

Supporting Information

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Experimental Section

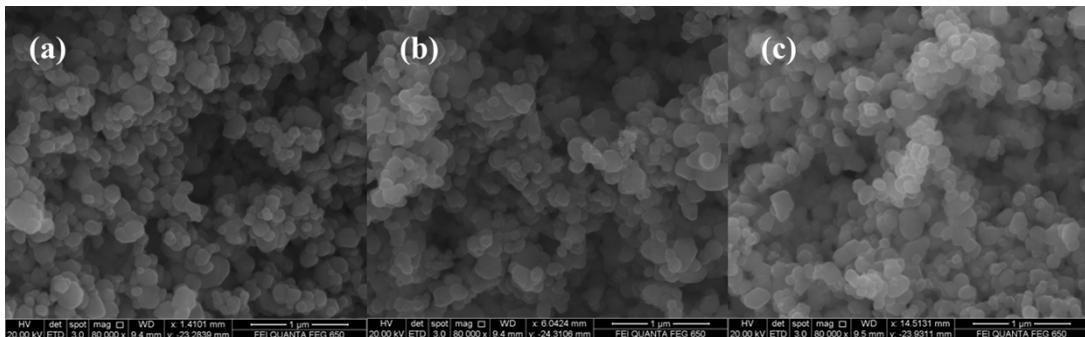


Figure S1. SEM image of (a) TiO₂ (b) Pd/TiO₂ and (c) Pt/TiO₂.

Summary Report

Surface Area

Single point surface area at P/P₀ = 0.254951315: 11.0895 m²/g

BET Surface Area: 12.2337 m²/g

Langmuir Surface Area: 51.2332 m²/g

t-Plot Micropore Area: 1.0105 m²/g

t-Plot external surface area: 11.2232 m²/g

BJH Adsorption cumulative surface area of pores between 1.7000 nm and 300.0000 nm diameter: 12.8044 m²/g

BJH Desorption cumulative surface area of pores between 1.7000 nm and 300.0000 nm diameter: 12.1893 m²/g

Pore Volume

t-Plot micropore volume: 0.000138 cm³/g

BJH Adsorption cumulative volume of pores between 1.7000 nm and 300.0000 nm diameter: 0.047246 cm³/g

BJH Desorption cumulative volume of pores between 1.7000 nm and 300.0000 nm diameter: 0.046204 cm³/g

Pore Size

Adsorption average pore diameter (4V/A by BET): 15.2674 nm

Desorption average pore diameter (4V/A by BET): 5.8623 nm

BJH Adsorption average pore diameter (4V/A): 14.7593 nm

BJH Desorption average pore diameter (4V/A): 15.1620 nm

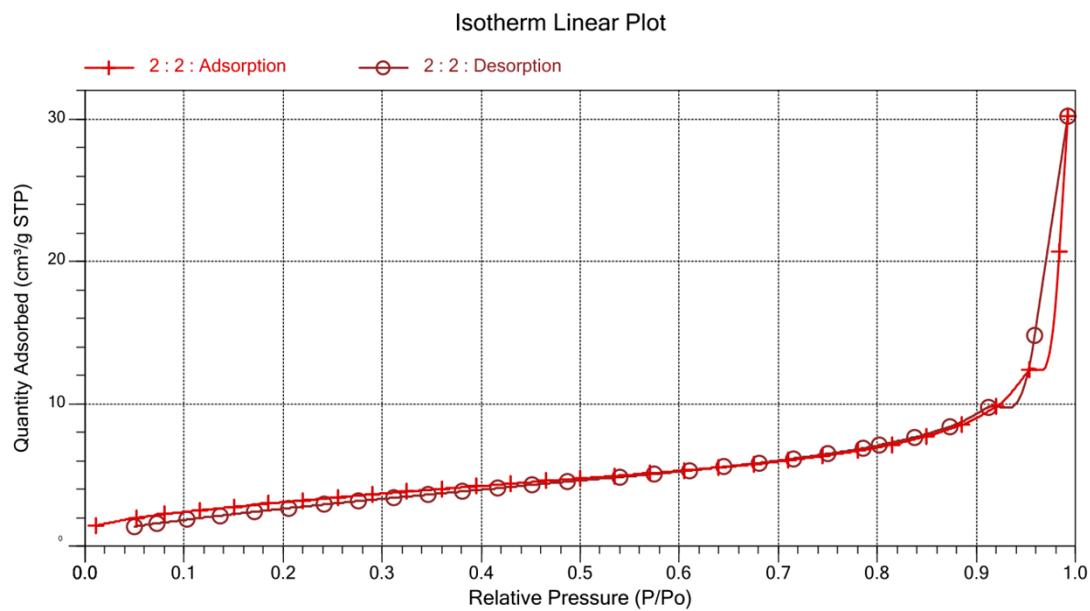


Figure S2. BET surface area and Nitrogen sorption isotherms at 77 K for $\text{Pd}_1\text{Pt}_1/\text{TiO}_2$

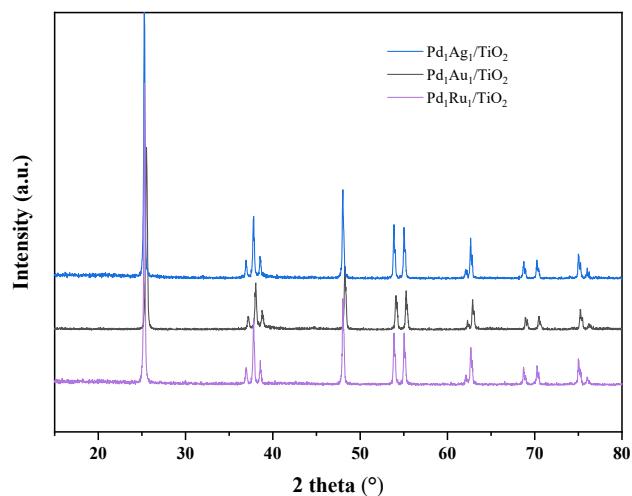


Figure S3. XRD diffraction patterns of other bimetallic photocatalysts.

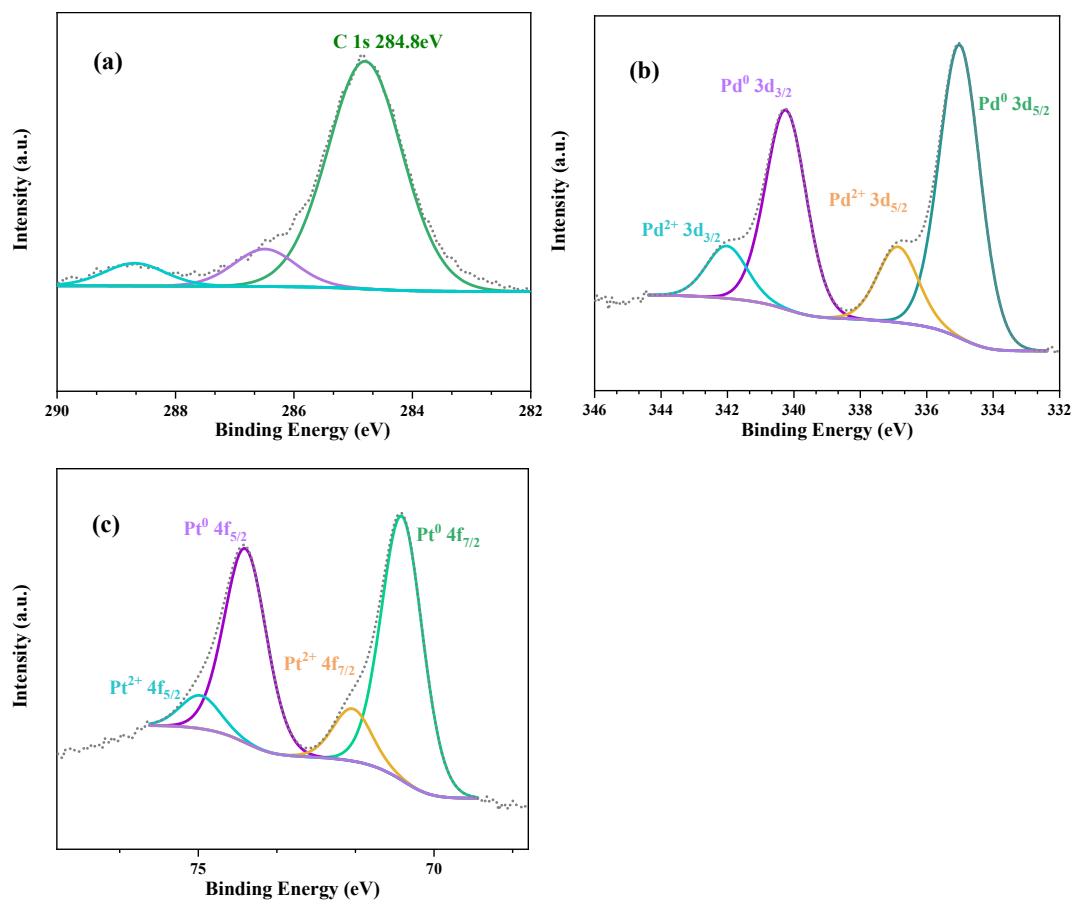
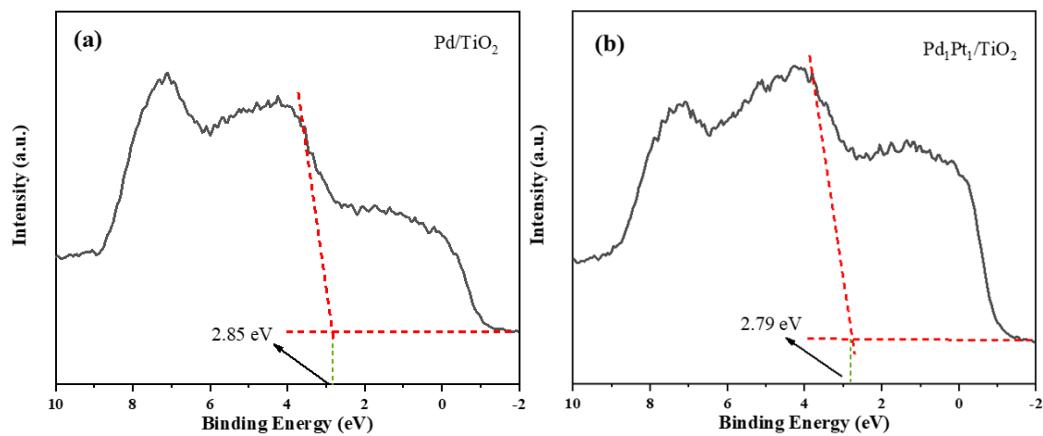


Figure S4. (a) core level spectra of C1s of Pd₁Pt₁/TiO₂. (b) XPS core level spectra of Pd 3d_{5/2} and 3d_{3/2} doublet region of Pd/TiO₂. (c) Pt 4f_{7/2} and 4f_{5/2} doublet region of Pt/TiO₂.



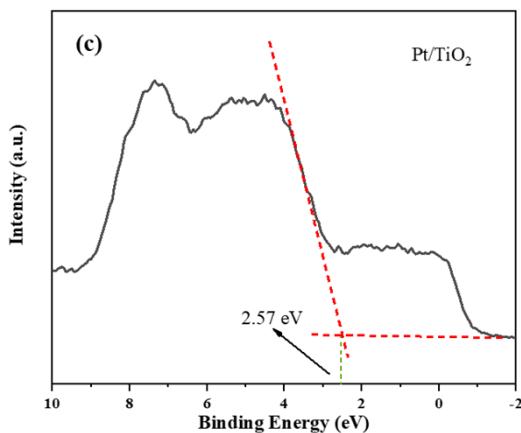


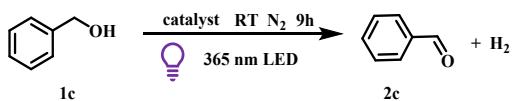
Figure S5. VB-XPS plots of (a) Pd/TiO₂ (b) Pd₁Pt₁/TiO₂ (c) Pt/TiO₂.

the EVB of the corresponding standard hydrogen electrode ($E_{VB, NHE}$) can be calculated according to the following formula: $E_{VB, NHE} = \varphi + E_{VB, XPS} - 4.44$, where φ is the work function of the instrument (4.5 eV).¹

Table S1. valence band edge potential (E_{VB}) and conductor band edge potential E_{CB} of various catalysts.

catalyst	E _{VB} (eV)	E _{CB} (eV)	Band gap (eV)
TiO ₂	2.932	-0.312	3.244
Pd/TiO ₂	2.906	-0.286	3.191
Pd ₂ Pt ₁ /TiO ₂	2.905	-0.285	3.189
Pd ₁ Pt ₁ /TiO ₂	2.891	-0.271	3.161
Pd ₁ Pt ₂ /TiO ₂	2.891	-0.271	3.161
Pt/TiO ₂	2.892	-0.271	3.163

Table S2. Benzaldehyde formed during photoreaction of benzyl alcohol with respective catalysts in the absence of amine.

