only trace amount of the desired product were obtained when 2,2,6,6-tetramethylpiperidin-1-yloxy (TEMPO) was added, whereas when BHT (2,6-di-tert-butyl-4-methylphenol) was added as in radical trapping agent, the reaction did not take place. When we replaced  $\beta$ -phenylketo acid by acetophenone, target product could not be observed too, which indicates that acetophenone is not an intermediate product of  $\beta$ -phenylketo acid.

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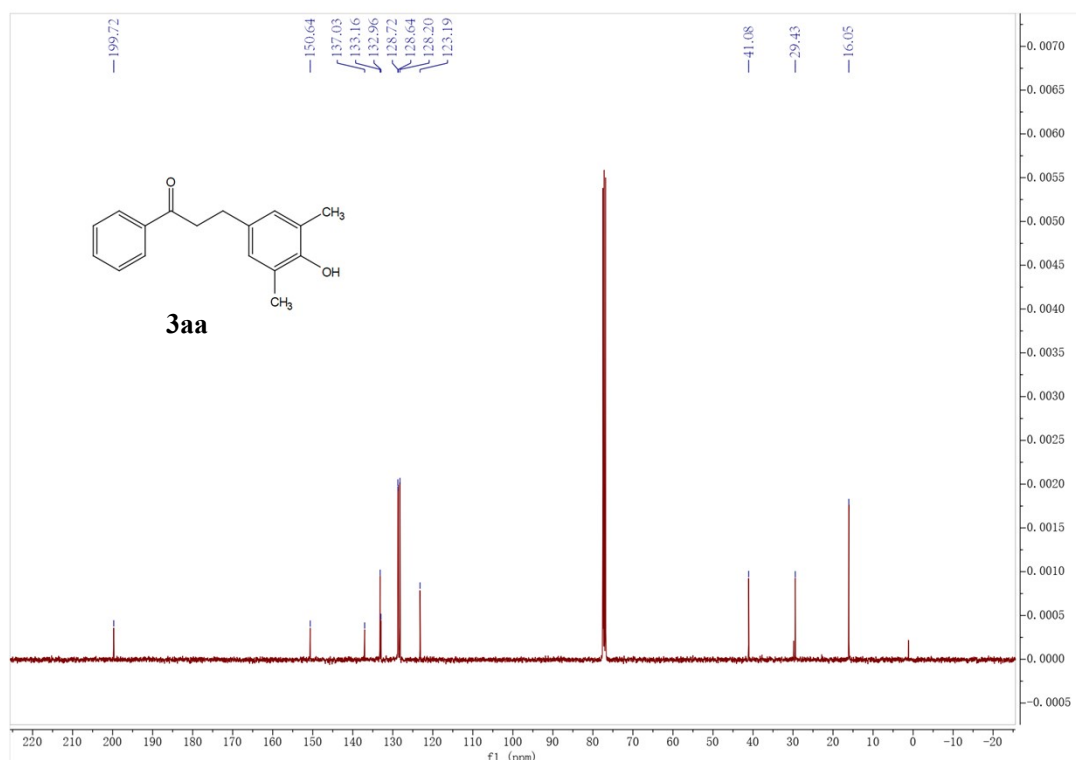
## 7. References

- [1] Xu X, Chen H, He J, et al. Copper-Catalysed Decarboxylative Trifluoromethylation of  $\beta$ -Ketoacids[J]. *Chinese Journal of Chemistry*, 2017, 35(11): 1665-1668
- [2] Jiang C, Chen Y, Huang G, et al. Scandium (III)-Catalysed Decarboxylative Addition of  $\beta$ -Ketoacids to para-Quinone Methides: Evidence for 1, 6-Addition and Base-Assisted Decarboxylation Tandem Process[J]. *Asian Journal of Organic Chemistry*, 2019, 8(2): 257-260.
- [3] Egami H, Ide T, Kawato Y, et al. Benzylic C–H trifluoromethylation of phenol derivatives[J]. *Chemical Communications*, 2015, 51(93): 16675-16678.
- [4] Luo S, Luo F X, Zhang X S, et al. Synthesis of Dibenzopyranones through Palladium-Catalyzed Directed C–H Activation/Carbonylation of 2-Arylphenols[J]. *Angewandte Chemie International Edition*, 2013, 52(40): 10598-10601.

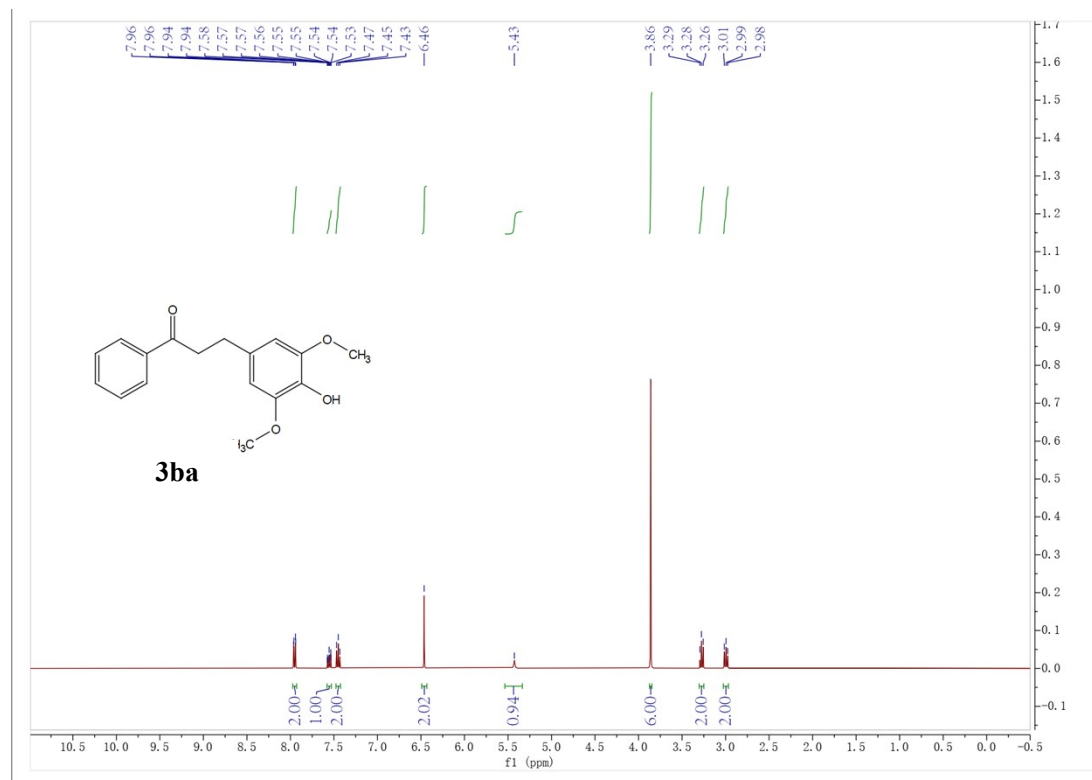
## 8. $^1\text{H}$ and $^{13}\text{C}$ -NMR spectra



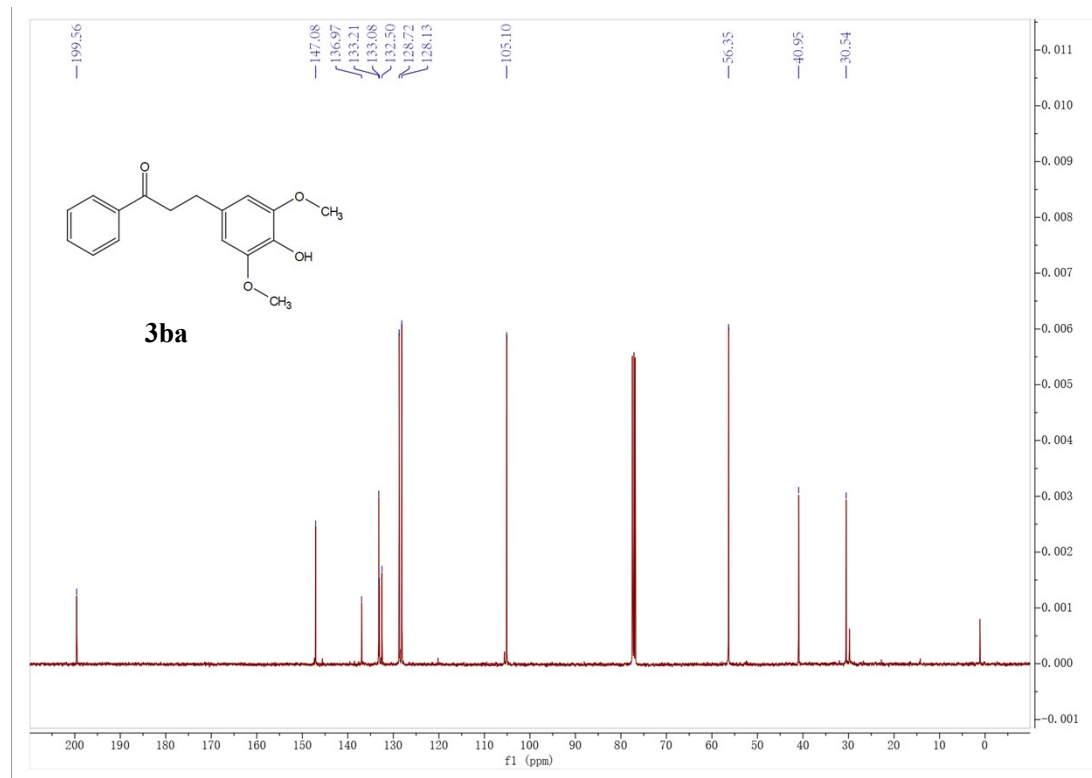
$^1\text{H}$  NMR spectrum of **3aa** ( $\text{CDCl}_3$ , 400 MHz)



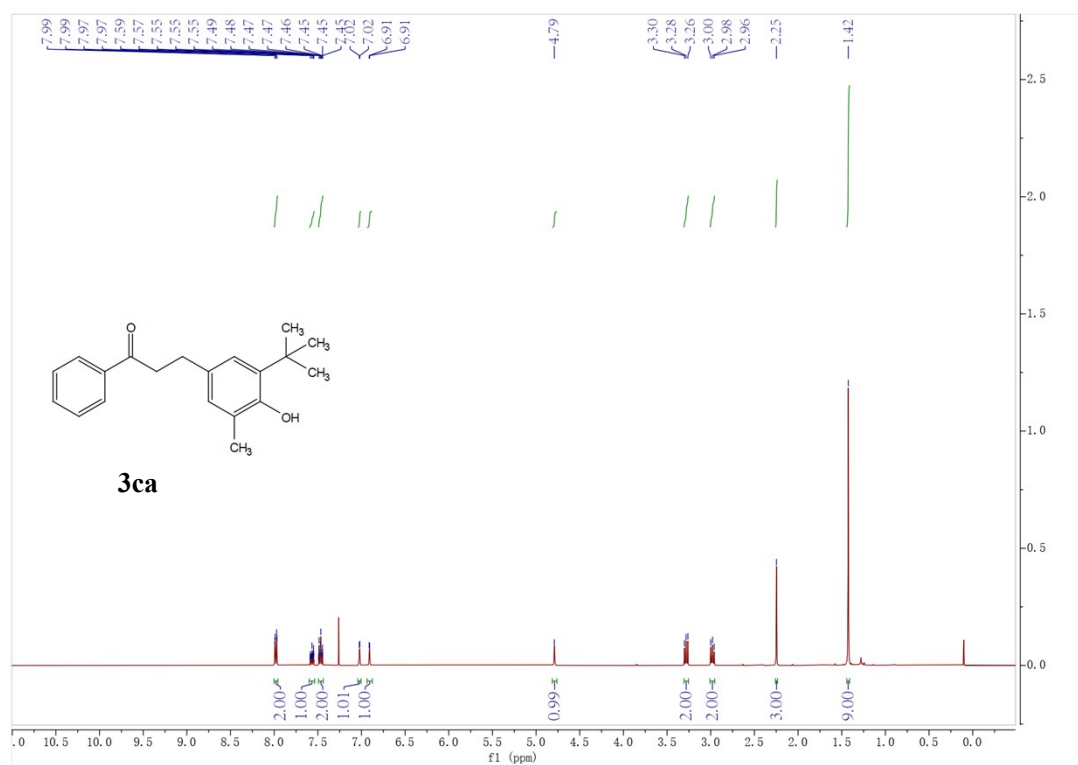
$^{13}\text{C}$  NMR spectrum of **3aa** ( $\text{CDCl}_3$ , 101 MHz)



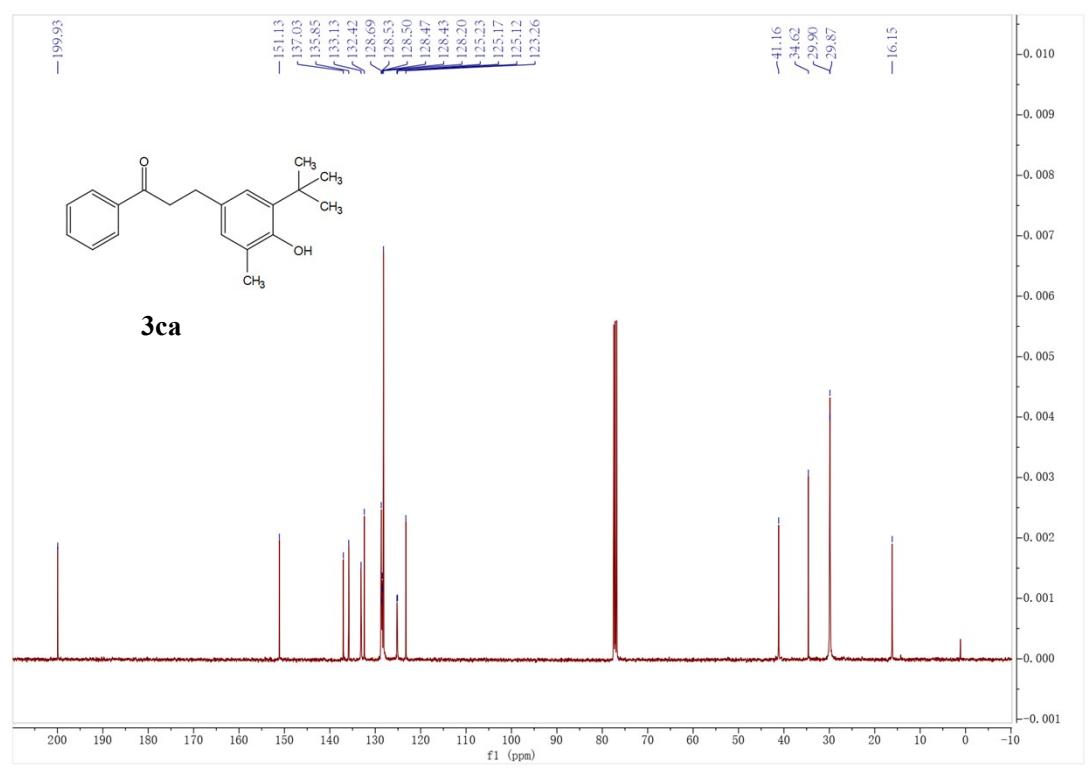
<sup>1</sup>H NMR spectrum of **3ba** (CDCl<sub>3</sub>, 400 MHz)



<sup>13</sup>C NMR spectrum of **3ba** (CDCl<sub>3</sub>, 101 MHz)

