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Electronic Supporting Information

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Electronically flexible PYA ligands for efficient palladium-catalyzed α-arylation of ketones

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S.1. Synthetic procedures

S.1.1 General information

Salts $[L_1H]I$, $[L_2H]I$, $[L_6H]I$ $[L_7H]I$ and the precursor of L_3 and L_4 , N-(3-methylpyridin-2yl)picolinamide, as well as complex 6 were prepared according to previously reported procedures.^{S1,S2} Acetonitrile and dichloromethane were dried by passage through a solvent purification column. All other reagents and solvents were commercially available and used as received. Unless otherwise stated all the reactions were run under exclusion of air using standard Schlenk techniques under an atmosphere of dry nitrogen. Benzotriazol-1-yloxytripyrrolidinophosphonium hexafluorophosphate (PyBOP) and all other chemicals were purchased from commercial vendors and used as received.

Unless otherwise specified, NMR spectra were recorded at 25 °C on Bruker spectrometers operating at 300 or 400 MHz (¹H NMR) and 75 or 100 MHz (¹³C {¹H} NMR), respectively. Chemical shifts (δ) are expressed in ppm downfield from SiMe₄ using the residual protonated solvent as an internal standard and coupling constants in Hz. Assignments are based on homo- and heteronuclear shift correlation spectroscopy.

High resolution mass spectrometry was carried out with a Thermo Scientific LTQ Orbitrap XL (ESI-TOF) or with a Thermo Scientific Q Exactive GC either directly used with direct injection probe (DIP) or coupled to a Trace 1310 Gas Chromatograph when electron impact ionization (EI-MS) was required to obtain satisfactory data. All the MS data were collected by the mass spectrometry group in the DCBP. Elemental analyses were performed on a Thermo Scientific Flash 2000 CHNS-O elemental analyzer by the microanalytic laboratory at the DCBP. GC-FID reaction follow-up was achieved using an Agilent technologies 7820A GC System. GC-MS reaction follow-up was carried out on a Perkin Elmer Clarus 500 GC coupled to Perkin Elmer Clarus 560S ESI mass spectrometer.

S.1.2 Synthesis of the pyridyl-pyridinium salt [L5H]I

Scheme S1. Synthetic pathway to ligand precursor **[L5H]I**

prec1. To a mixture of ethyl 6-bromopicolinate (200 mg, 0.87 mmol), (2,6 dimethylphenyl)boronic acid (261 mg, 1.74 mmol), Cs_2CO_3 (1.13 g, 3.48 mmol) and Pd(PPh₃)₄ (50.2 mg, 43.5 µmol) was added dry 1,4-dioxane (10 mL) and the resulting mixture was stirred for 16 h at 150 °C in a pressure tube. The resulting mixture was concentrated and purified by gradient flash chromatography (0 to 15%

EtOAc in n-hexane; SiO₂) to yield the pure title compound as a white solid (124 mg, 62%). ¹H NMR $(300 \text{ MHz}, \text{DMSO-}d_6, 298 \text{ K}): \delta = 8.14 - 8.00 \text{ (m, 2H, } CH_{\text{pyr}}), 7.58 \text{ (dd, } J_{HH} = 7.1, 1.7 \text{ Hz, } 1H, CH_{\text{pyr}}),$ 7.24 (t, ${}^{3}J_{HH}$ = 7.5 Hz, 1H, CH_{xyl}), 7.14 (d, ${}^{3}J_{HH}$ = 7.5, 2H, CH_{xyl}, 4.35 (q, ${}^{3}J_{HH}$ = 7.1, 2H, CH₂), 1.95 (s, 6H, xyl-CH₃), 1.32 (t, ³J_{HH} = 7.1, 3H, OCH₂CH₃) ppm. ¹³C{¹H} NMR (75 MHz, DMSO-d₆, 298 K): δ = 165.29 (*C*O), 159.58 (*C*pyr-xyl), 148.16 (*C*pyr-CO), 140.11 (*C*xyl-pyr), 138.50 (*C*Hpyr), 135.66 (2C*, C*_{xyl}CH₃), 128.48 (2C, *C*H_{xyl} + *C*H_{pyr}), 127.94 (2C, *C*H_{xyl}) 123.56 (*C*H_{pyr}), 61.71 (*C*H₂), 20.37 (2C, xyl-*C*H₃), 14.62 (CH₂*C*H₃) ppm. HR-MS (m/z): calculated for C₁₆H₁₈NO₂ [M+H]⁺ = 256.1338; found: 256.1337.

