

Supporting Information

AIE-Active Cyclometalated Iridium(III) Complexes for Detection of Lipopolysaccharide and Wash-Free Imaging of Bacteria

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2. Instruments

2.1 Thin layer chromatography (TLC) and column chromatography

TLC was performed on aluminium plates coated with silica gel mixed with fluorescent indicator. The purification of synthesized ligand and complexes were performed with silica gel (60-120 mesh) column chromatography.

2.2 NMR spectroscopy

^1H and ^{13}C NMR spectra were acquired on a Bruker 400 and 500 MHz spectrometer in CDCl_3 or DMSO-d_6 at ambient temperature with tetramethylsilane (TMS) as an internal standard. NMR standards used were as follows: (^1H -NMR) $\text{CDCl}_3 = 7.260$ ppm; $\text{DMSO-d}_6 = 2.50$ ppm. (^{13}C -NMR) $\text{CDCl}_3 = 77.16$ ppm; $\text{DMSO-d}_6 = 39.52$ ppm. All chemical shifts (δ) are reported in ppm relative to TMS. Spin multiplicities were reported as a singlet (s), doublet (d), triplet (t), quartet (q), doublet of doublets (dd), triplet of doublets (td), multiplet (m) and broad (br) with coupling constant (J) reported in Hz.

2.3 Mass spectrometry

Electrospray ionization mass spectrometry (ESI-MS) and high-resolution mass spectrometry (ESI-HRMS) were recorded on Xevo G2-XS QToF Quadrupole Time of Flight Mass Spectrometer from Waters India Pvt. Ltd.

2.4 FT-IR spectroscopy

Fourier transform-Infrared (FT-IR) spectra were measured using IR Affinity-1S (Shimadzu, Kyoto, Japan) FT-IR spectrophotometer equipped with a single reflection attenuated total reflectance (ATR) accessory. The IR spectra were recorded from 4000 to 450 cm^{-1} using a resolution of 4 cm^{-1} with 45 scans. In IR absorption spectra, the shapes and signal intensities (height) of peaks (bands) are denoted by the following abbreviations: br = broad, vs = very strong, s = strong, m = medium and w = weak.

2.5 UV-vis spectroscopy

UV-vis absorption spectra were measured using a SpectraMax M2 plate reader (Molecular Devices) and an Agilent Technologies Carry 100 spectrophotometer, respectively, at 298 K from 800 to 200 nm.

2.6 Emission spectroscopy

Emission spectra and quantum yields were carried out on Edinburgh Instruments F900 fluorescence spectrophotometer.

2.7 Dynamic light scattering (DLS)

DLS was carried out using Zetasizer Nano ZS90 (Malvern Instrument Ltd., Worcestershire, UK).

2.8 Transmission electron microscopy (TEM)

Transmission electron microscopy images were taken on a JEOL JEM-1400 electron microscope operated at an acceleration voltage of 120 kV.

Table S1 The LOD of LPS by the complexes (**Ir1-Ir3**) determined using fluorescence spectroscopy.

Complexes	LOD (nM)
Ir1	47 ± 5
Ir2	144 ± 8
Ir3	157 ± 5

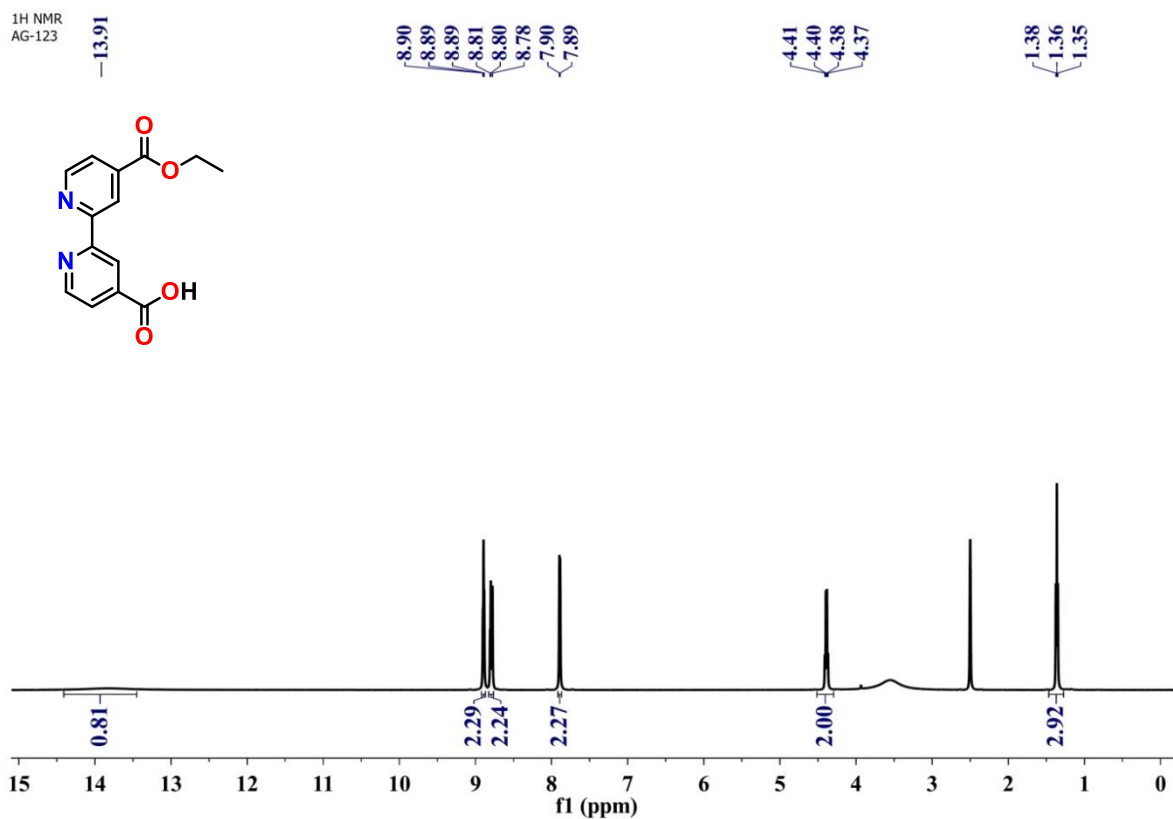


Figure S1 ¹H-NMR spectrum of compound **2** in DMSO-d₆ at 298K.

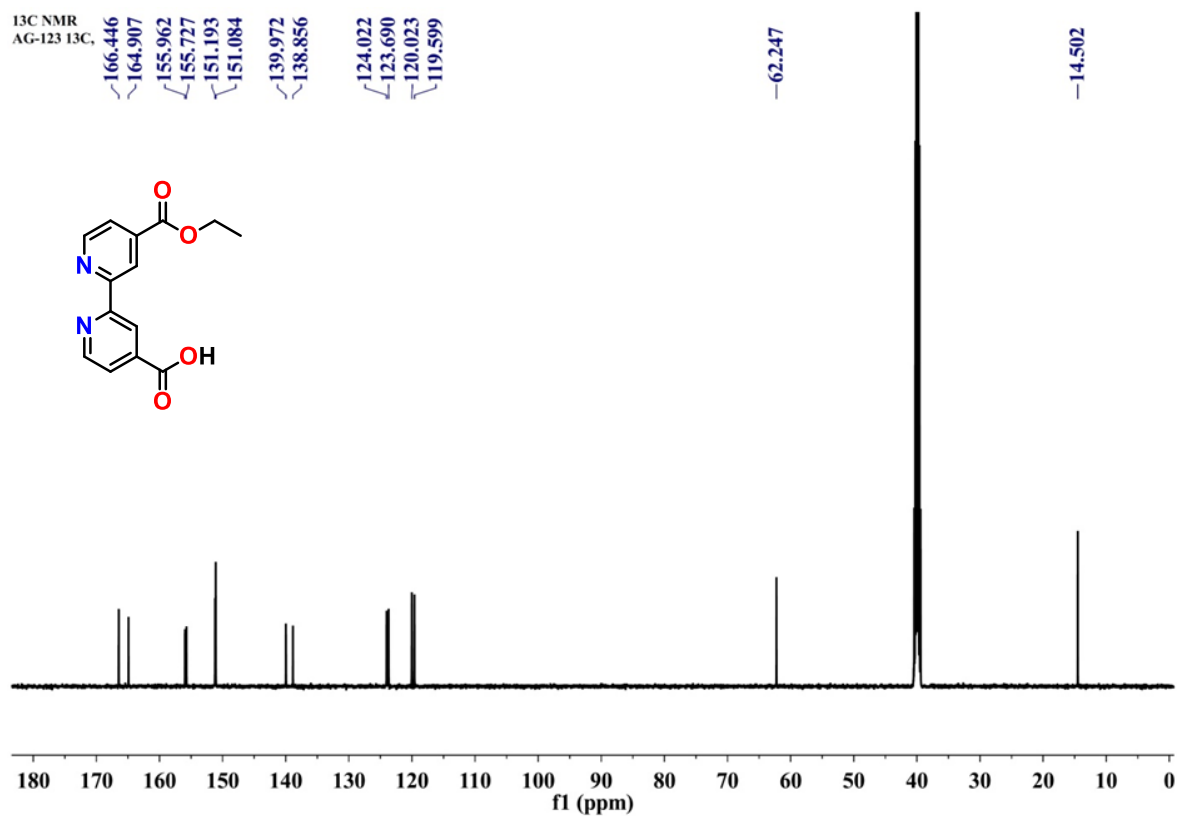


Figure S2 ¹³C-NMR spectrum of compound **2** in DMSO-d₆ at 298K.