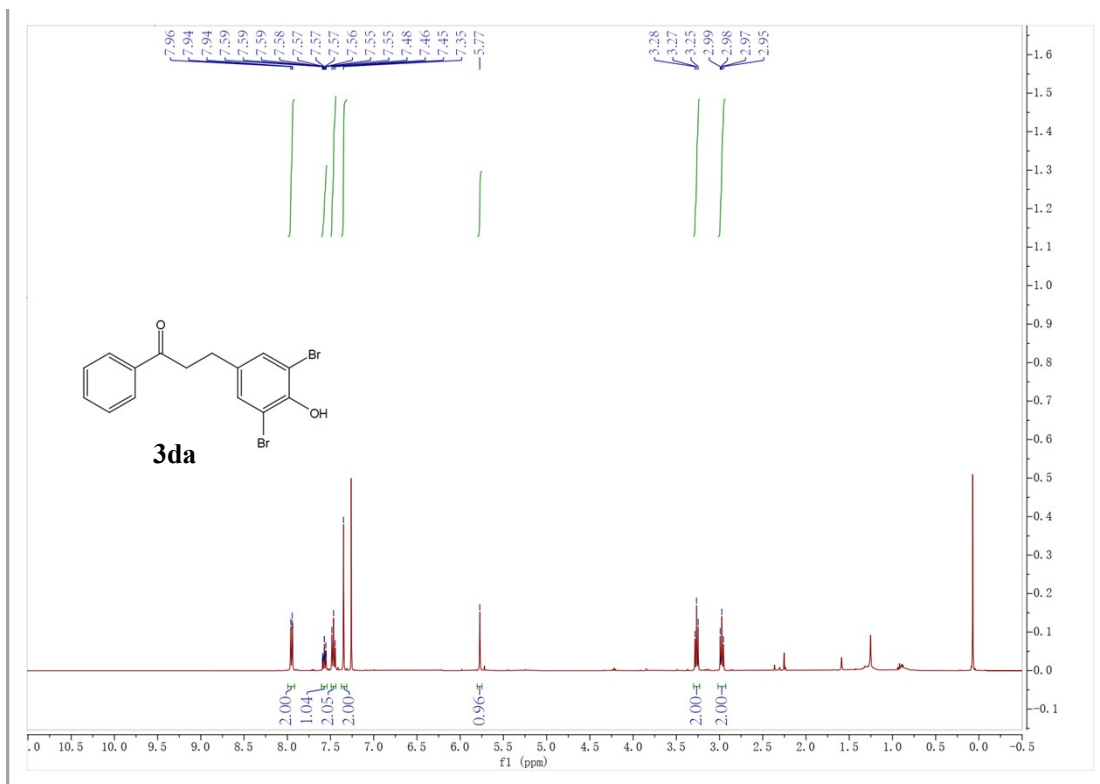


<sup>1</sup>H NMR spectrum of **3ca** (CDCl<sub>3</sub>, 400 MHz)

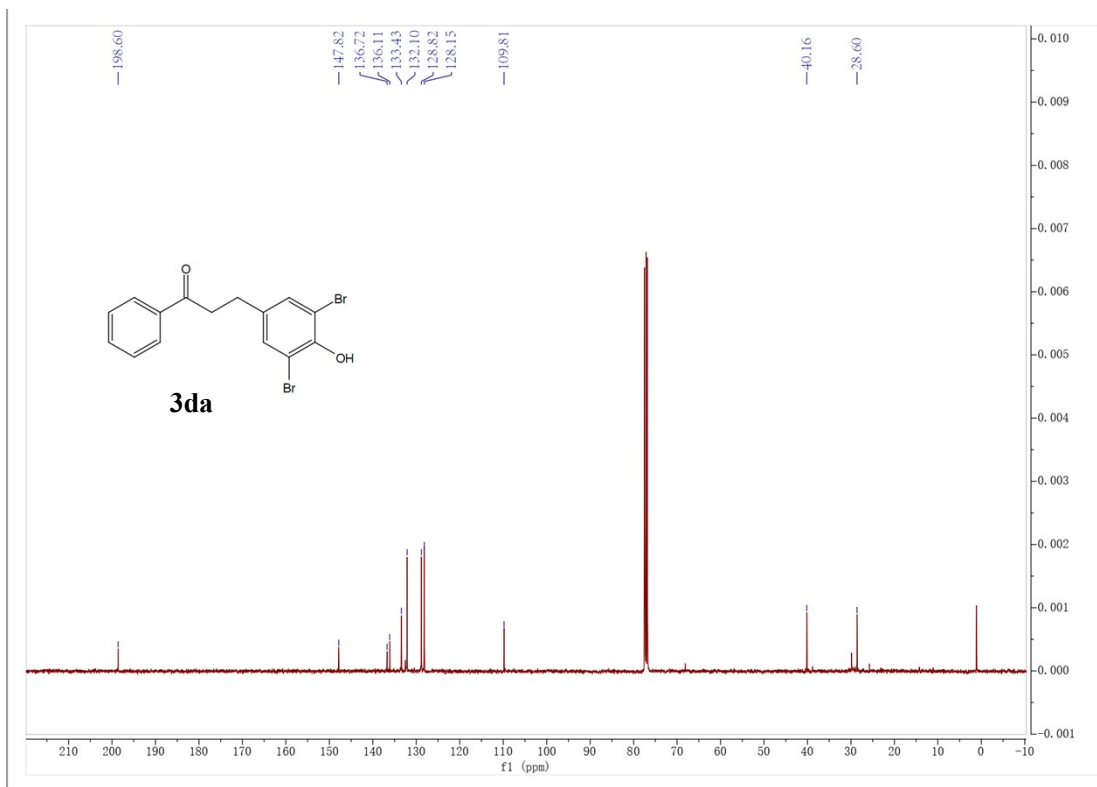


<sup>13</sup>C NMR spectrum of **3ca** (CDCl<sub>3</sub>, 101 MHz)

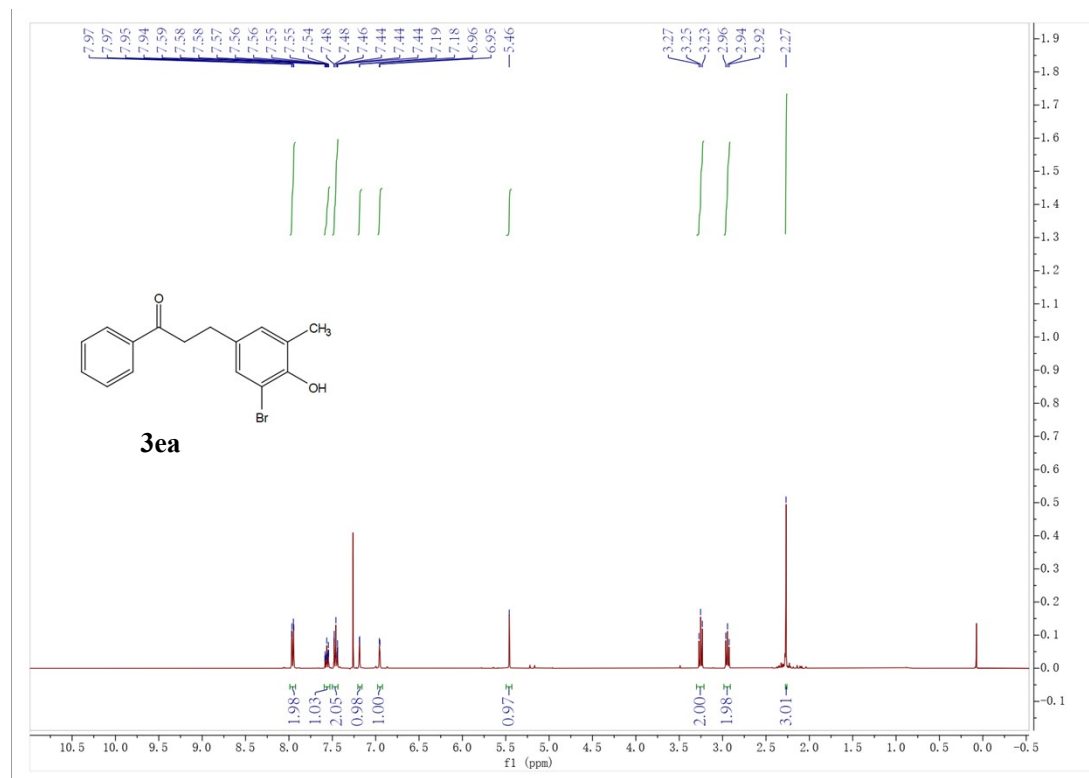




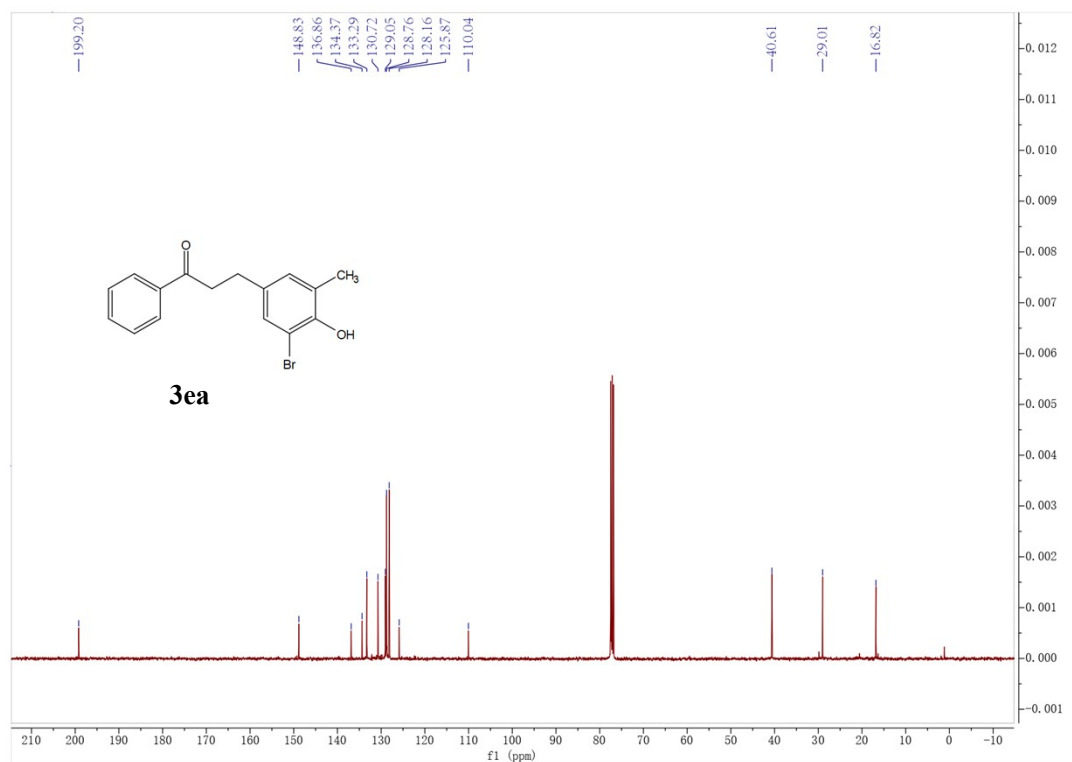
<sup>1</sup>H NMR spectrum of **3da** (CDCl<sub>3</sub>, 400 MHz)



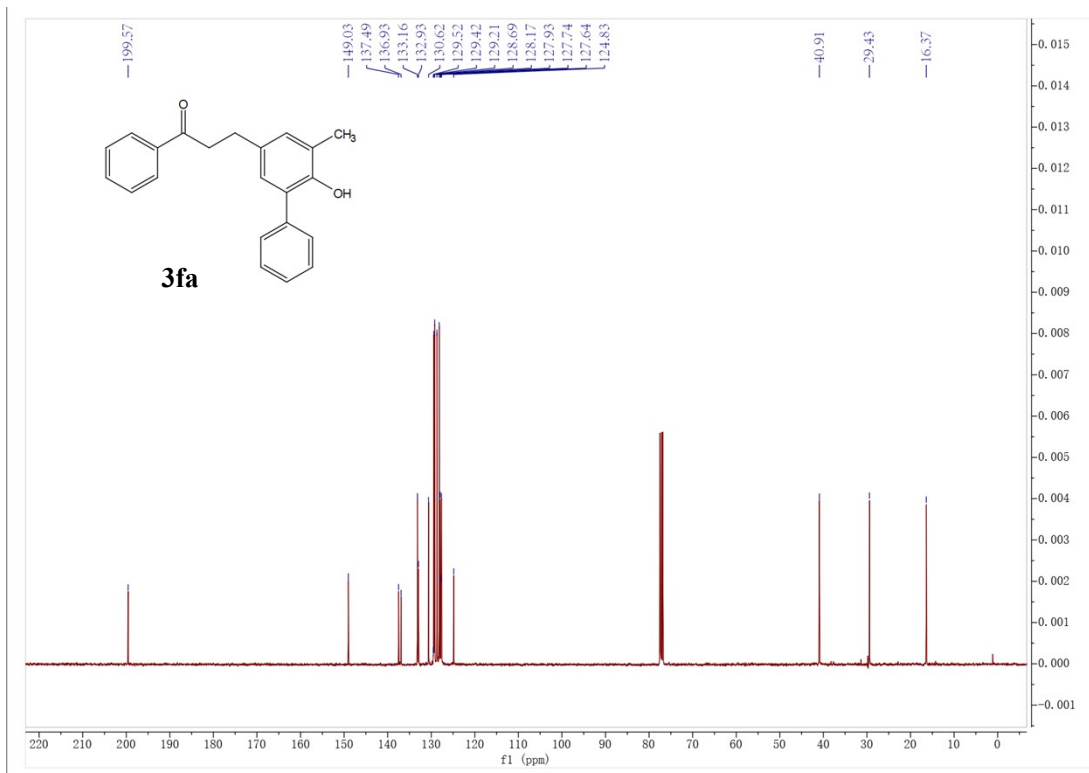
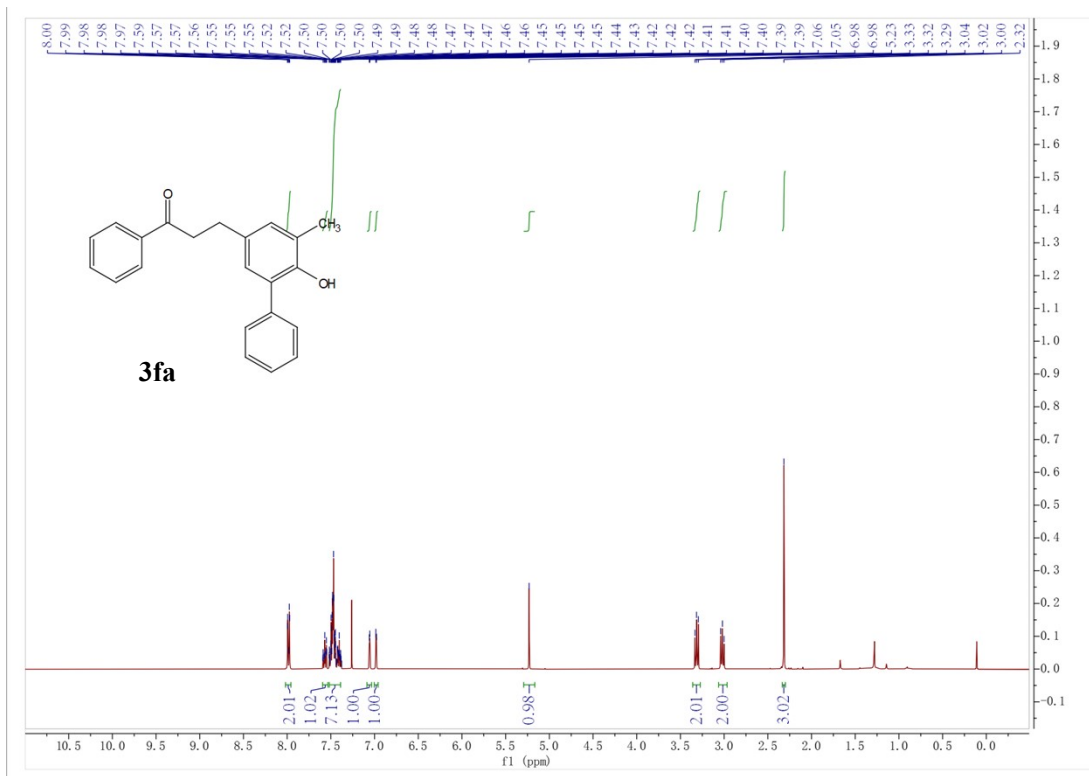
<sup>13</sup>C NMR spectrum of **3da** (CDCl<sub>3</sub>, 101 MHz)