Figure S1. ¹ H NMR spectrum (DMSO-*d6*, 298 K, 300 MHz) of **prec1**.

Figure S2. ¹³C $\{^1H\}$ NMR spectrum (DMSO- d_6 , 298 K, 75 MHz) of prec1.

prec2. To a solution of ethyl 6-(2,6-dimethylphenyl)picolinate (200 mg, 0.78 mmol) in MeOH (8 mL), EtOH (4 mL) and H₂O (4 mL) was added lithium hydroxide hydrate (66 mg, 1.56 mmol) and the resulting mixture was stirred for 1 h at 23°C. Conversion was checked by TLC and, aq. HCl (1M, 2.4 mL, 2.4 mmol) was directly added. The reaction mixture was stirred for 10 min at the same temperature and then extracted with CH_2Cl_2

 $(3 \times 20 \text{ mL})$. The combined organic layers were dried over MgSO₄ and concentrated under vacuum to yield the title compound as an off-white solid (178 mg, quant. yield). ¹H NMR (300 MHz, DMSO- d_6 , 298 K): $\delta = 8.13 - 7.99$ (m, 2H, CH_{pyr}), 7.56 (dd, $J_{HH} = 7.1$, 1.7 Hz, 1H, CH_{pyr}), 7.23 (t, ${}^{3}J_{HH} = 7.5$ Hz, 1H, CH_{xyl}), 7.14 (d, ${}^{3}J_{HH}$ = 7.5, 2H, CH_{xyl}), 1.96 (s, 6H, xyl-CH₃) ppm. ¹³C{¹H} NMR (75 MHz, DMSO*d6*, 298 K): δ = 166.27 (*C*O), 158.85 (*C*pyr-xyl), 148.30 (*C*pyr-CO), 139.70 (*C*xyl-pyr), 137.98 (*C*Hpyr), 135.23 (2C*, C*xylCH3), 127.96 (*C*Hxyl), 127.75 (*C*Hpyr) 127.43 (2C, *C*Hxyl), 122.91 (*C*Hpyr), 19.90 (2C, xyl-*C*H₃) ppm. HR-MS (m/z): calculated for $C_{14}H_{14}NO_2$ [M+H]⁺ = 228.1025; found: 228.1025.

Figure S3. ¹ H NMR spectrum (DMSO-*d6*, 298 K, 300 MHz) of **prec2**.

Figure S4. ¹³C $\{^1H\}$ NMR spectrum (DMSO- d_6 , 298 K, 75 MHz) of **prec2**.

prec3. To a solution of 6-(2,6-dimethylphenyl)picolinic acid (169 mg, 0.744 mmol), PyBOP (464 mg, 0.892 mmol) and 3-methylpyridin-2-amine (83 µL, 0.818 mmol) in THF (6 mL) was added iPr₂EtN (390 μ L, 2.23 mmol) and the resulting solution was stirred for 16 h at 50 °C. The reaction mixture was then concentrated under vacuum, dissolved in CH_2Cl_2 (15 mL), washed with brine (15 mL),