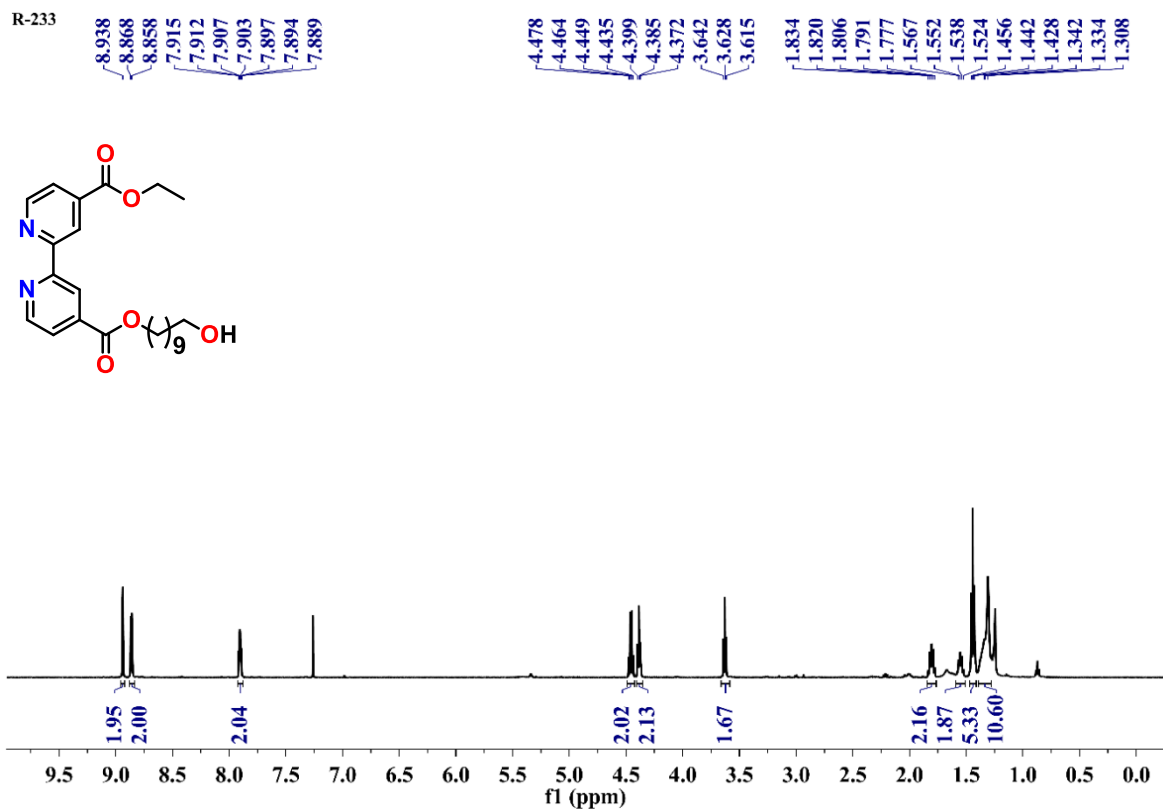


Figure S3 ^1H -NMR spectrum of compound **3** in CDCl_3 at 298K.

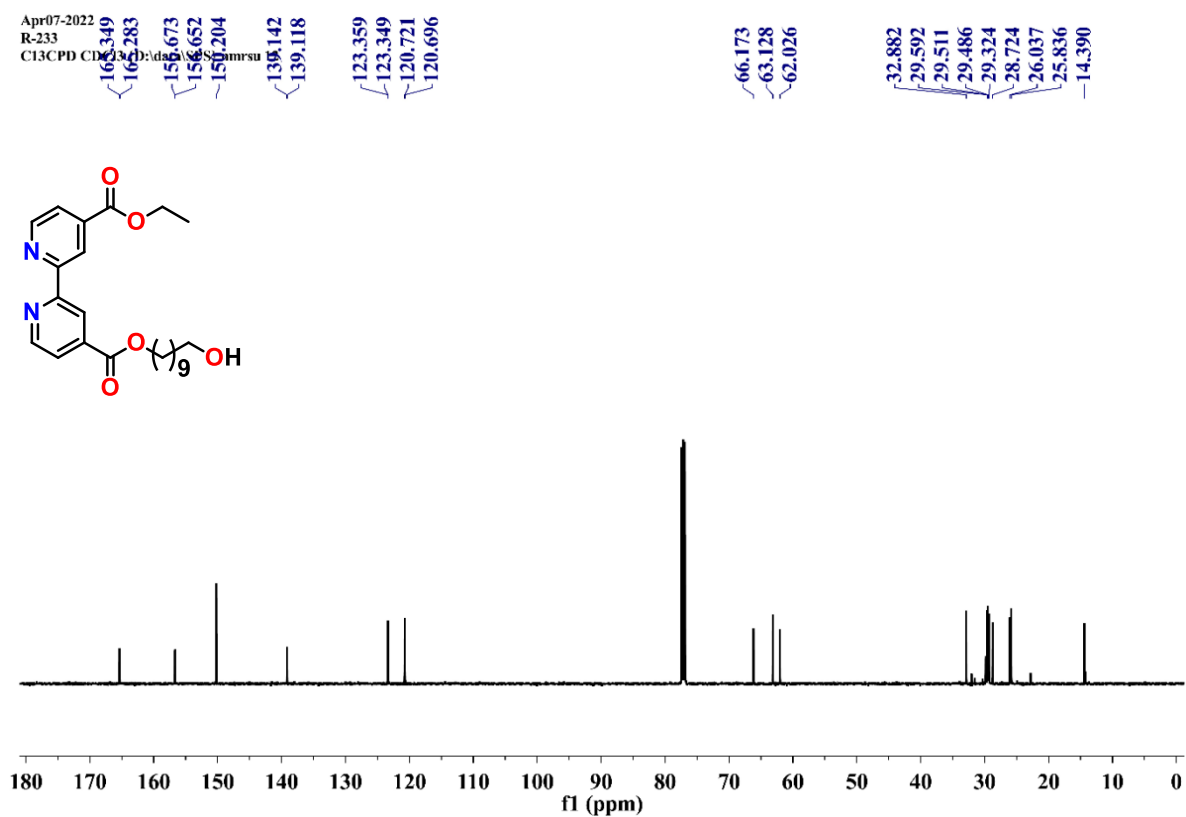


Figure S4 ^{13}C -NMR spectrum of compound **3** in CDCl_3 at 298K.

R-234

8.942
8.871
8.862
7.910
7.908
7.900
7.792
7.776
7.746
7.329

4.483
4.468
4.454
4.440
4.440
4.398
4.384
4.370
4.021
4.008
3.995

2.441
1.801
1.786
1.639
1.625
1.610
1.597
1.461
1.446
1.432
1.329
1.302
1.280

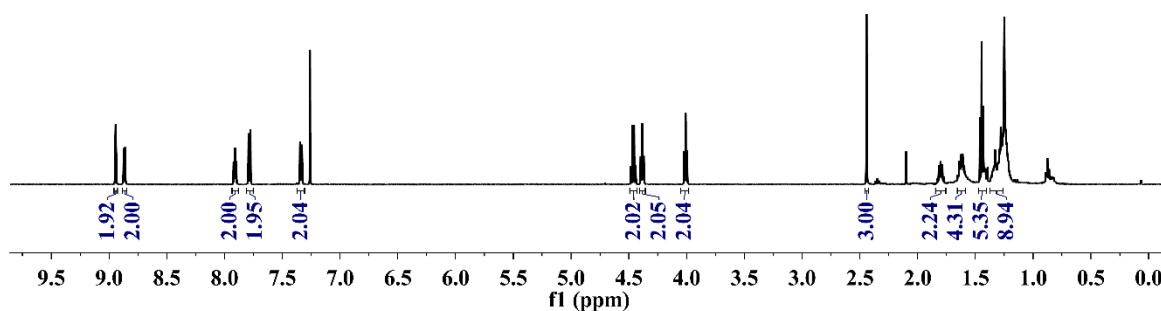
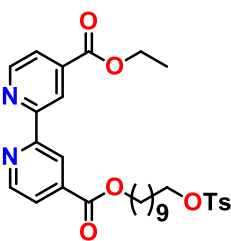


Figure S5 ¹H-NMR spectrum of compound 4 in CDCl₃ at 298K.

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R-234
C13CPD CDCl₃

165.338
165.272
158.712
158.680
158.197
143.699
138.136
133.494
129.899
129.688
127.988
127.170
123.341
120.690

