## **Supporting Information**

## One-step synthesis of 2D@3D hollow prussian blue analogue as

## high-performance bifunctional electrochemical sensor

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## **Experimental Section**

Materials. Cobalt nitrate hexahydrate (Co(NO<sub>3</sub>)<sub>2</sub>·6H<sub>2</sub>O), Anhydrous dextrose, Disodium hydrogen phosphate dodecahydrate (Na<sub>2</sub>HPO<sub>4</sub>·12H<sub>2</sub>O), Potassium chloride (KCl), Magnesium sulfate heptahydrate (MgSO<sub>4</sub>·7H<sub>2</sub>O), Ammonium chloride (NH<sub>4</sub>Cl), Sodium sulfate (NaSO<sub>4</sub>), Calcium chloride (CaCl<sub>2</sub>), Sodium acetate anhydrous (NaAC), Aodium citrate (C<sub>6</sub>H<sub>5</sub>Na<sub>3</sub>O<sub>7</sub>·2H<sub>2</sub>O), Potassium ferricyanide  $(K_3[Fe(CN)_6])$  and Sodium chloride (NaCl) were purchased from Sinopharma chemical reagent Co., Ltd. Dopamine hydrochloride (DA) was supplied by Shanghai Macklin Biochemical Co., Ltd. Goat serum, Ascorbic acid (AA) and Uric acid (UA) were acquired from Shanghai yuanye Bio-Technology Co., Ltd. Sodium nitroprusside (Na<sub>2</sub>[Fe(CN)<sub>5</sub>(NO)]·2H<sub>2</sub>O) and L-(+)-Lactic acid (LA) were acquired from Rhawn. Sodium dihydrogen Sodium nitrite  $(NaNO_2)$ and phosphate dihydrate (NaH<sub>2</sub>PO<sub>4</sub>·12H<sub>2</sub>O) were obtained from Macklin. All chemicals in the experiment were directly used without any purification.



Fig. S1 (a) TEM image of HTPBA-12 and (b) corresponding SAED pattern.



**Fig. S2** (a) SEM image of HTPBA-12 and (b) corresponding EDS spectrum (inset: The obtained elemental ratio).



Fig. S3 (a) SEM image of HTPBA-12 and (b-f) corresponding elemental mapping.

Element	Wt%	Atomic%
Na	0.00513	0.000223
Co	9.38	0.159
Fe	9.31	0.167

**Table S1.** ICP-OES results of HTPBA-12.



Fig. S4 The EIS curves of HTPBA/NF and the equivalent circuit.



Fig. S5 (a) Dependence of amperometric response at HTPBA/NF-12 on applied potential; (b) Amperometric responses of five identical HTPBA/NF-12 electrodes to 50  $\mu$ M glucose in 0.1 M NaOH; Amperometric response of the HTPBA/NF-12 electrode (c) to 20  $\mu$ M glucose in 0.1 M NaOH for a long running time and (d) measured by injecting 30  $\mu$ M glucose into 0.1 M NaOH every 3 days over 16 days.

Electrode	Sensitivity, (µA mM <sup>-1</sup> cm <sup>-2</sup> )	Linear Range, (µM)	LOD, (µM)	Working potential, (V)	Electrolyte	Ref.
CoFePBA/FTO	18.69	100~8200	67	1.15 (vs Ag/AgCl)	0.1 M PBS	5
Au@NiFePBA/Nafion	8.037	10~16000	4.686	0.22 (vs Ag/AgCl)	0.1 M NaOH	21
NiFePBA/NF	21040, 6570	2~263.3,263.3~650	0.2	0.5 (vs SCE)	0.1 M NaOH	22
PB-RGO	27.78	$\sim\sim$	7.94	-0.05 (vs Ag/AgCl)	0.05 M PB	36
Fe-doped NiCo <sub>2</sub> O <sub>4</sub>	3055.7	0.2~3100	0.19	0.5 (vs SCE)	0.1 M NaOH	37
Co-Ni(Fe)-MOF/PPy	1805	2~3000	1.13	0.6 (vs Hg/HgO)	0.1 M NaOH	38
Ni-MOF@Ni-HHTP-5	2124.9	0.5~2665.5	0.02	0.6 (vs Ag/AgCl)	0.1 M NaOH	39
Cu@Ni CSNPs/CNCs/NF	6905	1~1630	0.03	0.65 (vs Hg/HgO)	0.1 M NaOH	40
HTPBA/NF-12	21410, 3749	2~450, 450~1250	0.089	0.5 (vs Ag/AgCl)	0.1 M NaOH	This work

 Table S2. Comparison of HTPBA/NF-12 with other related materials for glucose detection.



**Fig. S6** The (a, c) XRD and (b, d) FT-IR pattern of HTPBA/NF-12 after long-term stability and reusability test.



**Fig. S7** SEM image of HTPBA/NF-12 after (a) long-term stability and (b) reusability test; (c) TEM image and (d) SAED pattern of HTPBA/NF-12 after long-term stability.



**Fig. S8** (a) The XPS survey spectra of HTPBA/NF-12; High-resolution XPS spectra for the (b) Co 2p, (c) Fe 2p, (d) C 1s, (e) N 1s and (f) O 1s of HTPBA/NF-12.



**Fig. S9** (a) The XPS survey spectra of HTPBA/NF-12 after long-term stability test; High-resolution XPS spectra for the (b) Co 2p, (c) Fe 2p, (d) C 1s, (e) N 1s and (f) O 1s of HTPBA/NF-12 after long-term stability test.



Fig. S10 (a) Optimization of detection potential; The (b) reproducibility and (c) reusability test of HTPBA/NF-12 by repeatedly measure current response to 40  $\mu$ M NaNO<sub>2</sub>.