evaporated and purified by FC (5 to 50% EtOAc/n-hexane; $rf = 0.6$ in 50% EtOAc/n-hexane) to yield the pure title compound as an off-white solid (189 mg, 80%). ¹H NMR (400 MHz, DMSO- d_6 , 298 K): δ = 10.39 (s, 1H, NH), 8.28 (dd, *J_{HH}* = 4.8, 1.8 Hz, 1H, C*H*_{PYA}), 8.19 – 8.09 (m, 2H, C*H*_{pyr}), 7.71 (dd, *JHH* = 7.1, 1.8 Hz, 1H, C*H*PYA), 7.61 (dd, *JHH* = 6.3, 2.6 Hz, 1H, C*H*pyr), 7.26 – 7.21 (m, 2H, C*H*xyl +CH_{PYA}), 7.16 (d, ${}^{3}J_{HH}$ = 7.5 Hz, 2H, CH_{xyl}), 2.23 (s, 3H, PYA-CCH₃), 2.05 (s, 6H, xyl-CH₃) ppm. ¹³C{¹H} NMR (101 MHz, DMSO-*d₆*, 298 K): δ = 162.64 (*C*O), 158.04 (*C*_{pyr}-xyl), 149.59 (*C*_{pyr}-CO or *C*PYA), 149.41 (*C*pyr-CO or *C*PYA), 145.97 (*C*Hpyr), 139.50 (*C*xyl-pyr), 139.38 (*C*HPYA), 138.60 (*C*Hpyr), 135.49 (2C, C_{xyl}CH₃), 129.18 (PYACCH₃)128.11 (CH_{xyl} or CH_{pyr}), 127.97 (CH_{xyl} or CH_{pyr}), 127.59 (2C, *CH_{xyl}*), 122.21 (*CH_{PYA}*), 120.64 (*CH_{pyr}*), 20.09 (PYAC*CH*₃), 17.59 (2C, xyl-*CH*₃) ppm. HR-MS (m/z): calculated for $C_{20}H_{19}N_3NaO$ $[M+Na]^+=340.1420$; found: 340.1420.

Figure S5. ¹H NMR spectrum (DMSO- d_6 , 298 K, 400 MHz) of prec3.

Figure S6. ¹³C $\{^1H\}$ NMR spectrum (DMSO- d_6 , 298 K, 101 MHz) of prec3.

[L5H]I. To a solution of 6-(2,6-dimethylphenyl)-N-(3-methylpyridin-2 yl)picolinamide (120 mg, 378 µmol) in MeCN (5 mL) was added methyl iodide (47 µL, 756 µmol) and the resulting mixture was stirred for 24 h at 60 °C. The crude mixture was concentrated to 1 mL MeCN then precipitated with Et₂O, filtered, washed with the same solvent and dried under vacuum to yield **[L5H]**I

as a pale-yellow solid (150 mg, 86%). ¹H NMR (400 MHz, CD₃CN, 298 K): δ = 10.48 (br s, 1H, NH), 8.62 (d, ${}^{3}J_{HH} = 6.1$ Hz, 1H, CH_{PYA}), 8.44 (d, ${}^{3}J_{HH} = 8.0$ Hz, 1H, CH_{PYA}), 8.21 (dd, $J_{HH} = 7.8$, 1.0 Hz, 1H, CH_{pyr}), 8.15 (t, ³ J_{HH} = 7.8 Hz, 1H, CH_{pyr}), 7.86 (dd, ³ J_{HH} = 8.0, 6.1 Hz, 1H, CH_{PYA}), 7.65 (dd, J_{HH} $= 7.8, 1.0$ Hz, 1H, CH_{pyr}), 7.27 (dd, $^3J_{HH} = 8.6, 6.5$ Hz, 1H, CH_{xyl}), 7.19 (d, $^3J_{HH} = 7.5$ Hz, 2H, CH_{xyl}), 4.20 (s, 3H NC*H*3), 2.43 (s, 3H, pyrC*H*3), 2.11 (s, 6H, xyl-C*H*3) ppm. 13C{1 H} NMR (101 MHz, CD3CN, 298 K): δ = 164.02 (*C*O), 159.58 (*C*pyr-xyl), 148.93 (*C*Hpyr),), 147.77 (*C*pyr-CO), 146.86 (*C*PYA), 144.96 (*C*HPYA), 139.88 (*C*xyl-pyr), 139.30 (*C*Hpyr), 138.72 (*C*pyr-Me), 136.58 (2C*, C*xylCH3), 130.03 (*C*Hpyr), 129.05 (CH_{xyl}), 128.31 (2C, CH_{xyl}), 126.18 (CH_{PYA}), 121.61 (CH_{pyr}), 45.97 (NCH₃), 20.10 (2C, xyl-*C*H₃), 17.77 (PYA-C*C*H₃) ppm. HR-MS (m/z): calculated for C₂₁H₂₂N₃O [M-I]⁺ = 332.1757; found: 332.1754.