70.757
66.121
62.001

42.121
29.433
29.388
29.292
29.003
28.953
28.737
26.031
25.438
21.715
14.387

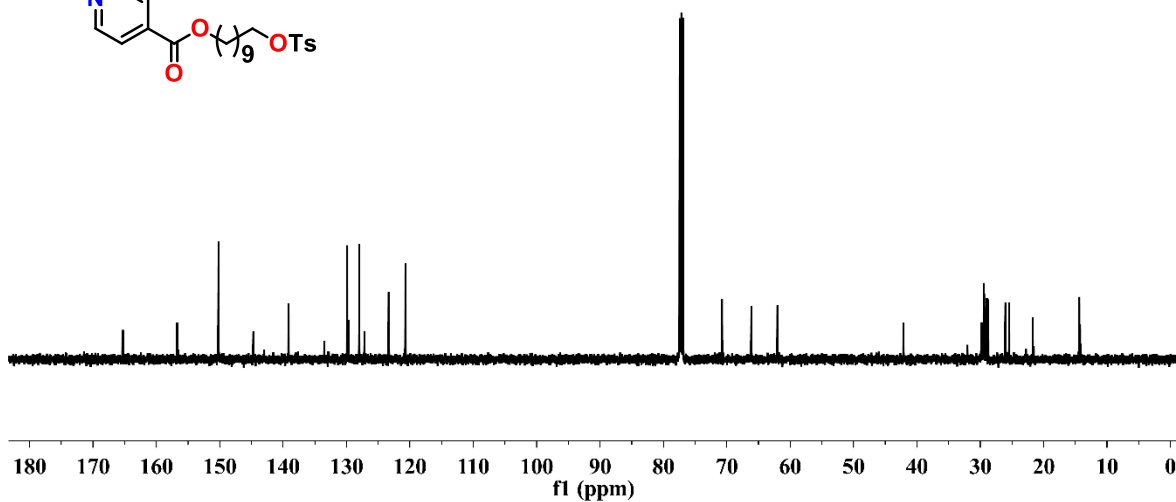
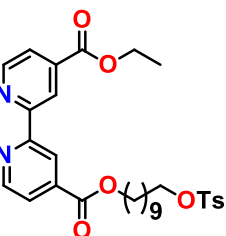
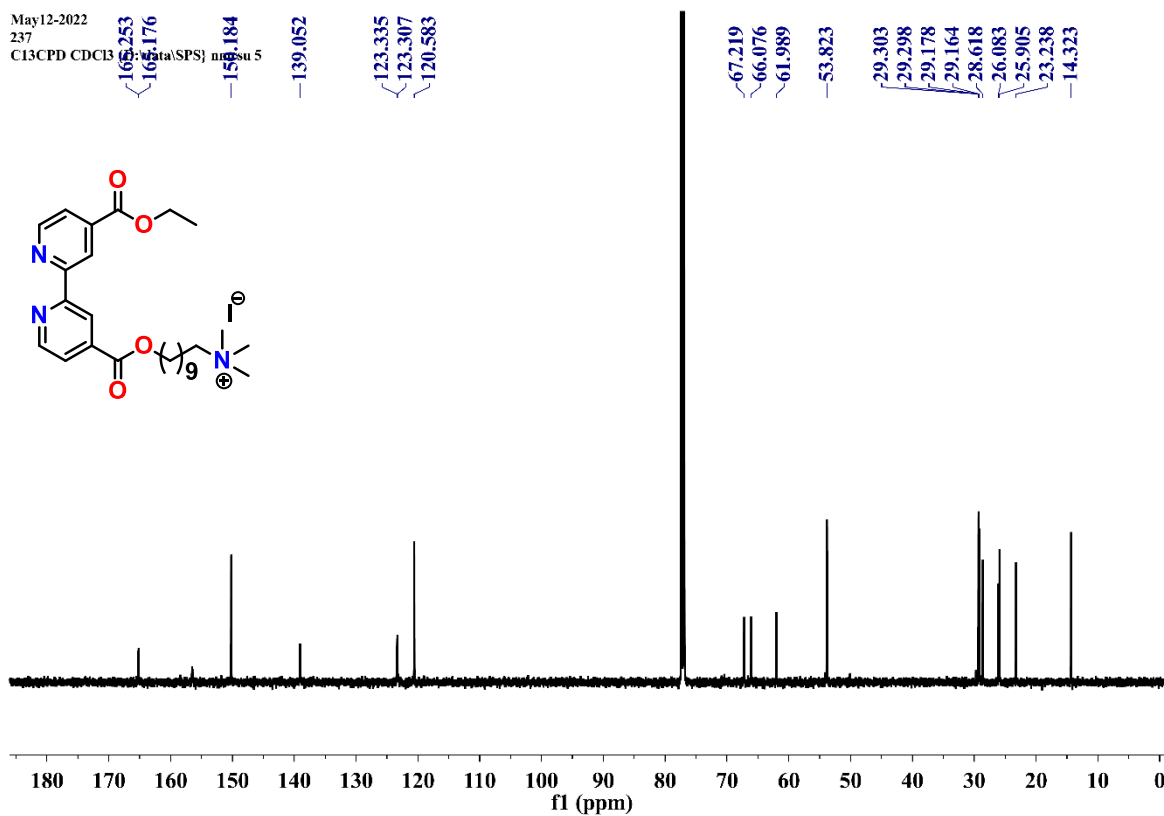
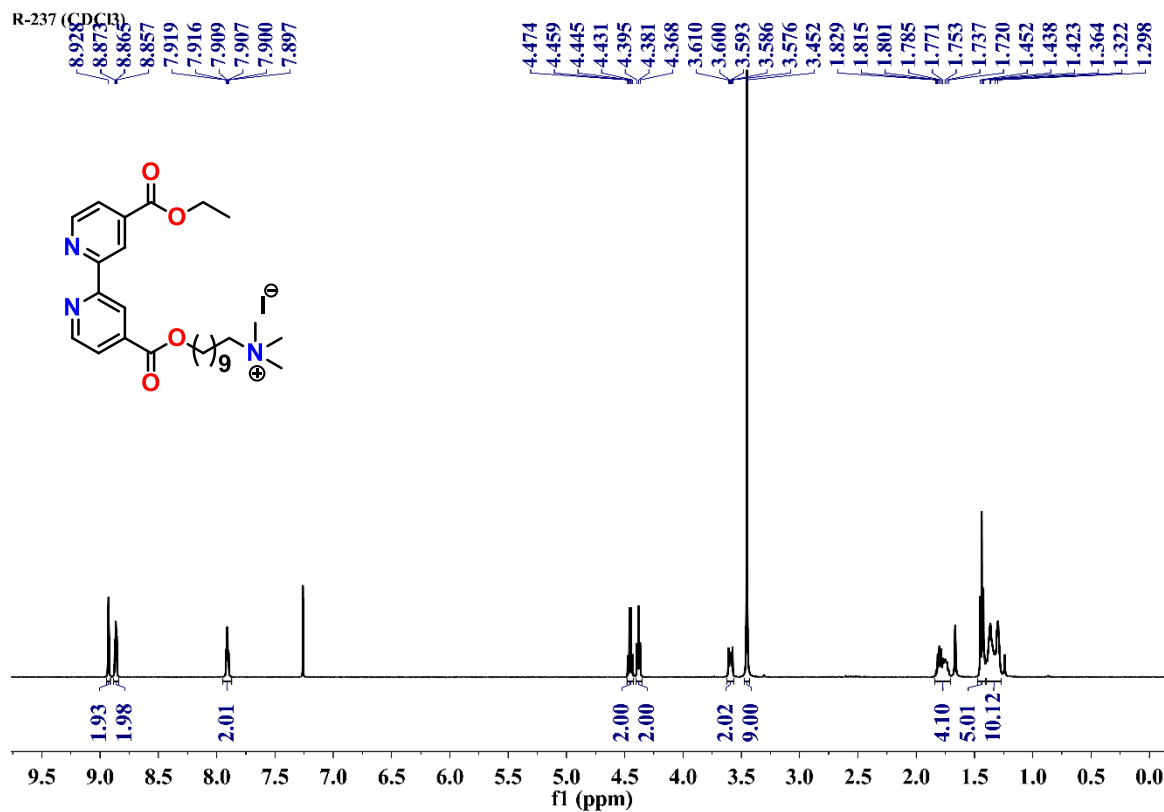


Figure S6 ¹³C-NMR spectrum of compound 4 in CDCl₃ at 298K.



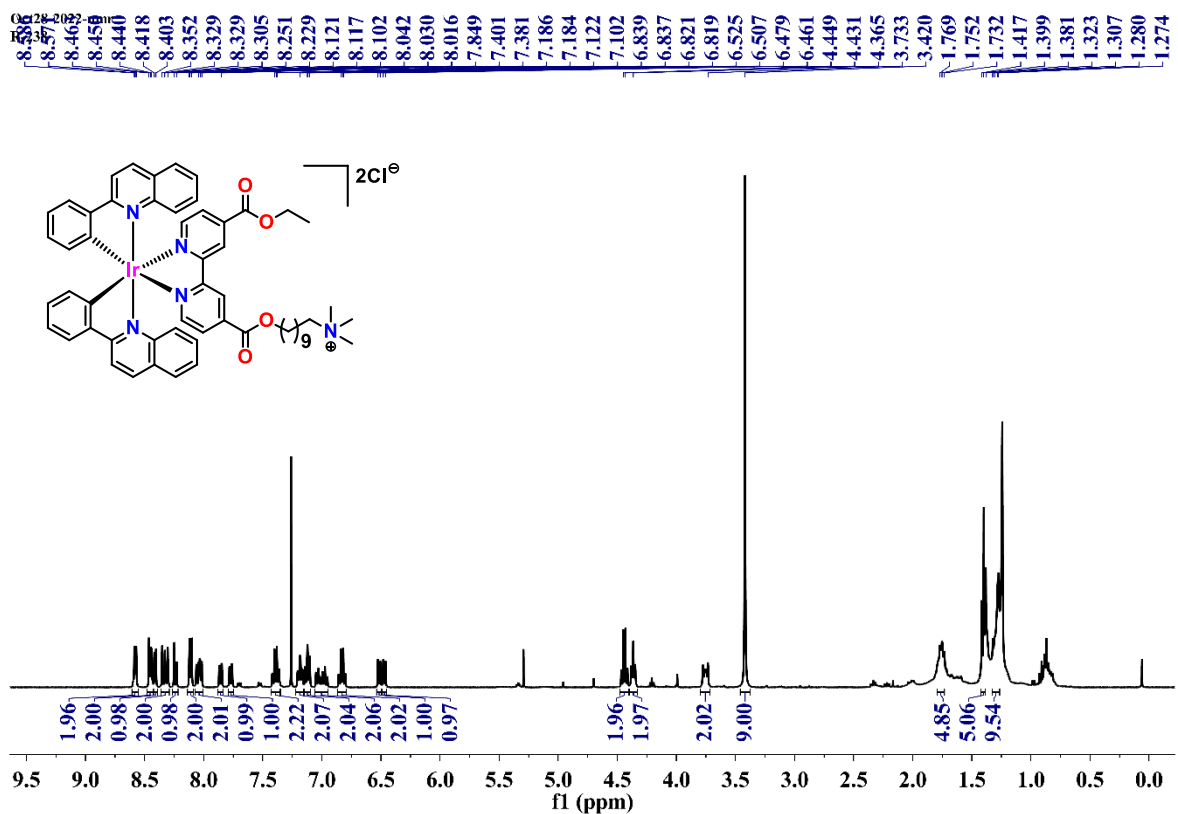


Figure S9 $^1\text{H-NMR}$ spectrum of Ir1 in CDCl_3 at 298K.

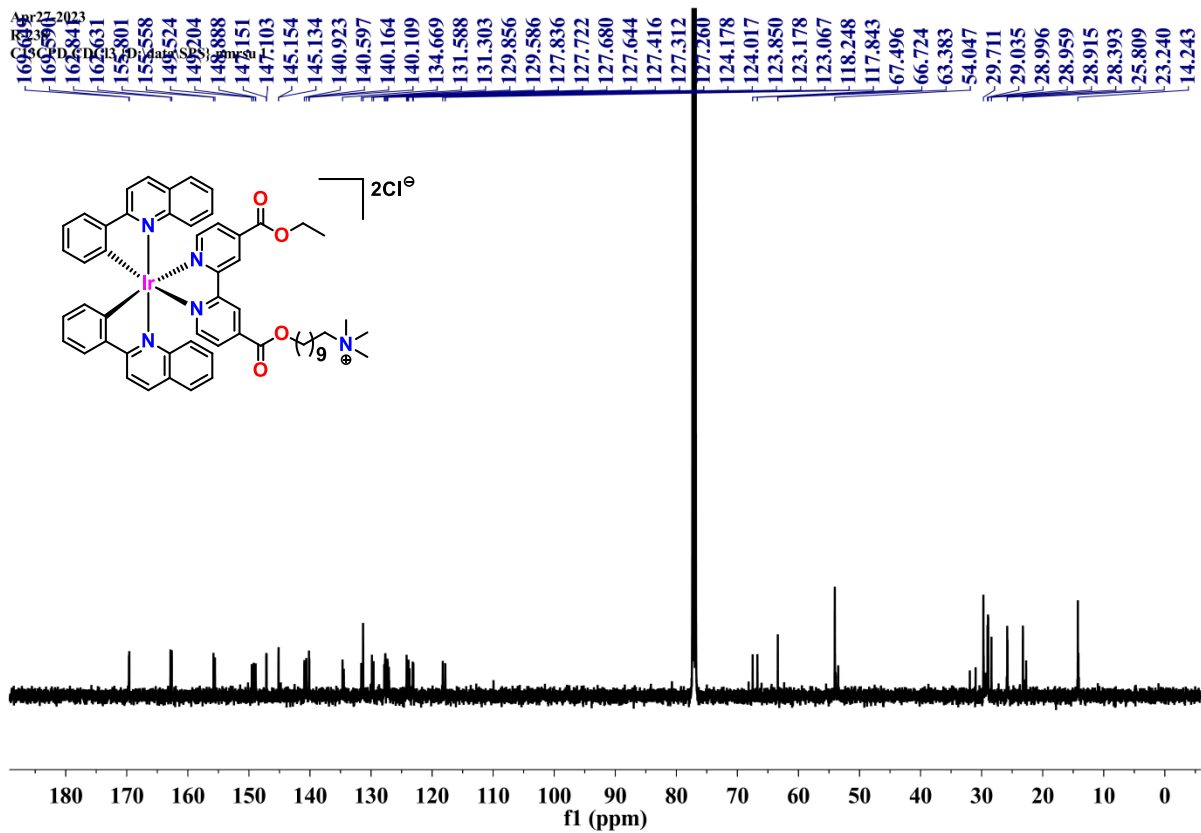


Figure S10 $^{13}\text{C-NMR}$ spectrum of Ir1 in CDCl_3 at 298K.

