Supporting Information

Iron-catalysed Highly Selective Hydroalkoxycarbonylation of Alkynes Using CO as C1 Source

Tanuja Tewari,^{a,b} Rohit Kumar,^{a,b} Samir H. Chikkali*,^{a,b}

^aPolymer Science and Engineering Division, CSIR-National Chemical Laboratory, Dr. Homi Bhabha Road, Pune-411008, INDIA. ^bAcademy of Scientific and Innovative Research (AcSIR), Sector 19, Kamla Nehru Nagar, Ghaziabad 201002, U. P., INDIA.

Table of Contents

1. General methods and materials:
2. General procedure for iron-catalysed, hydroalkoxycarbonylation of alkynes 2
3. Substrate Scope
3a. Alkyne Scope:
3b. Alcohol scope:
4. Characterization of P1-P40 by NMR spectroscopy:7
5. Sytnthetic application of product:
6. Further functionalization:
7. Deuterium labelling experiment: 100
8. Scalability test:
9. Mechanistic studies:
9a. Kinetic analysis:
9b. Control experiment:
9c. Radical trap experiment:
9d. Procedure for EPR analysis:
10. Investigation of possible metal contaminations:
11. References:

1. General methods and materials:

All air and moisture sensitive manipulations were carried out using standard vacuum line and Schlenk techniques, or in a M-Braun glove box containing a purified argon atmosphere. Tetrahydrofuran and toluene were distilled from sodium/benzophenone under argon atmosphere. [Fe₂(CO)₉] (99%) was purchased from Alfa Aesar, DABCO and phenyl acetylene derivatives were purchased from Sigma-Aldrich. Ligands L1-L4 were purchased from Sigma Aldrich, and Spectrochem. Ligand L5¹, L6², L7³, L8⁴ were prepared according to literature procedure. Different alcohols were purchased from Sigma Aldrich and Spectrochem.

¹H and ¹³C NMR spectra were recorded on a Bruker Avance/Varian NMR 400 and 500 MHz spectrometers at 298 K unless mentioned otherwise. All spectra were obtained at ambient temperature unless mentioned otherwise. The chemical shifts (δ) were recorded in parts per million (ppm) and the coupling constants (J) in Hertz (Hz). ¹H and ¹³C NMR multiplicity and coupling constants are reported wherever applicable. ¹H and ¹³C spectra were referenced to the residual deuterated solvent peak (CHCl₃ 7.27 ppm, 77.00 ppm). Mass spectrometry (HRMS) were recorded on a Thermo Scientific Q-Exactive spectrometer. EPR spectra were recorded on Bruker EMX microX A300 ESR spectrometer.

Gas Chromatography (GC) analysis for P1-P40 were performed on an Agilent 7890B GC system using HP-05 column (30 m × 320 μ m × 0.25 μ m), split ratio 30:1, column pressure 10 psi, injector temperature of 260 °C, detector temperature of 330 °C, argon carrier gas. Temperature program: Initial temperature 70 °C, hold for 1 min.; ramp 1: 4 °C/min. to 120 °C; ramp 2: 10 °C/min. to 250 °C; ramp 3: 20 °C/min. to 320 °C, hold for 2 min. The instrument was set to an injection volume of 1 μ L, an inlet split ratio of 10:1, and inlet and detector temperatures of 250 and 320 °C, respectively. UHP-grade argon was used as carrier gas with a flow rate of 30 mL/min. Response factors for all the necessary compounds with respect to standard n-dodecane were calculated from the average of three independent GC runs.

2. General procedure for iron-catalysed, hydroalkoxycarbonylation of alkynes

In a glove box, an oven-dried vial (4 mL) containing a stirring bar was charged with [Fe₂(CO)₉] (0.005 mmol, 1.8 mg), L1-L8 (0.01 mmol), DABCO (0.89 mmol, 100 mg), aryl alkyne (0.2 mmol), and alcohol (4 mmol). The vial was then sealed with a PTFE/white rubber septum and was taken out from the glove box. Subsequently, toluene (0.38 mL), and THF (0.3 mL) were added by syringe under argon. The vial was placed in a beaker, which was transferred into a stainless steel autoclave (450 mL) and pierced with a needle. After flushing the autoclave three times with CO, a pressure of 10 bar of CO was adjusted at ambient temperature. Then, the mixture was stirred for 36 hrs at 110 °C. After that, the autoclave was cooled down to room temperature and the pressure was released carefully. The conversion was determined by the GC and product was purified by column chromatography.





Entry	[M]/L	base	time	Temp./Press.	%Conv. ^b
					(A/B/C). %
1.	FeCl ₂ / L7	DABCO	36	110/10	11/0/0
2.	FeCl ₃ / L7	DABCO	36	110/10	0
3.	Fe ₃ (CO) ₁₂ / L7	DABCO	36	110/10	5/0/0
4.	BDAFe(CO) ₃ /L7	DABCO	36	110/10	72/0/0
5.	HFe(CO) ₄ SiPh ₃ /L7	DABCO	36	110/10	58/0/0
6.	[HFe(CO) ₄]PPN/L7	DABCO	36	110/10	2/0/0

7.	Fe ₂ (CO) ₉ /L7	DABCO	36	110/10	97/0/0
8.	Fe ₂ (CO) ₉ / L1	DABCO	36	110/10	80/0/0
9.	Fe ₂ (CO) ₉ / L2	DABCO	36	110/10	12/0/0
10.	Fe ₂ (CO) ₉ / L3	DABCO	36	110/10	79/0/0
11.	Fe ₂ (CO) ₉ /L4	DABCO	36	110/10	82/0/0
12.	Fe ₂ (CO) ₉ / L5	DABCO	36	110/10	92/0/0
13.	Fe ₂ (CO) ₉ / L6	DABCO	36	110/10	90/0/0
14.	Fe ₂ (CO) ₉ / L8	DABCO	36	110/10	70/0/0
15.	Fe ₂ (CO) ₉ /L7	Cs ₂ CO ₃	36	110/10	0
16.	Fe ₂ (CO) ₉ /L7	K ₂ CO ₃	36	110/10	7/0/0
17.	Fe ₂ (CO) ₉ /L7	NaOSiMe ₃	36	110/10	25/0/0
18.	Fe ₂ (CO) ₉ /L7	NaO ^t Bu	36	110/10	72/0/0
19.	Fe ₂ (CO) ₉ /L7	DBU	36	110/10	24/0/0
20.	Fe ₂ (CO) ₉ /L7	Et ₃ N	36	110/10	59/0/0
21.	/L7	DABCO	36	110/10	0
22.	Fe ₂ (CO) ₉ /	DABCO	36	110/10	76/0/0
23.	Fe ₂ (CO) ₉ /L7	DABCO	16	110/10	36/0/0
24.	Fe ₂ (CO) ₉ /L7	DABCO	24	110/10	78/0/0
25. °	Fe ₂ (CO) ₉ /L7	DABCO	36	110/10	62/0/0

26.	Fe ₂ (CO) ₉ /L7	DABCO	36	110/5	80/0/0
27.	Fe ₂ (CO) ₉ /L7	DABCO	36	110/20	92/0/0
28.	Fe ₂ (CO) ₉ /L7	DABCO	36	80/10	45/0/0
29.	Fe ₂ (CO) ₉ /L7	DABCO	36	120/10	97/0/0
30.	Fe ₂ (CO) ₉ /	DABCO	48	110/10	84/0/0
31.	Fe ₂ (CO) ₉ /	DABCO	36	120/10	86/0/0

Conditions: a) [Fe] = 0.0018 g (0.005 mmol) (5 mol%), L = 0.01 mmol, DABCO = 4.5 equiv, Phenyl acetylene = 0.02 mL (0.2 mmol), Solvent = Tol.:THF, IPA = 0.3 ml (3.9 mmol); b) conversion determined by GC; c) 2.2 equiv DABCO. IPA: Isopropyl alcohol; Tol: Toluene; THF: Tetrahydrofuran.