Figure S7. ¹ H NMR spectrum (CD3CN, 298 K, 400 MHz) of **[L5H]I**.

Figure S8. ¹³C $\{^1H\}$ NMR spectrum (CD₃CN, 298 K, 101 MHz) of [L₅H]I.

S.1.3 General procedure for the preparation of neutral pyridylidene amides L1- L5, L8

Scheme S2. Preparation of the neutral PYA ligand **L1**

The pyridinium salt (2 mmol) was dissolved in aq. KOH (2 M, 10 mL, 20 mmol) and the resulting solution was briefly sonicated and stirred at 23 °C for 10 min. The resulting slurry was extracted with CH_2Cl_2 (5 x 15 mL), the combined organic fractions were dried over MgSO₄, filtered and concentrated under vacuo to yield the pure pyridylidene amide (PYA) ligand.

S.1.3.1 Synthesis of neutral PYA ligand L1

Following the general procedure, reaction of $[L_1H]I$ (750 mg, 2.1 mmol) yielded compound L_1 as a pale-yellow solid (432 mg, 90%). Crystals suitable for Xray diffraction were grown by slow diffusion of Et₂O in a CH₂Cl₂ solution. ¹H NMR (300 MHz, CD₂Cl₂, 298 K): δ = 8.61 (d, ³J_{HH} = 4.6 Hz, 1H, CH_{pyr}), 8.18 (d, ${}^{3}J_{HH}$ = 7.4 Hz, 1H, CH_{pyr}), 7.76 (t, ${}^{3}J_{HH}$ = 7.4 Hz, 1H, CH_{pyr}), 7.66 (d, ${}^{3}J_{HH}$ = 6.6 Hz, 1H, CH_{PYA}), 7.57 (d, ${}^{3}J_{HH}$ = 7.2 Hz, 1H, CH_{PYA)}, 7.31 (dd, ${}^{3}J_{HH}$ = 7.4, 4.6 Hz, 1H, CH_{pyr}), 6.73 (dd, ${}^{3}J_{HH}$ = 7.2, 6.7 Hz, 1H, C*H*PYA), 3.86 (s, 3H, PYA-NC*H*3), 2.20 (s, 3H, PYA-CC*H*3) ppm. 13C{1 H} NMR (75 MHz, CD₂Cl₂, 298 K): δ = 166.73 (*C*O), 161.26 (*C*_{PYA}), 156.28 (C_{pyr}), 149.06 (*C*H_{pyr}), 140.49 (*C*H_{PYA}), 137.85 (*C*HPYA), 136.47 (*C*Hpyr), 132.51 (*C*PYA–Me), 124.63 (*C*Hpyr) 124.13 (*C*Hpyr), 113.27 (*C*HPYA), 42.83 (PYA-NCH₃), 19.34 (PYA-CCH₃) ppm. HR-MS (m/z): calculated for C₁₃H₁₄N₃O [M+H]⁺ = 228.1137; found: 228.1126. Note: The PF_6 analog $[L_1H]PF_6$ was prepared by anion metathesis according to a previously reported procedure.^{S1} All analytical data in accordance with reported values.

Figure S9. ¹ H NMR spectrum (CD2Cl2, 298 K, 300 MHz) of **L1**.

Figure S10. ¹³C $\{^1H\}$ NMR spectrum (CD₂Cl₂, 298 K, 75 MHz) of L₁.