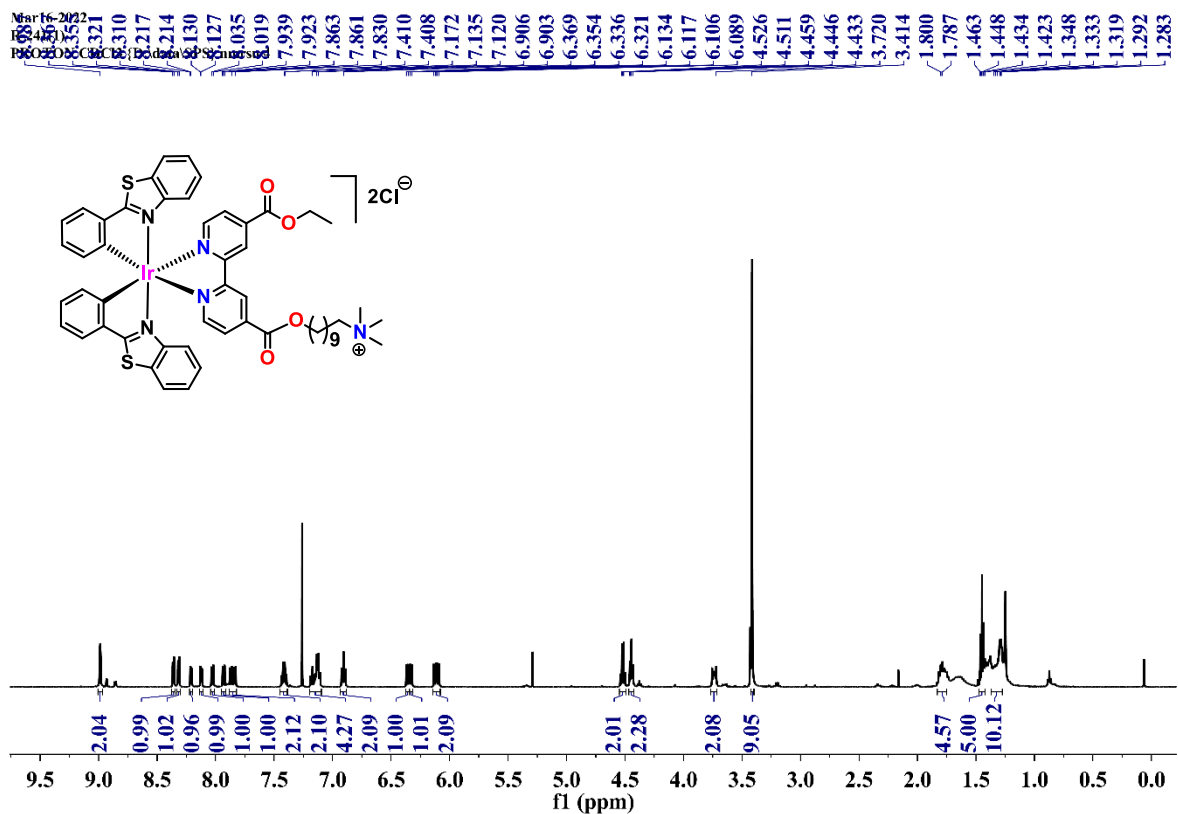


Figure S11 ¹H-NMR spectrum of Ir2 in CDCl₃ at 298K.

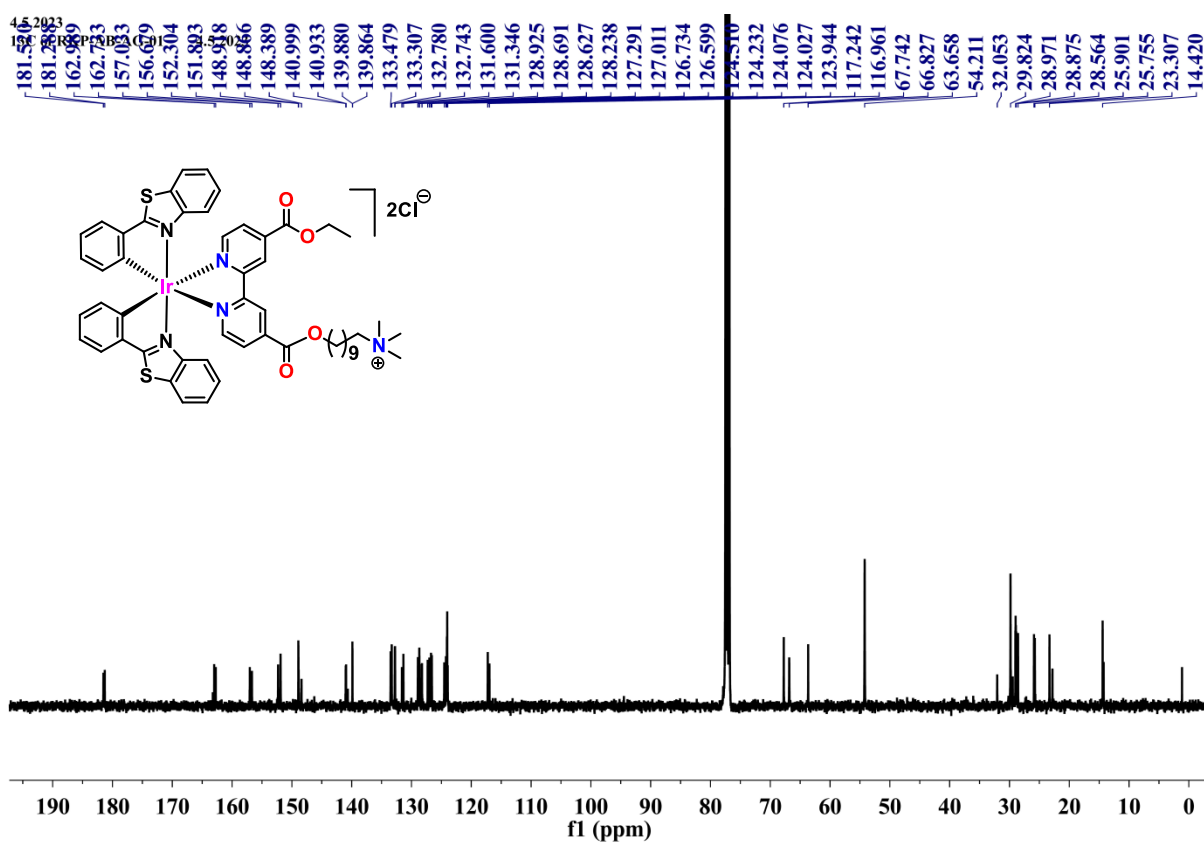


Figure S12 ¹³C-NMR spectrum of Ir2 in CDCl₃ at 298K.

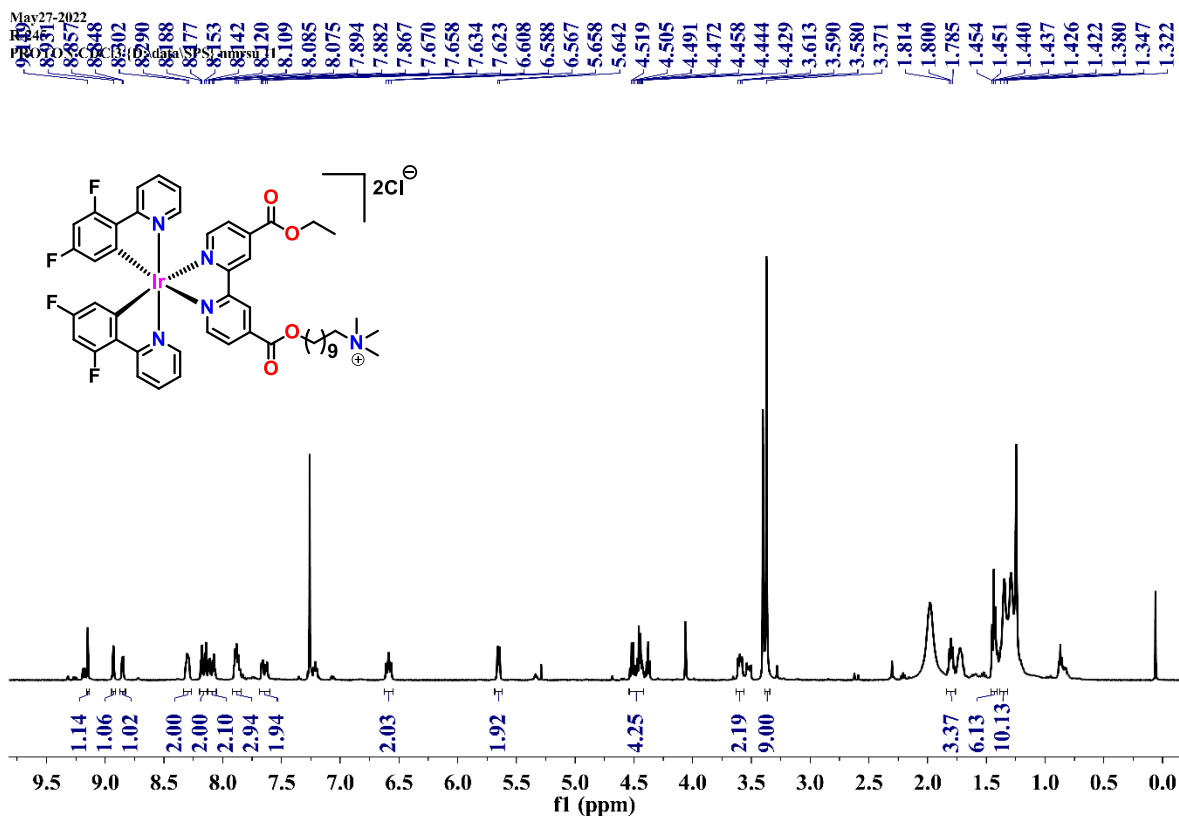


Figure S13 $^1\text{H-NMR}$ spectrum of **Ir3** in CDCl_3 at 298K.