Figure S1: Various ligands for hydroalkoxycarbonylation of alkyne

3. Substrate Scope

3a. Alkyne Scope:



Figure S2A: A set of hydroalkoxycarbonylated products from various alkyne demonstrating the scope of the reaction. **Conditions:** a) $[Fe_2(CO)_9] = 0.0018 \text{ g} (0.005 \text{ mmol}) (5 \text{ mol}\%), L = 0.005 \text{ g} (0.01 \text{ mmol}), DABCO = 4.5 equivalent, Phenyl acetylene = 0.02 mL (0.2 mmol), Solvent = Tol.:THF, IPA = 0.3 ml (3.9 mmol); b) Conversion is determined by GC and isolated yield given in parenthesis.$

3b. Alcohol scope:



Figure S2B: A set of hydroalkoxycarbonylated products with various alcohol demonstrating the scope of the reaction. **Conditions:** a) $[Fe_2(CO)_9] = 0.0018 \text{ g} (0.005 \text{ mmol}) (5 \text{ mol}\%), L7 = 0.005 \text{ g} (0.01 \text{ mmol}), DABCO = 4.5 equiv, Phenyl acetylene = 0.02 mL (0.2 mmol); b) conversion determined by GC and isolated yield given in parenthesis.$

4. Characterization of P1-P40 by NMR spectroscopy:

Isopropyl cinnamate P1:



Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 90% yield (34.2 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P1 = 15.5 min.

¹H NMR (500 MHz, CDCl₃): $\delta = 7.62$ (d, J = 16.01 Hz, 1H), 7.46-7.43 (m, 2H), 7.30-7.29 (m, 3H), 6.37 (d, J = 16.01 Hz, 1H), 5.14-5.05 (m, 1H), 1.26 (d, J = 6.32 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 166.4$, 144.3, 134.5, 130.1, 128.8, 120.0, 118.8, 67.7, 21.9. The above data is in accordance with previous reports for compound P1.⁵



Figure S3: ¹H NMR spectrum of P1 in CDCl₃.



Figure S4: ¹³C NMR spectrum of P1 in CDCl₃.

Isopropyl (E)-3-(p-tolyl)acrylate P2:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 64% yield (26.15 mg). GC retention time for Ethynyl toluene = 3.26 min.; hydroalkoxycarbonylated product P2 = 17.38 min.



¹**H NMR** (400 MHz, CDCl₃): $\delta = 7.62$ (d, J = 15.99 Hz, 1H), 7.40 (d, J = 8.15 Hz, 2H), 7.16 (d, J = 7.98 Hz, 2H), 6.35 (d, J = 15.98 Hz, 1H), 5.12 (sep., J = 6.25 Hz, 1H), 2.35 (s, 3H), 1.29 (d, J = 6.27 Hz, 6H). ¹³C **NMR** (100 MHz, CDCl₃): $\delta = 166.8$ (C_q), 144.4, 140.6, 131.9, 129.7, 128.1, 117.9, 67.8, 22.1, 21.6 (CH₃). The above data is in accordance with previous reports for compound P2.⁶



Figure S5: GC trace of P2.



Figure S6: ¹H NMR spectrum of P2 in CDCl₃.



Figure S7: ¹³C NMR spectrum of P2 in CDCl₃.

Isopropyl (E)-3-(4-pentylphenyl)acrylate P3:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 84% yield (43.7 mg). GC retention time for 4-pentyl phenyl acetylene = 12.67 min.; hydroalkoxycarbonylated product P3 = 22.43 minutes.

¹**H NMR** (**500 MHz**, **CDCl**₃): $\delta = 7.65$ (d, J = 15.99 Hz, 1H), 7.43 (d, J = 8.12 Hz, 2H), 7.18 (d, J = 8.09 Hz, 2H), 6.37 (d, J = 15.98 Hz, 1H), 5.14 (sep, J = 12.51 Hz, 1H), 2.61 (t, J = 15.48 Hz, 2H), 1.65-1.58 (m, 2H), 1.33-1.30 (m, 10H), 0.89 (t, J = 6.90 Hz, 3H). ¹³**C NMR** (**125 MHz**, **CDCl**₃): $\delta = 166.9$ (C_q), 145.7, 144.5, 132.2, 129.1, 128.2, 117.8, 67.8, 35.9, 31.6, 31.1, 22.7, 22.1, 14.1 (CH₃).

Figure S8: GC trace of P3.

Figure S9: ¹H NMR of P3 in CDCl₃.

Figure S10: ¹³C NMR of P3 in CDCl₃.

Isopropyl (E)-3-(4-methoxyphenyl)acrylate P4:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 74% yield (32.5 mg). GC retention time for 4methoxy phenylacetylene = 5.93 min.; product P4 = 19.57 min.

hydroalkoxy carbonylated

¹**H** NMR (400 MHz, CDCl₃): $\delta = 7.61$ (d, J = 15.94 Hz, 1H), 7.46 (d, J = 8.41 Hz, 2H), 6.88 (d, J = 8.47 Hz, 2H), 6.27 (d, J = 15.94 Hz, 2H), 5.15-5.09 (m, 1H), 3.82 (s, 3H), 1.29 (d, J = 6.17 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.0$ (C_q), 161.4, 144.1, 129.8, 127.4, 116.5, 114.4, 67.7, 55.5, 22.1 (CH₃). The above data is in accordance with previous reports for compound P4.⁷

Figure S11: GC trace of P4.

Figure S12: ¹H NMR spectrum of P4 in CDCl₃.

Figure S13: ¹³C NMR spectrum of P4 in CDCl₃.

Isopropyl (E)-3-(2-methoxyphenyl)acrylate P5:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 88% yield (38.7 mg). GC retention time for 2-methoxyphenylacetylene = 6.02 min.; hydroalkoxycarbonylated product P5 = 19.14 min.

¹**H NMR (400 MHz, CDCl₃):** δ = 7.96 (d, *J* = 16.03 Hz, 1H), 7.47 (d, *J* = 7.33 Hz, 1H), 7.31 (t, *J* = 7.79 Hz, 1H), 6.94-6.87 (m, 2H), 6.49 (d, *J* = 15.57 Hz, 1H), 5.14-5.09 (m, 1H), 3.85 (s, 3H), 1.29 (d, *J* = 5.04 Hz, 6H). ¹³**C NMR (100 MHz, CDCl₃):** δ = 167.1 (C_q), 158.4, 139.8, 131.4, 129.0, 123.6, 120.8, 119.4, 111.2, 67.6, 55.5, 22.1 (CH₃).

/0044/1	94.55	2077311	95.50
8107912	100.00	2807697	100.00
	8107912	8107912 100.00	8107912 100.00 2807697

Figure S14: GC trace of P5.

Figure S15: ¹H NMR spectrum of P5 in CDCl₃.

Figure S16: ¹³C NMR spectrum of P5 in CDCl₃.

Isopropyl (E)-3-(3,4-dimethoxyphenyl)acrylate P6:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 85% yield (42.5 mg). GC retention time for 3,4-dimethoxyphenylacetylene = 11.82 min.; hydroalkoxycarbonylated product P6 = 21.72 min.

¹**H NMR** (**500 MHz**, **CDCl**₃): $\delta = 7.60$ (d, J = 15.91 Hz, 1H), 7.08 (d, J = 10.17 Hz, 1H), 7.04 (s, 1H), 6.85 (d, J = 8.27 Hz, 1H), 6.28 (d, J = 15.90 Hz, 1H), 5.13 (sep., J = 12.50 Hz, 1H), 3.90 (s, 6H), 1.30 (d, J = 6.26 Hz, 6H). ¹³**C NMR** (**125 MHz**, **CDCl**₃): $\delta = 166.8$ (C_q), 151.1, 149.2, 144.3, 127.6, 122.6, 116.5, 111.1, 109.6, 67.6, 56.0, 55.9, 22.0. The above data is in accordance with previous reports for compound P6.⁸

Front Signal Results				
Retention Time	Area	Area %	Height	Height %
11.820	106177	2.99	34303	2.36
15.355	157974	4.45	40134	2.76
21.726	3289710	92.57	1379158	94.88
Totals				
	3553861	100.00	1453595	100.00

Figure S17: GC trace of P6.

Figure S18A: ¹H NMR spectrum of P6 in CDCl₃.

Figure S18B: ¹³C NMR spectrum of P6 in CDCl₃.

Isopropyl (E)-3-(3,5-dimethoxyphenyl)acrylate P7:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 66% yield (33.3 mg). GC retention time for 3,5 di-methoxy phenylacetylene = 15.5 min.; hydroalkoxycarbonylated product P7 = 21.65 min.

¹**H** NMR (400 MHz, CDCl₃): $\delta = 7.57$ (d, J = 15.94 Hz, 1H), 6.65 (br s, 2H), 6.47 (s, 1H), 6.37 (d, J = 15.94 Hz, 1H), 5.13 (sep, J = 12.47 Hz, 1H), 3.79 (s, 6H), 1.30 (d, J = 6.25 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.5$ (C_q), 161.1, 144.4, 136.5, 119.4, 106.0, 102.6, 67.9, 55.5, 22.1(CH₃).

Figure S19: GC trace of P7.

Figure S20: ¹H NMR spectrum of P7 in CDCl₃.