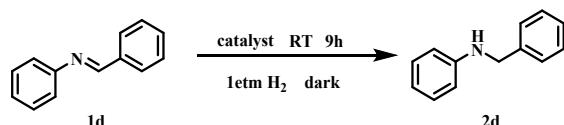


Entry	Catalyst	2c Yields. (%)
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1	Pd/TiO ₂	28
2	Pd ₂ Pt ₁ /TiO ₂	43
3	Pd ₁ Pt ₁ /TiO ₂	56
4	Pd ₁ Pt ₂ /TiO ₂	65
5	Pt/TiO ₂	71

Reaction conditions: benzyl alcohol (2.1 mmol), MeCN (5 ml), catalyst (10 mg), N₂, r.t., light irradiation (365 nm LED, 18 mW/cm²), 9 h. The yields were obtained by HPLC.

Table S3. The yields of secondary amine 2d obtained during hydrogenation of imine 1d in methanol with respective catalysts.



Entry	Catalyst	2d Yields. (%)
1	Pd/TiO ₂	75
2	Pd ₂ Pt ₁ /TiO ₂	86
3	Pd ₁ Pt ₁ /TiO ₂	60
4	Pd ₁ Pt ₂ /TiO ₂	43
5	Pt/TiO ₂	10

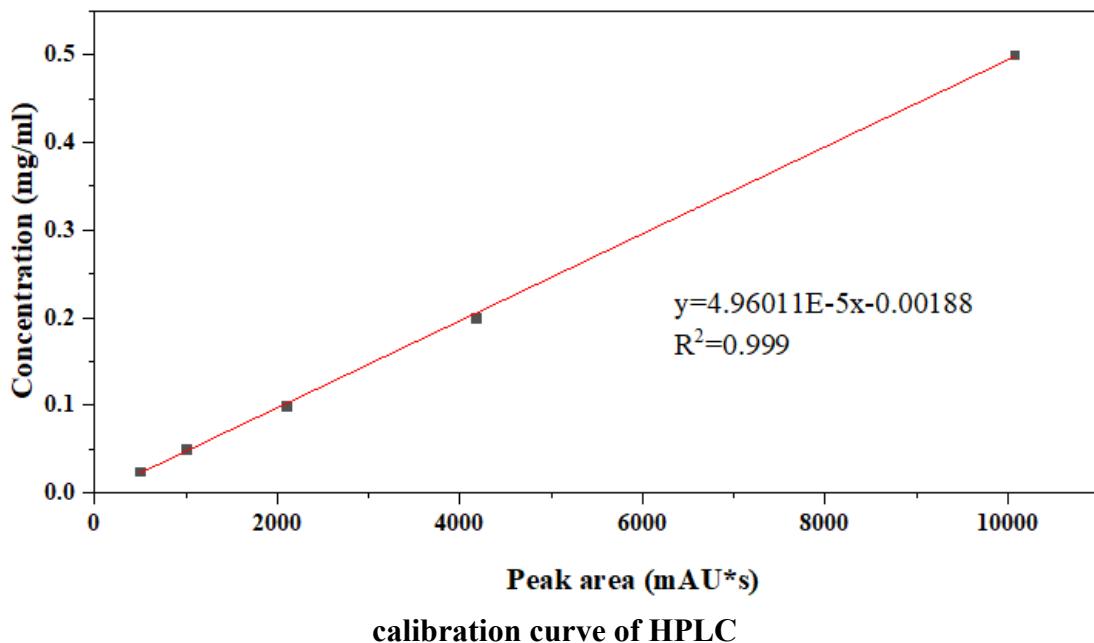
Reaction conditions: N-Benzylideneaniline (2.1 mmol), methanol (5 ml), catalyst (10 mg), 1 atm H₂, r.t., 9 h. The yields were obtained by HPLC.

The details of product analysis and drawing of calibration curve

For aniline products, HPLC was used with calibration (C18 column, 30 °C, λ=254 nm, MeOH mobile phase, 1 mL min⁻¹), and the calibration curve was drawn by measuring the peak area of 0.025, 0.05, 0.1, 0.2, 0.5 mg/ml of standard sample. The calibration curve is as follows, and the HPLC yields were calculated as following formula:

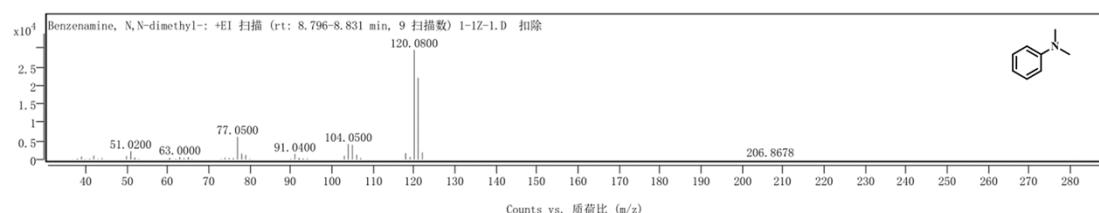
$$HPLC \text{ yield} = \frac{c \times 75 \times 5}{1.4 \times M}$$

Where the “c” represents for concentration obtained from calibration curve, “75” represent for dilution factor; “5” represent for volume of reaction solution; “M” represents for molecular weight of the product. Other products were analyzed by GC-FID using area normalization method.