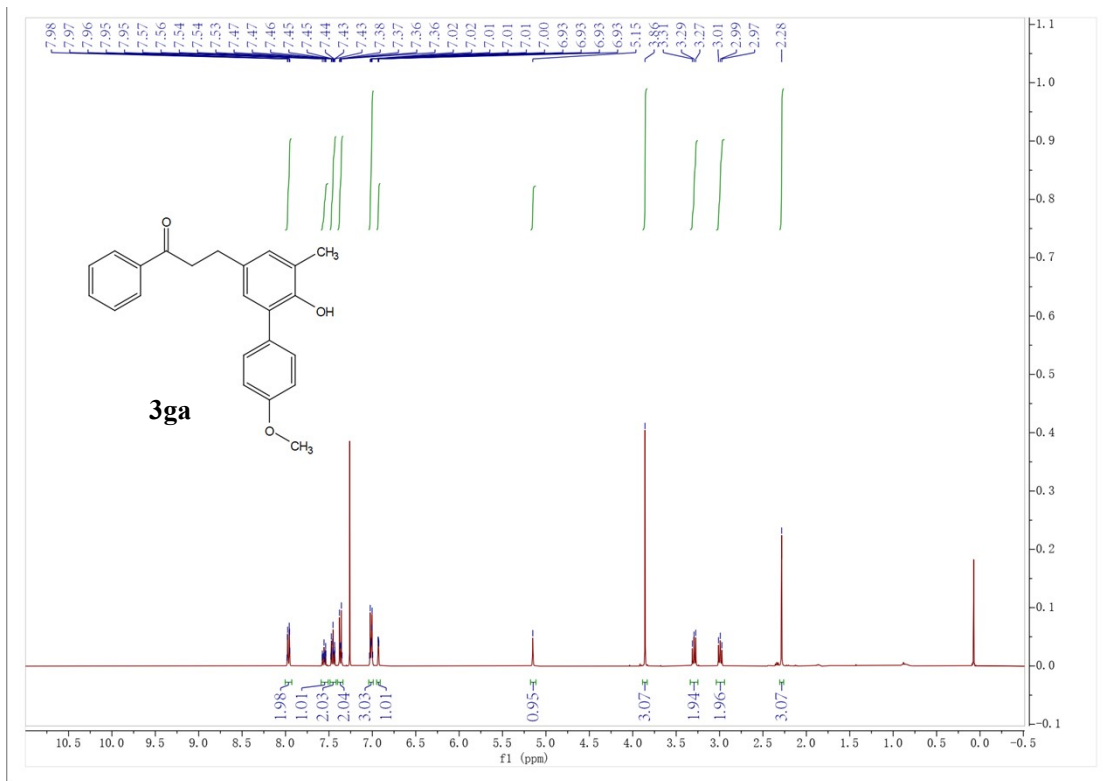


<sup>1</sup>H NMR spectrum of **3ea** (CDCl<sub>3</sub>, 400 MHz)

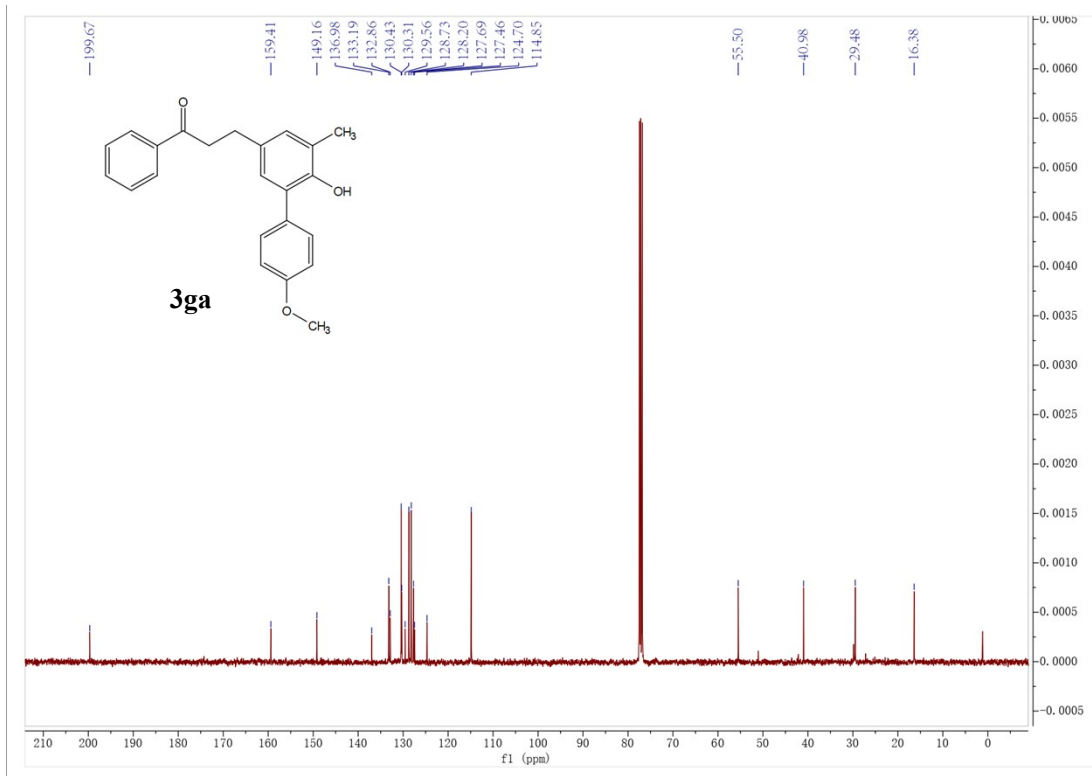


<sup>13</sup>C NMR spectrum of **3ea** (CDCl<sub>3</sub>, 101 MHz)

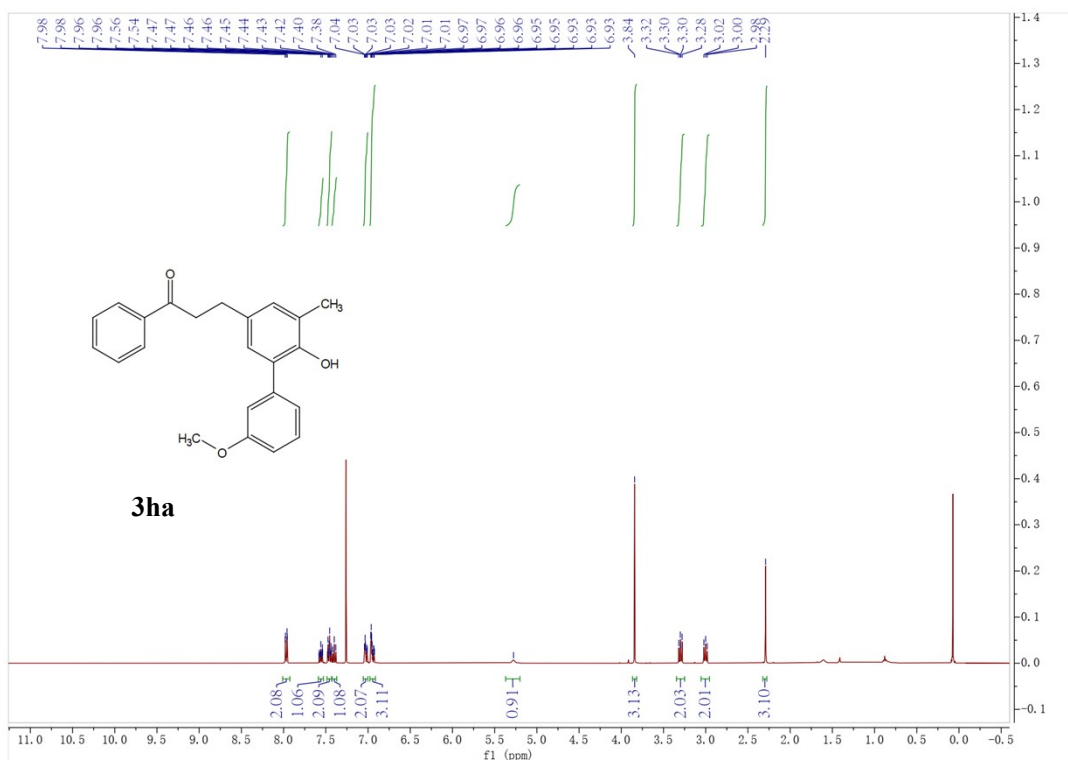




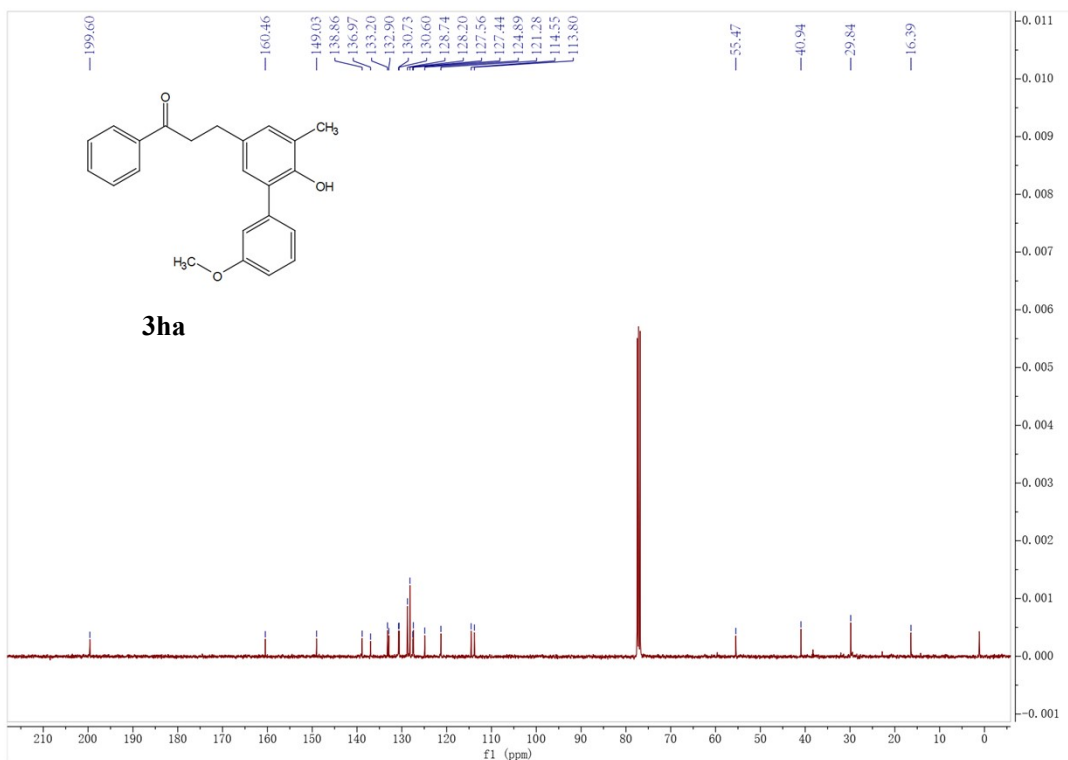
<sup>1</sup>H NMR spectrum of **3ga** (CDCl<sub>3</sub>, 400 MHz)



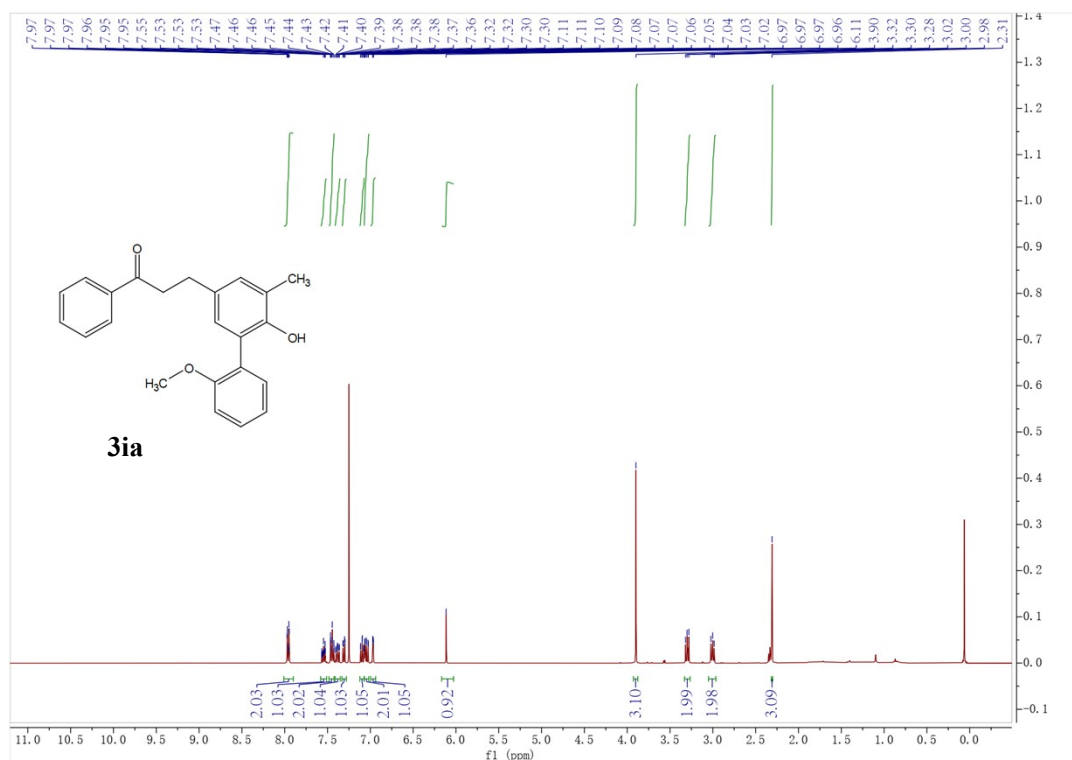
<sup>13</sup>C NMR spectrum of **3ga** (CDCl<sub>3</sub>, 101 MHz)



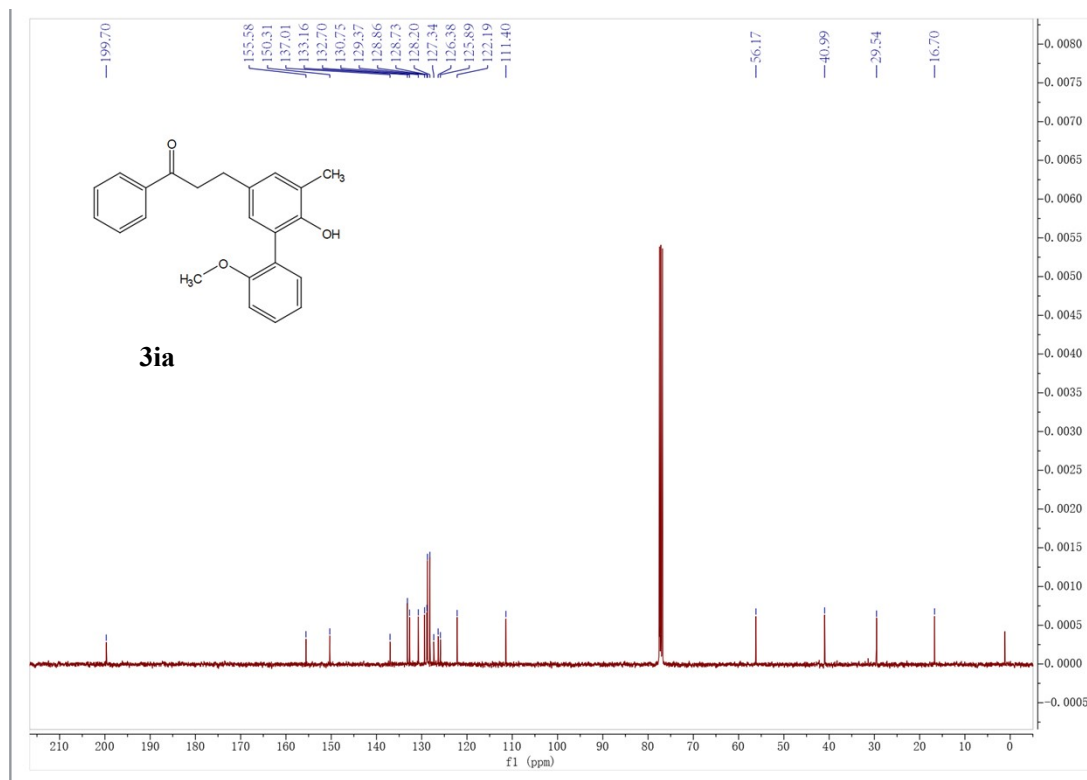
<sup>1</sup>H NMR spectrum of **3ha** (CDCl<sub>3</sub>, 400 MHz)