Figure S21: ¹³C NMR spectrum of P7 in CDCl_{3.}

Isopropyl (E)-3-(4-phenoxyphenyl)acrylate P8:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 70% yield (39.5 mg). GC retention time for 4-phenoxy phenyl acetylene = 17.24 min.; hydroalkoxycarbonylated product P8 = 24.49 min.

¹H NMR (400 MHz, CDCl₃): δ = 7.61 (d, *J* = 15.97 Hz, 1H), 7.46 (d, *J* = 8.50 Hz, 2H), 7.34 (t, J = 7.77 Hz, 2H), 7.13 (m, 1H), 7.02 (d, *J* = 7.85 Hz, 2H), 6.95 (d, *J* = 8.48 Hz, 2H), 6.30 (d, *J* = 15.96 Hz, 1H), 5.12 (sep, *J* = 6.20 Hz, 1H), 1.29 (d, *J* = 6.21 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃): δ = 166.77 (C_q), 159.5, 156.3, 143.7, 130.1, 129.8, 129.4, 124.2, 119.8, 118.5, 117.6, 67.8, 22.1 (CH₃).

Figure S22: GC trace of P8.

Figure S23: ¹H NMR spectrum of P8 in CDCl_{3.}

Figure S24: ¹³C NMR spectrum of P8 in CDCl₃.

Isopropyl (E)-3-(4-(dimethylamino)phenyl)acrylate P9:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 89% yield (41.5 mg). GC retention time for 4-Dimethylaminophenylacetylene = 11.53 min.; hydroalkoxycarbonylated product P9 = 22.23 min.

¹**H NMR** (**500 MHz**, **CDCl**₃): $\delta = 7.60$ (d, J = 15.84 Hz, 1H), 7.41 (d, J = 8.85 Hz, 2H), 6.66 (d, J = 8.87 Hz, 2H), 6.20 (d, J = 15.83 Hz, 1H), 5.12 (sep, J = 12.51 Hz, 1H), 3.01 (s, 6H), 1.29 (d, J = 6.26 Hz, 6H). ¹³**C NMR** (**125 MHz**, **CDCl**₃): $\delta = 167.6$ (C_q), 151.8, 145.0, 129.8, 122., 113.3, 111.9, 67.4, 40.3, 22.2 (CH₃).

Figure S25: GC trace of P9.

Figure S27: ¹³C NMR spectrum of P9 in CDCl_{3.}

Isopropyl (E)-3-(4-fluorophenyl)acrylate P10A:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 85% yield (35.4 mg). GC retention time for 4-fluoro phenyl acetylene = 2.15 min.; hydroalkoxycarbonylated product P10 = 15.32 min.

¹H NMR (400 MHz, CDCl₃): $\delta = 7.61$ (d, J = 16.00 Hz, 1H), 7.51-7.47 (m, 2H), 7.05(t, J = 8.63 Hz, 2H), 6.32 (d, J = 15.98 Hz, 1H), 5.12 (sep, J = 12.52 Hz, 1H), 1.30 (d, J = 6.27 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.5$ (C_q), 163.9 (Cq, d, ¹*J*_{C-F} = 251.04 Hz), 143.1,

130.9, 130.0 (d, ${}^{3}J_{C-F} = 8.45$ Hz), 118.7, 116.1 (d, ${}^{2}J_{C-F} = 22.07$ Hz), 68.0, 22.1 (CH₃). The above data is in accordance with previous reports for compound P10.⁹

Figure S28: GC trace of P10A.

Figure S29A: ¹H NMR spectrum of P10A in CDCl₃.

Isopropyl (E)-3-(4-(trifluoromethyl)phenyl)acrylate P10B:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a yellow liquid was isolated in 26% yield (13.4 mg). GC retention time for 4-trifluoromethyl phenyl acetylene = 2.5 min.; hydroalkoxycarbonylated product P10b = 15.32 min.

¹H NMR (500 MHz, CDCl₃): δ = 7.68-7.60 (m, 3H), 7.52-7.37 (m, 2H), 6.48 (d, J = 16.01 Hz, 1H), 5.18-5.13 (m, 1H), 1.32 (d, J = 6.17 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃): δ = 166.1, 144.5, 142.6, 129.0, 128.3, 126.0 (*J*_{C-F} = 3.69 Hz), 121.6, 119.0, 68.4, 22.1.

Kesults				
Retention Time	Area	Area %	Height	Height %
2.597	561296	14.12	134863	15.09
12.599	2179627	54.84	447701	50.09
15.291	1233645	31.04	311250	34.82
Totals				
	3974568	100.00	893814	100.00

Figure S30A: GC trace of P10B.

Figure S30B: ¹H NMR spectrum of P10B in CDCl₃.

Figure S30C: ¹³C NMR spectrum of P10B in CDCl₃.

Isopropyl (E)-3-(4-chlorophenyl)acrylate P11:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a yellow orange liquid was isolated in 86% yield (38.6 mg).

¹**H NMR (400 MHz, CDCl₃):** δ = 7.60 (d, *J* = 16.01 Hz, 1H), 7.44 (d, *J* = 8.38 Hz, 2H), 7.34 (d, J = 8.51 Hz, 2H), 6.38 (d, J = 16.00 Hz, 1H), 5.13 (sep, J = 12.51 Hz, 1H), 7.30 (d, J = 6.26 Hz, 6H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.4, 143.0, 136.2, 133.2, 129.3, 129.3, 119.6,$ 68.1, 22.1. The above data is in accordance with previous reports for compound P11.¹⁰

Figure S31: ¹H NMR spectrum of P11 in CDCl₃.

Isopropyl (E)-3-(naphthalen-1-yl)acrylate P12:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 72% yield (34.6 mg). GC retention time for 1-ethynyl naphthalene = 12.61 min.; hydroalkoxycarbonylated product P12 = 22.04 min.

¹**H NMR** (**500 MHz**, **CDCl**₃): $\delta = 8.50$ (d, J = 15.74 Hz, 1H), 8.19 (d, J = 8.35 Hz, 1H), 7.87 (t, J = 13.66 Hz, 2H), 7.74 (d, J = 7.22 Hz, 1H), 7.59-7.45 (m, 3H), 6.50 (d, J = 15.74 Hz, 1H), 5.19 (sep, J = 12.49 Hz, 1H), 1.35 (d, J = 6.26 Hz, 6H). ¹³**C NMR** (**125 MHz**, **CDCl**₃): $\delta = 166.6, 141.5, 133.8, 132.0, 131.5, 130.5, 128.8, 126.9, 126.3, 125.6, 125.1, 123.6, 121.6, 68.1, 22.1.$

Figure S33: GC trace of P12.

8.52 8.48 8.48 8.21 8.21 7.176 7.7.76 7.7.76 7.7.57 7.57 7.57 7.57 7	5.22 5.20 5.19 5.14 5.14	1.36 1.34
	° C	
		hul
9 8 7 6	5 4	3 2 1 0 ppm

Figure S34: ¹H NMR spectrum of P12 in CDCl₃.

Figure S35: ¹³C NMR spectrum of P12 in CDCl₃.

Isopropyl (E)-3-(naphthalen-2-yl)acrylate P13:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 82% yield (39.4 mg). GC retention time for 2-ethynylnaphthalene = 12.56 min.; hydroalkoxycarbonylated product P13 = 22.54 min.

¹**H NMR (400 MHz, CDCl₃):** δ = 7.85-7.74 (m, 5H), 7.59 (d, *J* = 8.36 Hz, 1H), 7.45-7.43 (m, 2H), 6.48 (d, *J* = 15.96 Hz, 1H), 5.17-5.10 (m, 1H), 1.29 (d, *J* = 6.23 Hz, 6H).

Figure S36: GC trace of P13.

Figure S37: ¹H NMR spectrum of P13 in CDCl₃.
Isopropyl (E)-3-(6-methoxynaphthalen-2-yl)acrylate P14:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 84% yield (45.4 mg). GC retention time for 2-ethynyl-6-methoxynaphthalene = 18.03 min.; hydroalkoxycarbonylated product P14 = 25.02 min.



¹**H** NMR (500 MHz, CDCl₃): $\delta = 7.79-7.57$ (m, 5H), 7.12 (d, J = 8.87 Hz, 1H), 7.07 (br s, 1H), 6.45 (d, J = 15.91 Hz, 1H), 5.14 (sep. J = 12.45 Hz, 1H), 3.87 (s, 3H), 1.30 (d, J = 6.24 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 166.8$ (C_q), 158.8, 144.6, 135.7, 130.2, 130.0, 129.7, 128.8, 127.6, 124.3, 119.5, 117.9, 106.1, 67.8, 55.5, 22.1 (CH₃).