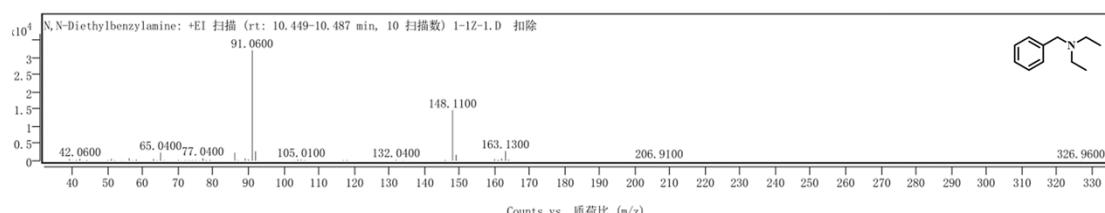


MS spectra of all products

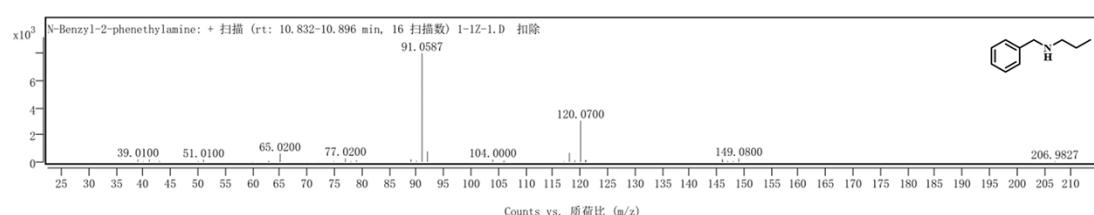
N,N-Dimethylaniline



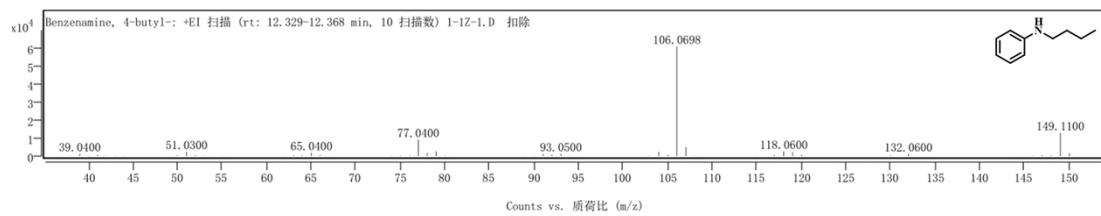
N-Benzyldiethylamine



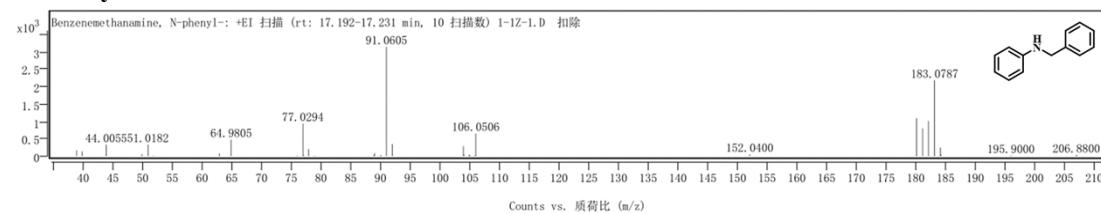
N-Benzylpropanamine



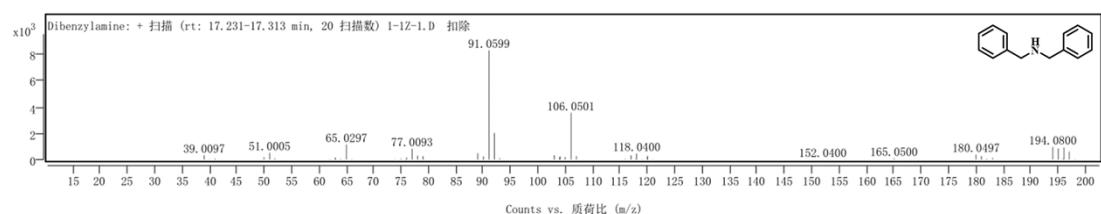
N-Butylaniline



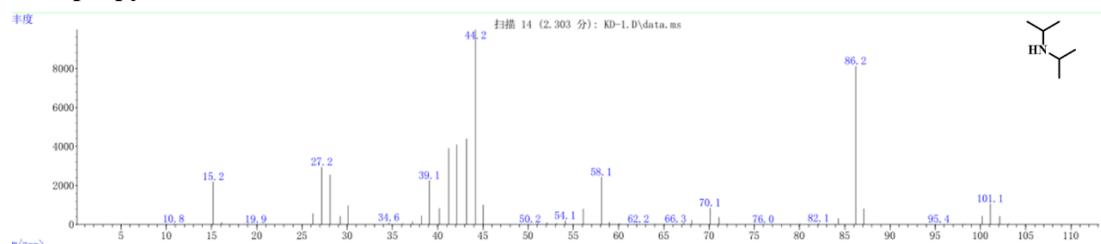
N-Benzylaniline



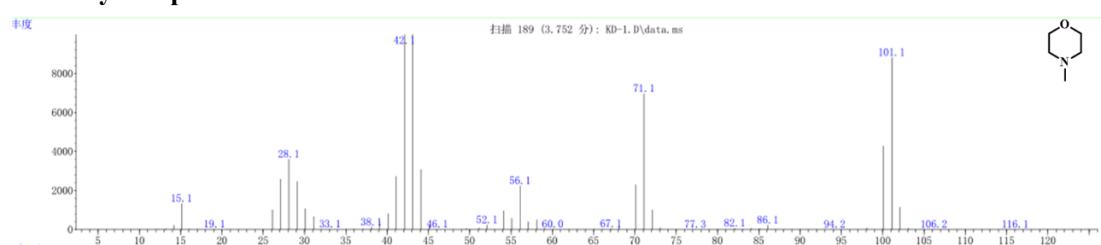
Dibenzylamine



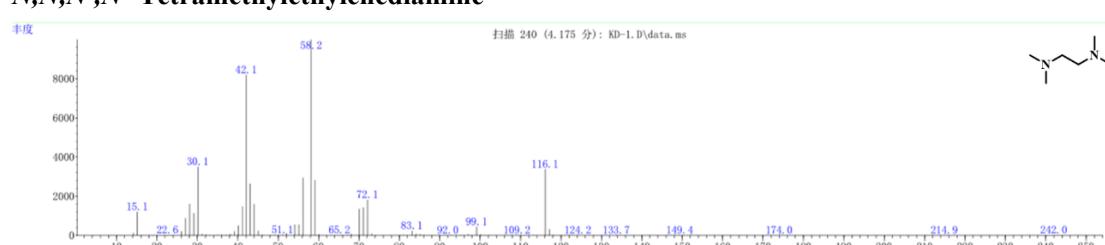
Diisopropylamine



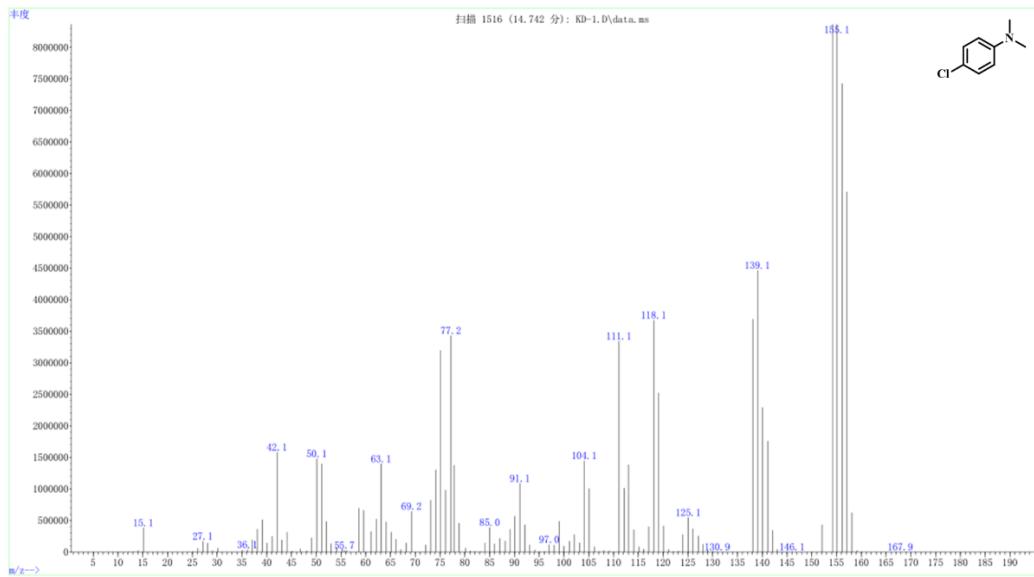
4-Methylmorpholine



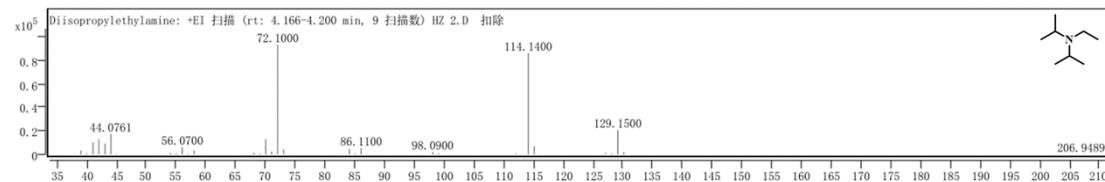
N,N,N',N'-Tetramethylethylenediamine



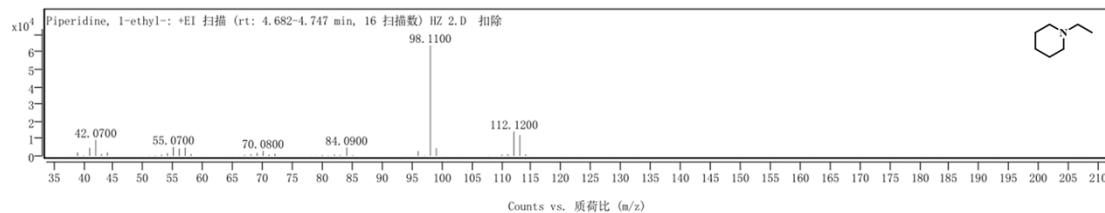
4-chloro-N,N-dimethylaniline



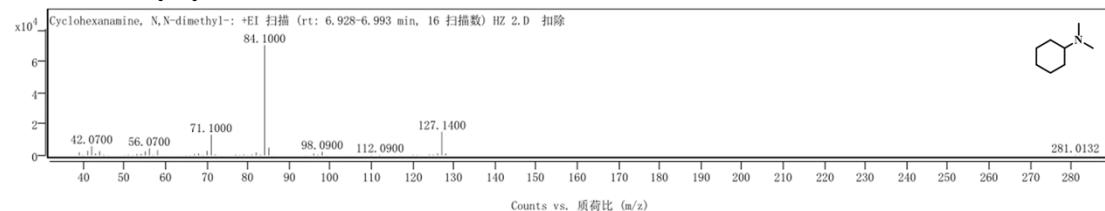
N,N-Diisopropylethylamine



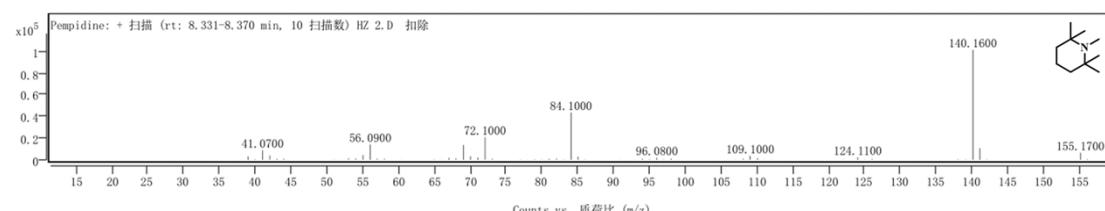
1-Ethylpiperidine



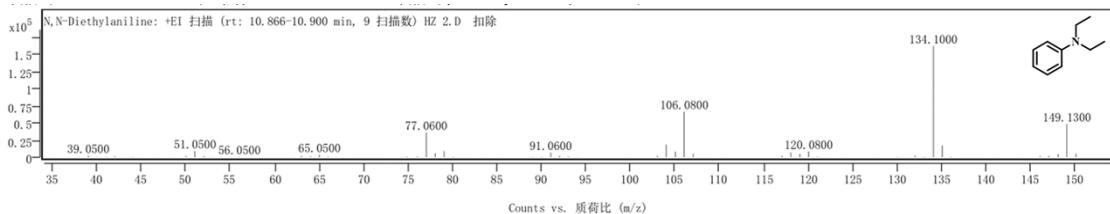
N,N-Dimethylcyclohexanamine



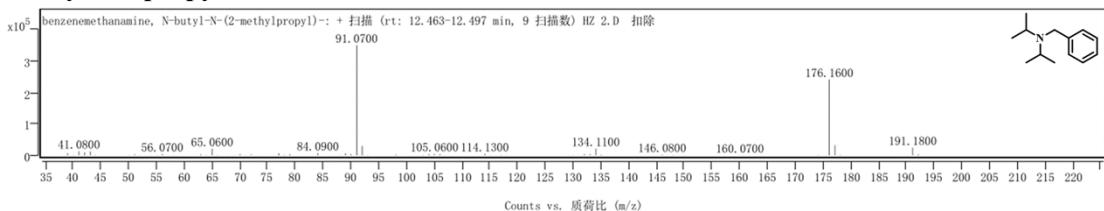
1,2,2,6,6-Pentamethylenepiperidine



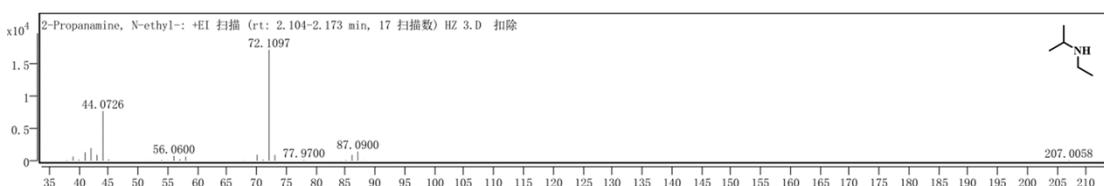
N,N-Diethylaniline



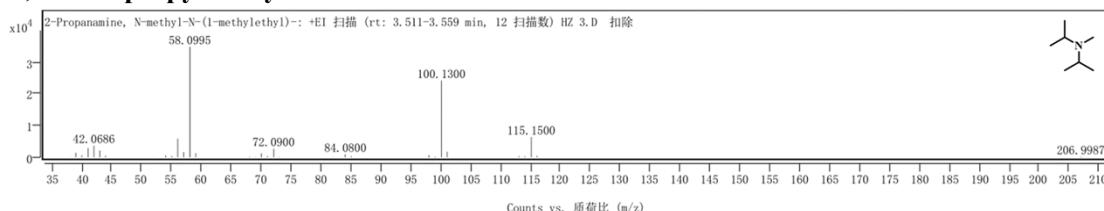
Benzylidisopropylamine



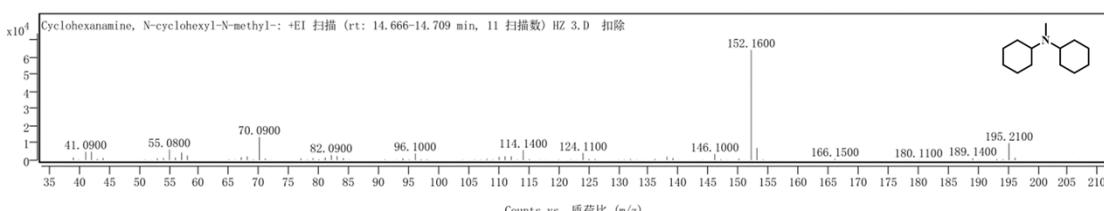
N-isopropylethylamine



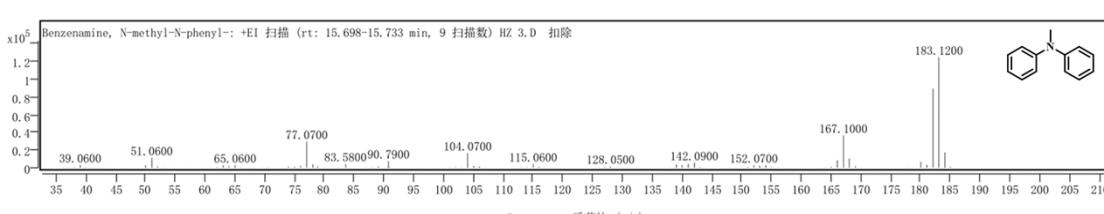
N,N-Diisopropylmethylamine



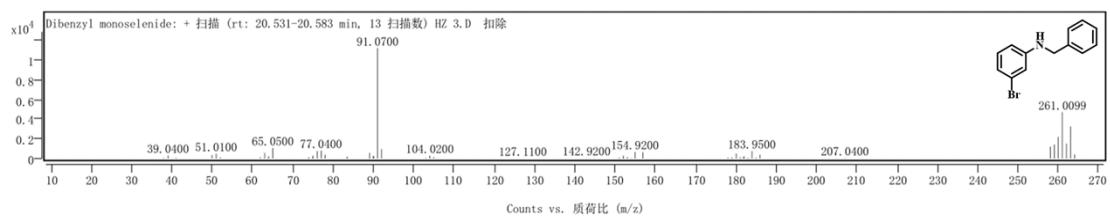