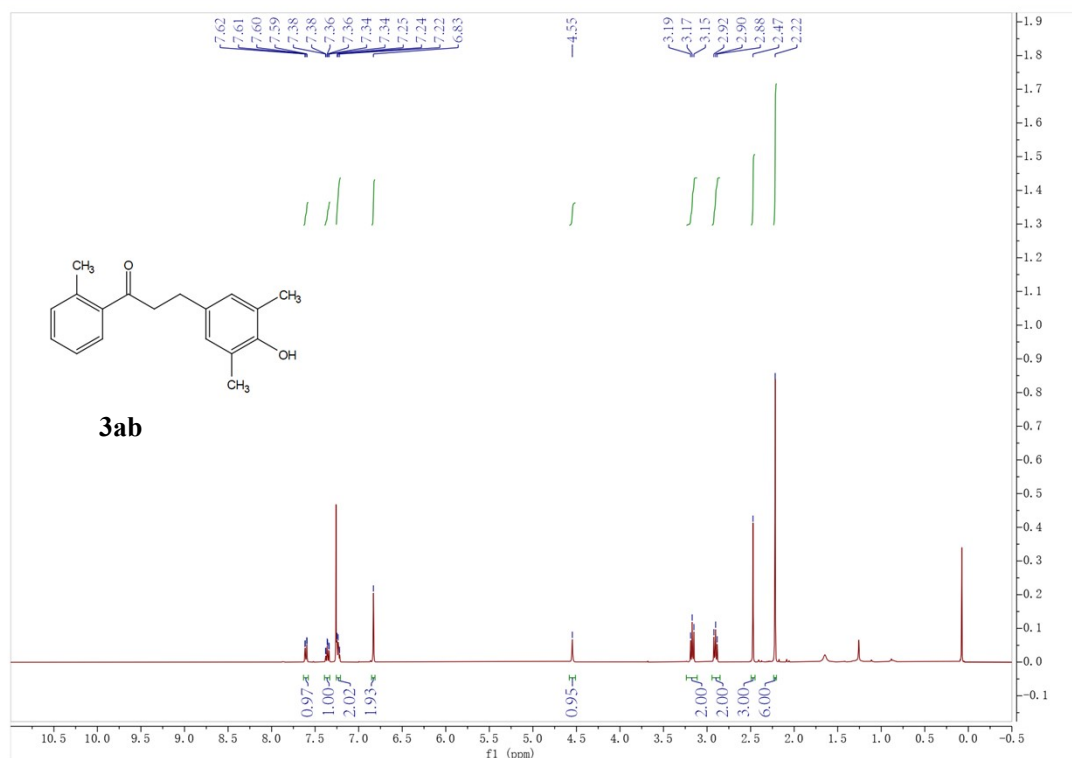
<sup>13</sup>C NMR spectrum of **3ha** (CDCl<sub>3</sub>, 101 MHz)



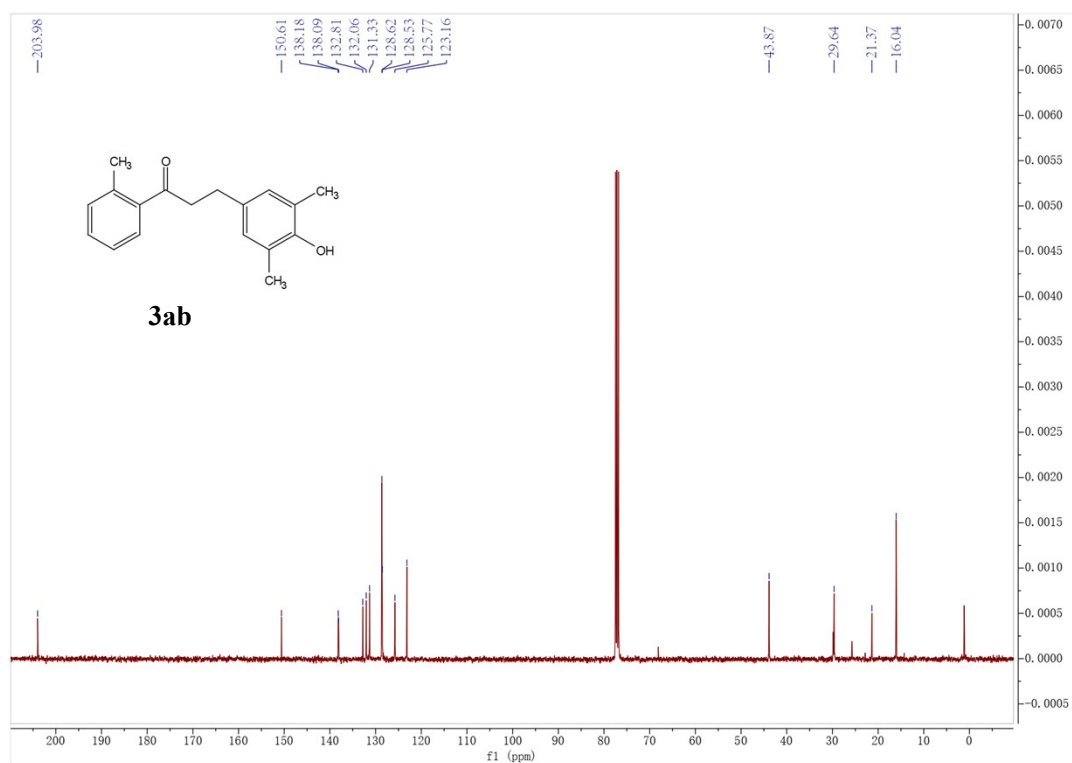
$^1\text{H}$  NMR spectrum of **3ia** ( $\text{CDCl}_3$ , 400 MHz)



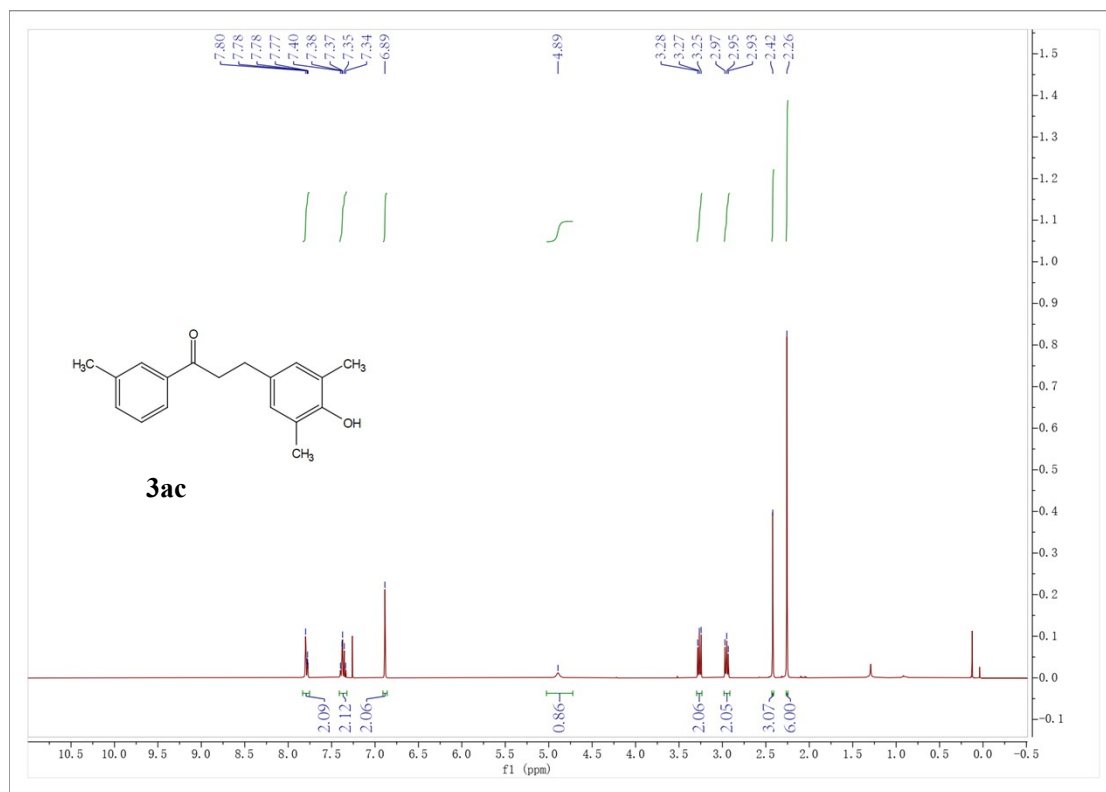
$^{13}\text{C}$  NMR spectrum of **3ia** ( $\text{CDCl}_3$ , 101 MHz)



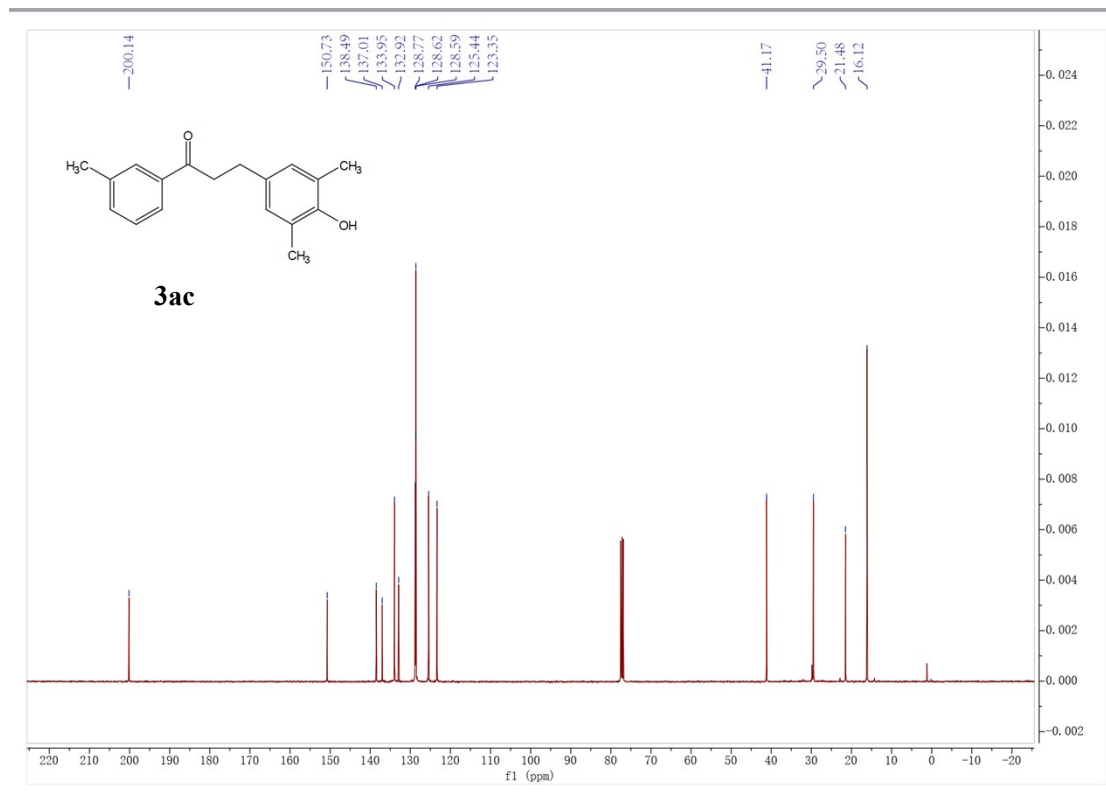
<sup>1</sup>H NMR spectrum of **3ab** (CDCl<sub>3</sub>, 400 MHz)



<sup>13</sup>C NMR spectrum of **3ab** (CDCl<sub>3</sub>, 101 MHz)

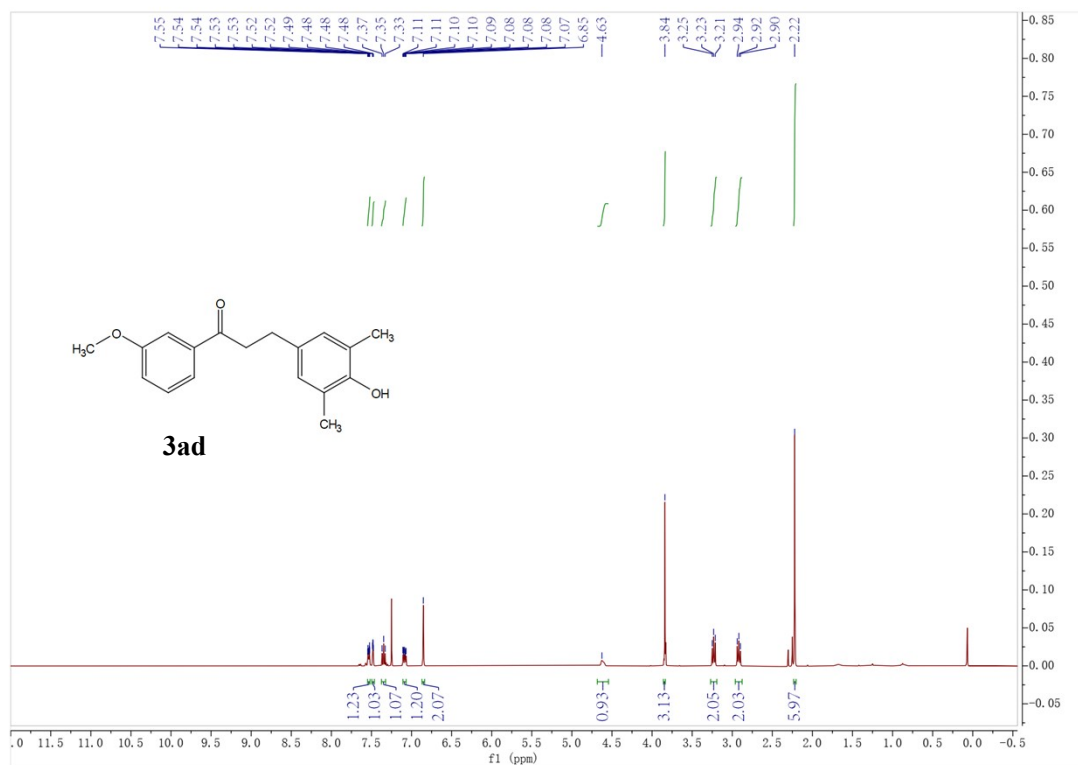


<sup>1</sup>H NMR spectrum of **3ac** (CDCl<sub>3</sub>, 400 MHz)

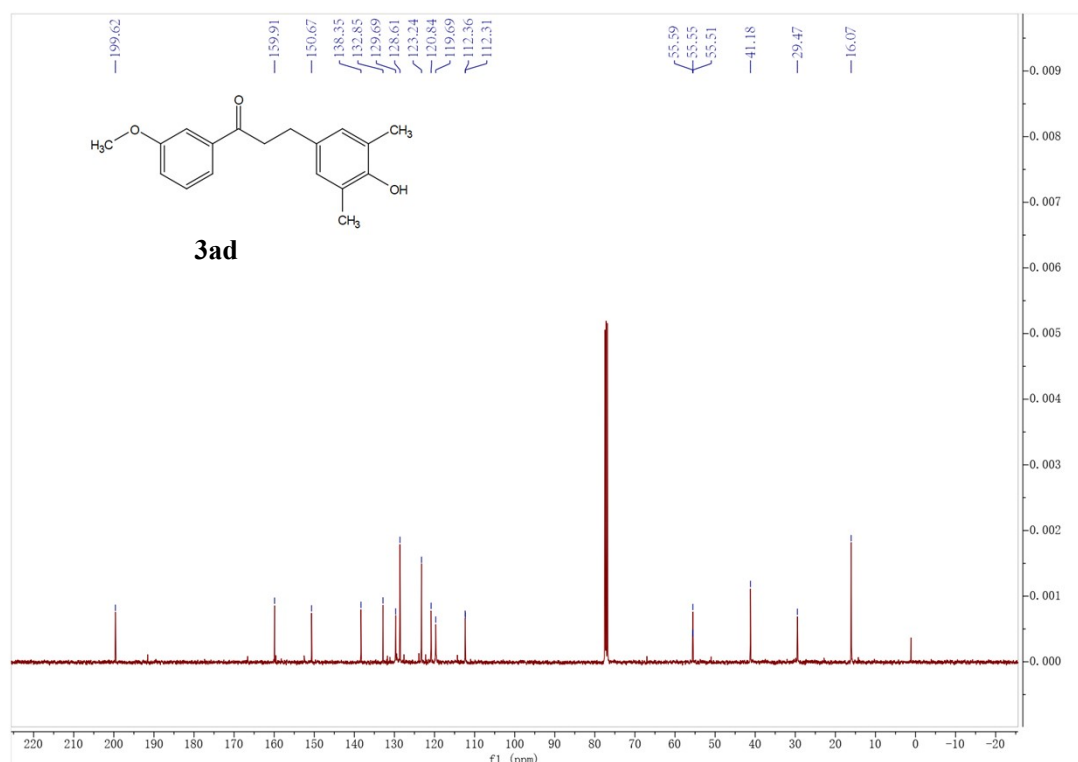


<sup>13</sup>C NMR spectrum of **3ac** (CDCl<sub>3</sub>, 101 MHz)