Figure S38: GC trace of P14.



Figure S39A: ¹H NMR spectrum of P14 in CDCl₃.



Figure S39B: ¹³C NMR spectrum of P14 in CDCl₃.

2-methyl-3-phenylacrylaldehyde or 2-phenylbut-2-enal P15A:

GC retention time for methylphenylacetylene = 4.56 min.; hydroalkoxycarbonylated product P15 = 16.35 min.





Figure S40A: GC trace of P15.

isopropyl 2-methyleneoctanoate or isopropyl (E)-non-2-enoate P15B:

GC retention time for 1-octyne = 1.8 min.; hydroalkoxycarbonylated product P15B = 12.2 min.





Retention Time	Area	Area %
1.784	946402	8.74
2.598	1166483	10.77
12.233	8713289	80.48
Totals		
	10826174	Active to Wind
		Go to Settings to a

Figure S40B: GC trace of P15B.



Figure S40C: GC-MS chromatogram of P15B.

Methyl cinnamate P16:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 55% yield (17.7 mg). GC retention time for phenyl acetylene = 2.2 min.; hydroalkoxycarbonylated product P16 = 12.24 min.



¹**H** NMR (500 MHz, CDCl₃): $\delta = 7.69$ (d, J = 16.03 Hz, 1H), 7.53-7.51 (m, 2H), 7.39-7.37 (m, 3H), 6.44 (d, J = 16.03 Hz, 1H), 3.81 (s, 3H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.6$ (C_q), 145.1, 134.6, 130.5, 129.1, 128.2, 117.9, 51.9 (CH₃). The above data is in accordance with previous reports for compound P16.⁵



Figure S41: GC trace of P16.



Figure S42: ¹H NMR spectrum of P16 in CDCl₃.



Figure S43: ¹³C NMR spectrum of P16 in CDCl₃.

Ethyl cinnamate P17:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 78% yield (27.4 mg). GC retention time for phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P17 = 14.53 min.



¹H NMR (400 MHz, CDCl₃): $\delta = 7.67$ (d, J = 16.02 Hz. 1H), 7.52-7.49 (m, 2H), 7.37-7.35 (m, 3H), 6.42 (d, J = 16.02 Hz, 1H), 4.24 (q, J = 7.13 Hz, 2H), 1.32 (t, J = 7.13 Hz, 3H). The above data is in accordance with previous reports for compound P17.⁵



Results				
Retention Time	Area	Area %	Height	Height %
2.207	1919756	12.31	510980	14.44
14.533	13674606	87.69	3027233	85.56
Totals				
	15594362	100.00	3538213	100.00

Figure S44: GC trace of P17.



Figure S45: ¹H NMR spectrum of P17 in CDCl₃.



n-butyl cinnamate P18:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 75% yield (30.6 mg). GC retention time for phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P18 = 18.14 min.



¹H NMR (500 MHz, CDCl₃): $\delta = 7.68$ (d, J = 16.02 Hz, 1H), 7.53-7.51 (m, 2H), 7.38-7.37 (m, 3H), 6.44 (d, J = 16.01 Hz, 1H), 4.21 (t, J = 6.69 Hz, 2H), 1.73-1.66 (m, 2H), 1.49-1.39 (m, 2H), 0.96 (t, J = 7.39 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.2$ (C_q), 144.7, 134.6, 130.3, 129.0, 128.2, 118.4, 64.6, 30.9, 19.3, 13.9 (CH₃). The above data is in accordance with previous reports for compound P18.¹¹



100.00

7339534

100.00

25131079

Figure S47: GC trace of P18.



Figure S48: ¹H NMR spectrum of P18 in CDCl₃.



Figure S49: ¹³C NMR spectrum of P19 in CDCl₃.

Isopentyl cinnamate P19:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 51% yield (22.26 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P19 = 18.92 min.



¹**H** NMR (500 MHz, CDCl₃): $\delta = 7.66$ (d, J = 16.00 Hz, 1H), 7.50 (s, 2H), 7.35 (s, 3H), 6.42 (d, J = 15.99 Hz, 1H), 4.22 (t, J = 6.64 Hz, 2H), 1.75-1.70 (m, 1H), 1.61-1.56 (m, 2H), 0.94 (d, J = 6.46 Hz, 6H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.2$ (C_q), 144.7, 134.6, 130.3, 128.9, 128.2, 118.4, 63.3, 37.6, 25.2, 22.6. The above data is in accordance with previous reports for compound P19.⁵



Figure S50: GC trace of P19.



Figure S51: ¹H NMR spectrum of P19 in CDCl₃.



Figure S52: ¹³C NMR spectrum of P19 in CDCl₃.

2-ethylhexyl cinnamate P20:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 86% yield (44.8 mg). GC retention time for phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P20 = 21.95 min.



¹**H** NMR (500 MHz, CDCl₃): $\delta = 7.68$ (d, J = 16.01 Hz, 1H), 7.55-7.22 (m, 2H), 7.39-7.38 (m, 3H), 6.45 (d, J = 16.01 Hz, 1H), 4.13 (d, J = 2.50 Hz, 1H), 4.11 (d, J = 2.75 Hz, 1H) 1.59

(br s, 4H), 1.32-1.31 (m, 6H), 0.92-0.90 (m, 5H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.4$ (C_q), 144.7, 134.6, 130.4, 129.0, 128.2, 118.5, 67.2, 39.0, 30.6, 29.1, 24.0, 23.2, 14.3, 11.2 (CH₃). The above data is in accordance with previous reports for compound P20.¹²



Figure S53: GC trace of P20.



Figure S54: ¹H NMR spectrum of P20 in CDCl₃.



Figure S55: ¹³C NMR spectrum of P20 in CDCl₃.

3-phenylpropyl cinnamate P21:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 79% yield (42.1 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P21 = 24.50 min.



¹H NMR (400 MHz, CDCl₃): $\delta = 7.63$ (d, J = 16.02 Hz, 1H), 7.46-7.44 (m, 2H), 7.30-7.29 (m, 3H), 7.25-7.21 (m, 2H), 7.15-7.11 (m, 3H), 6.39 (d, J = 16.01 Hz, 1H), 4.17 (t, J = 6.54 Hz, 2H), 2.68 (t, J = 15.35, Hz, 2H), 1.97 (m, 2H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.9$ (C_q), 144.7, 141.3, 134.5, 130.3, 128.9, 128.5, 128.5, 128.1, 126.1, 118.2, 63.9, 32.3, 30.3. The above data is in accordance with previous reports for compound P21.¹³



Figure S56: GC trace of P21.



Figure S57: ¹H NMR spectrum of P21 in CDCl₃.



Phenyl cinnamate P22:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 62% yield (27.8 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P22 = 21.56 min.



¹H NMR (400 MHz, CDCl₃): $\delta = 7.86$ (d, J = 15.97 Hz, 1H), 7.57 (m, 2H), 7.41-7.40 (m, 5H), 7.24 (t, J = 7.14 Hz, 1H), 7.16 (d, J = 7.89 Hz, 2H), 6.63 (d, J = 15.97 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 165.56$ (C_q), 150.9, 146.7, 134.4, 130.9, 129.6, 129.2, 128.5, 125.9, 121.8, 117.5. The above data is in accordance with previous reports for compound P22.¹⁴



Figure S59: GC trace of P22.



Figure S60: ¹H NMR spectrum of P22 in CDCl₃.



Figure S61: ¹³C NMR spectrum of P22 in CDCl₃.

[1,1'-biphenyl]-4-yl cinnamate P23:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 92% yield (55.3 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P23 = 28.09 min.



¹H NMR (500 MHz, CDCl₃): $\delta = 7.89$ (d, J = 16.01 Hz, 1H), 7.62-7.56 (m, 6H), 7.45-7.41 (m, 5H), 7.36-7.32 (m, 1H), 7.25-7.23 (m, 2H), 6.65 (d, J = 16.00 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 165.6$ (C_q), 150.4, 146.9, 140.6, 139.1, 134.3, 130.9, 129.2, 128.9, 128.5, 128.4, 127.5, 127.3, 122.1, 117.4. The above data is in accordance with previous reports for compound P23.¹⁵



Figure S62: GC trace of P23.





Figure S63: ¹H NMR spectrum of P23 in CDCl₃.



Figure S64: ¹³C NMR spectrum of P23 in CDCl₃.

Benzyl cinnamate P24:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 86% yield (40.9 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P24 = 22.65 min.