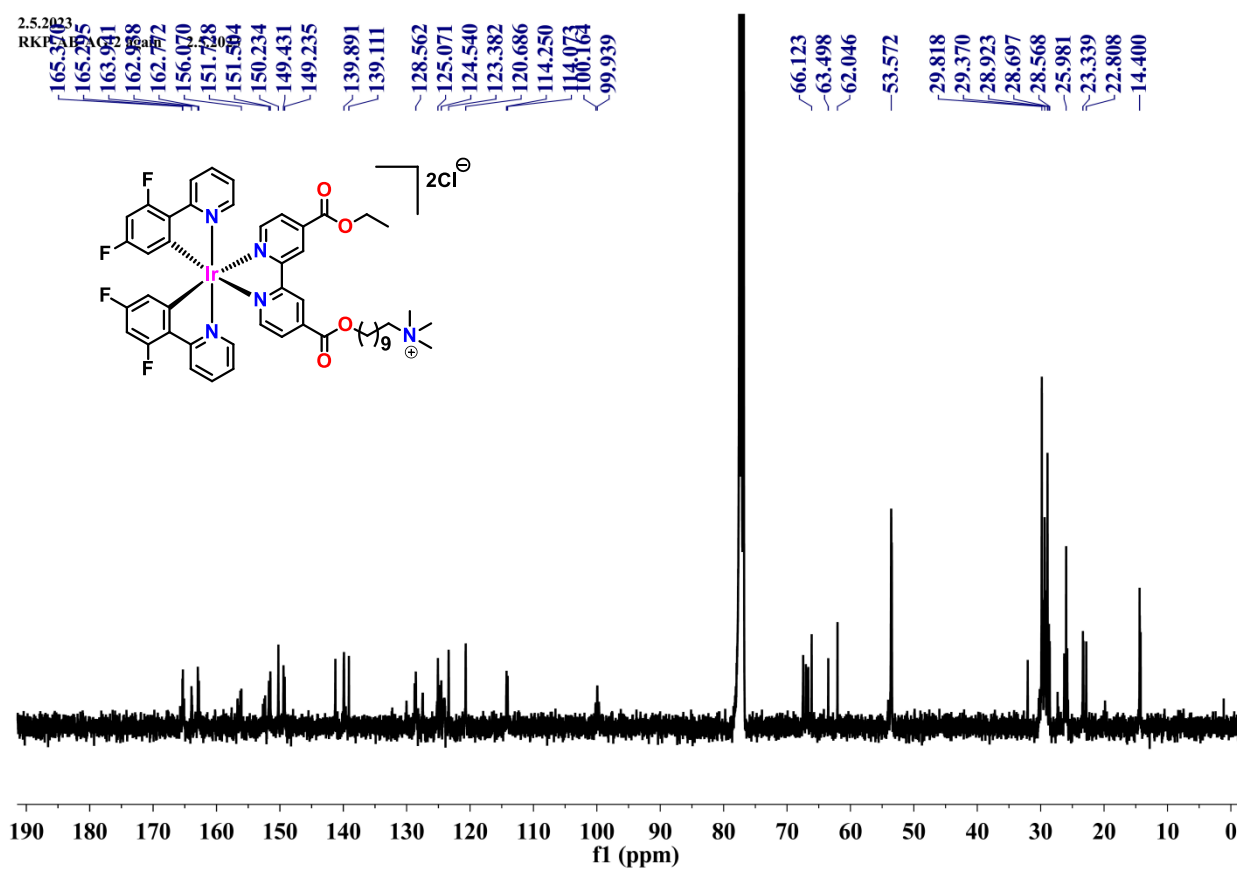


Figure S14 $^{13}\text{C-NMR}$ spectrum of **Ir3** in CDCl_3 at 298K.

Sample Name	JNU	Position	P1-A1	Instrument Name	Instrument 1	User Name	
Inj Vol	1	InjPosition		SampleType	Sample	IRM Calibration Status	Success
Data Filename	AG-123.d	ACQ Method	UNION POS.m	Comment		Acquired Time	20-12-2021 12:21:01

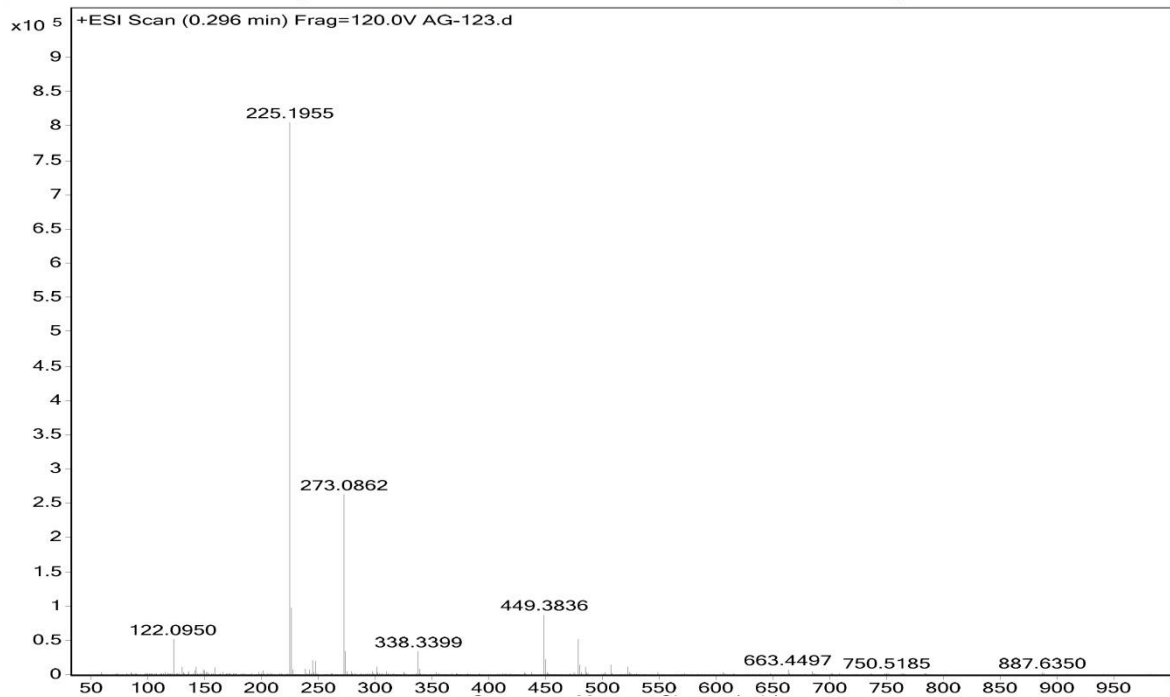


Figure S15 ESI-HRMS of compound **2** in DCM/MeOH showing the peak at 273.0862 (m/z) assignable to $[M+H]^+$ at 298K.

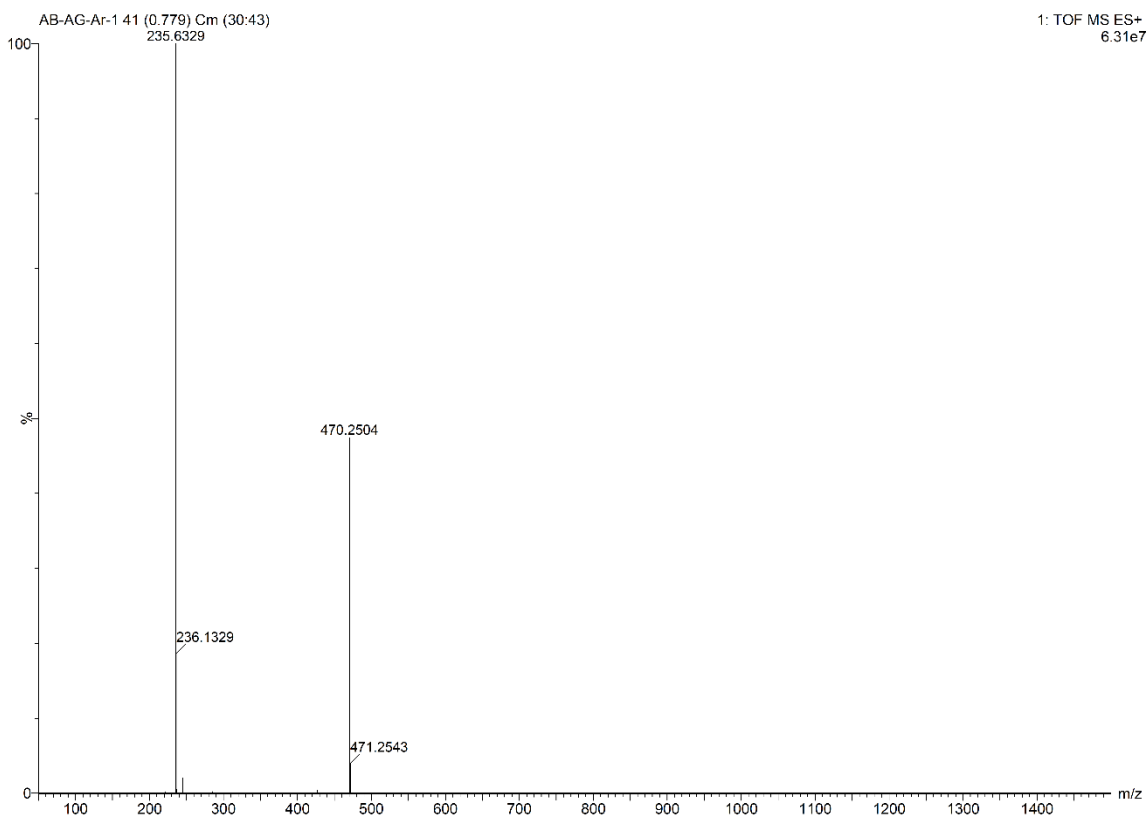


Figure S16 ESI-HRMS of compound **L** in DCM/MeOH showing the peak at 470.2504 (m/z) assignable to $[M]^+$ at 298K.