N,N-Dimethylcyclohexanamine



N-Methyldiphenylamine



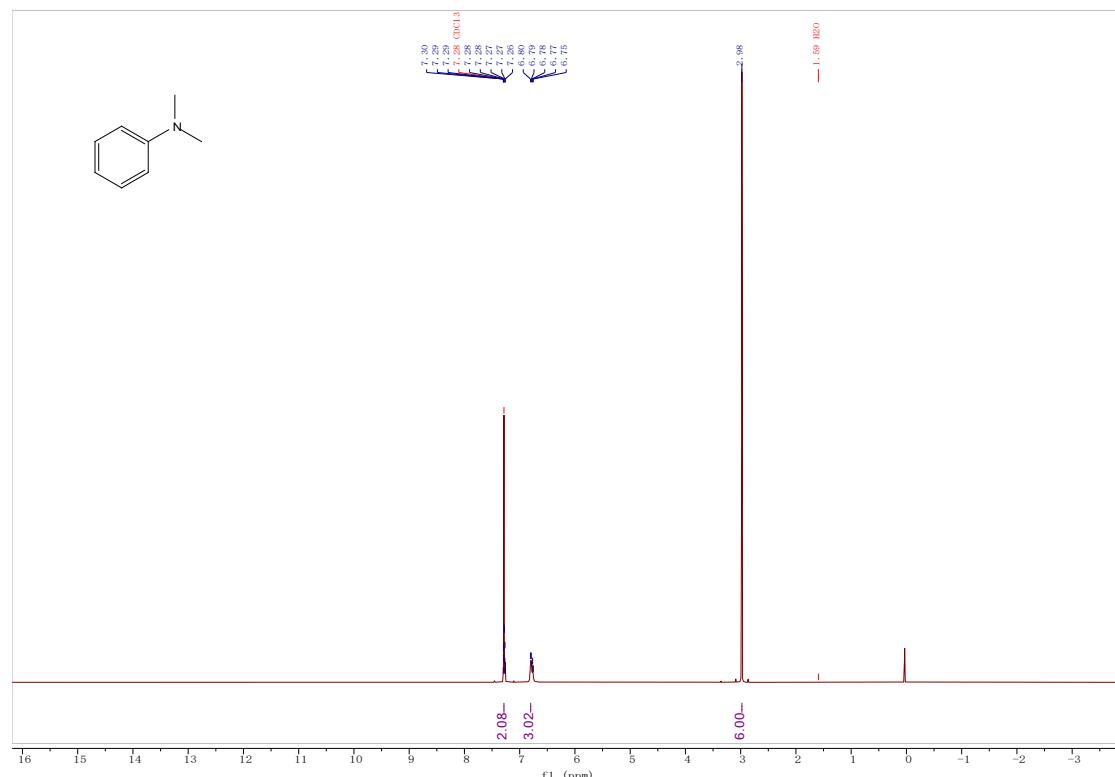
N-benzyl-3-bromoaniline



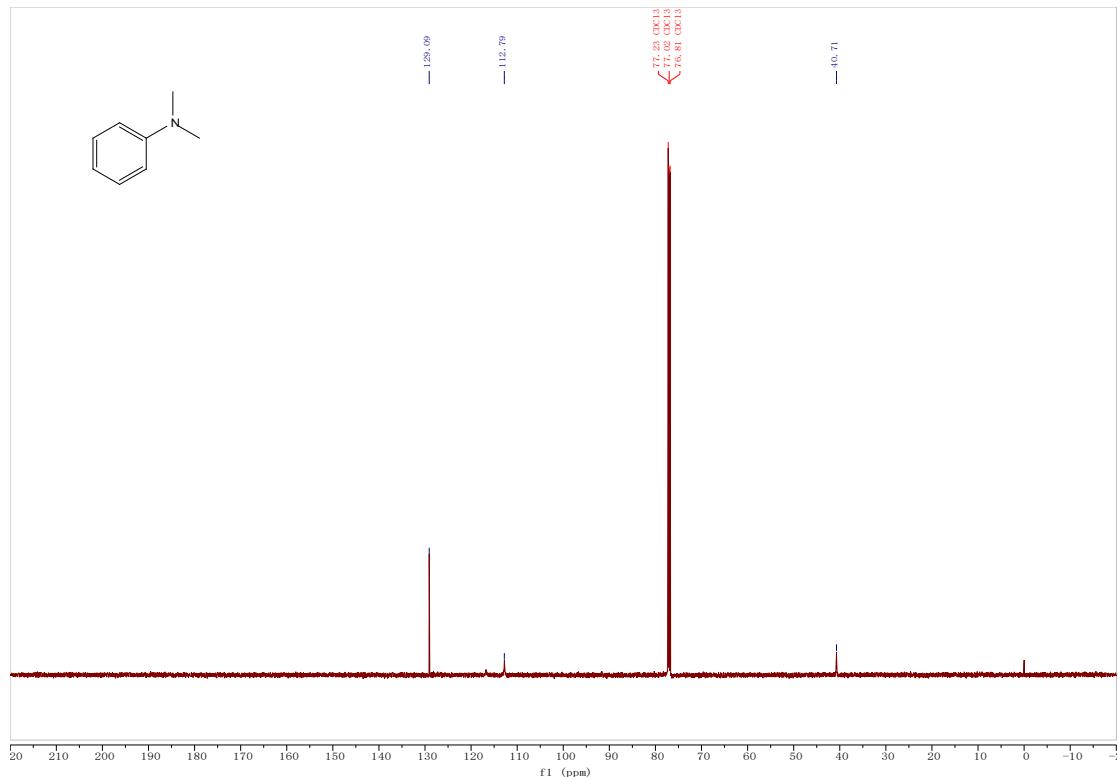
NMR spectra of selected products

N,N-Dimethylaniline

^1H NMR (600 MHz, Chloroform-*d*) δ 7.28 (d, $J = 5.4$ Hz, 2H), 6.78 (dd, $J = 21.9, 7.8$ Hz, 3H), 2.98 (s, 6H). ^{13}C NMR (151 MHz, Chloroform-*d*) δ 129.09, 112.79, 40.71.



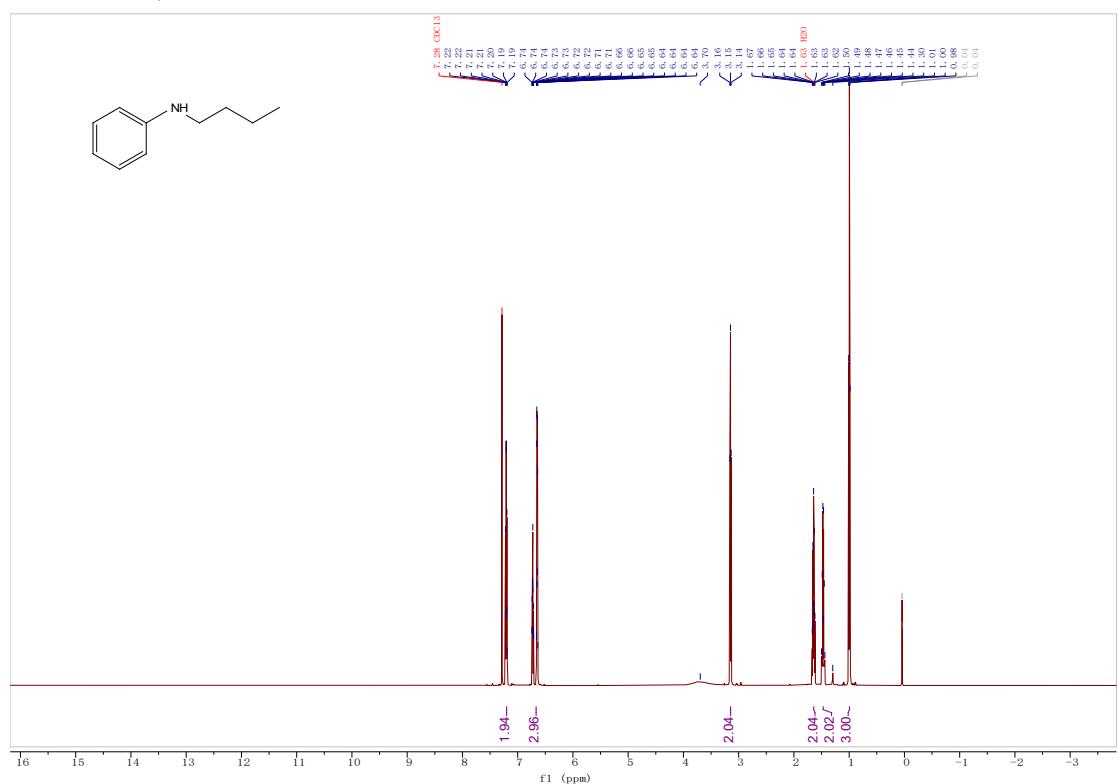
^1H NMR spectra of *N,N*-Dimethylaniline



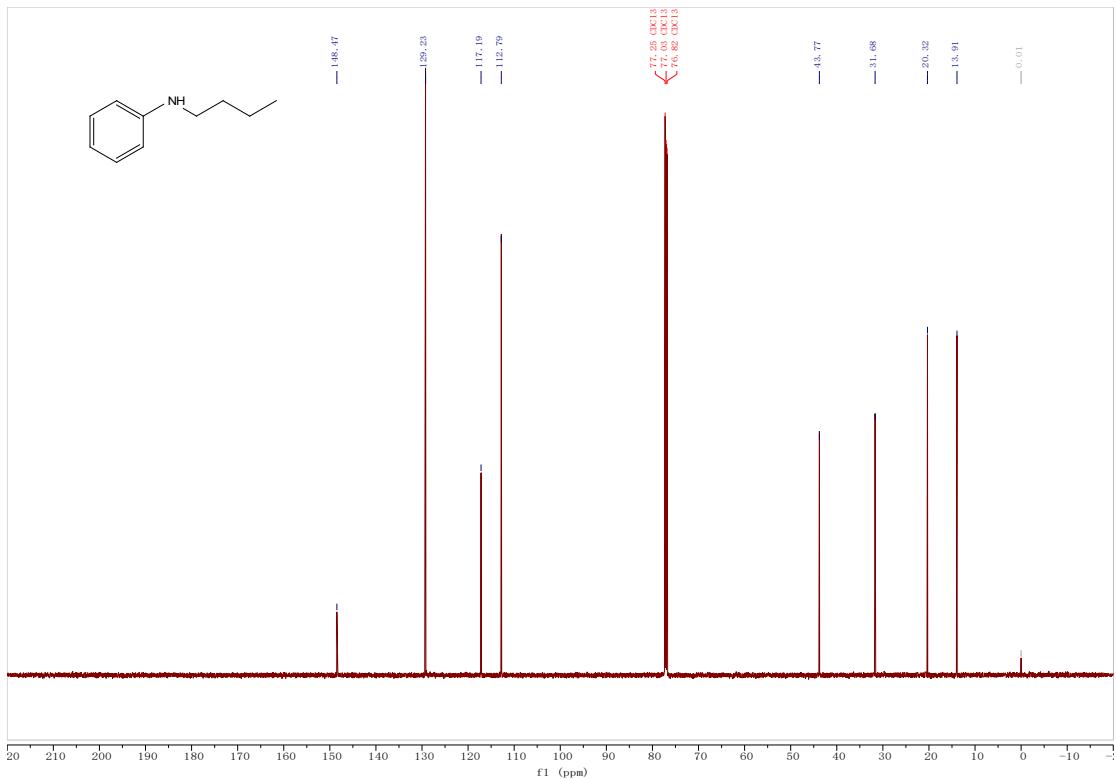
¹³C NMR spectra of N,N-Dimethylaniline

N-Butylaniline

¹H NMR (600 MHz, Chloroform-*d*) δ 7.26 – 7.14 (m, 2H), 6.82 – 6.51 (m, 3H), 3.15 (t, *J* = 7.1 Hz, 2H), 1.64 (p, *J* = 7.2 Hz, 2H), 1.47 (h, *J* = 7.4 Hz, 2H), 1.00 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 148.47, 129.23, 117.19, 112.79, 43.77, 31.68, 20.32, 13.91.



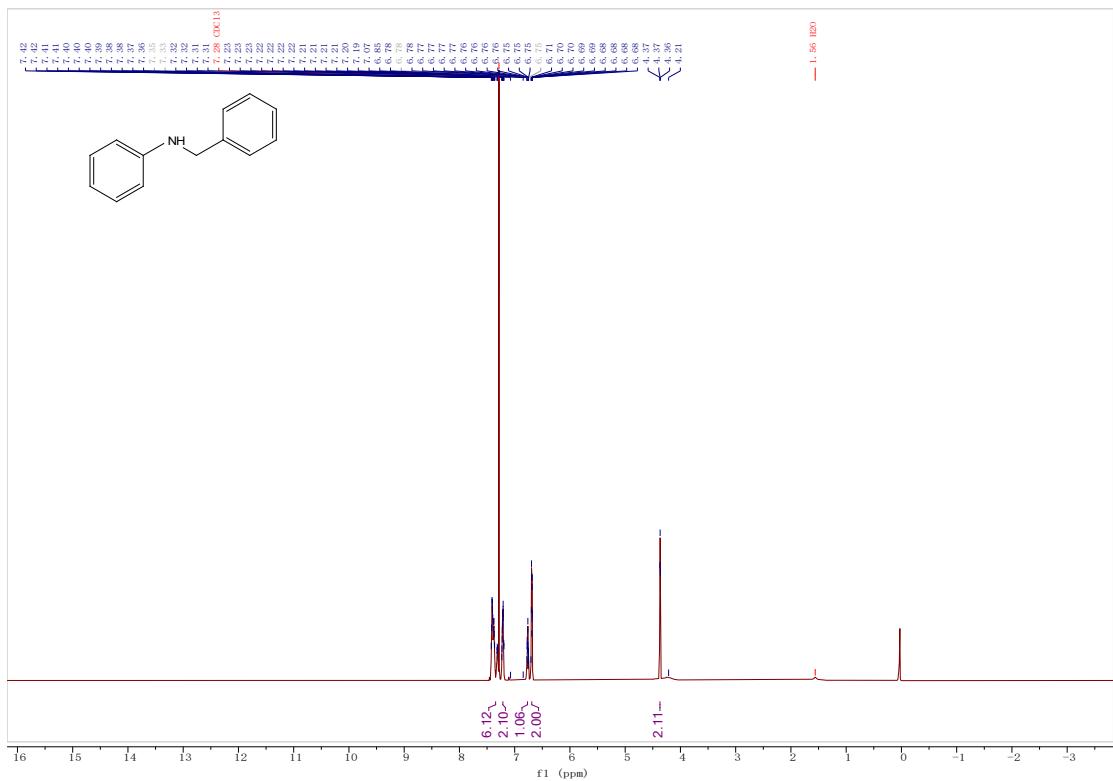
¹H NMR spectra of N-Butylaniline

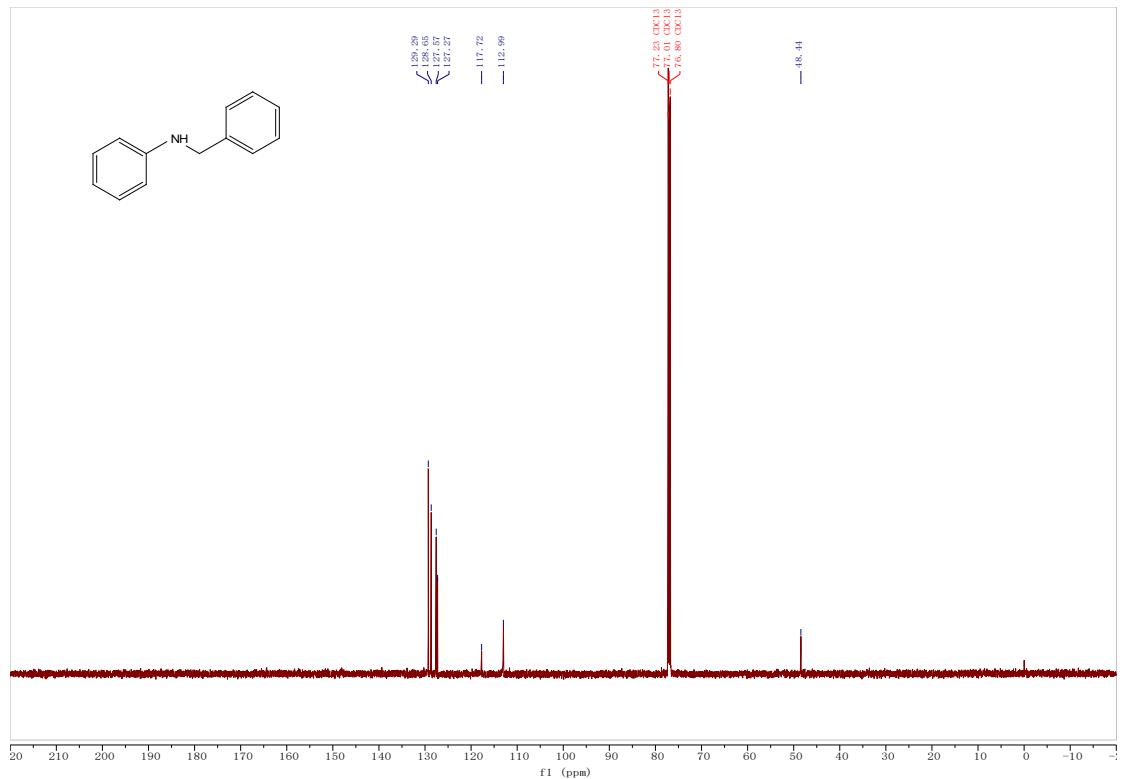


¹³C NMR spectra of N-Butylaniline

N-Benzylaniline

¹H NMR (600 MHz, Chloroform-*d*) δ 7.41 – 7.28 (m, 6H), 7.24 – 7.18 (m, 2H), 6.76 (dd, *J* = 7.3, 6.2, 3.1, 1.8 Hz, 1H), 6.72 – 6.67 (m, 2H), 4.38 – 4.36 (m, 2H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 129.29, 128.65, 127.57, 127.27, 117.72, 112.99, 48.44.