¹H NMR (400 MHz, CDCl₃): $\delta = 7.73$ (d, J = 16.02 Hz, 1H), 7.533-7.51 (m, 2H), 7.44-7.37 (m, 8H), 6.49 (d, J = 16.06 Hz, 1H), 5.26 (s, 2H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.9$ (C_q), 136.2, 134.6, 130.5, 129.1, 128.8, 128.6, 128.5, 128.3, 118.1, 65.5. The above data is in accordance with previous reports for compound P24.⁵



22.050	41001452	94.84	990/903	95.05
Totals				
	43929332	100.00	10488907	100.00

Figure S65: GC trace of P24.



Figure S66: ¹H NMR spectrum of P24 in CDCl₃.



Figure S67: ¹³C NMR spectrum of P24 in CDCl₃.

4-methoxybenzyl cinnamate P25:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 72% yield (38.6 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P25 = 25.01 min.



¹H NMR (500 MHz, CDCl₃): $\delta = 7.68$ (d, J = 16.02 Hz, 1H), 7.48-7.45 (m, 2H), 7.34-7.32 (m, 5H), 6.88 (d, J = 8.69 Hz, 2H), 6.43 (d, J = 16.01 Hz, 1H), 5.16 (s, 2H), 3.77 (s, 3H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 166.9$ (C_q), 159.8, 145.1, 134.5, 130.4, 130.3, 128.9, 128.3, 128.2, 118.1, 114.1, 66.3, 55.4. The above data is in accordance with previous reports for compound P25.⁵



Figure S68: GC trace of P25.



Figure S69: ¹H NMR spectrum of P25 in CDCl₃.



Figure S70: ¹³C NMR spectrum of P25 in CDCl₃.

4-(trifluoromethyl)benzyl cinnamate P26:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 88% yield (54 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P26 = 22.45 min.



¹H NMR (200 MHz, CDCl₃): $\delta = 7.74$ (d, J = 16.03 Hz, 1H), 7.66-7.61 (m, 2H), 7.54-7.50 (m, 4H), 7.40-7.37 (m, 3H), 6.49 (d, J = 16.02 Hz, 1H), 5.29 (s, 2H). The above data is in accordance with previous reports for compound P26.¹⁶



Figure S71: GC trace of P26.



Figure S72: ¹H NMR spectrum of P26 in CDCl₃.

2-bromobenzyl cinnamate P27:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 87% yield (55.2 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P27 = 25.14 min.



¹H NMR (500 MHz, CDCl₃): $\delta = 7.72$ (d, J = 16.02 Hz, 1H), 7.55 (d, J = 7.96 Hz, 1H), 7.49-7.47 (m, 2H), 7.43 (d, J = 7.58 Hz, 1H), 7.35-7.33 (m, 3H), 7.29 (t, J = 7.51 Hz, 1H), 7.15 (t, J = 7.61 Hz, 1H), 6.48 (d, J = 16.0 Hz, 1H), 5.29 (s, 2H). ¹³C NMR (500 MHz, CDCl₃): $\delta = 166.6, 145.6, 135.4, 134.3, 132.9, 130.5, 129.9, 129.8, 128.9, 128.2, 127.6, 123.5, 117.6, 65.9.$ The above data is in accordance with previous reports for compound P27.⁵



Figure S73: GC trace of P27.



Figure S74: ¹H NMR spectrum of P27 in CDCl₃.



Figure S75: ¹³C NMR spectrum of P27 in CDCl₃.

Benzhydryl cinnamate P28:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 71% yield (44.6 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P28 = 27.20 min.



¹**H** NMR (400 MHz, CDCl₃): $\delta = 7.73$ (d, J = 15.98 Hz, 1H), 7.50 (br s, 2H), 7.37-7.27 (m, 13H), 7.01 (s, 1H), 6.54 (d, J = 15.99 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.1$ (C_q), 145.6, 140.4, 134.5, 130.5, 129.0, 128.7, 128.3, 128.1, 127.3, 118.2, 77.1. The above data is in accordance with previous reports for compound P28.⁵



Figure S76: GC trace of P28.


Figure S77: ¹H NMR spectrum of P28 in CDCl₃.



Figure S78: ¹³C NMR spectrum of P28 in CDCl₃.

2,2,2-trifluoroethyl cinnamate P29:



GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P29 = 11.20 min.



Figure S79: GC chromatogram for P29.

Pyridin-2-ylmethyl cinnamate P30:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 70% yield (33.5 mg). GC retention time for Phenylacetylene = 2.17 min.; hydroalkoxycarbonylated product P30 = 23.01 min.



¹**H** NMR (400 MHz, CDCl₃): $\delta = 8.61$ (br s, 1H), 7.76 (d, J = 16.02 Hz, 1H), 7.71 (t, J = 7.70 Hz, 1H), 7.54-7,51 (m, 2H), 7.41-7.37 (m, 4H), 7.26-7.22 (m, 1H), 6.54 (d, J = 16.02 Hz, 1H), 5.37 (s, 2H). ¹³C NMR (400 MHz, CDCl₃): $\delta = 166.7$, 156.1, 149.6, 145.7, 136.9, 134.4, 130.6, 129.1, 128.3, 123.0, 122.0, 117.7, 66.9. The above data is in accordance with previous reports for compound P30.^{13b}



Figure S80: GC trace of P30.



Figure S81: ¹H NMR spectrum of P30 in CDCl₃.



Figure S82: ¹³C NMR spectrum of P30 in CDCl₃.

3,7-dimethyloct-6-en-1-yl cinnamate P31:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 71% yield (40.6 mg). GC retention time for Phenylacetylene =2.2 min.; hydroalkoxycarbonylated product P31 = 23.93 min.



¹H NMR (500 MHz, CDCl₃): $\delta = 7.68$ (d, J = 16.02 Hz, 1H), 7.54-7.51 (m, 2H), 7.39-7.37 (m, 3H), 6.44 (d, J = 16.02 Hz, 1H), 5.10 (t, J = 7.11 Hz, 1H), 4.29-4.19 (m, 2H), 2.08-1.92 (m, 2H), 1.79-1.72 (m, 1H), 1.68 (s, 3H), 1.61 (s, 3H), 1.59-1.48 (m, 2H), 1.43-1.17 (m, 3H), 0.95 (d, J = 6.54 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.3$ (C_q), 144.7, 134.7, 131.5, 130.4, 129.1, 128.2, 124.8, 118.5, 63.3, 37.2, 35.7, 29.7, 25.9, 25.6, 19.6, 17.8. The above data is in accordance with previous reports for compound P31.¹⁷



Figure S83: GC trace of P31.



Figure S84: ¹H NMR spectrum of P31 in CDCl₃.



Figure S85: ¹³C NMR spectrum of P31 in CDCl₃.

4-allyl-2-methoxyphenyl cinnamate P32:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 60% yield (35 mg).



¹**H** NMR (400 MHz, CDCl₃): $\delta = 7.88$ (d, J = 15.99 Hz, 1H), 7.58 (s, 2H), 7.42 (s, 3H), 7.03 (d, J = 7.90 Hz, 1H), 6.83-6.79 (m, 2H), 6.67 (d, J = 15.99 Hz, 1H), 6.04-5.94 (m, 1H), 5.13 (d, J = 12.95 Hz, 2H), 3.83 (s, 3H), 3.40 (d, J = 6.47 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 165.3$ (C_q), 151.2, 146.6, 139.1, 138.2, 137.2, 134.5, 130.7, 129.1, 128.4, 122.8, 120.9,

117.3, 116.3, 112.9, 56.1, 40.3. The above data is in accordance with previous reports for compound P32.¹⁸



Figure S86: ¹H NMR spectrum of P32 in CDCl₃.



Figure S87: ¹³C NMR spectrum of P32 in CDCl₃.

(3S,8S,9S,10R,13R,14S,17R)-10,13-dimethyl-17-((R)-6-methylheptan-2-yl)-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1H-cyclopenta[a]phenanthren-3-yl cinnamate P33:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 40% yield (41.3 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P33 = 29.35 min.



¹**H NMR** (**500 MHz, CDCl₃**): $\delta = 7.67$ (d, J = 16.00 Hz, 1H), 7.53-7.50 (m, 2H), 7.38-7.36 (m, 3H), 6.42 (d, J = 16.00 Hz, 1H) 5.41-5.39 (br s, 1H), 4.78-4.70 (m, 1H), 2.40 (d, J = 7.68 Hz, 2H), 2.03-1.08 (m, 26H), 1.04 (s, 3H), 0.92 (d, J = 6.53 Hz, 3H), 0.87-0.85 (d, J = 6.61 Hz, J = 1.77 Hz 6H), 0.68 (s, 3H). ¹³**C NMR** (**125 MHz, CDCl₃**): $\delta = 166.6$ (C_q), 144.6, 139.9, 134.7, 130.3, 129.0, 128.2, 122.9, 118.9, 74.3, 56.9, 56.3, 50.2, 42.5, 39.9, 39.7, 38.4, 37.2, 36.8, 36.4, 35.9, 32.1, 28.4, 28.2, 28.1, 24.0, 23.0, 22.7, 21.2, 19.5, 18.9, 12.0. The above data is in accordance with previous reports for compound P33.¹⁹



Figure S88: GC trace of P33.