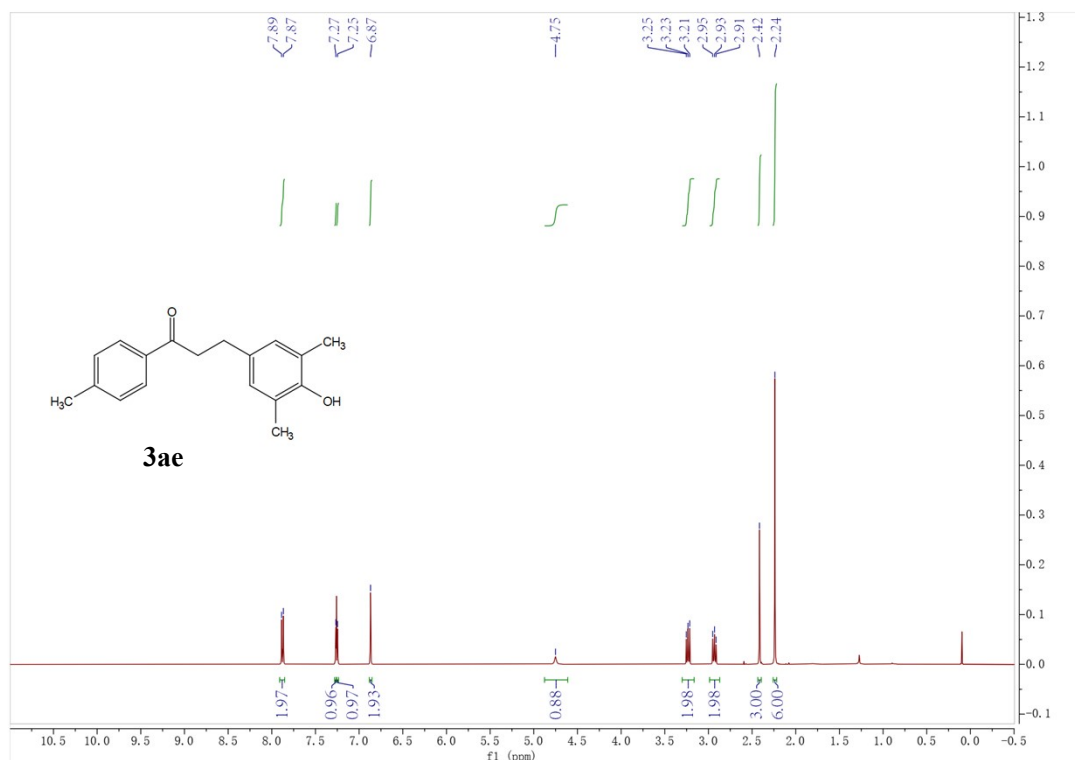




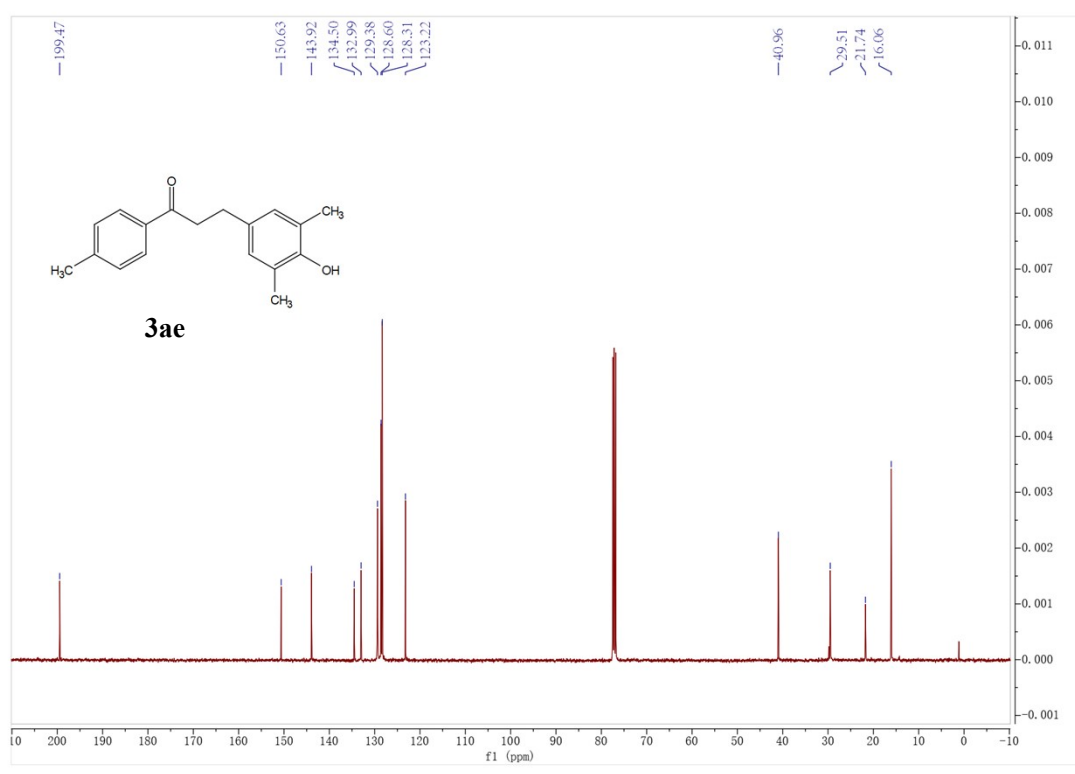
<sup>1</sup>H NMR spectrum of **3ad** (CDCl<sub>3</sub>, 400 MHz)



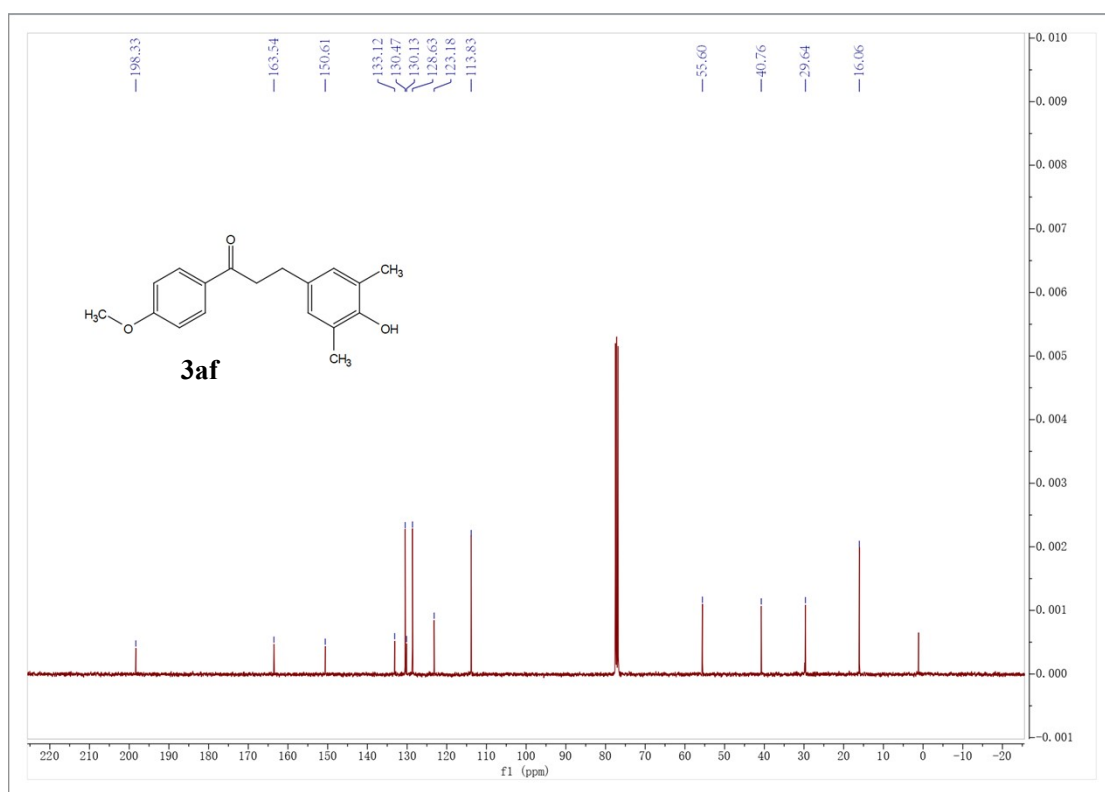
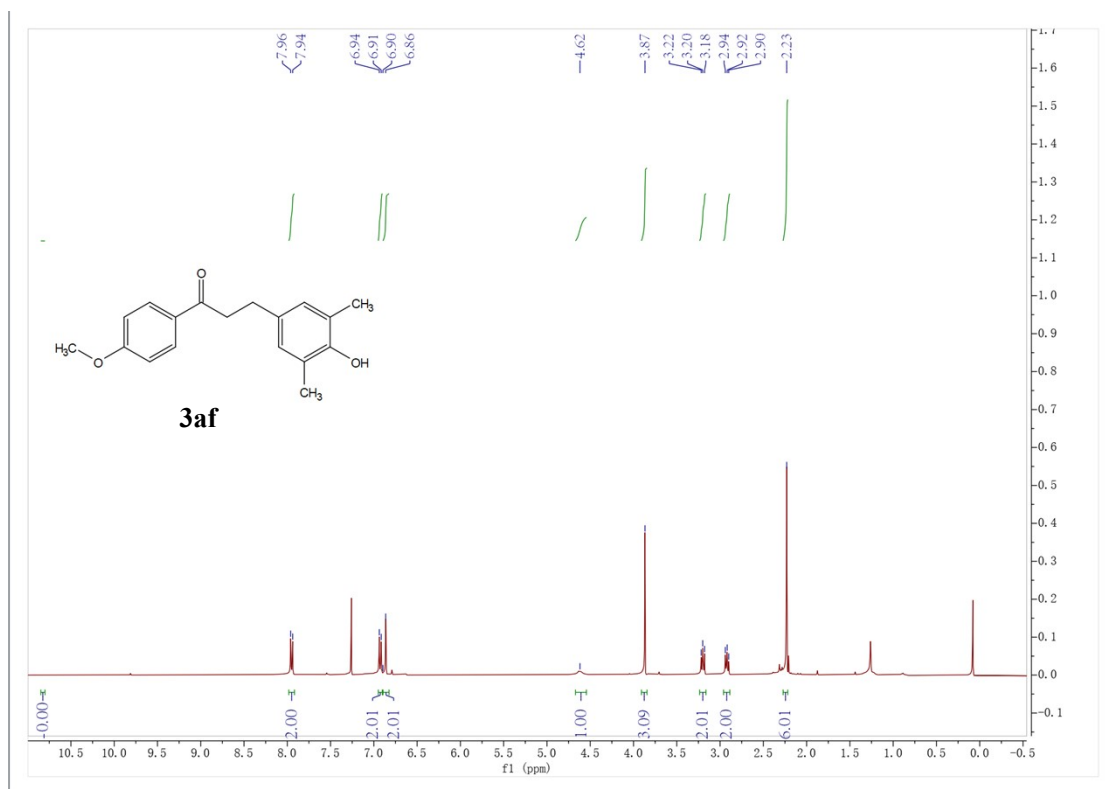
<sup>13</sup>C NMR spectrum of **3ad** (CDCl<sub>3</sub>, 101 MHz)

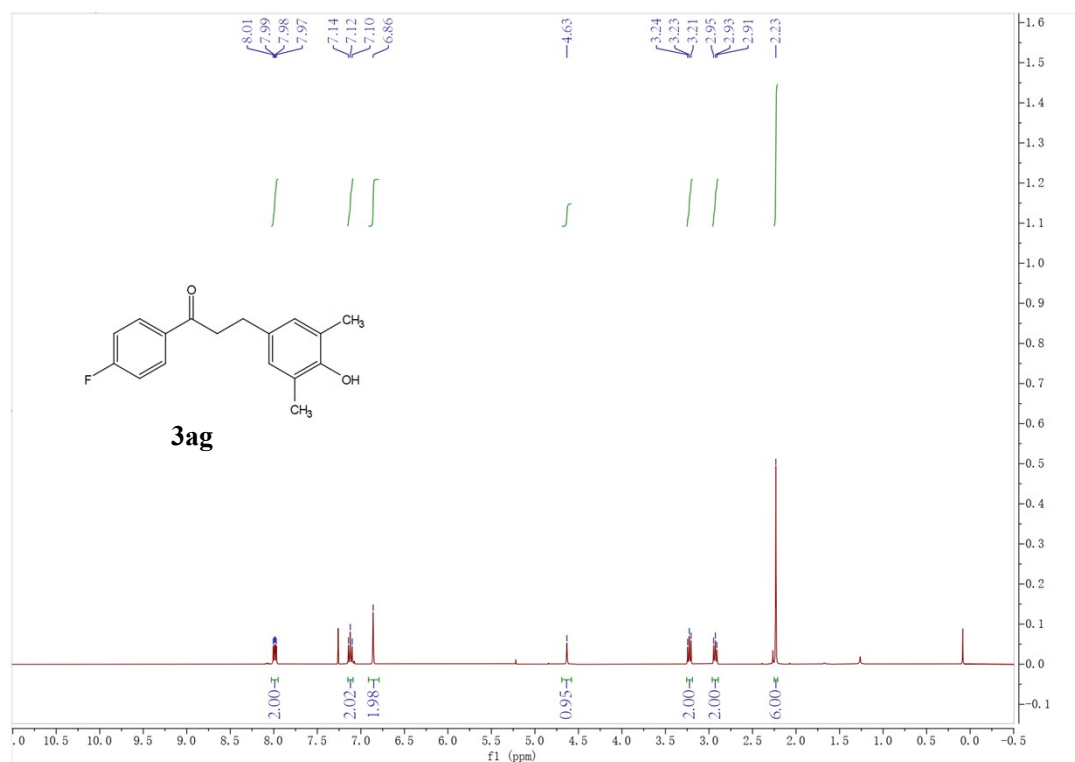


$^1\text{H}$  NMR spectrum of **3ae** ( $\text{CDCl}_3$ , 400 MHz)

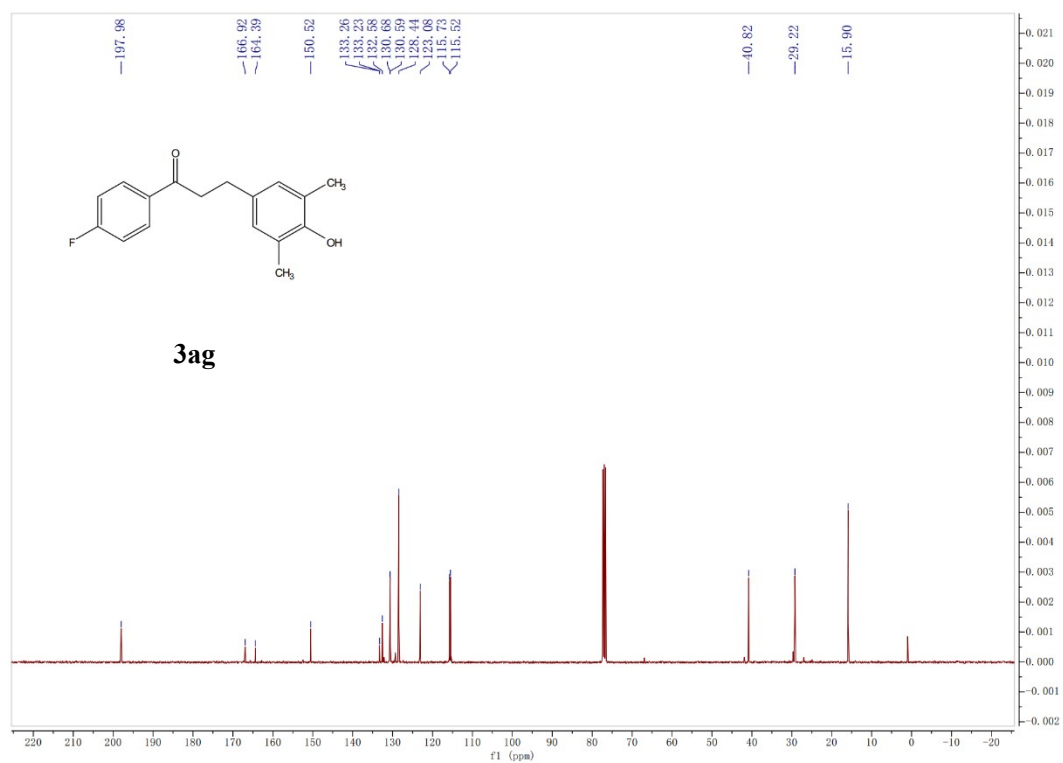


$^{13}\text{C}$  NMR spectrum of **3ae** ( $\text{CDCl}_3$ , 101 MHz)