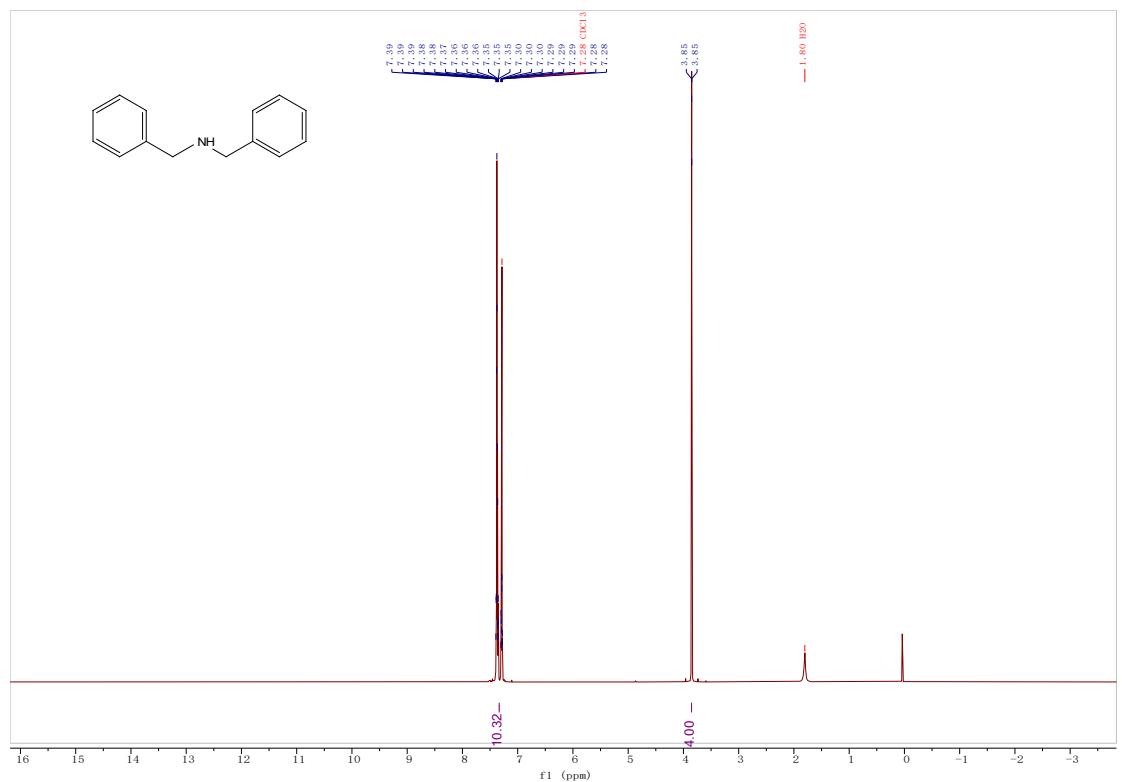




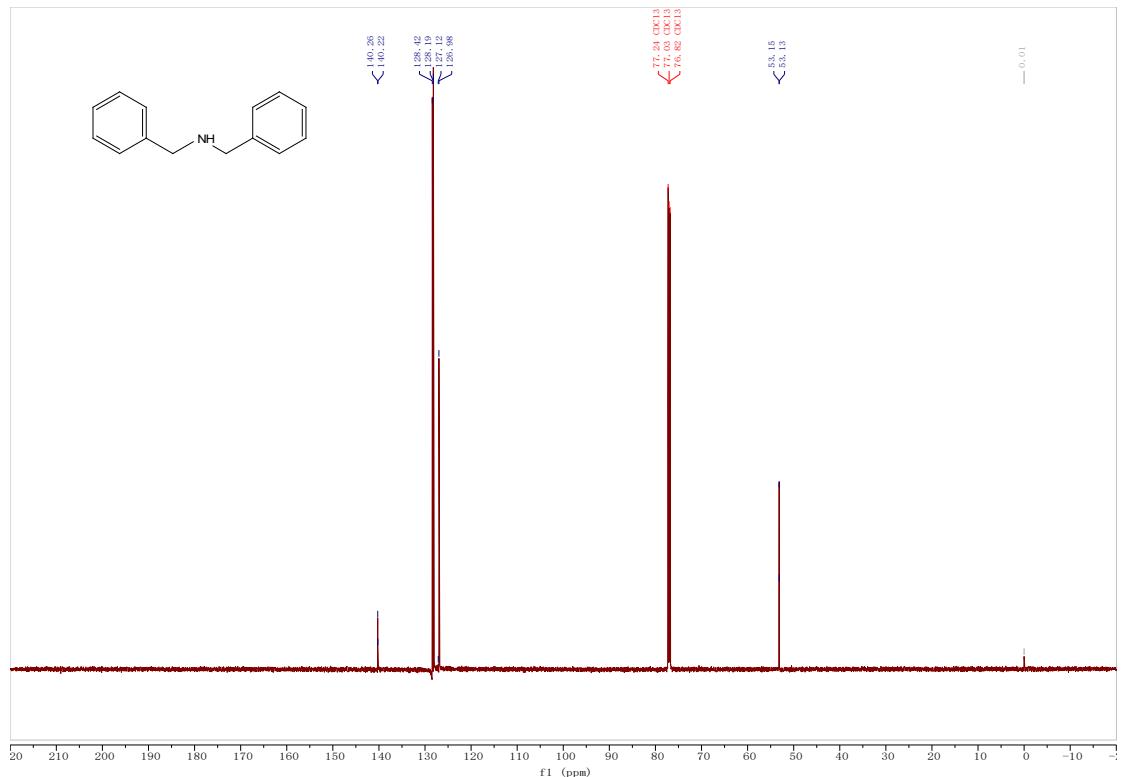
¹³C NMR spectra of N-Benzylaniline

Dibenzylamine

¹H NMR (600 MHz, Chloroform-*d*) δ 7.40 – 7.27 (m, 10H), 3.85 (d, *J* = 1.2 Hz, 4H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 140.26, 140.22, 128.42, 128.19, 126.98, 53.15, 53.13.



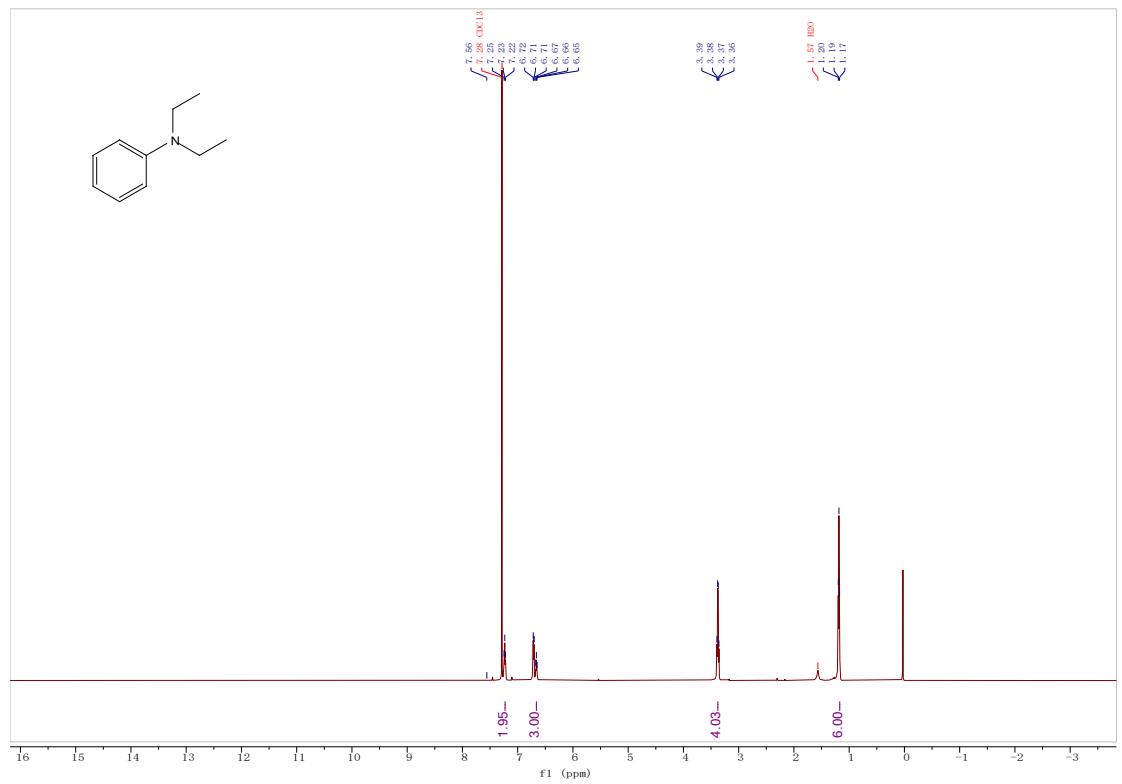
¹H NMR spectra of Dibenzylamine



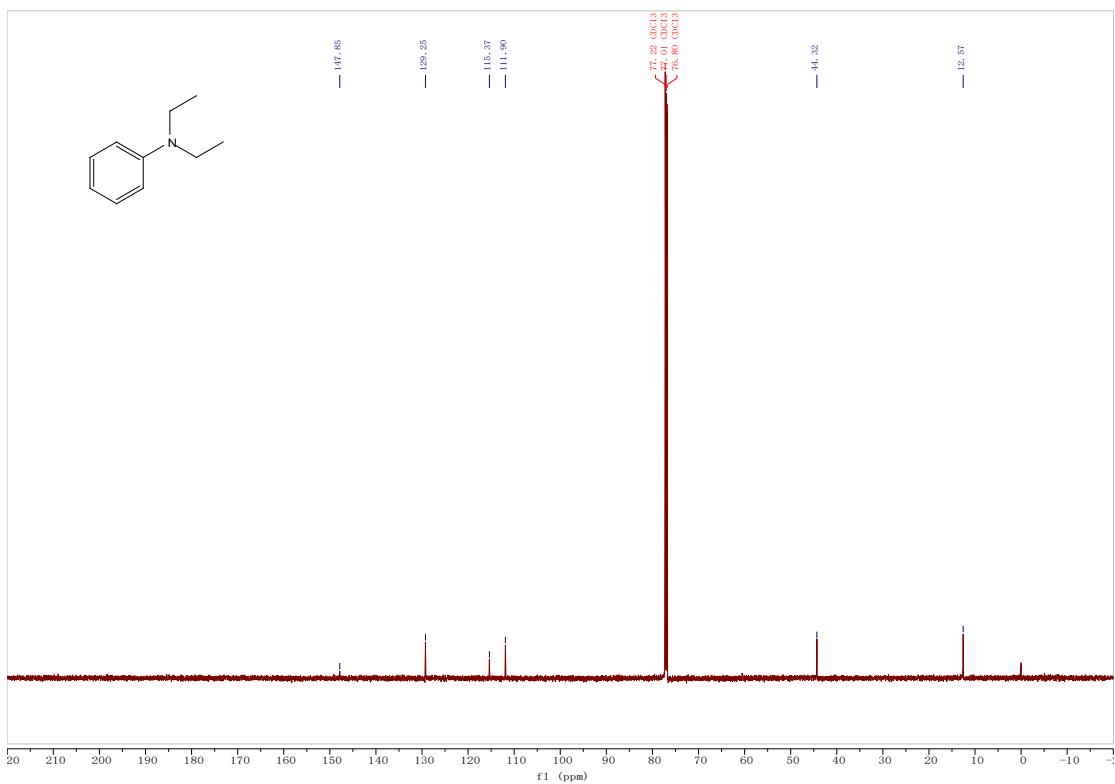
¹³C NMR spectra of Dibenzylamine

***N,N*-Diethylaniline**

¹H NMR (600 MHz, Chloroform-*d*) δ 7.23 (t, *J* = 7.7 Hz, 2H), 6.77 – 6.55 (m, 3H), 3.38 (q, *J* = 7.0 Hz, 4H), 1.19 (t, *J* = 7.0 Hz, 6H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 147.85, 129.25, 115.37, 111.90, 44.32, 12.57.