Figure S89: ¹H NMR spectrum of P33 in CDCl₃.



Figure S90: ¹³C NMR spectrum of P33 in CDCl₃.

2'-hydroxy-[1,1'-binaphthalen]-2-yl cinnamate P34:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 12% yield (10 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxycarbonylated product P34 = 27.11 min.



¹**H NMR** (**500 MHz**, **CDCl**₃): $\delta = 8.06$ (d, J = 8.87 Hz, 1H), 7.97-7.92 (m, 2H), 7.86-7.83 (m, 2H), 7.81-7.78 (m, 2H), 7.50-7.44 (m, 1H), 7.35 (br s, 1H), 7.34-7.32 (m, 2H), 7.29-7.27 (m, 5H), 7.13-7.08 (m, 2H), 6.20 (d, J = 15.97 Hz, 1H), 5.33 (s, 1H). ¹³**C NMR** (**125 MHz**, **CDCl**₃): $\delta = 166.4, 152.9, 152.0, 148.3, 147.2, 134.0, 133.7, 133.6, 132.4, 131.5, 131.0, 130.9, 130.5, 129.6, 129.2, 129.0, 128.6, 128.4, 128.2, 127.6, 126.8, 126.4, 125.9, 124.8, 124.4, 124.2, 123.6, 123.4, 122.0, 118.5, 117.9, 116.3, 114.2, 111.1. The above data is in accordance with previous reports for compound P34.²⁰$



Figure S91: GC trace of P34.



Figure S92: ¹H NMR spectrum of P34 in CDCl₃.



Figure S93: ¹³C NMR spectrum of P34 in CDCl₃.

Isosorbide-2-O-cinnamate P35:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 38 % yield (20.7 mg).



¹**H** NMR (400 MHz, CDCl₃): $\delta = 7.71$ (d, J = 15.84 Hz, 1H), 7.52 (br s, 2H), 7.39 (s, 3H), 6.43 (d, J = 16.01 Hz, 1H), 5.36 (br s, 1H), 4.68 (s, 1H), 4.56 (s, 1H), 4.34 (s, 1H), 4.09 (m, 2H), 3.91 (t, J = 15.02 Hz, 1H), 3.59 (t, J = 14.99 Hz, 1H), 2.68 (S, 1H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 166.0$ (C_q), 146.2, 134.2, 130.8, 129.1, 128.3, 117.3, 85.9, 82.2, 78.6, 73.8, 73.7, 72.5. The above data is in accordance with previous reports for compound P35.²¹



Figure S94: ¹H NMR spectrum of P35 in CDCl₃.



Figure S95: ¹³C NMR spectrum of P35 in CDCl₃.

2-hydroxyethyl cinnamate P36A:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 72% yield (27 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxyated product P36A = 18.66 min.



¹H NMR (500 MHz, CDCl₃): $\delta = 7.69$ (d, J = 16.00 Hz, 1H), 7.47 (br s, 2H), 7.34 (s, 3H), 6.44 (d, J = 15.99 Hz, 1H), 4.32 (s, 2H), 3.87 (s, 2H), 2.84 (s, 1H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.4$, 145.5, 134.3, 130.5, 128.9, 128.2, 117.6, 66.2, 61.2. The above data is in accordance with previous reports for compound P36A.²²



Figure S96: GC trace of P36A and P36B.



Figure S97: ¹H NMR spectrum of P36A in CDCl₃.



Figure S98: ¹³C NMR spectrum of P36A in CDCl₃.

2-(((E)-2-oxo-4-phenylbut-3-en-1-yl)oxy)ethyl cinnamate P36B:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 10% yield (6.4 mg). GC retention time for Phenylacetylene = 2.2 min.; hydroalkoxyated product P36B = 27.93 min.



¹**H NMR (200 MHz, CDCl₃):** δ = 7.73 (d, *J* = 15.94 Hz, 2H), 7.55 (s, 4H), 7.40 (s, 6H), 6.48 (d, *J* = 16.03 Hz, 2H), 4.49 (s, 4H). The above data is in accordance with previous reports for compound P36B.²³



Figure S99: ¹H NMR spectrum of P36B in CDCl₃.

Methyl (E)-3-(4-methoxyphenyl)acrylate P37:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (97:3). After column, a pale yellow liquid was isolated in 85% yield (32.6 mg).



¹H NMR (400 MHz, CDCl₃): $\delta = 7.64$ (d, J = 15.97 Hz, 1H), 7.46 (d, J = 8.68 Hz, 2H), 6.89 (d, J = 8.68 Hz, 2H), 6.30 (d, J = 15.96 Hz, 1H), 3.82 (s, 3H), 3.78 (s, 3H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 167.9$, 161.5, 144.7, 129.9, 127.2, 115.3, 114.4, 55.5, 51.7. The above data is in accordance with previous reports for compound P37.⁷



Figure S100: ¹H NMR spectrum of P37 in CDCl₃.



Figure S101: ¹³C NMR spectrum of P37 in CDCl₃.

Ethyl (E)-3-(4-methoxyphenyl)acrylate P38:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 92% yield (37.9 mg).



¹**H** NMR (500 MHz, CDCl₃): $\delta = 7.58$ (d, J = 15.95 Hz, 1H), 7.39 (d, J = 8.73 Hz, 2H), 6.83 (d, J = 8.74 Hz, 2H), 6.25 (d, J = 15.96 Hz, 1H), 4.19 (q, J = 7.13 Hz, 2H), 3.74 (s, 3H), 1.27 (t, J = 7.13 Hz, 3H). ¹³C NMR (125 MHz, CDCl₃): $\delta = 167.2$ (C_q), 161.3, 144.2, 129.6, 127.1, 115.7, 114.3, 60.2, 55.2, 14.3 (CH₃). The above data is in accordance with previous reports for compound P38.⁷



Figure S102: ¹H NMR spectrum of P38 in CDCl₃.



Figure S103: ¹³C NMR spectrum of P38 in CDCl₃.

Methyl (E)-3-(4-chlorophenyl)acrylate P39A and Methyl 3-(4-chlorophenyl)propanoate P39B:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (96:4). After column, a pale yellow liquid was isolated in 67% yield (26.3 mg). GC retention time for 4 chlorophenylacetylene = 4.21 min.; hydroalkoxycarbonylated product P39 = 14.49 min. The NMR investigations suggest that both P39A and P39B exist in solution and the ratio between them is about 1:1. Iron catalyzed direct and transfer hydrogenation of different alkene has been reported. Accordingly, the 4-chloro phenyl acetylene after hydroalkoxycarbonylation, might further undergo hydrogenation (of P39A) to form P39B.



¹**H NMR** (400 MHz, CDCl₃): $\delta = 7.63$ (d, J = 16.03 Hz, 1H), 7.46-7.34 (m, 4H), 7.24 (d, J = 8.54 Hz, 2H), 7.12 (d, J = 8.33 Hz, 2H), 6.40 (d, J = 16.02 Hz, 1H), 3.80 (s, 3H), 3.65 (s, 3H), 2.91 (t, J = 15.37 Hz, 2H), 2.60 (t, J = 15.38 Hz, 2H). ¹³C NMR (100 MHz, CDCl₃): $\delta = 173.2$

 (C_q) , 167.3 (C_q) , 143.6, 139.1, 136.4, 133.0, 132.2, 129.8, 129.4, 129.3, 128.8, 118.6, 51.9 (CH₃), 51.8 (CH₃), 35.7 (CH₂), 30.4 (CH₂). The above data is in accordance with previous report for compound P39.¹⁰



Figure S104: GC trace of P39.



Figure S105: ¹H NMR spectrum of P39 in CDCl₃.



Figure S106: ¹³C NMR spectrum of P39 in CDCl₃.

2-ethylhexyl (E)-3-(4-methoxyphenyl)acrylate P40:

Purified by silica gel chromatography using petroleum ether and ethyl acetate (98:2). After column, a pale yellow liquid was isolated in 88% yield (51 mg). GC retention time for 4-methoxyphenylacetylene = 6.1 min.; hydroalkoxycarbonylated product P40 = 24.8 min.