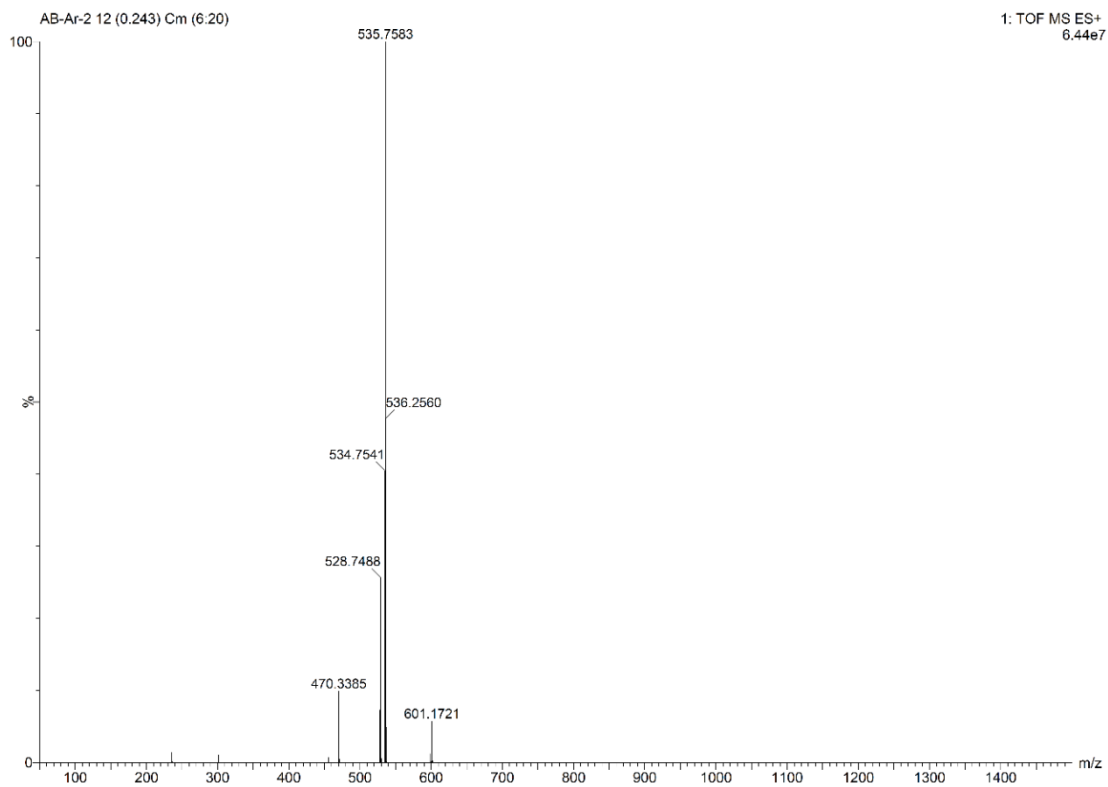


Figure S17 ESI-HRMS of **Ir1** in DCM/MeOH showing the peak at 535.7583 (m/z) assignable to $[M]^{2+}$ at 298K.

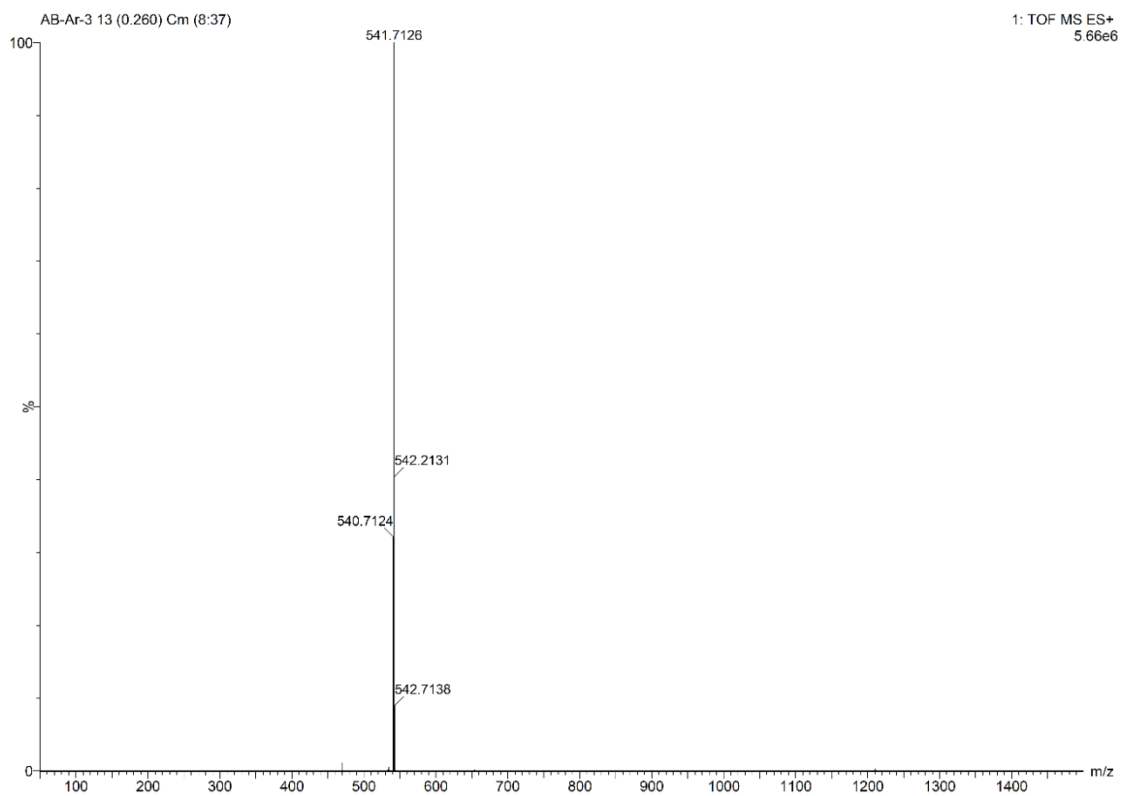


Figure S18 ESI-HRMS of **Ir2** in DCM/MeOH showing the peak at 541.7126 (m/z) assignable to $[M]^{2+}$ at 298K.

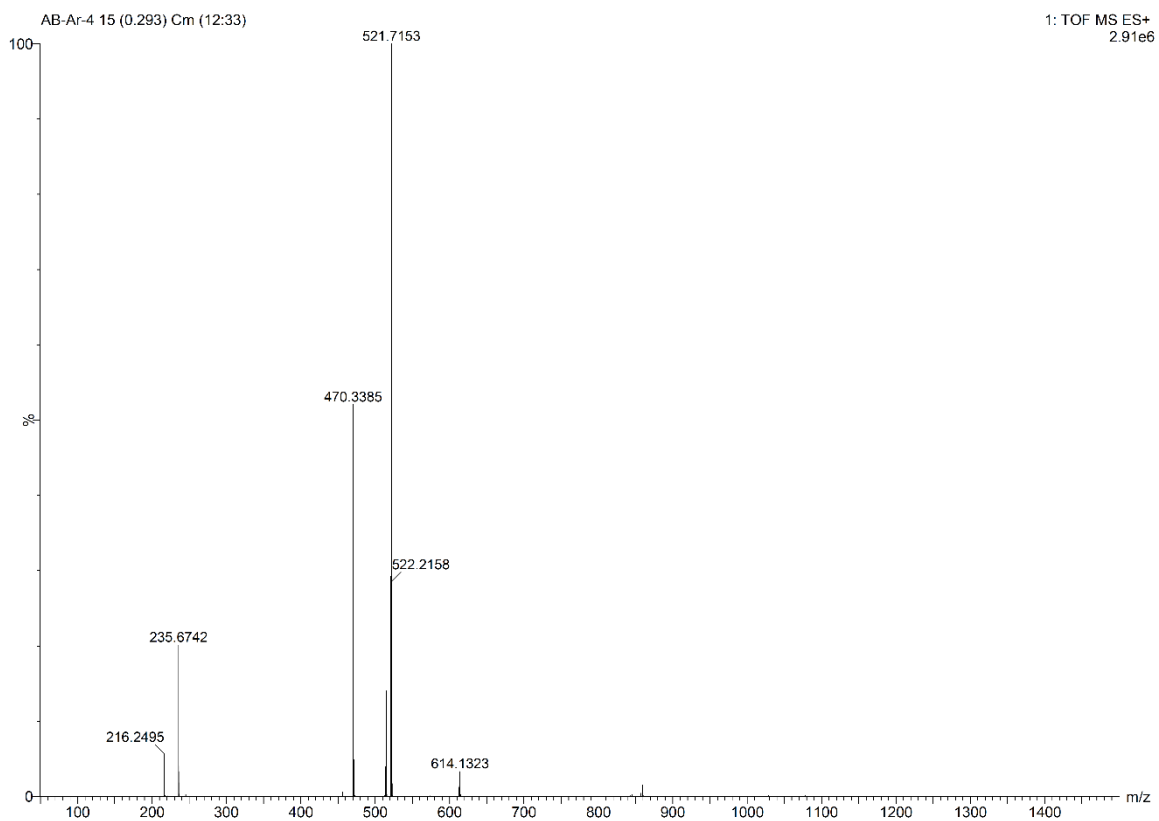


Figure S19 ESI-HRMS of **Ir3** in DCM/MeOH showing the peak at 521.7153 (m/z) assignable to $[M]^{2+}$ at 298K.

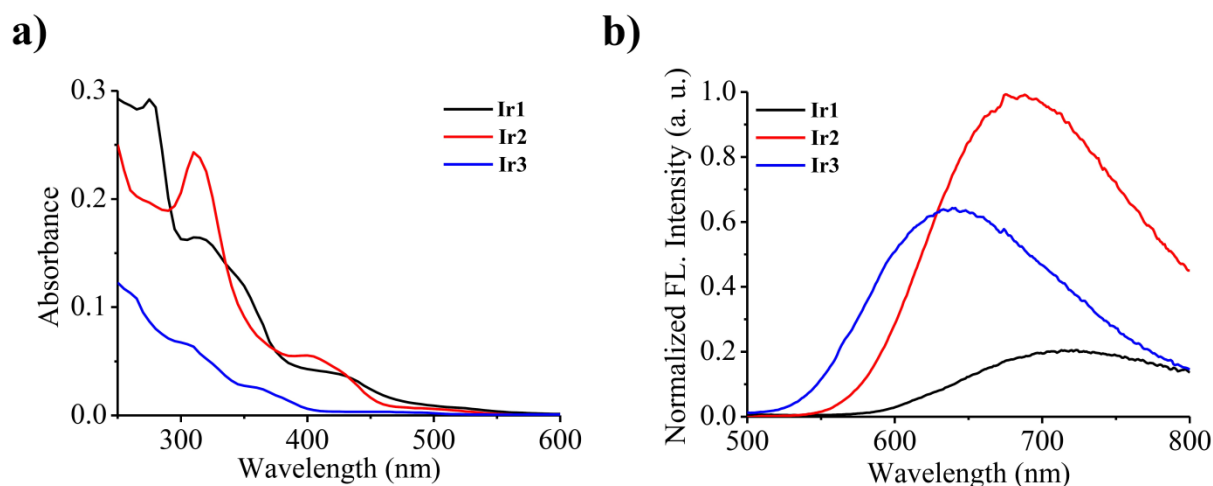


Figure S20 (a) Absorption (10 μM) and (b) normalized emission (50 μM) spectra of the complexes (**Ir1-Ir3**) were recorded in water containing 1% DMSO at RT. The emission spectra were measured upon excitation at 425 nm.

