

Syntax for search in Scopus

((TITLE-ABS-KEY (carrier) OR TITLE-ABS-KEY (support) OR TITLE-ABS-KEY (matrix) OR TITLE-ABS-KEY (matrices) OR TITLE-ABS-KEY (framework))) AND (TITLE-ABS-KEY (enzyme)) AND ((TITLE-ABS-KEY (immobiliz*) OR TITLE-ABS-KEY (immobilis*) OR TITLE-ABS-KEY (stabiliz*) OR TITLE-ABS-KEY (stabilis*) OR TITLE-ABS-KEY (adsorption) OR TITLE-ABS-KEY (adsorb*) OR TITLE-ABS-KEY (encapsulat*) OR TITLE-ABS-KEY (entrap*) OR TITLE-ABS-KEY (cross-link*) OR TITLE-ABS-KEY (covalent* bound*) OR TITLE-ABS-KEY (covalent* bind*) OR TITLE-ABS-KEY (covalent* bond*))) AND ((TITLE-ABS-KEY ("in silico") OR TITLE-ABS-KEY (bioinformatic*) OR TITLE-ABS-KEY (comput*) OR TITLE-ABS-KEY (dock*) OR TITLE-ABS-KEY (simulat*) OR TITLE-ABS-KEY (comput* model*)))

Syntax for search in Web of Science

(TS=(carrier) OR TS=(support) OR TS=(matrix) OR TS=(matrices) OR TS=(framework)) AND (TS=(enzyme)) AND (TS=(immobiliz*) OR TS=(immobilis*) OR TS=(stabiliz*) OR TS=(stabilis*) OR TS=(adsorption) OR TS=(adsorb*) OR TS=(encapsulat*) OR TS=(entrap*) OR TS=(cross-link*) OR TS=(covalent* bound*) OR TS=(covalent* bind*) OR TS=(covalent* bond*)) AND (TS=("in silico") OR TS=(bioinformatic*) OR TS=(comput*) OR TS=(dock*) OR TS=(simulat*) OR TS=(comput* model*))

Supplementary Tables

Table S1. Docking method and its detail in the included studies.

Software	Type of docking	Reference
Autodock 4.2	-	1
Autodock vina	Flexible ligand docking	2, 3
Dock v.6.522	Rigid body docking	4
Autodock 4.2	-	5
Hex 8.0	Flexible-ligand docking	6
AutoDock 3.05	-	7
AutoDock 4.2	Flexible-Ligand docking	8
-	-	9
AutodockVina	-	10
PatchDock server & FireDock server	Rigid body docking	11, 12
AutoDock Vina	Enzyme: flexible and rigid Surface modifier: flexible and rigid	13
Haddock online tool & Firedock (for refinement of the results)	-	14
Autodock vina	Flexible docking	15, 16

Table S2. The employed tools for the analysis of enzyme structure and surface.

Software/Tool and its function	Reference
PyMol: Visualization software	17
PyMol: Visualization software	18
- GetArea tool: Calculation the solvent accessible surface	19
- APBS plugin of visual molecular dynamics (VMD): Visualization of surface charges	
- PDB2PQR server: Determination of enzyme protonation state	20
- APBS software: To solve the PBE	
- PyMol software: Visualization of surface charges	
- FUZZPRO available at EBI SRS server: Searching protein motifs to recognize N-glycosylation sites	21
- GRID program version 22: Calculation of the molecular fields which are produced through interaction between the enzyme and hydrophilic/hydrophobic probes	
- DSSP program: Computation of solvent accessibility	22
- RASMOL: Visualization	
- A neural network-based program: Prediction the cysteine tendency for disulfide bond formation	
- MarvinSketch V6.2.0: Prediction of physicochemical properties	11
- MSMS package: Displaying hydrophilic surface	
- PropKa and PDB2PQR in UCSF Chimera: Determination of enzyme protonation state	
- APBS: Computation of surface electrostatic potential of enzyme	
- SwissPdb Viewer 4.1.0 software: Identification of exposed residues on the enzyme surface	23
- Maestro 10.3 software: Visualization of the hydrophobic and charged residues	
- PropKa program: Determination of enzyme protonation state	24
- APBS plugin in PyMol: Visualization of surface charges	
APBS function of PyMol: Visualization of surface charges	25
- GRASP program: Computation of surface electrostatic potential of the enzymes	26
- Maestro: Visualization of the hydrophobic and charged surface	27
- PyMol: Visualization software	28
- Calculation the solvent accessible surface via a web server (available at: http://cib.cf.ocha.ac.jp/bitool/ASA/)	