¹**H NMR** (**500 MHz**, **CDCl**₃): $\delta = 7.62$ (d, J = Hz, 1H), 7.47 (d, J = Hz, 2H), 6.88 (d, J = Hz, 2H), 6.31 (d, J = Hz, 1H), 4.14-4.06 (m, 2H), 3.82 (s, 3H), 1.66-1.59 (m, 1H), 1.42-1.24 (m, 8H), 0.93-0.87 (m, 6H). ¹³**C NMR** (**125 MHz**, **CDCl**₃): $\delta = 167.7$, 161.4, 144.3, 129.8, 127.3, 115.9, 114.4, 66.9, 55.5, 38.9, 30.6, 29.1, 23.9, 23.1, 14.2, 11.2. The above data is in accordance with previous reports for compound P40.²⁴



Retention Time	Area	Area Area %		Height %
6.108	2190191	10.70	637304	10.33
24.817 18270495		89.30	5534230	89.67
Totals				
	20460686	100.00	6171534	100.00

Figure S107: GC trace of P40.



Figure S108: ¹H NMR spectrum of P40 in CDCl_{3.}



Figure S109: ¹³C NMR spectrum of P40 in CDCl_{3.}

5. Sytnthetic application of product:



Figure S110: Practical utility of esters.

6. Further functionalization:

Product [A] and [B] were prepared by following previous literature procedure.²⁵



Scheme 1: Deprotection of Methyl-4-methoxy cinnamate and polymerization of 4-HCA



Figure S111: ¹H NMR spectrum of [A] in DMSO.



Figure S112: Molecular weight of polymer [B] by GPC in DMF

7. Deuterium labelling experiment:

Mw/Mn



In a glove box, an oven-dried vial (4 mL) containing a stirring bar was charged with $[Fe_2(CO)_9]$ (0.005 mmol, 1.8 mg), L7 (0.01 mmol, 5 mg), DABCO (0.89 mmol, 100 mg), phenyl acetylene (0.2 mmol, 0.02 mL), and CD₃OD (1 mmol, 36.06 mg). The vial was then sealed with a PTFE/white rubber septum and was taken out from the glove box. Then, PhMe (0.3 mL), and THF (0.3 mL) were added by syringe under argon. The vial was placed in a beaker, which was

1.003

transferred into a stainless steel autoclave (450 mL). After flushing the autoclave three times with CO, a pressure of 10 bar of CO was adjusted at ambient temperature. Then, the mixture was stirred for 36 h at 110 °C. After the reaction was complete, the autoclave was cooled down to room temperature and the pressure was released carefully. The conversion was determined by the GC and was purified by column chromatography on silica gel (P.E./EA) to obtain the desired product.



Figure S113: ²H-NMR spectrum of deuterated methyl cinnamate in CHCl₃.



Figure S114: ¹³C-NMR spectrum of deuterated methyl cinnamate in CDCl₃.



In a glove box, an oven-dried conical flask (100 mL) containing a stirring bar was charged with $[Fe_2(CO)_9]$ (0.489 mmol, 88.9 mg), L7 (0.489 mmol, 244.5 mg), DABCO (48.9 mmol, 5.48 g), Phenyl acetylene (9.79 mmol, 1 g), and IPA (97.9 mmol, 7.48 mL). Then, Toluene (14 mL), and THF (14 mL) were added. Next, the content was transferred into a stainless steel autoclave (450 mL). After flushing the autoclave three times with CO, a pressure of 10 bar of CO was adjusted at ambient temperature. Then, the mixture was stirred for 36 h at 110 °C. After the reaction was complete, the autoclave was cooled down to room temperature and the pressure was released carefully. The conversion was determined by the GC and product was purified by column chromatography on silica gel (P.E./EA).



Figure S115: GC chromatogram of isopropyl cinnamate

9. Mechanistic studies:

9a. Kinetic analysis:

In a glove box, an oven-dried vial (4 mL) containing a stirring bar was charged with $[Fe_2(CO)_9]$ (0.005 mmol, 1.8 mg), L7 (0.01mmol, 5 mg), DABCO (0.89 mmol, 100 mg), aryl alkyne (0.2 mmol), and alcohol (4 mmol). The vial was then sealed with a PTFE/white rubber septum and removed from the glovebox. Then, Toluene (0.38 mL), and THF (0.3 mL) were added by syringe under argon. The vial was placed in a beaker, which was transferred into a stainless steel autoclave (450 mL). After flushing the autoclave three times with CO, a pressure of 10 bar of CO was adjusted at ambient temperature. Then, the mixture was stirred for 0.5/1/1.5/2/2.5 hrs at 110 °C. After that, the autoclave was cooled down to room temperature and the pressure was released carefully. The product yield was determined in the presence of internal standard (dodecane 45.4 µL, 0.2 mmol) by GC.

Sr. No.	Time (min.)	Conc.of Product P1 (Mmin ⁻¹)
1.	10	0.0011
2.	20	0.0047
3.	30	0.0058
4.	60	0.03
5.	90	0.046
6.	120	0.072
7.	150	0.089



Figure S116A: Time-dependent formation of α - aryl aldehyde P1 using iron catalyst.

Sr. No.	Time (min.)	Conc.of Product P1 [M]		
		In presence of ligand	In absence of ligand	
1.	30	0.0058	0.0018	
2.	60	0.03	0.013	
3.	90	0.046	0.022	
4.	120	0.072	0.031	
5.	150	0.089	0.048	



Figure S116B: Initial rate comparison for the formation of α - aryl aldehyde P1 in presence and absence of ligand.





Figure S116C: GC chromatogram for hydroalkoxycarbonylation of phenyl acetylene in absence of ligand



Figure S116D: GC chromatogram for hydroalkoxycarbonylation of phenyl acetylene in presence of ligand

In the presence of ligand we obtained 41% of product P1, while, in the absence of ligand, we observed 34% of product P1 with respect to internal standard and the corresponding GC yield is 71% and 51% respectively.

		Conc. of P1 [M]			
	Catalyst mol %	1	2.5	5	7.5
(0.5	0.00557	0.00657	0.0058	0.0184
(µ)	1	0.00836	0.0225	0.03	0.0472
me	1.5	0.0129	0.03012	0.046	0.088
Ï	2	0.02896	0.0482	0.072	0.115



Figure S117A: Time-dependent formation of P1 at different concentration of Iron catalyst.

log [cat.]	log rate
-3	-1.95
-2.6	-1.65
-2.3	-1.48
-2.12	-1.25



Figure S117B: Plot of log(rate) versus log(conc Fe₂(CO)₉.

	Conc. of P1 [M]				
	Substrate				
	Conc.				
	[S1]	0.2M	0.4M	0.6M	0.8M
	0.5	0.0058	0.0078	0.011	0.015
0	1	0.03	0.03	0.043	0.049
ime	1.5	0.046	0.064	0.0541	0.07
Ĥ	2	0.072	0.078	0.067	0.075


Figure S118A: Time-dependent formation of **P1** at different Substrate (S1 = Phenyl acetylene) concentration.

Conc. of S1	Initial rate	
0.2	0.033	
0.4	0.035	
0.6	0.038	
0.8	0.04	



Figure S118B: Plot of initial rate versus substrate concentration.

9b. Control experiment:

Presence of Iron hydride:

An oven dried Schlenk tube was charged with $[Fe_2(CO)_9]$ (0.1 mmol, 36.3 mg, DABCO (0.1 mmol, 11.12 mg), MeOH (0.6 mmol, 24µL), further THF (0.5 mL) was added into it and the content was stirred for 1.5h/10h at room temperature. The presence of iron hydride has been determined by ¹H NMR spectroscopy.

$$[Fe_2(CO)_9] + DABCO + MeOH \longrightarrow [Fe-H]$$



Figure S119: ¹H NMR of reaction mixture



Figure S120: Proposed catalytic cycle

9c. Radical trap experiment:

To confirm the involvement of radical species in the reaction, the reaction was performed in the presence diphenylethylene which hardly affected the reaction and gives product with slight less yield (89%).²⁶



Scheme S2: Radical trap experiment for hydroalkoxycarbonylation reaction

In a glove box, an oven-dried vial (4 mL) containing a stirring bar was charged with Fe₂(CO)₉ (0.005 mmol, 1.8 mg), L7 (0.01 mmol, 5 mg), DABCO (0.89 mmol, 100 mg), phenyl acetylene (0.2 mmol, 0.02 mL) and DPE (0.2 mmol, 36.05 mg). The vial was then sealed and removed from the glovebox. Then IPA (4 mmol, 0.3 mL), Toluene (0.3 mL), and THF (0.3 mL) were added by syringe under argon. The vial was placed in a beaker, which was transferred into an autoclave. After flushing the autoclave three times with CO, a pressure of 10 bar of CO was adjusted at ambient temperature. Then, the mixture was stirred for 36 h at 110 °C. After the reaction was complete, the autoclave was cooled down to room temperature and the pressure was released carefully. The reaction mixture was then analyzed by GC and the result is shown below.