Electrode	Sensitivity, (µA mM <sup>-1</sup> cm <sup>-2</sup> )	Linear Range, (µM)	LOD, (µM)	Working potential, (V)	Electrolyte (pH)	Ref.
PANI-MnO <sub>2</sub> nanocomposite	225	19.98~732.17	1.08	0.85 (vs SCE)	0.1 M PBS (5.0)	55
NiCo <sub>2</sub> O <sub>4</sub> /GCE	1030	10~300	1.04	0.75 (vs Ag/AgCl)	0.1 M PBS (7.0)	59
AgNC@NCS	~~	1.12~1400	0.38	~~	0.1 M PB (5.2)	62
ZnLX <sub>2</sub> /SPCE	~~	2~500, 500~4838	0.78	0.85 (vs Ag/AgCl)	0.01 M PBS (4.4)	63
PPy/UiO-66/GCE	297.7	0.05~1055.5	0.037	1.05 (vs SCE)	0.1 M PBS (6.5)	64
LaAlO3@GO/GCE	1132, 1176	0.01~1540.5	0.0041	0.9 (vs Ag/AgCl)	0.1 M PB (7.0)	65
Ni/MoS <sub>2</sub> /GCE	72.47	5~800	2.48	~~	0.1 M PBS (4.0)	66
Near-spherical ZnO/GCE	785	0.6~220, 460~5500	0.39	1.05 (vs SCE)	0.1 M PBS (7.4)	67
GO-PANI-AuNPs/GCE	~~	0.5~240, 240~2580	0.17	0.9 (vs SCE)	0.1 M PBS (6.0)	68
HTPBA/NF-12	1248, 705.8	5~1280, 1280~3380	0.38	0.9 (vs Ag/AgCl)	0.1 M PB (7.5)	This work

Table S3. The comparison of sensing performance toward NaNO<sub>2</sub> between this work and other previous works.



Fig. S11 The SEM image of HTPBA/NF-12 after (a) long-term stability and (b) reusability test.



**Fig. S12** The (a) XRD and (b) FT-IR pattern of HTPBA/NF-12 after long-term stability and reusability test.

**Table S4.** The Comparison of sensing performances between HTPBA/NF-12 and cubic CoFePBA towards glucose and NaNO<sub>2</sub> detection.

Detection target	Electrode	Sensitivity, (µA mM <sup>-1</sup> cm <sup>-2</sup> )	Linear Range, (µM)	LOD, (µM)
glucose	Cubic CoFePBA/NF	2644, 1331	10~480, 480~1280	0.38
	HTPBA/NF-12	21410, 3749	2~450, 450~1250	0.089
NaNO <sub>2</sub>	Cubic CoFePBA/NF	458.4, 373.6	5~1530, 1530~3330	1.4
	HTPBA/NF-12	1248, 705.8	5~1280, 1280~3380	0.38



Fig. S13 (a-b) SEM image, (c) XRD and (d) FT-IR pattern of cubic CoFePBA/NF.



Fig. S14 (a-b) Potential optimization for cubic CoFePBA/NF towards glucose; (c-d)



**Fig. S15** (a) Chronoamperometric response of cubic CoFePBA/NF under different applied potential; (b) the plots of peak current density vs. NaNO<sub>2</sub> concentration; (c) Amperometric responses of cubic CoFePBA/NF injecting various concentration NaNO<sub>2</sub> in 0.1 M PB at 0.9 V (inset: the current response under a low NaNO<sub>2</sub> concentration); (d) Corresponding calibration curve.

Another CoFePBA material was synthesized based on similar synthetic process as HTPBA/NF-12, except that the same amount of potassium ferricyanide  $(K_3[Fe(CN)_6]$  was used instead of sodium nitroprusside  $(Na_4[Fe(CN)_5(NO)]$  in solution A. The reaction solution was maintained at 90 °C for 2 h. The obtained CoFePBA material was namely as cubic CoFePBA. SEM results as shown in Fig. S13a-b also demonstrate its cubic morphology with particle sizes of about 100-200 nm. Then, XRD and FT-IR as shown in Fig. S13c-d confirm its successful synthesis of CoFePBA material. As shown in Fig. S13c, all diffraction peaks in its XRD pattern match well with the characteristic peaks of  $K_2CoFe(CN)_6$  (JCPDS 75-0038). In FT-IR diagram, the peak at 2098 cm<sup>-1</sup> can be attributed to the stretching vibration of C $\equiv$ N, that is the characteristic peak of PBA material, suggesting the successful preparation of cubic CoFePBA on the surface of nickel foam.

As comparison, the sensing performances of cubic CoFePBA/NF towards glucose and nitrite were tested under the optimal test conditions. As shown in Fig. S14, its sensing sensitivity as glucose sensor was calculated to be 2644 and 1331  $\mu$ A mM<sup>-1</sup> cm<sup>-2</sup> in the linear intervals of 10~480 and 480~1280  $\mu$ M, respectively. The corresponding LOD was calculated to be 0.38  $\mu$ M. As nitrite sensor as shown Fig. S15, its sensitivity toward low and high concentration of NO<sub>2</sub><sup>-</sup> was calculated to be 458.4 (5~1530  $\mu$ M) and 373.6  $\mu$ A mM<sup>-1</sup> cm<sup>-2</sup> (1530~3330  $\mu$ M), respectively, and the LOD was 1.4  $\mu$ M.



Fig. S16  $N_2$  adsorption-desorption isotherm at 77 K of (a) HTPBA/NF-12 and (b) cubic CoFePBA/NF (inset: the pore size distribution).