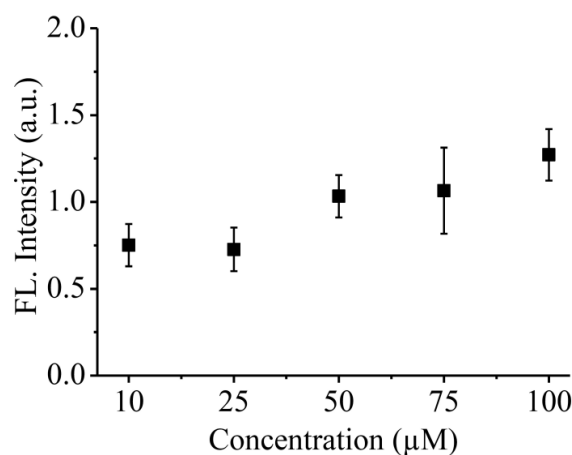


Figure S21 Emission intensity ($\lambda_{\text{ex}} = 425 \text{ nm}$, $\lambda_{\text{em}} = 720 \text{ nm}$) of complex **Ir1** as a function of concentration.

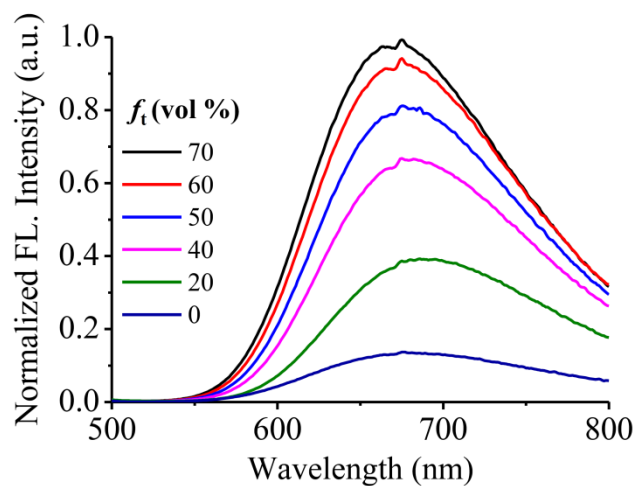


Figure S22 Emission spectra of **Ir2** (50 μM) recorded in a water-THF mixture with different THF fractions (f_t).

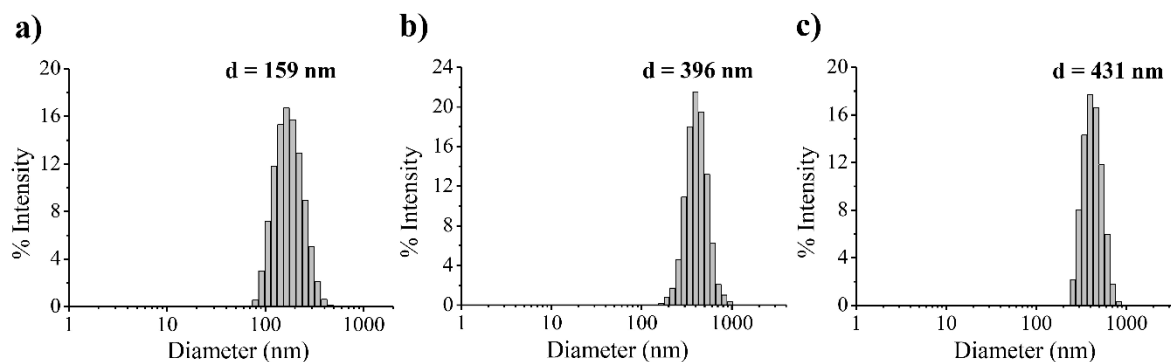


Figure S23 Hydrodynamic diameter (d) and particle size distribution of the complex **Ir1** ($50 \mu\text{M}$) in the absence (a) and in the presence of 25% THF (b) and 50% THF (c) in a water-THF mixture.

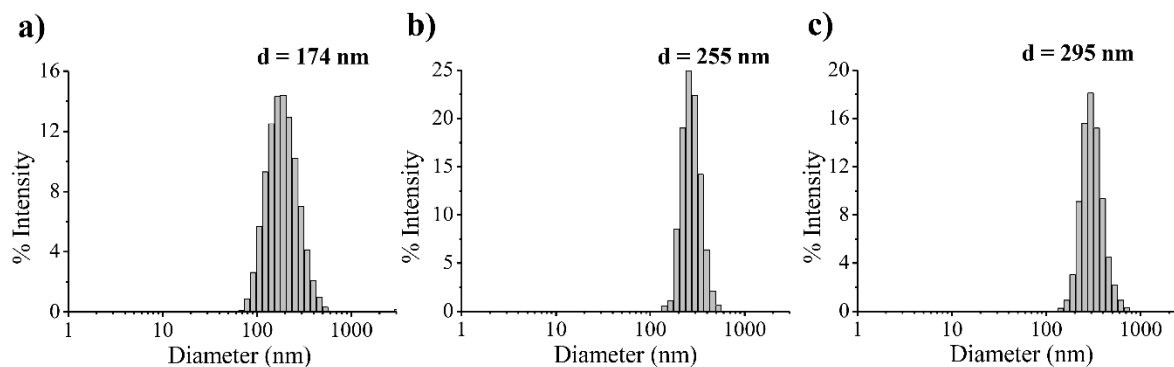


Figure S24 Hydrodynamic diameter (d) and particle size distribution of the complex **Ir2** ($50 \mu\text{M}$) in the absence (a) and in the presence of 25% THF (b) and 50% THF (c) in a water-THF mixture.

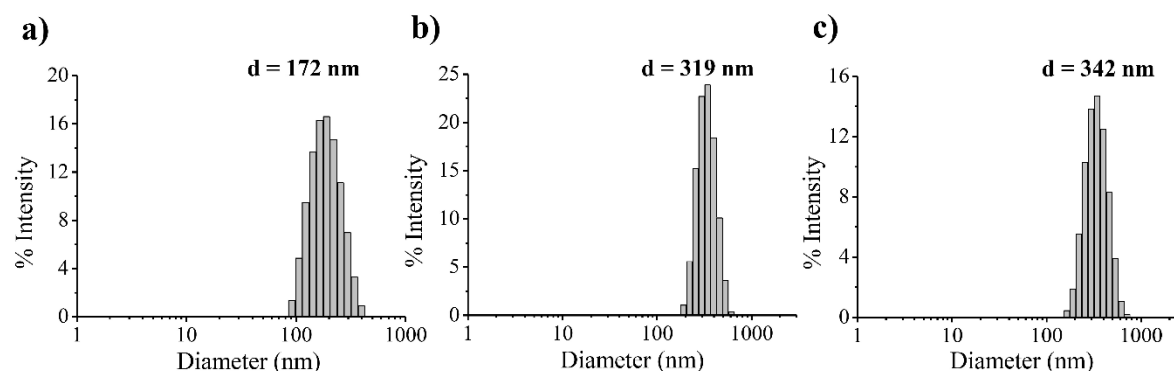


Figure S25 Hydrodynamic diameter (d) and particle size distribution of the complex **Ir3** ($50 \mu\text{M}$) in the absence (a) and in the presence of 25% THF (b) and 50% THF (c) in a water-THF mixture.

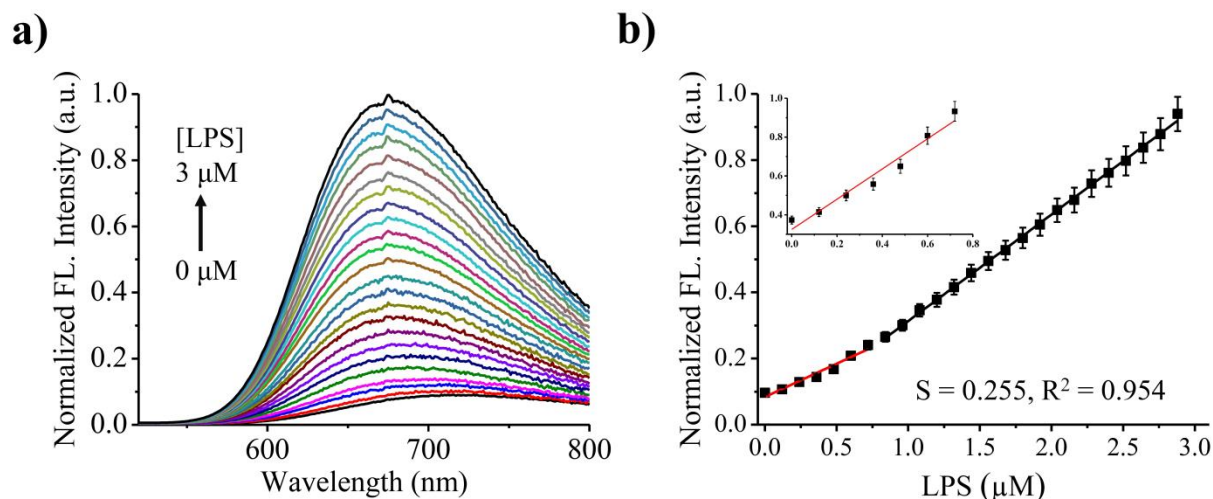


Figure S26 (a) The emission titration spectra of complex **Ir1** (50 μM) upon gradual addition of LPS (0-3 μM). (b) The plot of emission intensity of **Ir1** at 720 nm as a function of the concentration of LPS (0-3 μM). (Inset) The plot and linear fitting of the emission intensity of **Ir1** at 720 nm with different concentrations of LPS (0 to 0.84 μM).

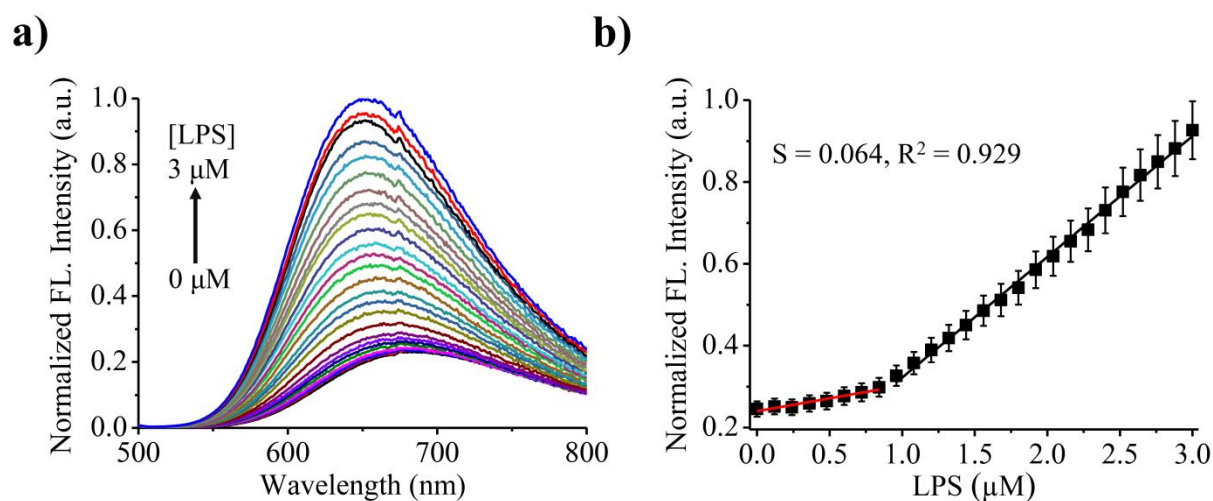


Figure S27 (a) The emission titration spectra of complex **Ir2** (50 μM) upon gradual addition of LPS (0-3 μM). (b) The plot of emission intensity of **Ir2** at 688 nm as a function of LPS showing a linear dynamic range between 0 to 0.84 μM .