S.1.3.2 Synthesis of neutral PYA ligand L2

Following the general procedure, reaction of **[L2H]I** (750 mg, 2.2 mmol) yielded compound \mathbf{L}_2 as a pale red solid that crystallized under vacuo (412 mg, 88%). ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ = 8.66 (d, *J*_{HH} = 4.6 Hz, 1H, CH_{pyr}), 8.32 – 8.27 (m, 2H, CH_{PYA} + CH_{pyr}), 7.76 (td, *J*_{HH} = 7.7, 1.8 Hz, 1H, CH_{pyr}), 7.67 (dd, *J*_{HH} = 6.7, 1.9 Hz, 1H, C*H*PYA), 7.60 (ddd, *JHH* = 9,0, 6.7, 1.9 Hz, 1H, C*H*PYA), 7.31 (ddd, *JHH* = 7.7, 4.6, 1.3 Hz, 1H, C*H*PYA), 6.59 (td, J_{HH} = 6.7, 1.5 Hz, 1H, CH_{PYA}), 3.87 (s, 3H, CH₃) ppm. ¹³C{¹H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 172.46 (*C*O), 160.08 (*C*PYA), 156.67 (*C*pyr), 149.38 (*C*Hpyr), 140.03 (*C*HPYA), 137.84 (*C*HPYA), 136.49 (*C*Hpyr), 129.94 (*C*Hpyr), 124.62 (*C*Hpyr), 121.20 (*C*HPYA), 111.78 (*C*HPYA), 41.92 (N*C*H3) ppm. HR-MS (m/z): calculated for $C_{12}H_{12}N_3O$ $[M+H]^+$ = 214.0980; found: 214.0969.

Figure S11. ¹H NMR spectrum (CD₂Cl₂, 298 K, 400 MHz) of **L**₂.

Figure S12. ¹³C $\{^1H\}$ NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of **L**₂.

S.1.3.3 Synthesis of neutral PYA ligand L3

To a solution of N-(3-methylpyridin-2-yl)picolinamide (1.04g, 4.88 mmol) in dry MeCN (16 mL) in a pressure tube was added nBuI (2.8 mL, 24.4 mmol). The resulting mixture was stirred 24 h at 120 °C. Solvent amount was then reduced to 5 mL and $Et₂O$ was added. The suspension was filtered and dried. The crude yellow

solid was dissolved in a KOH solution (2 M, 25.0 mL, 50.0 mmol). The resulting slurry was stirred 5 min at 23 °C and then extracted with CH_2Cl_2 (3 x 20 mL). The combined organic fractions were concentrated under vacuum and purified by gradient flash chromatography (0 to 10% MeOH in CH_2Cl_2) on Al₂O₃; rf 0.7 in 10% MeOH/ CH₂Cl₂) to yield **L**₃ as a pale brown waxy solid (1.28 g, 66%) ¹H NMR $(400 \text{ MHz}, \text{CD}_2\text{Cl}_2, 298 \text{ K}): \delta = 8.63 \text{ (dd, } J_{\text{HH}} = 4.8, 1.8 \text{ Hz}, 1H, \text{ } CH_{\text{pyr}}), 8.16 \text{ (d, } {}^3J_{\text{HH}} = 7.8 \text{ Hz}, 1H,$ CH_{pyr}), 7.76 (td, J_{HH} = 7.8, 1.8 Hz, 1H, CH_{pyr}), 7.61 (d, ${}^{3}J_{HH}$ = 6.6 Hz, 1H, CH_{PYA}), 7.56 (d, ${}^{3}J_{HH}$ = 7.0 Hz, 1H, C*H*PYA), 7.31 (dd, *³ JHH* = 7.8, 4.8 Hz, 1H, C*H*pyr), 6.74 (dd, *³ JHH* = 7.8, 6.6 Hz, 1H, C*H*PYA), 4.30 $(t, {}^{3}J_{HH} = 7.4 \text{ Hz}, 2H, \text{NCH}_2)$, 2.20 (s, 3H, PYA-CCH₃), 1.82 (apparent quintet, ${}^{3}J_{HH} = 7.4 \text{ Hz}, 2H,$ NCH₂CH₂), 1.36 (apparent sextet, ³ J_{HH} = 7.4 Hz, 2H, CH₂CH₃), 0.91 (t, ³ J_{HH} = 7.4 Hz, 3H, CH₂CH₃)ppm. ¹³C{¹H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 166.89 (*C*O), 161.20 (*C*_{PYA}), 156.52 (C_{pyr}), 149.20 (*C*Hpyr), 140.56 (*C*HPYA), 137.00 (*C*HPYA), 136.41 (*C*Hpyr), 132.79 (*C*PYA–Me), 124.54 (*C*Hpyr) 124.17 (*C*Hpyr), 113.34 (*C*HPYA), 54.88 (N*C*H2), 31.92 (NCH2*C*H2), 20.21 (*C*H2CH3), 19.46 (PYA-C*C*H3), 13.79 (CH₂CH₃) ppm. HR-MS (m/z): calculated for C₁₆H₂₀N₃O [M+H]⁺ = 270.1606; found: 270.1596.