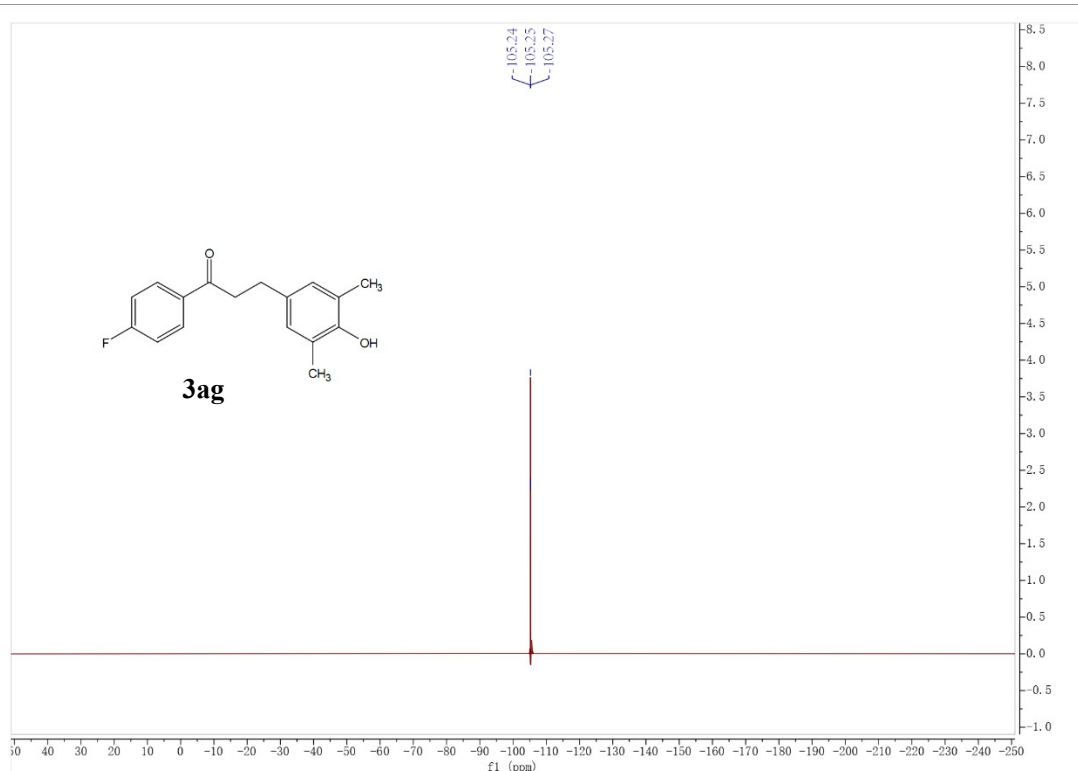




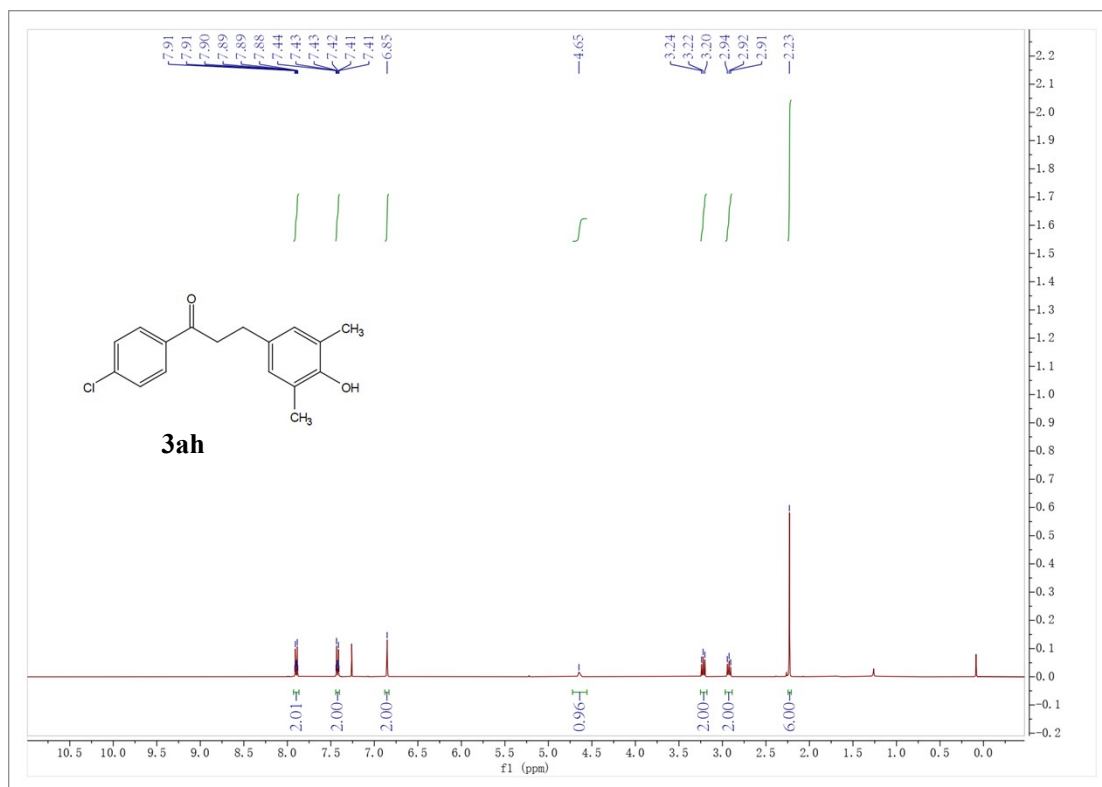
<sup>1</sup>H NMR spectrum of **3ag** (CDCl<sub>3</sub>, 400 MHz)



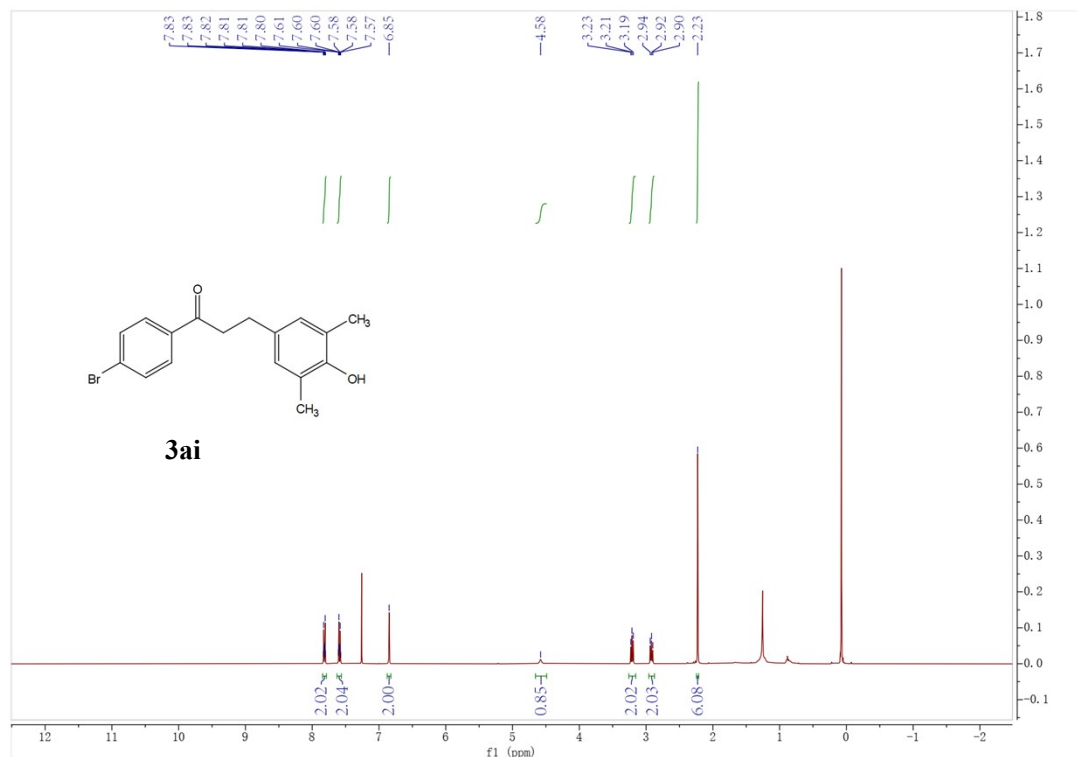
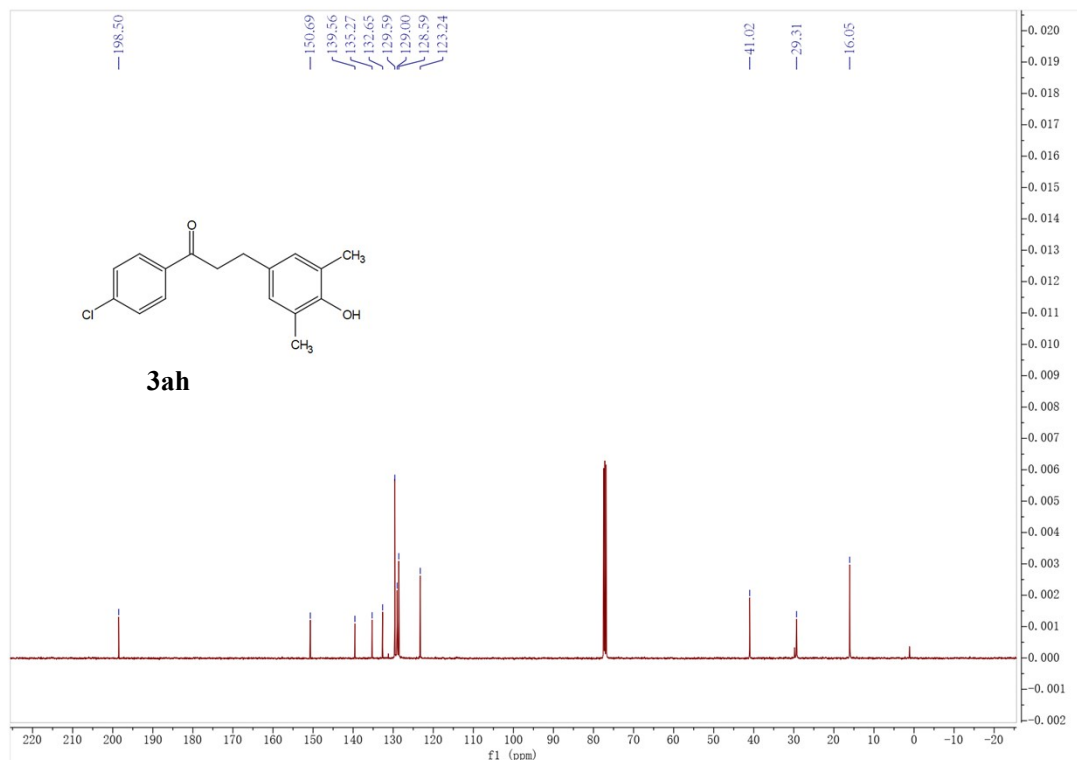
<sup>13</sup>C NMR spectrum of **3ag** (CDCl<sub>3</sub>, 101 MHz)

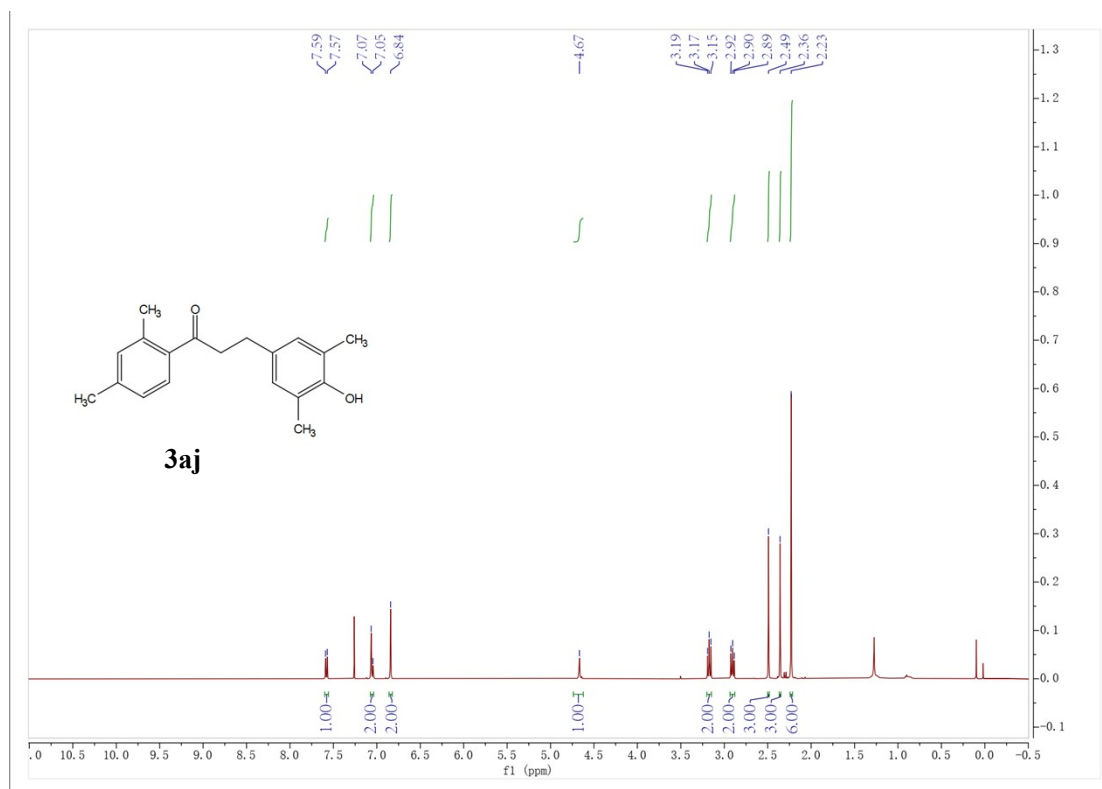
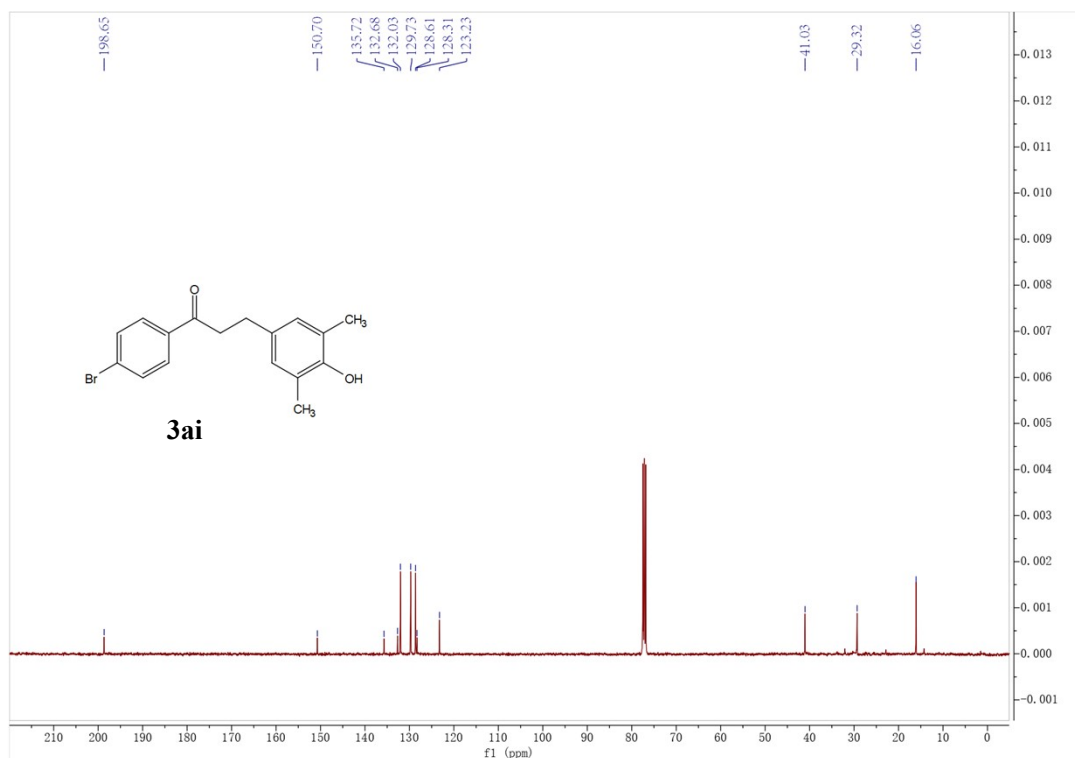