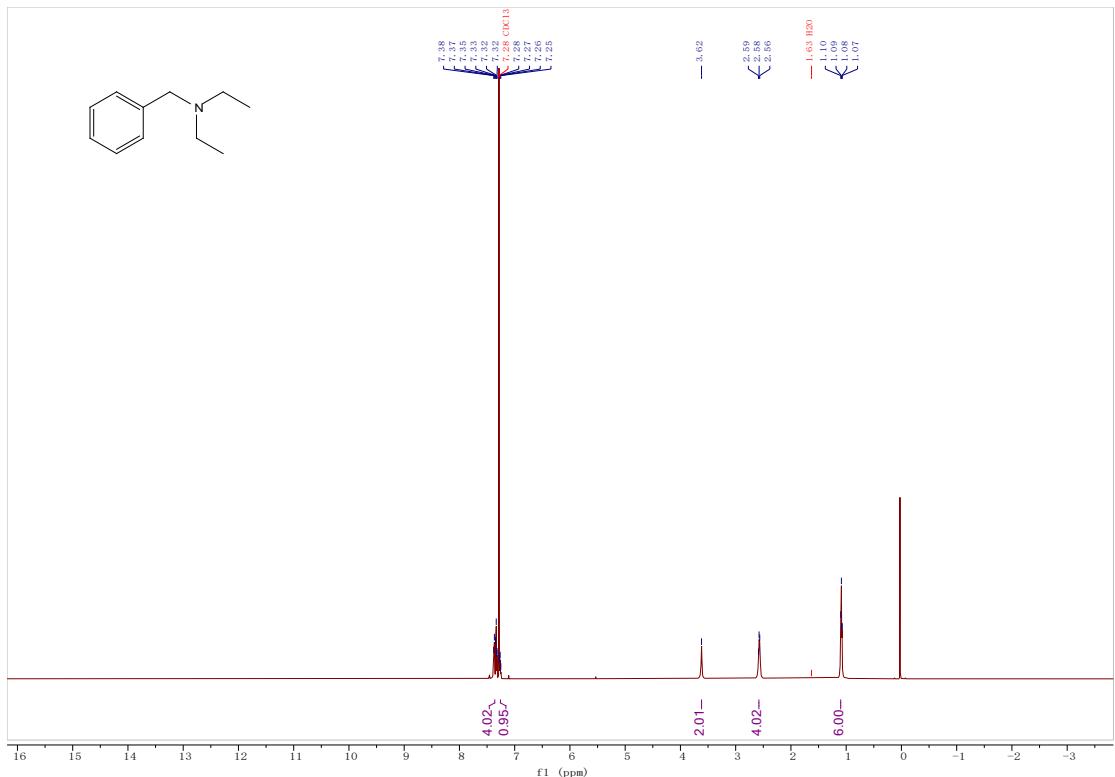
¹H NMR spectra of N,N-Diethylaniline



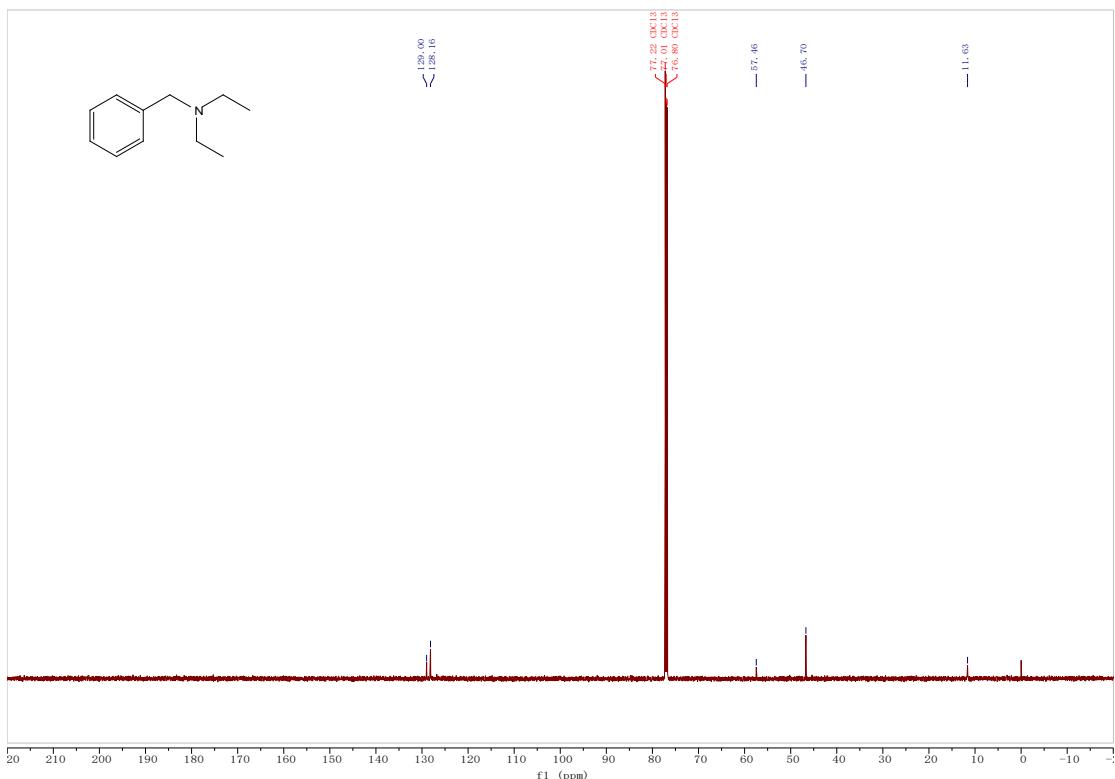
¹³C NMR spectra of N,N-Diethylaniline

N-Benzyldiethylamine

¹H NMR (600 MHz, Chloroform-*d*) δ 7.42 – 7.30 (m, 4H), 7.28 – 7.24 (m, 1H), 3.62 (s, 2H), 2.66 – 2.50 (m, 4H), 1.09 (t, J = 6.6 Hz, 6H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 129.00, 128.16, 57.46, 46.70, 11.63.



¹H NMR spectra of N-Benzyl-diethylamine

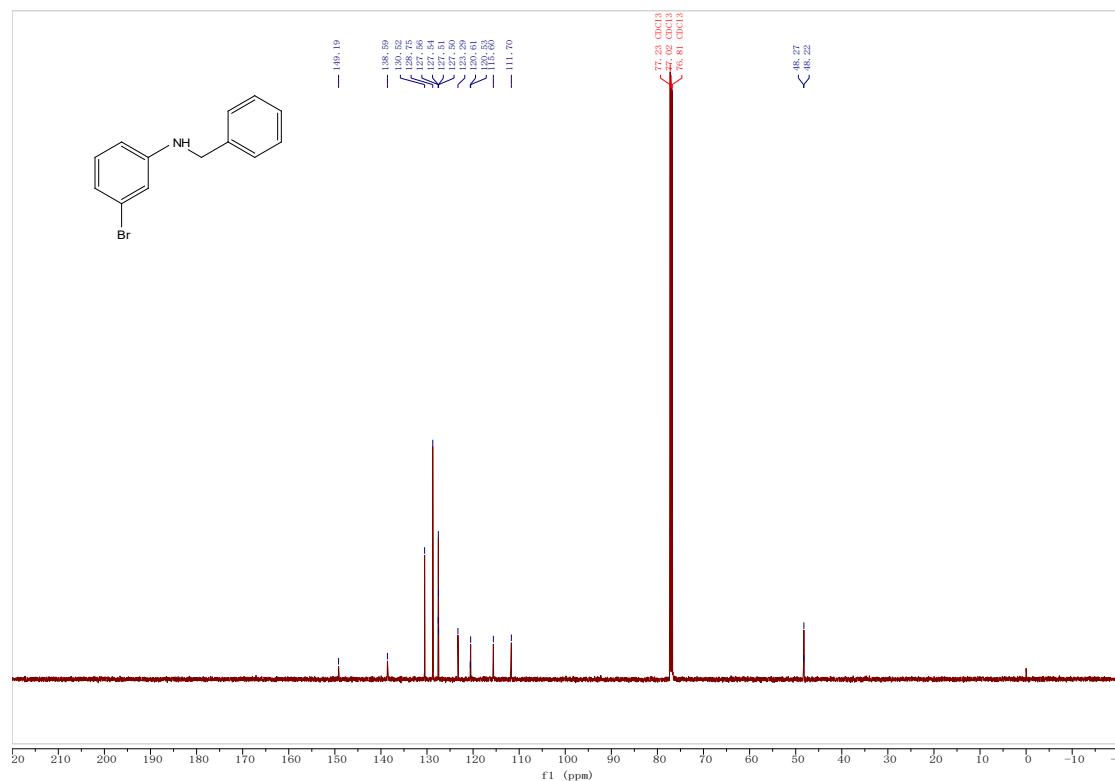
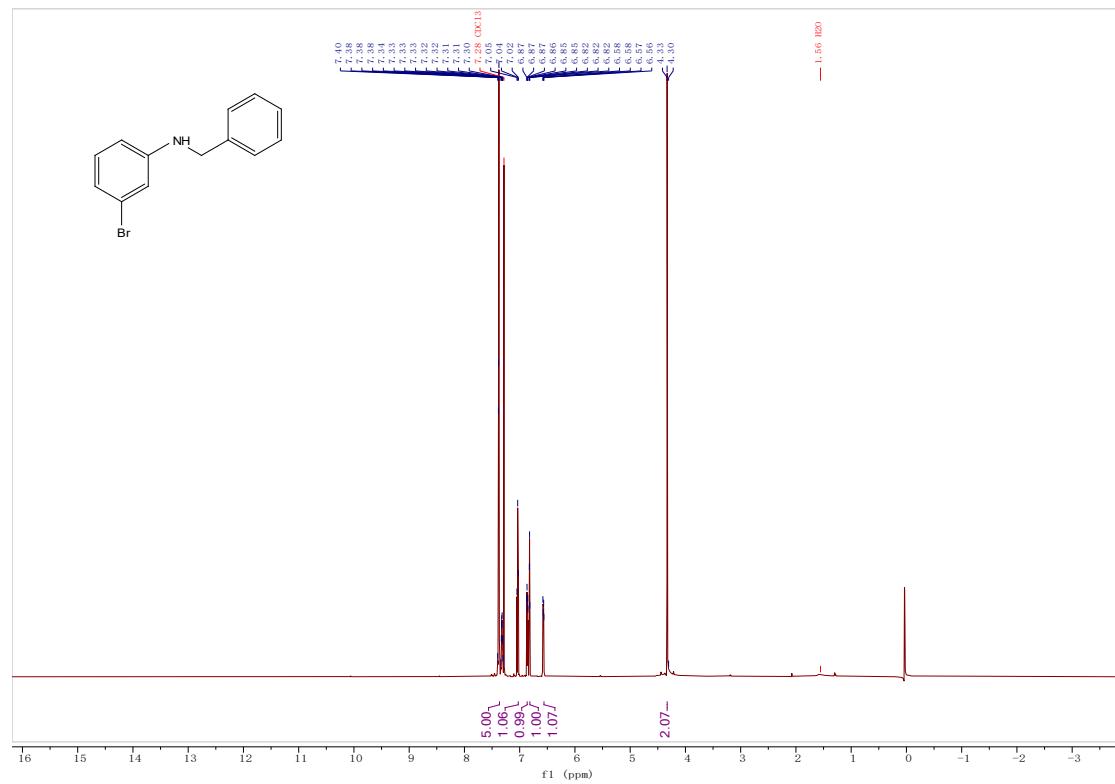