Gas Chromatography (GC) analysis for P1 was performed on an Agilent 7890B GC system using HP-05 column (30 m × 320 μ m × 0.25 μ m), split ratio 30:1, column pressure 5 psi, injector temperature of 260 °C, detector temperature of 300 °C, argon carrier gas. Temperature program: Initial temperature 70 °C, hold for 1 min.; ramp 1: 4 °C/min. to 120 °C; ramp 2: 6 °C/min. to 160 °C; ramp 3: 10 °C/min. to 250 °C ramp 4: 20 °C/min. to 320 °C, hold for 2 min. GC retention time for Phenyl acetylene = 3.9 min.; ester product P1 = 19.8 min.



Figure S121: GC data of P1 in the presence of DPE.



Figure S122 : Effect of DPE concentration in hydroalkoxycarbonylation of alkyne

DPE equiv	Product conversion (%)	
0.25	87	
0.5	89	
0.75	92	
1.00	89	

Table S2: Effect of DPE loading on hydroakoxycarbonylation of alkyne

9d. Procedure for EPR analysis:



Figure S123: EPR spectra of the control experiments.

Reaction vials (4 mL) containing stirring bars were charged with $[Fe_2(CO)_9]$ (0.05 mmol, 18.2 mg), L7 (0.05 mmol, 25 mg), DABCO (0.2 mmol, 22.4 mg), isopropyl alcohol (0.4 mmol, 30 μ L), alkyne (0.1 mmol, 10 μ L) then toluene:THF (0.3:0.1 mL) was added in the glove box. The vials were then sealed with a PTFE caps and taken out from the glove box. Then, mixtures were heated at 110 °C under 10 bar CO pressure in autoclave for 45 min. then content was cooled down and then reaction mixture was transferred to EPR tubes in glove box and was frozen at 77 K, which was then subjected to the EPR measurement.

As seen in figure S123, the blue line (Fe+base) and red line (Fe+base+ROH) does not show any EPR signal. The green line (Fe+base+alkyne+ROH) disclosed a broad hump. However, there is no clear signal to conclude the existence of radical species. A similar observation is made by Wu and co-workers and it was concluded that such hump can not be assigned as an EPR signal. In line with this report, figure S123 does not show any clear EPR signal and the possibility of existence of a radical species is very dim.

10. Investigation of possible metal contaminations:



Entry	Catalyst [M]	Conversion (%) ^b
1.	Co ₂ (CO) ₈	0.88
2.	PdCl ₂	0.55
3.	CuBr	0.3
4.	CuCl	0
5.	Ni(COD) ₂	0.6
6.	Pd ₂ (DBA) ₃	0.8
7.	Ru ₃ (CO) ₁₂	0.8

Cu(OAc)₂

Table S3: Testing of other metals.

8.

Conditions: a) [M] = 0.0006 mmol (0.3 mol%), L = 0.0006 mmol, DABCO = 4.5 equiv, Phenyl acetylene = 0.02 mL (0.2 mmol); b) conversion determined by GC.

0.6

11. References:

- 1. M. Niehues, G. Erker, G. Kehr, P. Schwab, R. Froehlich, O. Blacque, H. Berke, *Organometallics* 2002, **21**, 2905–2911.
- 2. J. M. Kliegman, R. K. Barnes, Tetrahedron 1970, 26, 2555-2560
- 3. A. Paulovicova, U. El-Ayaan, K. Shibayama, T. Morita, Y. Fukuda, *Eur. J. Inorg. Chem.* 2001, **10**, 2641–2646.
- 4. Y.-Y. Zhu, C. Cui, N. Li, B.-W. Wang, Z.-M. Wang, S. Gao, *Eur. J. Inorg. Chem.* 2013, 3101–3111.
- 5. A.B. Lutjen, M. A. Quirk, A. M. Barbera, E. M. Kolonko, *Bioorganic and Medicinal Chemistry* 2018, **26**, 5291–5298.
- 6. L. Wang, Y. Wang, C. Liu, A. Lei, Angew. Chem. Int. Ed. 2014, 53, 5657–5661.
- 7. H. Wang, Q.-Yi Wei, H. Jiang, Z.-H. Jiang, *Res Chem Interme* 2012, **38**, 207–213.
- 8. Z. Zhang, J. Liu, F. Wu and L. Zhao, *Letters in Drug Design & Discovery*, 2013, **10**, 529-534.
- H.-F. Lu, C. Xie, J. Chang, G.-Q. Lin, X. Sun, European Journal of Medicinal Chemistry, 2011, 46(5), 1743-1748.
- R. H. N. Silva, A. C. M. Andrade, Diego F. Nóbrega, Ricardo D. de Castro, Hilzeth L.
 F. Pessôa, Nidhi Rani, Damião P. de Sousa, *BioMed Research International*, 2019, 2019, Article ID 3941242.
- R. S. Galaverna, L. P. Fernandes, V. H. Menezes da Silva, A. de Siervo, and J. C. Pastre, *Eur. J. Org. Chem.* 2022, e202200376.
- 12. J. Li, L. Liu, Y.-Y. Zhou and S.-N. Xu, RSC Adv., 2012, 2, 3207–3209
- A) Y. Okuno, S. Isomura, A. Sugamata, K. Tamahori, A. Fukuhara, M.Kashiwagi, Y. Kitagawa, E. Kasai, K. Takeda, *ChemSusChem* 2015, 8, 3587–3589. B) B. Zhang, P. Feng, Y. Cui and N. Jiao, *Chem. Commun.*, 2012, 48, 7280.
- 14. N. Devarajan and P. Suresh, Org. Chem. Front., 2018, 5, 2322.
- V. Palermo, Diego M. Ruiz, Juan C. Autino, Patricia G. Vázquez, and Gustavo P. Romanelli, *Pure and Applied Chemistry*, 2011, 84 (3), 529-540.

- J.A.D. Vale, M.P. Rodrigues, A.M.A. Lima, S.S. Santiago, G.D.A. Lima, A. A. Almeida, L.L. Oliveira, G.C. Bressan, R.R. Teixeira, M. Machado-Neves, *Biomedicine & Pharmacotherapy* 2022, 148, 112689
- Mei-Ling Wang, H. Xu, H.-Y. Li, B. Ma, Z.-Yu Wang, X. Wang, and H.-X. Dai *Org. Lett.* 2021, 23(6), 2147–2152
- S. Tawata, S. Taira, N. Kobamoto, J. Zhu, M. Ishihara, S. Toyama, *Bioscience, Biotechnology, and Biochemistry*. 1996, 60(5), 909-910.
- 19. X. Liu and T. Werner, *Adv. Synth. Catal.*, 2021, **363**, 1096–1104.
- 20. S. Lu, Si Bei Poh, and Yu Zhao, Angew. Chem. Int. Ed. 2014, 53, 11041 –11045
- D. Ragno, C. Leonardi, G.Di Carmine, O. Bortolini, A. Brandolese, C. De Risi, and A. Massi, ACS Sustainable Chemistry & Engineering 2021, 9 (24), 8295-8305
- 22. H. Aman, Y. H. Wang, G. J. Chuang, ACS Omega 2020, 5, 918–925.
- 23. Y. Jiang, Q. Ma, X. Zhang, J. Li, S. Liao, Polym. Chem. 2022, 13, 2538.
- T. Monhaphol, B. Albinsson and S. P. Wanichwecharungruang, J. Pharm.
 Pharmacol, 2007, T.H. Thi, M. Matsusaki, D. Shi, T. Kaneko, M. Akashi, J. Biomater.
 Sci. Polym. Ed. 2008, 19(1), 75-85. doi: 10.1163/156856208783227668. PMID: 18177555.59, 279–288.
- T.H. Thi, M. Matsusaki, D. Shi, T. Kaneko, M. Akashi, J. Biomater. Sci. Polym. Ed. 2008, 19(1), 75-85. doi: 10.1163/156856208783227668. PMID: 18177555.
- 26. Han-Jun Ai, B. N. Leidecker, P. Dam, C. Kubis, J. Rabeah, X.-Feng Wu, *Angew. Chem. Int. Ed.* 2022, **61**(43), e202211939.