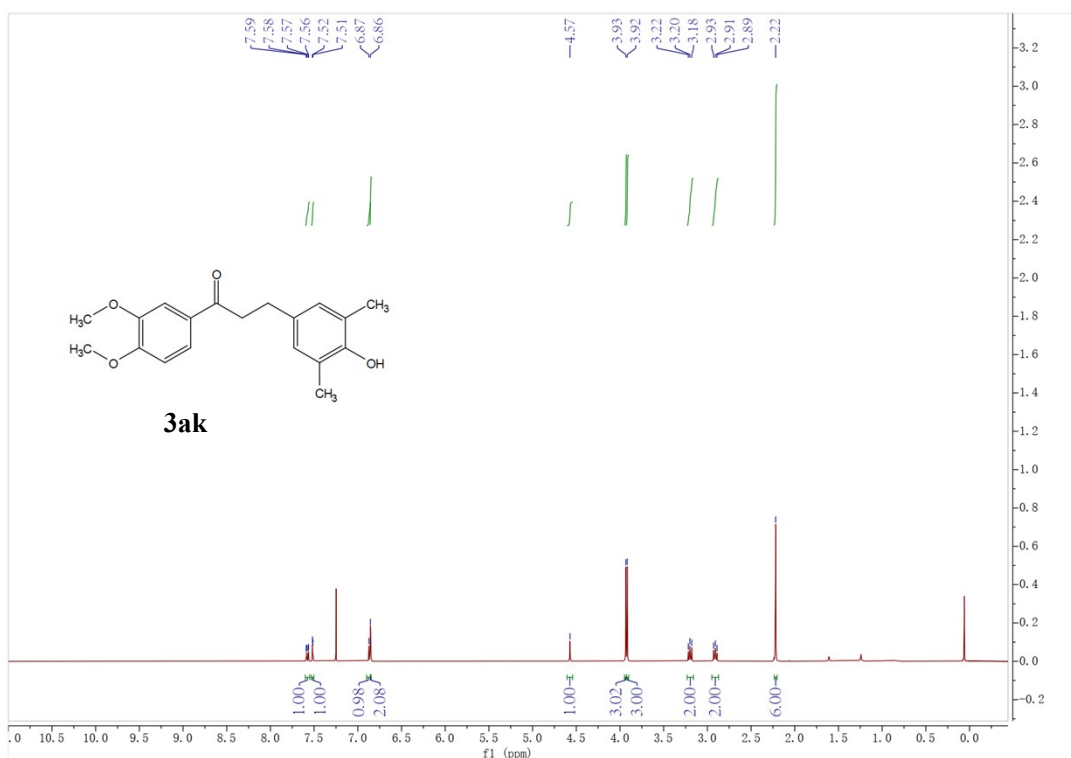
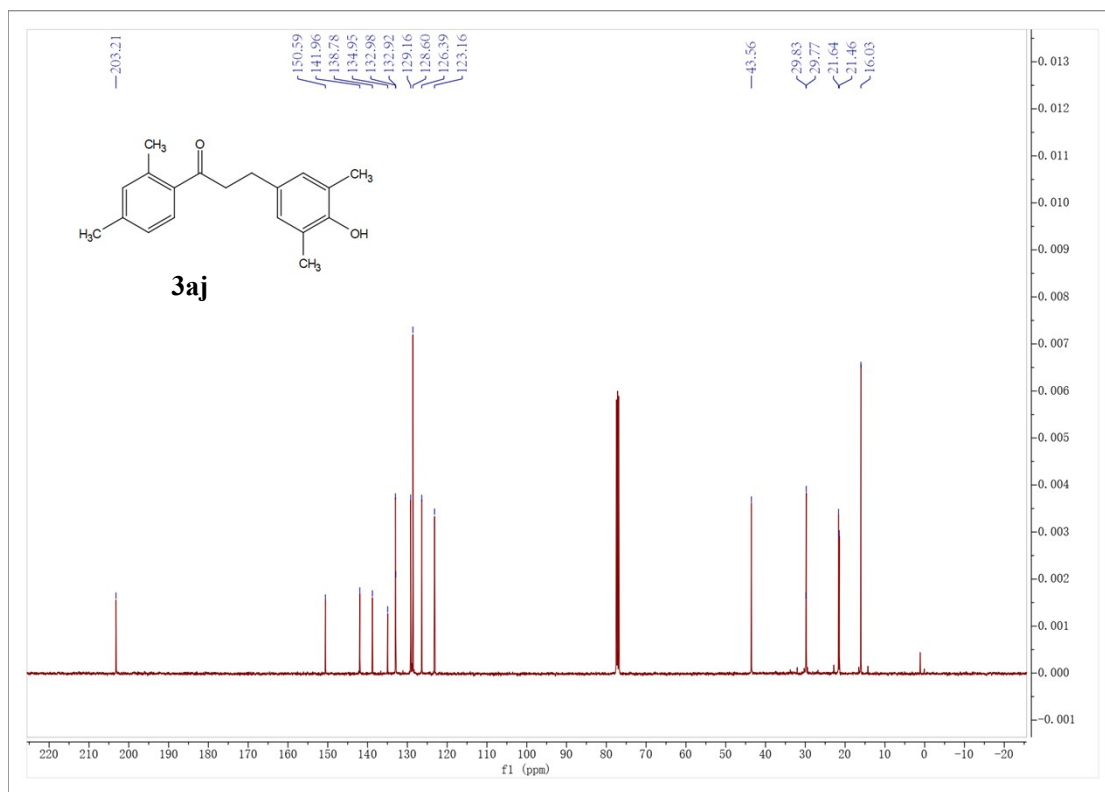
<sup>19</sup>F NMR spectrum of **3ag** (CDCl<sub>3</sub>, 376 MHz)



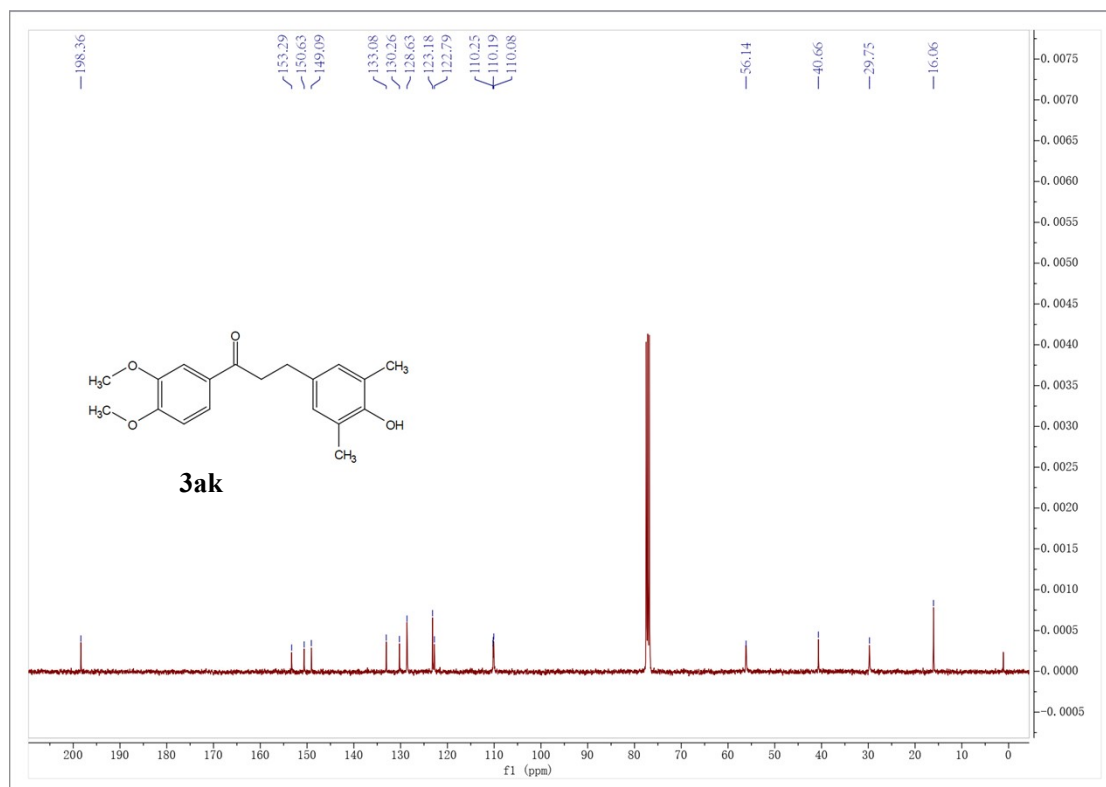
<sup>1</sup>H NMR spectrum of **3ah** (CDCl<sub>3</sub>, 400 MHz)



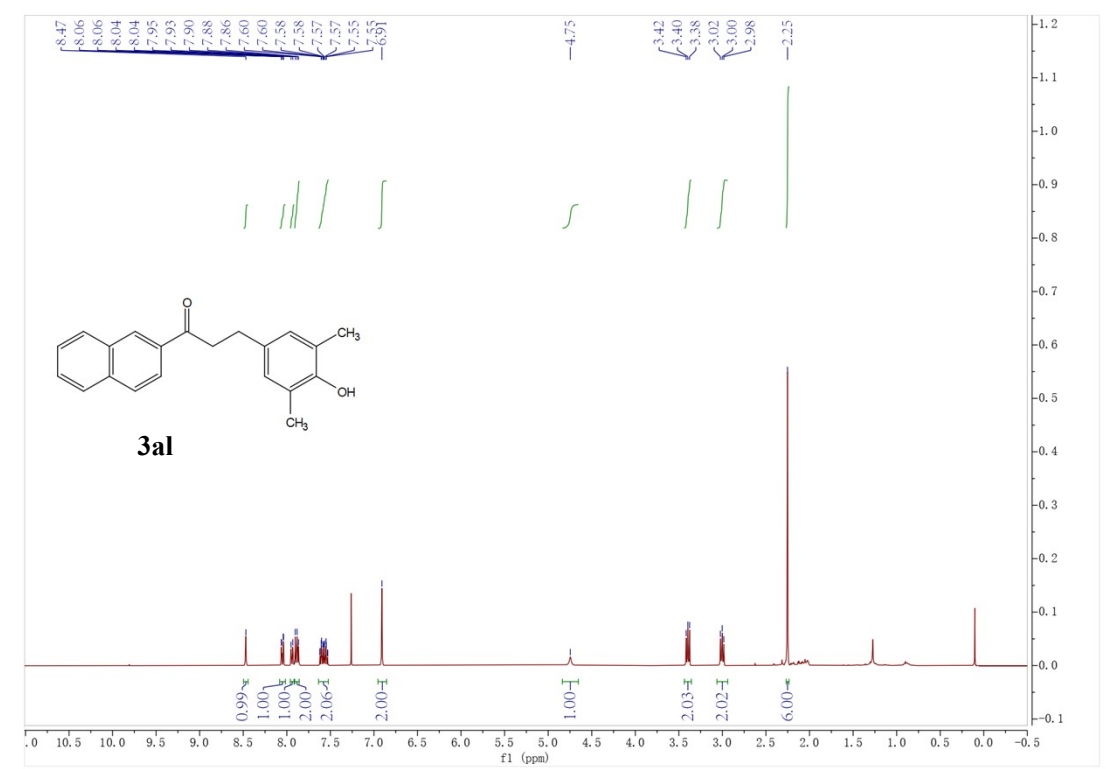




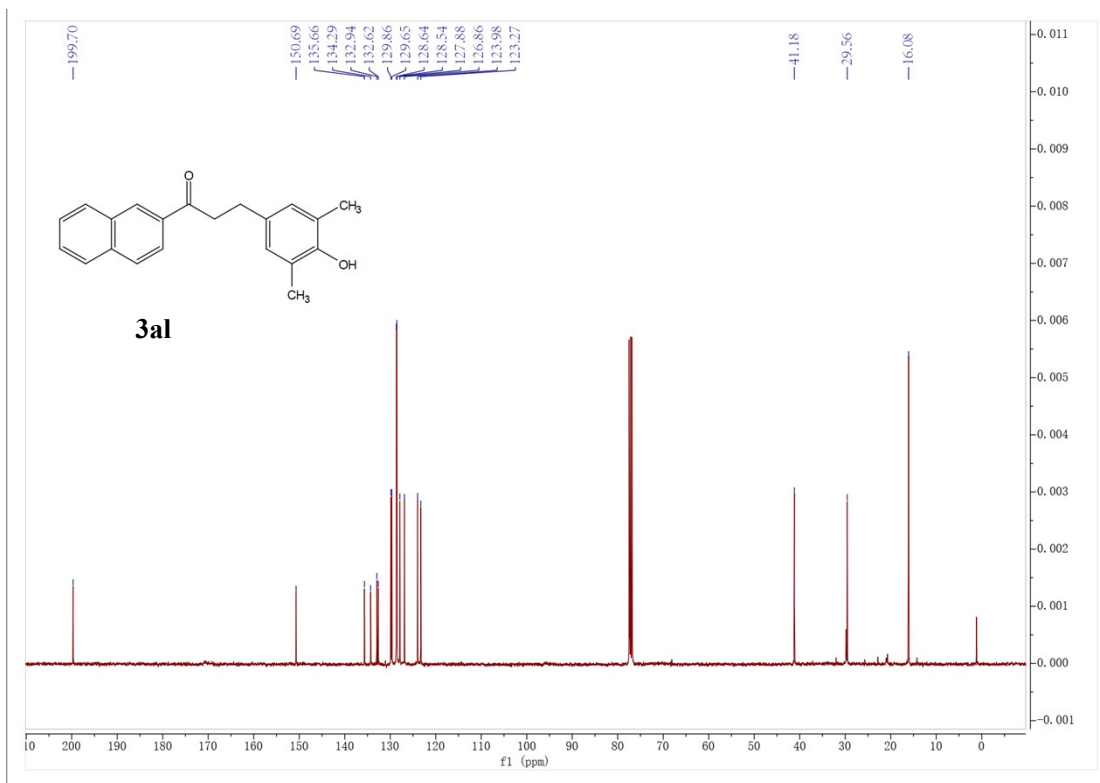




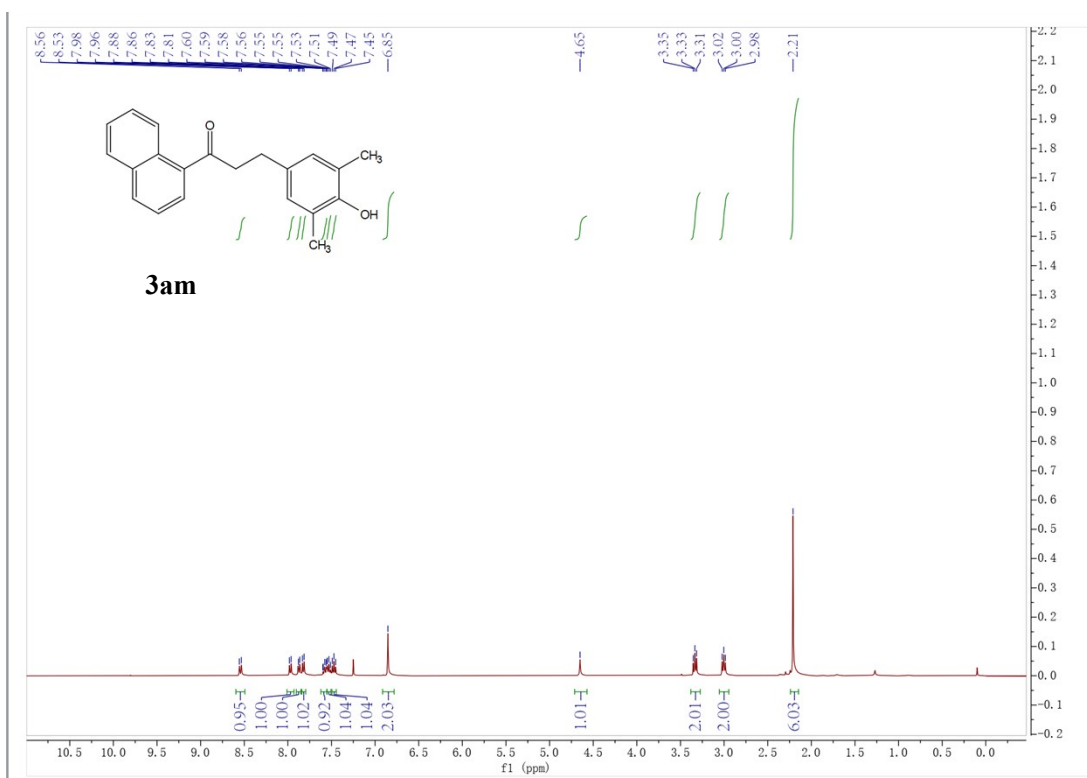
<sup>13</sup>C NMR spectrum of **3ak** (CDCl<sub>3</sub>, 101 MHz)