¹³C NMR spectra of N-Benzyl-diethylamine

N-benzyl-3-bromoaniline

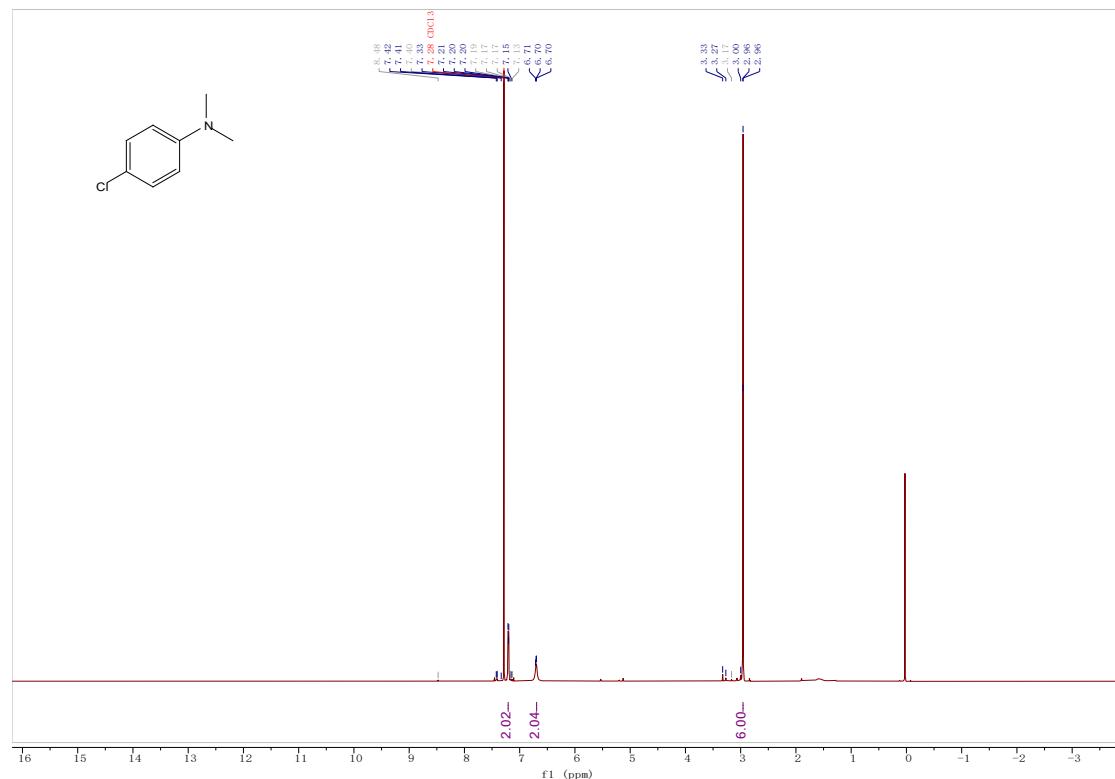
¹H NMR (600 MHz, Chloroform-*d*) δ 7.43 – 7.30 (m, 5H), 7.04 (t, *J* = 8.0 Hz, 1H), 6.86 (dt, *J* = 7.8, 1.2

Hz, 1H), 6.82 (t, J = 2.1 Hz, 1H), 6.57 (dd, J = 8.2, 2.4 Hz, 1H), 4.33 (s, 2H). ^{13}C NMR (151 MHz, Chloroform-*d*) δ 149.19, 138.59, 130.52, 128.75, 127.56, 127.54, 127.51, 127.50, 123.29, 120.61, 120.53, 115.60, 111.70, 48.27, 48.22.

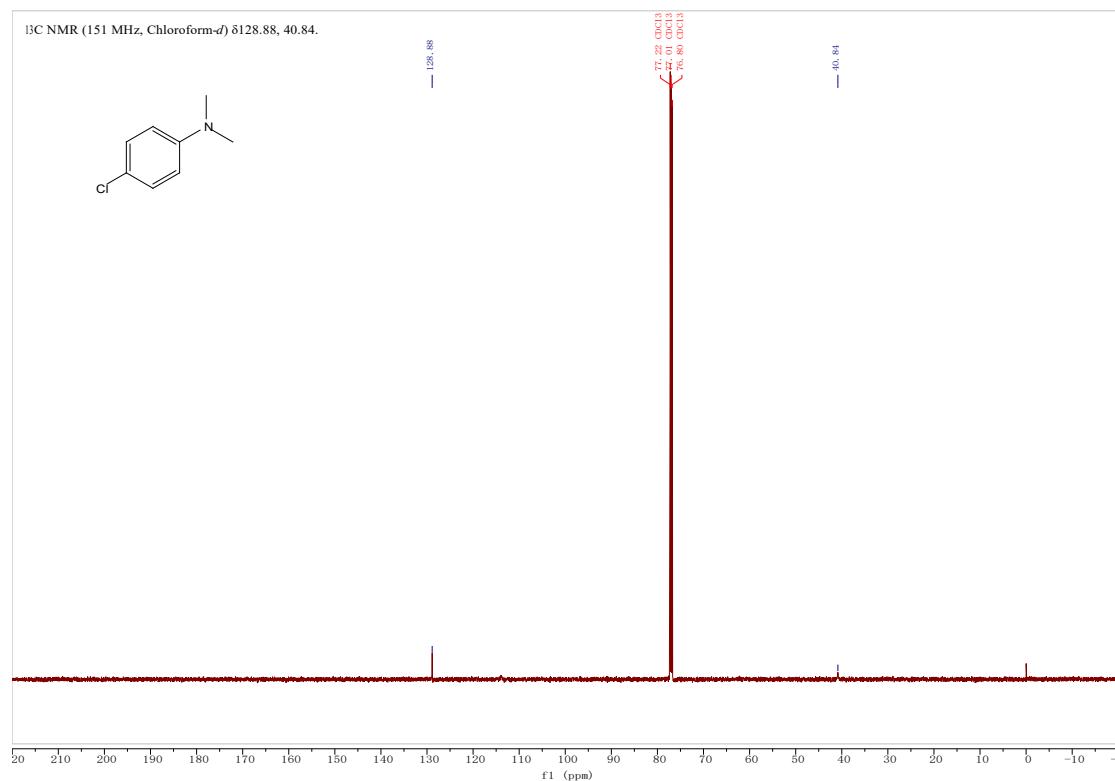


4-chloro-N,N-dimethylaniline

^1H NMR (600 MHz, Chloroform-*d*) δ 7.24 – 7.17 (m, 2H), 6.70 (t, J = 4.4 Hz, 2H), 2.96 (d, J = 1.4 Hz, 6H). ^{13}C NMR (151 MHz, Chloroform-*d*) δ 128.88, 40.84.



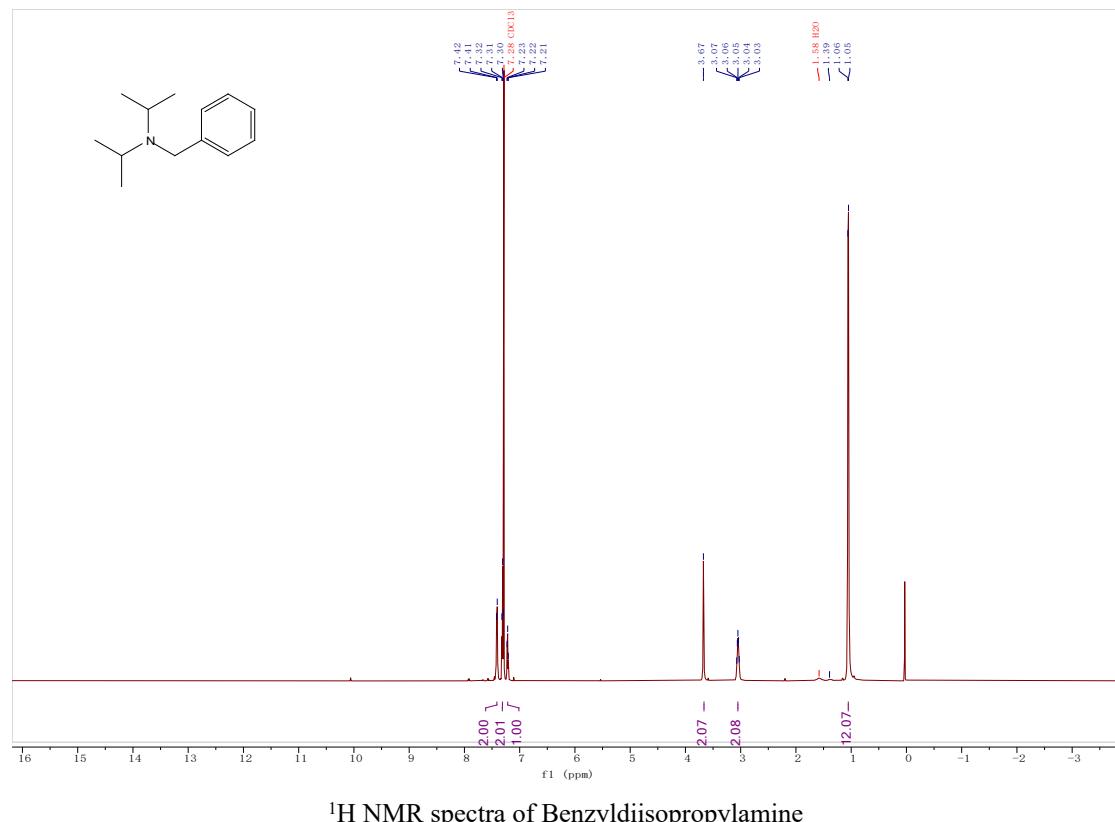
^1H NMR spectra of 4-chloro-N,N-dimethylaniline



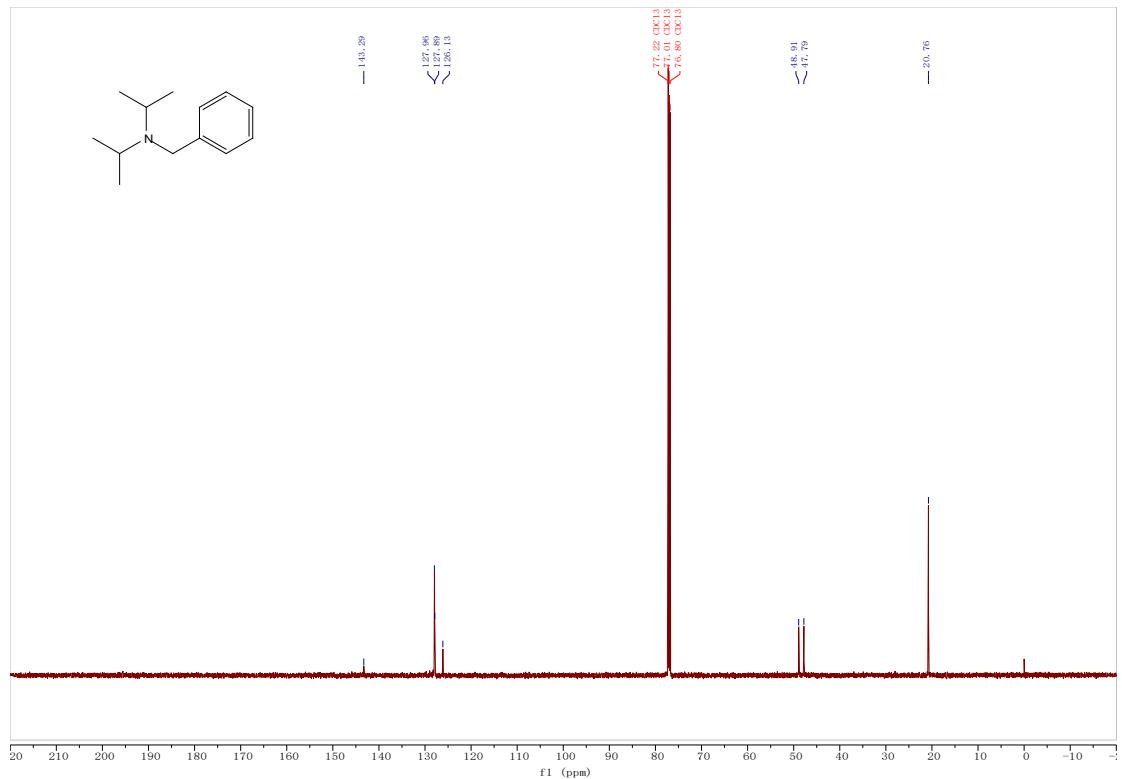
^{13}C NMR spectra of 4-chloro-N,N-dimethylaniline

Benzylidiisopropylamine

^1H NMR (600 MHz, Chloroform-*d*) δ 7.41 (d, $J = 7.5$ Hz, 2H), 7.31 (t, $J = 7.5$ Hz, 2H), 7.22 (t, $J = 7.3$ Hz, 1H), 3.67 (s, 2H), 3.06 (dd, $J = 12.9, 6.7$ Hz, 2H), 1.05 (d, $J = 6.6$ Hz, 12H). ^{13}C NMR (151 MHz, Chloroform-*d*) δ 143.29, 127.96, 127.89, 126.13, 48.91, 47.79, 20.76.