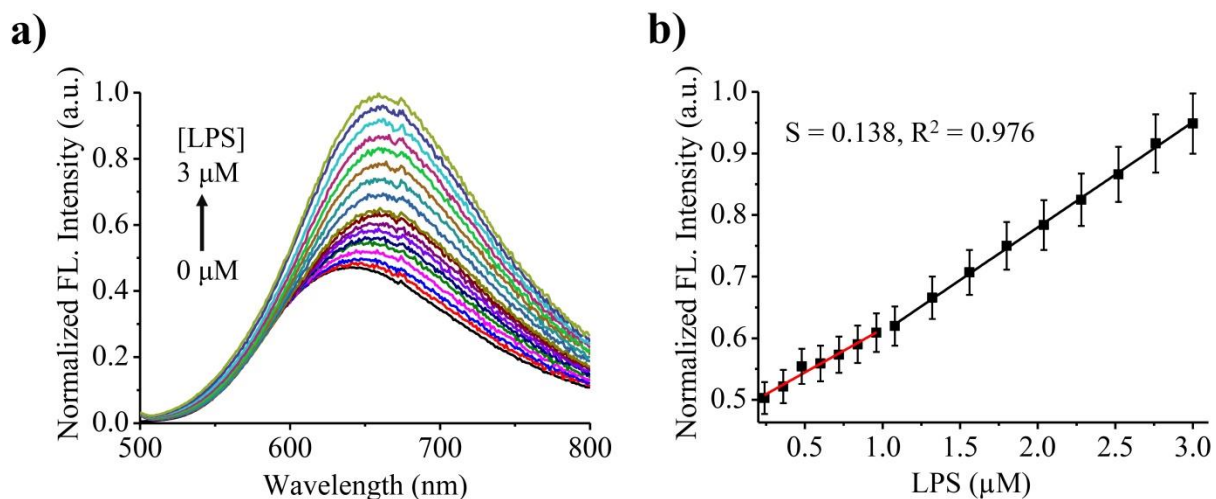


Figure S28 (a) The emission titration spectra of complex **Ir3** (50 μM) upon gradual addition of LPS (0-3 μM). (b) The plot of emission intensity of **Ir3** at 640 nm as a function of LPS showing a linear dynamic range between 0 to 0.96 μM .

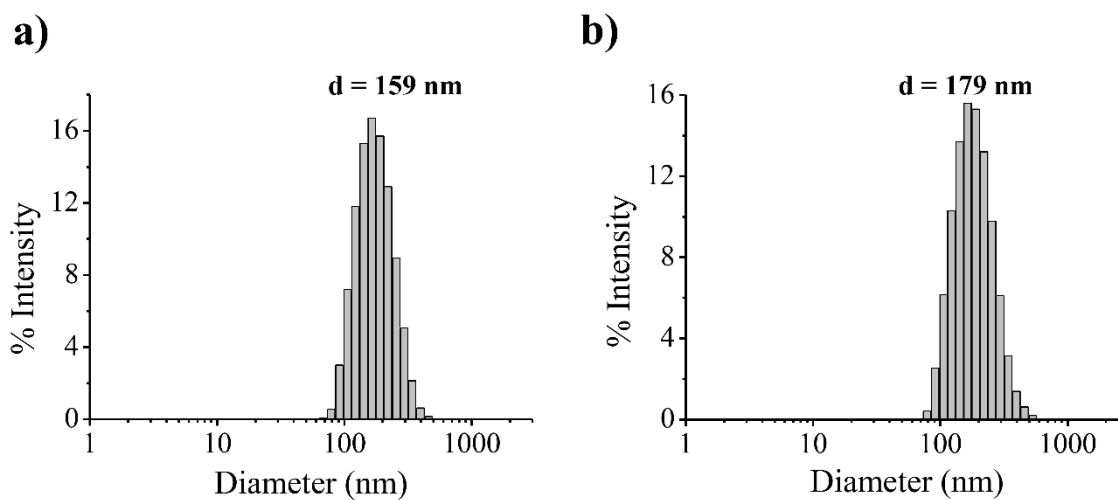


Figure S29 Hydrodynamic diameter (d) and particle size distribution of **Ir1** (50 μM) in aqueous media in the (a) absence and (b) presence of LPS (3 μM).

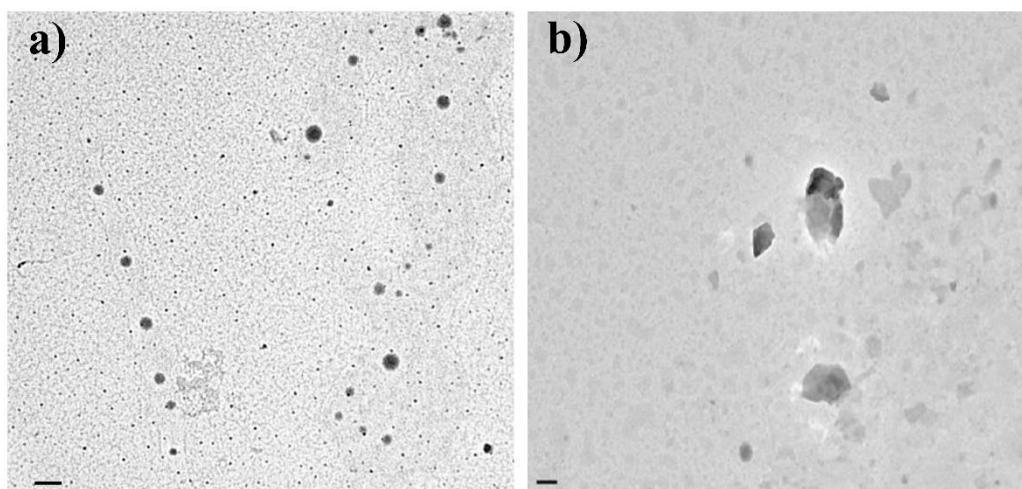


Figure S30 TEM images of complex **Ir1** (50 μM) in water containing 1% MeCN in the (a) absence and (b) presence of 3 μM LPS. Scale bar: 100 nm.

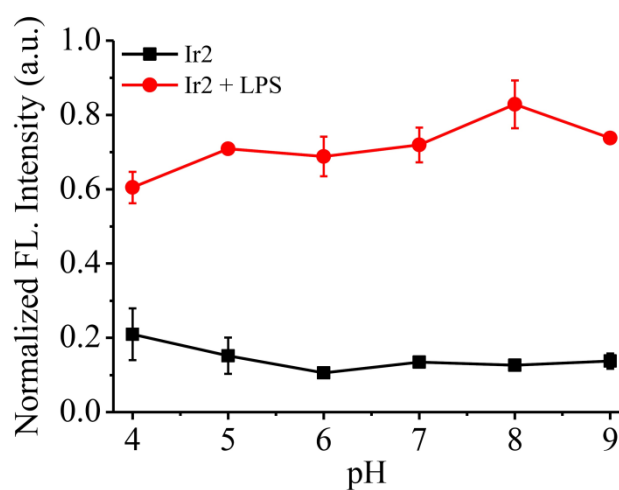


Figure S31 The emission intensity of **Ir2** (50 μM) at 688 nm in 10 mM HEPES buffer in the absence (■) and presence (●) of LPS (6 μM) in different pH solutions at room temperature.

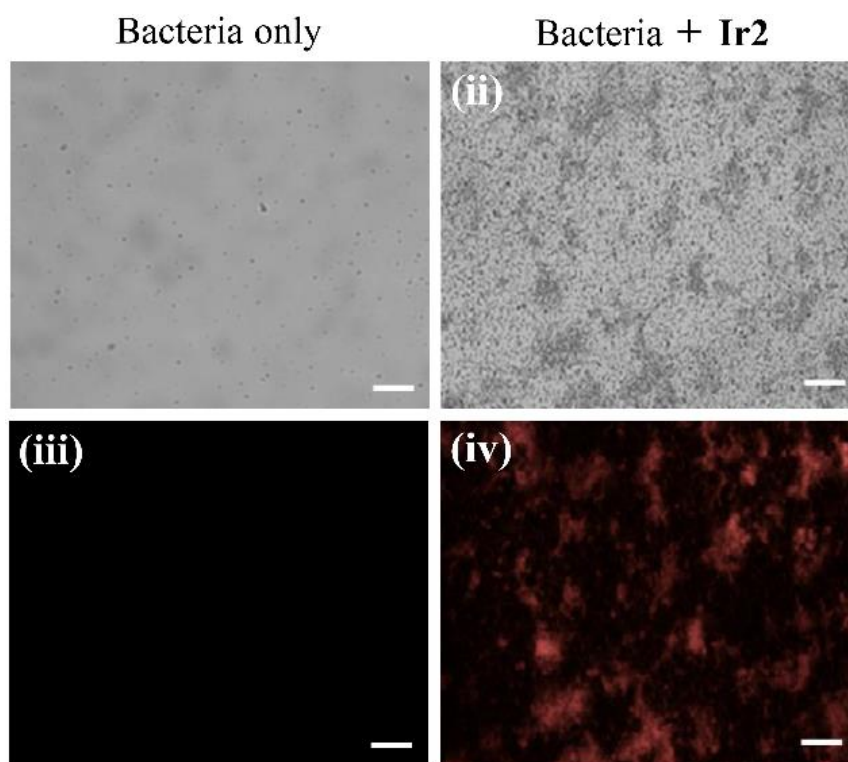


Figure S32 Optical microscopy images of *E. coli* (10^8 CFU/mL) after incubation with 400 μ M of complex **Ir2** and the results were observed in complementary bright-field (i,ii) and fluorescence (iii,iv) modes ($\lambda_{\text{ex}} = 450\text{-}480$ nm). Here, left panels (i,iii) indicate results for bacteria only and right panels (ii,iv) for bacteria treated with complex **Ir2**. Scale bar: 5 μ m.