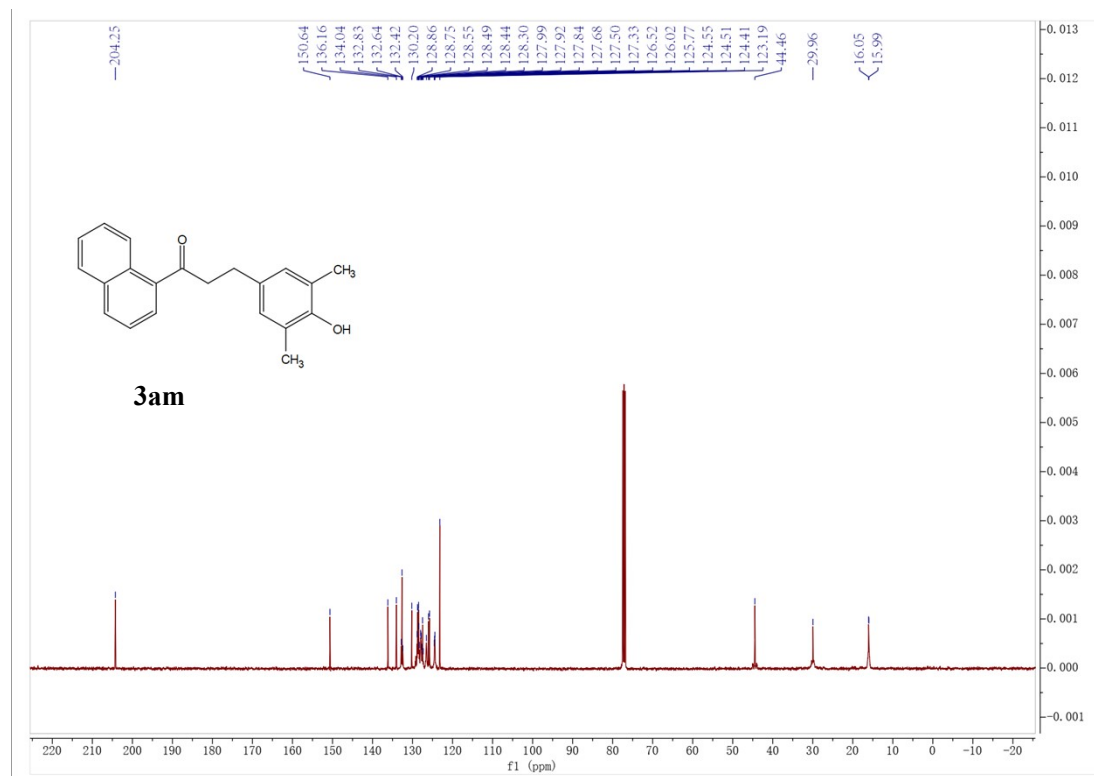
<sup>1</sup>H NMR spectrum of **3al** (CDCl<sub>3</sub>, 400 MHz)



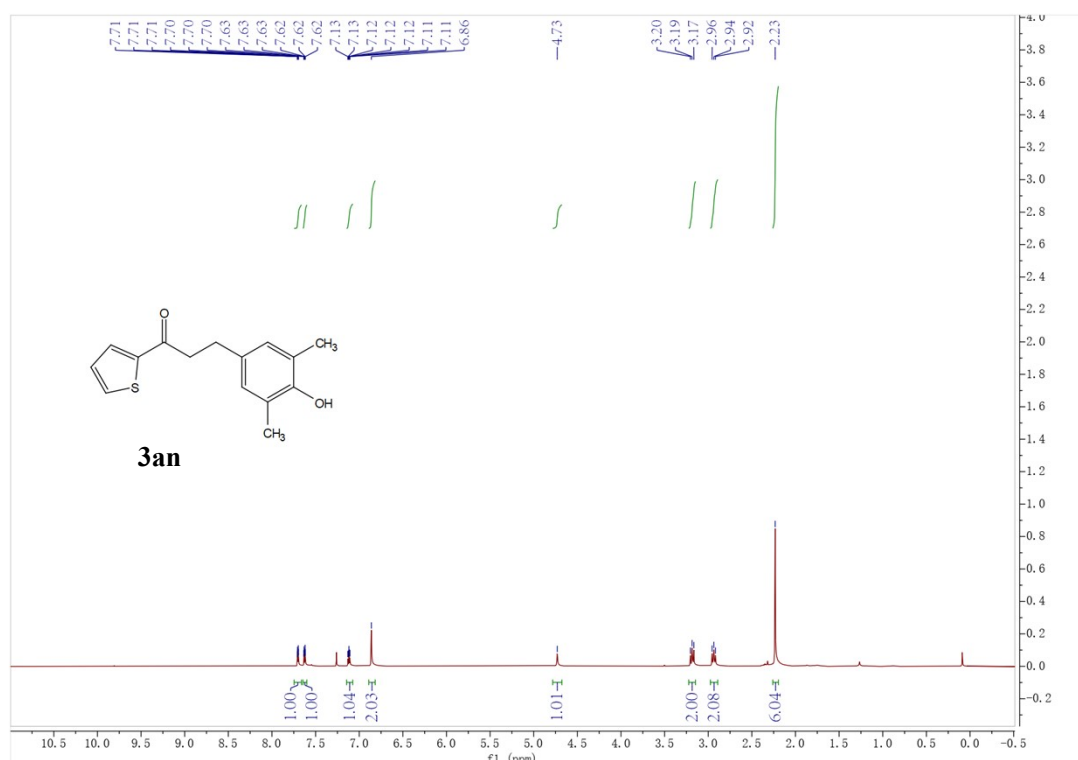
<sup>13</sup>C NMR spectrum of **3al** (CDCl<sub>3</sub>, 101 MHz)



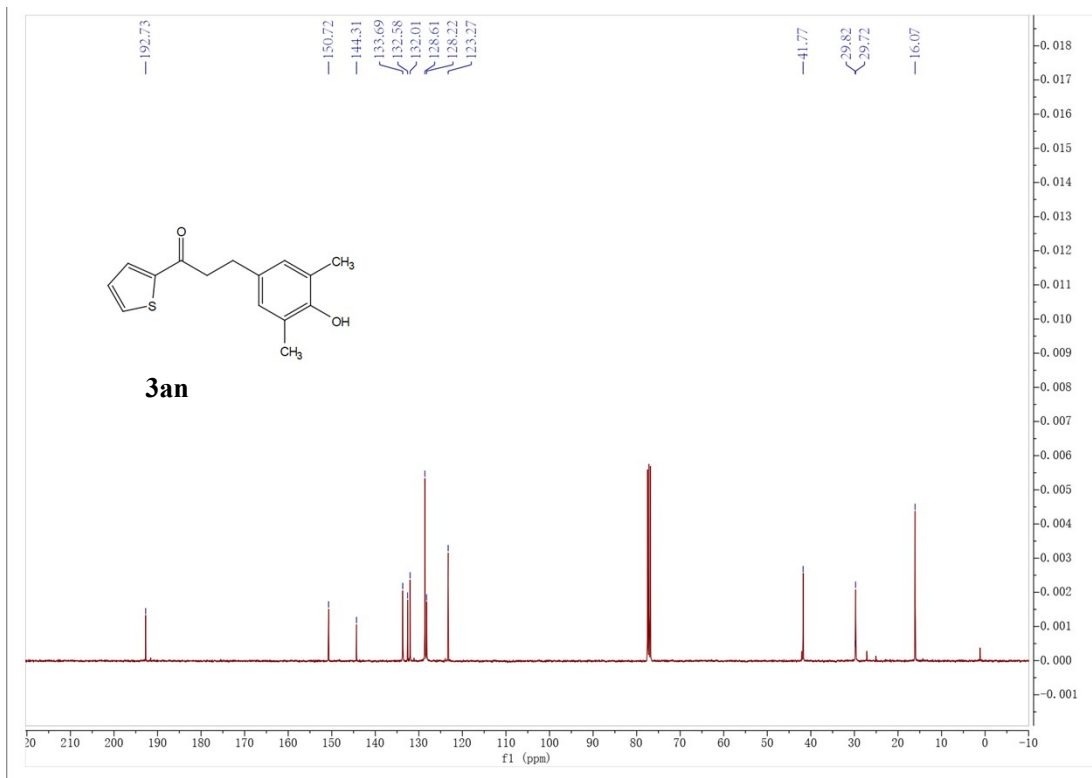
<sup>1</sup>H NMR spectrum of **3am** (CDCl<sub>3</sub>, 400 MHz)



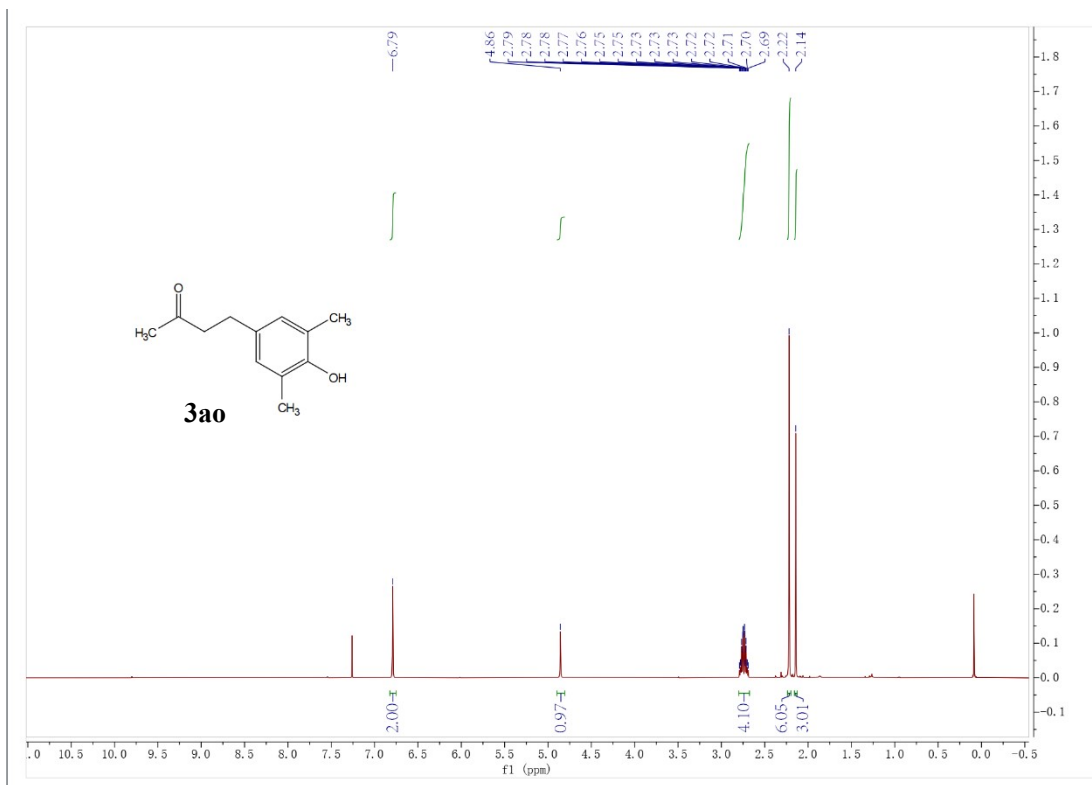
<sup>13</sup>C NMR spectrum of **3am** (CDCl<sub>3</sub>, 101 MHz)



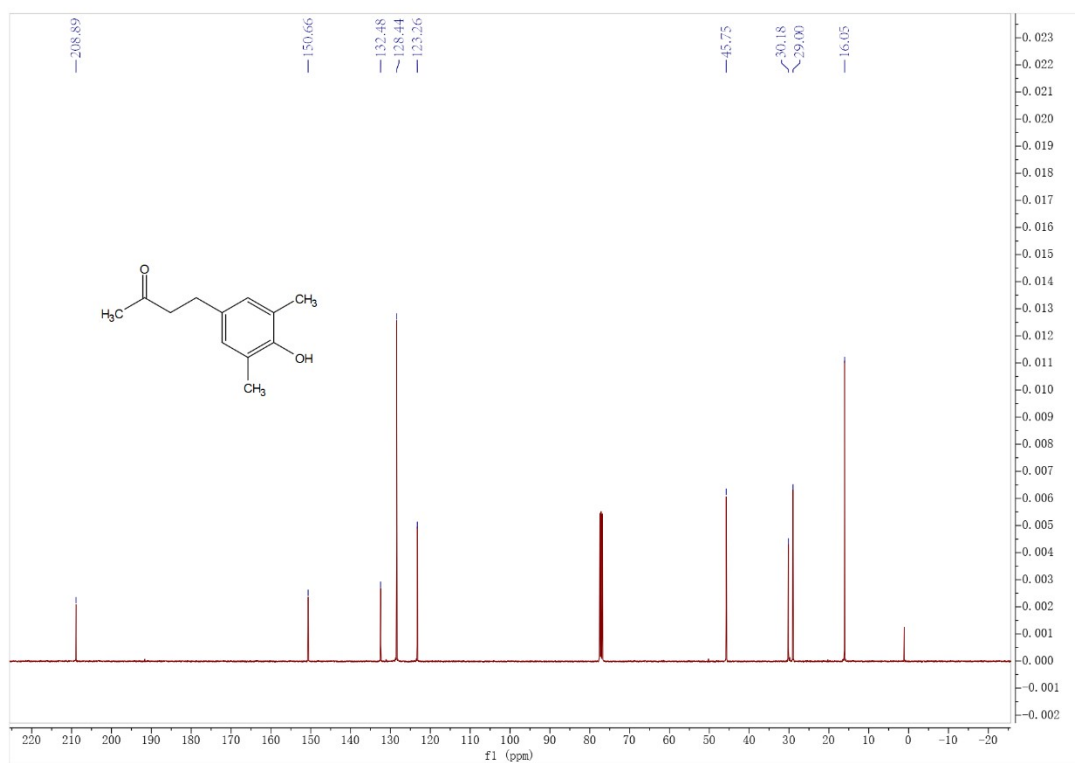
<sup>1</sup>H NMR spectrum of **3an** (CDCl<sub>3</sub>, 400 MHz)



<sup>13</sup>C NMR spectrum of **3an** (CDCl<sub>3</sub>, 101 MHz)



<sup>1</sup>H NMR spectrum of **3ao** (CDCl<sub>3</sub>, 400 MHz)



$^{13}\text{C}$  NMR spectrum of **3ao** ( $\text{CDCl}_3$ , 101 MHz)