References

1. S. Bhapkar, N. Kumbhar, P. Sharma, S. Jagtap, R. Gacche, V. T. Barvkar, D. Sonune, K. D. Sonawane and U. Jadhav, *Journal of Biomolecular Structure & Dynamics*, 2021, DOI: 10.1080/07391102.2021.1975566, 11.
2. S. Ariaeenejad, H. Lanjanian, E. Motamedi, K. Kavousi, A. A. Moosavi-Movahedi and G. Hosseini Salekdeh, *Bioconjugate Chemistry*, 2020, **31**, 2158-2171.
3. M. Holyavka, D. Faizullin, V. Koroleva, S. Olshannikova, N. Zakhartchenko, Y. Zuev, M. Kondratyev, E. Zakharova and V. Artyukhov, *International Journal of Biological Macromolecules*, 2021, **180**, 161-176.
4. S. A. Ansari, M. A. Jafri, R. Satar, S. I. Ahmad and S. Chibber, *Oriental Journal of Chemistry*, 2018, **34**, 820-824.
5. M. Ali, Q. Husain, S. Sultana and M. Ahmad, *Chemosphere*, 2018, **202**, 198-207.
6. M. Khan, Q. Husain and N. Ahmad, *Journal of Chemical Technology and Biotechnology*, 2019, **94**, 837-849.
7. S. M. Lane, Z. Kuang, J. Yom, S. Arifuzzaman, J. Genzer, B. Farmer, R. Naik and R. A. Vaia, *Biomacromolecules*, 2011, **12**, 1822-1830.
8. T. Taghizadeh, A. Ameri, A. Talebian-Kiakalaieh, S. Mojtavavi, A. Ameri, H. Forootanfar, S. Tarighi and M. A. Faramarzi, *International Journal of Biological Macromolecules*, 2021, **166**, 1301-1311.
9. D. Weiser, F. Nagy, G. Bánóczy, M. Oláh, A. Farkas, A. Szilágyi, K. László, Á. Gellért, G. Marosi, S. Kemény and L. Poppe, *Green Chemistry*, 2017, **19**, 3927-3937.
10. Y. N. Franco and M. Mesa, *International Journal of Biological Macromolecules*, 2021, **192**, 736-744.
11. P. R. Lino, J. Leandro, M. Amaro, L. M. D. Gonçalves, P. Leandro and A. J. Almeida, *Pharmaceutics*, 2021, **13**, 1-17.
12. E. Shen, Y. Qu, H. Zhou, C. Kong, Q. Ma, X. Zhang and J. Zhou, *Cuihua Xuebao/Chinese Journal of Catalysis*, 2013, **34**, 723-733.
13. I. Matanovic, S. Babanova, M. S. Chavez and P. Atanassov, *Journal of Physical Chemistry B*, 2016, **120**, 3634-3641.
14. G. Murugappan and K. J. Sreeram, *Colloids and Surfaces B: Biointerfaces*, 2021, **197**.
15. M. G. Holyavka, M. S. Kondratyev, A. A. Samchenko, A. V. Kabanov, V. M. Komarov and V. G. Artyukhov, *Computers in Biology and Medicine*, 2016, **71**, 198-204.

16. M. G. Holyavka, M. S. Kondratyev, V. V. Terentyev, A. A. Samchenko, A. V. Kabanov, V. M. Komarov and V. G. Artyukhov, *Biophysics (Russian Federation)*, 2017, **62**, 5-11.
17. W. Tang, T. H. Ma, L. N. Zhou, G. Y. Wang, X. L. Wang, H. J. Ying, C. Chen and P. Wang, *Catalysis Science & Technology*, 2019, **9**, 6015-6026.
18. W. Tang, C. Chen, W. Sun, P. Wang and D. Z. Wei, *International Journal of Biological Macromolecules*, 2019, **128**, 814-824.
19. J. A. Bulos, R. Guo, Z. Wang, M. A. Delessio, J. G. Saven and I. J. Dmochowski, *Biochemistry*, 2021, DOI: 10.1021/acs.biochem.1c00515.
20. A. De Vasconcellos, J. Bergamasco Laurenti, A. H. Miller, D. A. Da Silva, F. R. De Moraes, D. A. G. Aranda and J. G. Nery, *Microporous and Mesoporous Materials*, 2015, **214**, 166-180.
21. A. Basso, P. Braiuca, S. Cantone, C. Ebert, P. Linda, P. Spizzo, P. Caimi, U. Hanefeld, G. Degrassi and L. Gardossi, *Advanced Synthesis and Catalysis*, 2007, **349**, 877-886.
22. H. El-Sherif, P. L. Martelli, R. Casadio, M. Portaccio, U. Bencivenga and D. G. Mita, *Journal of Molecular Catalysis B-Enzymatic*, 2001, **14**, 15-29.
23. F. A. Sakibaev, M. G. Holyavka, S. M. Makin and V. G. Artyukhov, *Biophysics (Russian Federation)*, 2019, **64**, 323-330.
24. Y. Qu, C. Kong, H. Zhou, E. Shen, J. Wang, W. Shen, X. Zhang, Z. Zhang, Q. Ma and J. Zhou, *Journal of Molecular Catalysis B: Enzymatic*, 2014, **99**, 136-142.
25. Y. Pan, S. Neupane, J. Farmakes, M. Bridges, J. Froberg, J. Rao, S. Y. Qian, G. Liu, Y. Choi and Z. Yang, *Nanoscale*, 2017, **9**, 3512-3523.
26. S. Hudson, E. Magner, J. Cooney and B. Kieran, *Journal of Physical Chemistry B*, 2005, **109**, 19496-19506.
27. M. H. D. Cavalcanti, L. B. Alves, A. Duarte, A. A. Mendes, J. da Silva, N. J. F. da Silveira, M. T. Escote and L. S. Virtuoso, *Chemical Engineering Journal*, 2022, **431**, 13.
28. W. Tang, H. X. Li, W. Zhang, T. H. Ma, J. F. Zhuang, P. Wang and C. Chen, *Acs Sustainable Chemistry & Engineering*, 2022, **10**, 5384-5395.