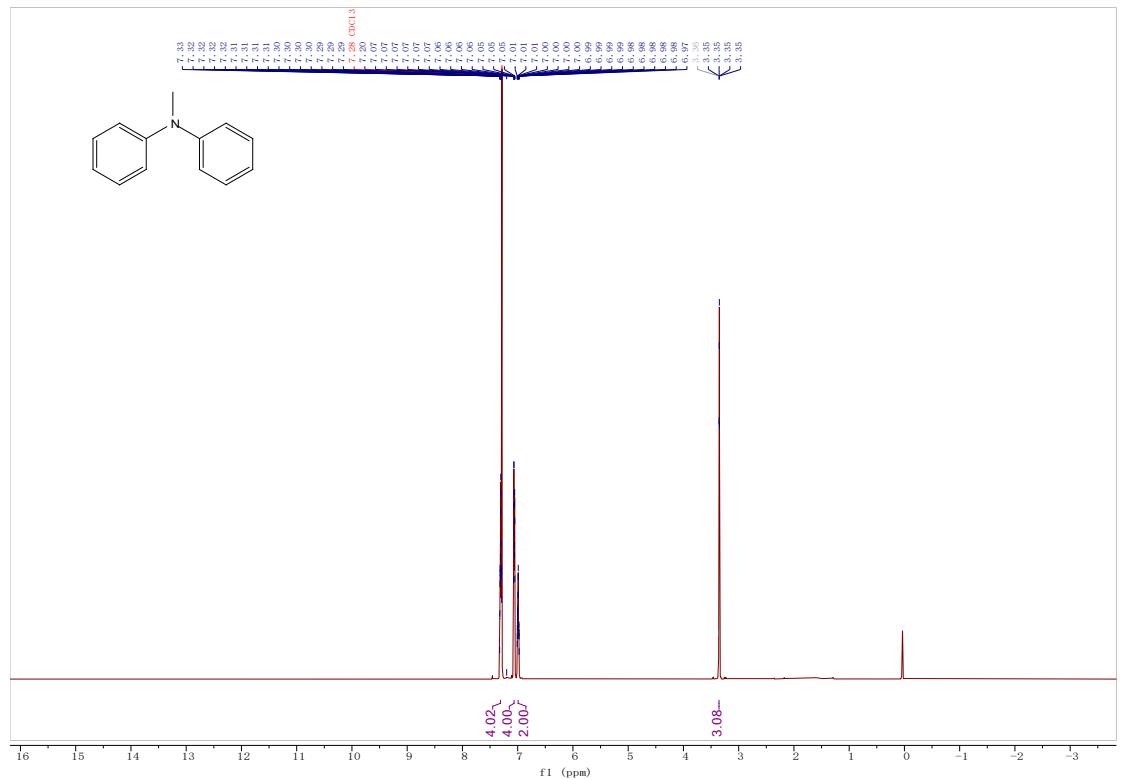
^1H NMR spectra of Benzylidiisopropylamine



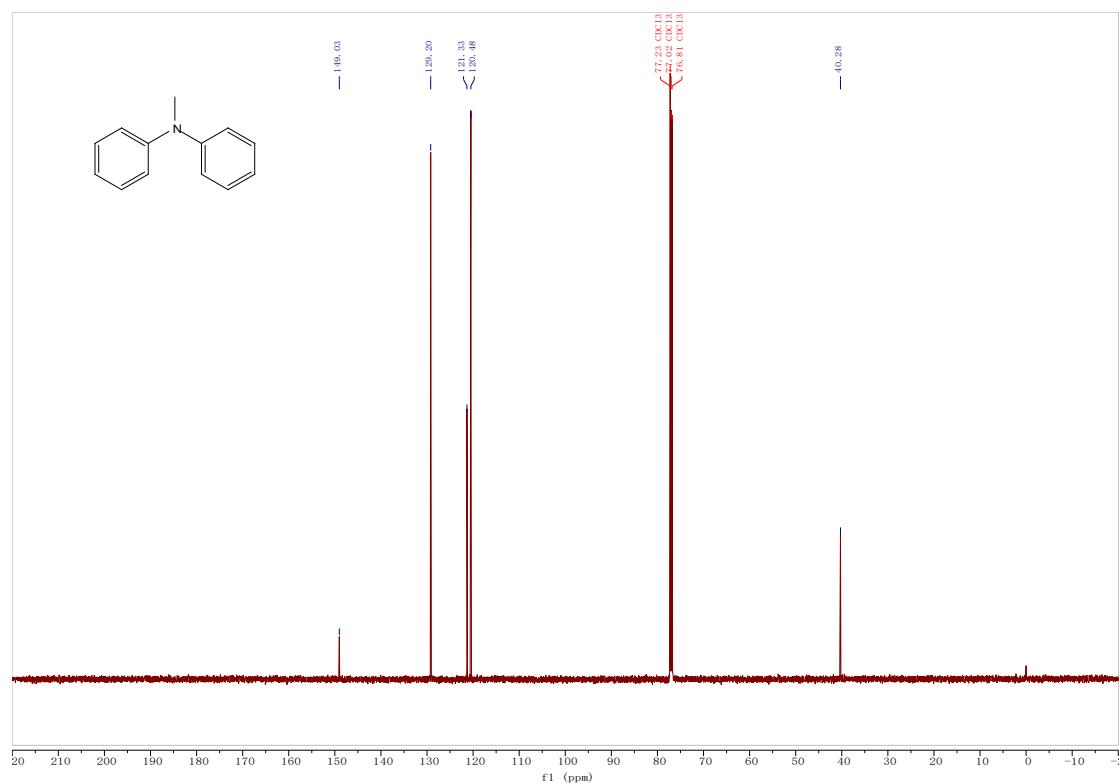
¹³C NMR spectra of Benzylidiisopropylamine

***N*-Methyldiphenylamine**

¹H NMR (600 MHz, Chloroform-*d*) δ 7.31 (dddt, *J* = 8.7, 7.5, 2.8, 1.6 Hz, 4H), 7.10 – 7.03 (m, 4H), 6.99 (dddd, *J* = 9.7, 7.2, 2.5, 1.3 Hz, 2H), 3.35 (dd, *J* = 2.8, 1.5 Hz, 3H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 149.03, 129.20, 121.33, 120.48, 40.28.



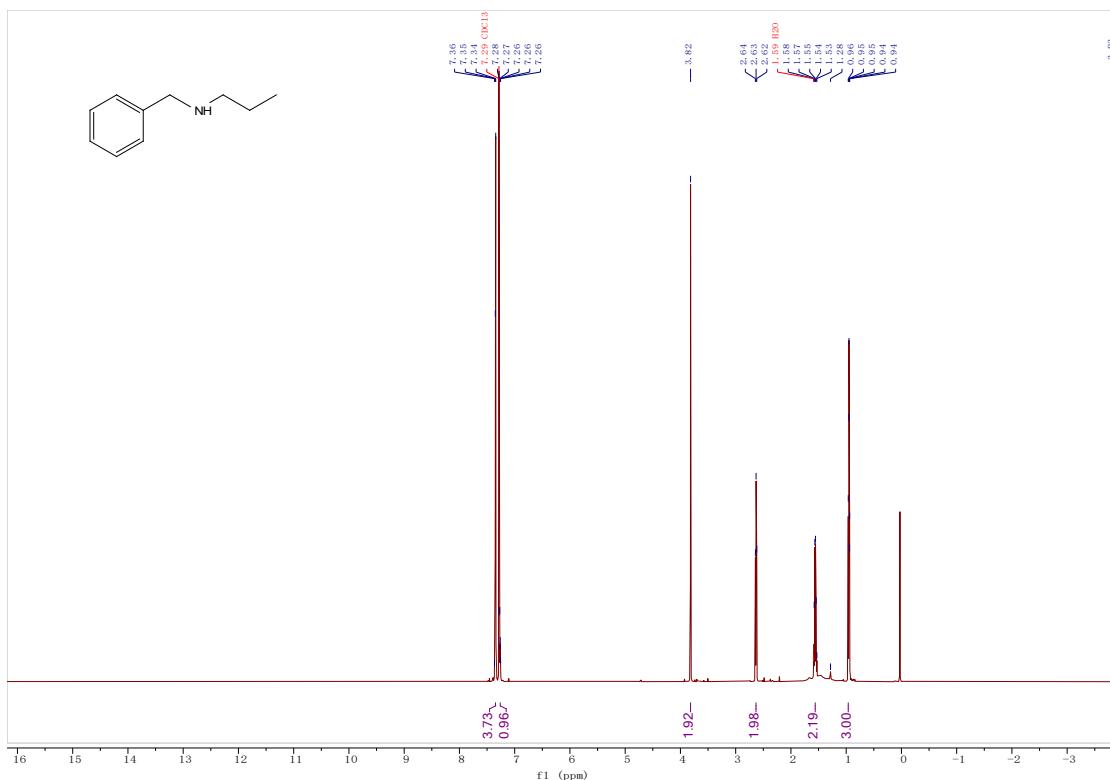
¹H NMR spectra of N-Methyldiphenylamine



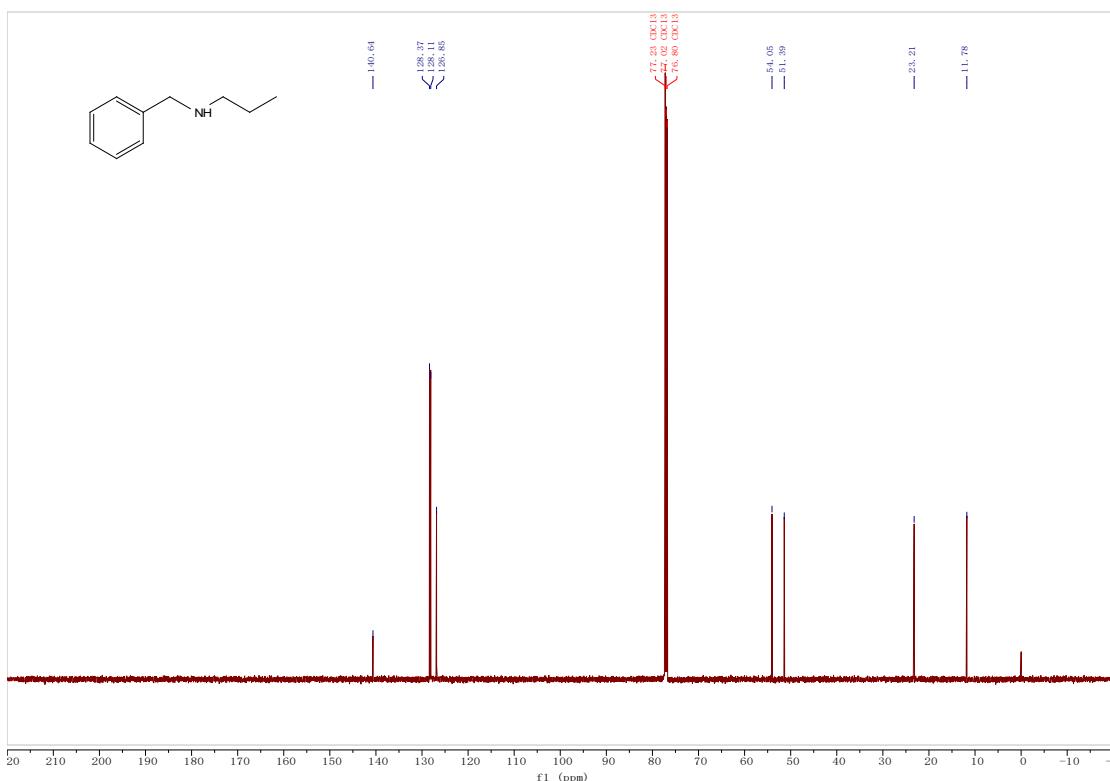
¹³C NMR spectra of N-Methyldiphenylamine

N-Benzylpropanamine

¹H NMR (600 MHz, Chloroform-*d*) δ 7.35 (d, J = 4.4 Hz, 4H), 7.28 – 7.24 (m, 1H), 3.82 (s, 2H), 2.63 (t, J = 7.2 Hz, 2H), 1.55 (p, J = 7.3 Hz, 2H), 1.04 – 0.89 (m, 3H). ¹³C NMR (151 MHz, Chloroform-*d*) δ 140.64, 128.37, 128.11, 126.85, 54.05, 51.39, 23.21, 11.78.



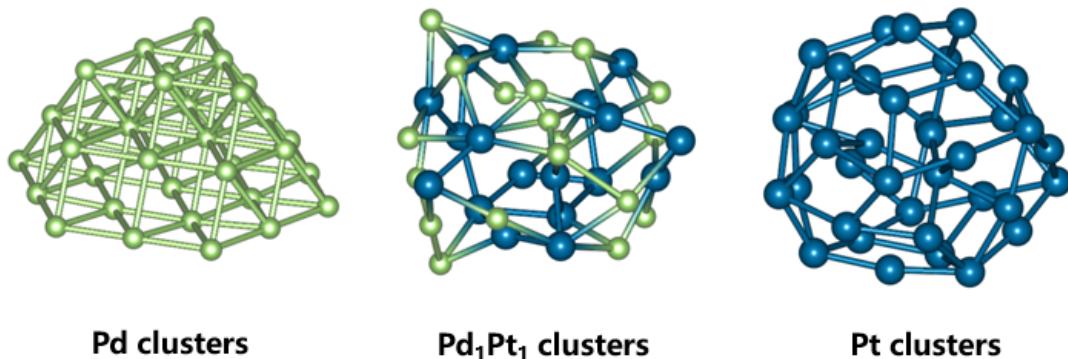
¹H NMR spectra of N-Benzylpropanamine



¹³C NMR spectra of N-Benzylpropanamine

Computational Section

Figure S6. Stick model for Pd, Pd₁Pt₁ and Pt clusters



Computational Details

The structural optimization and the frequency analysis were performed at the GFN-xTB level of the xTB package.^{2,3} Stationary points were optimized without symmetry constraint, and their nature was confirmed by vibrational frequency analysis.⁴ To achieve this, a Gaussian interface to the xTB code “gau_xtb” was employed.⁵ Analysis on the molecular orbitals was performed at the PBE0/def2-SV(P) level of theory using ORCA 4.2 package.⁶

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