Figure S13. ¹H NMR spectrum (CD₂Cl₂, 298 K, 400 MHz) of L₃.

Figure S14. ¹³C $\{^1H\}$ NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of **L**₃.

S.1.3.4 Synthesis of neutral PYA ligand L4

To a solution of N-(3-methylpyridin-2-yl)picolinamide (1.0 g, 4.68 mmol) in dry MeCN (20 mL) in a pressure tube was added benzyl chloride (10.8 mL, 93.8 mmol). The resulting mixture was stirred for 3 days at 85 °C. The solvent was evaporated and the residue was triturated into Et₂O, filtered and recrystallized

from MeCN/Et₂O at -20 °C. The precipitate was filtered, washed with Et₂O and dried to yield a paleyellow solid, which was dissolved in a KOH solution (2 M, 25 mL, 50 mmol). The resulting slurry was stirred for 5 min at 23 °C and then extracted with CH_2Cl_2 (3 x 20 mL). The combined organic fractions were dried over MgSO4, filtered and concentrated under vacuum to yield **L4** as a pale brown waxy solid (653 mg, 46%) that precipitated upon drying for several days under vacuum. ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ = 8.63 (dd, *J*_{HH} = 4.7, 1.5 Hz, 1H, C*H*_{pyr}), 8.12 (dd, *J*_{HH} = 7.9, 1.1 Hz, 1H, C*H*_{pyr}), 7.74 (td, J_{HH} = 7.7, 1.8 Hz, 1H, CH_{pyr}), 7.62 (d, ${}^{3}J_{HH}$ = 6.8 Hz, 1H, CH_{PYA}), 7.56 (d, ${}^{3}J_{HH}$ = 6.8 Hz, 1H, CH_{PXA} , 7.46 - 7.22 (m, 6H, CH_{pyr} + 5 CH_{phenyl}), 6.72 (t, ${}^{3}J_{HH}$ = 6.8 Hz, 1H, CH_{PYA}), 5.53 (s, 2H, NC*H*₂), 2.22 (s, 3H, PYA-CCH₃) ppm. ¹³C{¹H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 167.20 (*C*O), 161.16 (*C*PYA), 156.21 (Cpyr), 149.21 (*C*Hpyr), 140.69 (*C*HPYA), 136.71 (*C*Hpyr), 136.48 (*C*HPYA), 135.86 (Cphenyl), 132.79 (PYA-*C*CH3), 129.33 (2C, CHphenyl), 128.91 (2C, CHphenyl), 128.73 (CHphenyl), 124.69 (*C*Hpyr) 124.27 (*C*Hpyr), 113.35 (*C*HPYA), 56.97 (N*C*H2), 19.70 (PYA-C*C*H3), ppm. HR-MS (m/z): calculated for $C_{19}H_{18}N_3O$ $[M+H]^+$ = 304.1450; found: 304.1441.

Figure S15. ¹ H NMR spectrum (CD2Cl2, 298 K, 400 MHz) of **L4**.

Figure S16. ¹³C $\{^1H\}$ NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of L₄.

S.1.3.5 Synthesis of neutral PYA ligand L5

Following the general procedure, reaction of **[L5H]I** (120 mg, 0.26 mmol) yielded compound L_5 as an off-white solid (72 mg, 83%). ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ = 8.14 (dd, J_{HH} = 7.6, 1.1 Hz, 1H, CH_{pyr}), 7.85 (t, ${}^{3}J_{HH}$ = 7.6 Hz, 1H, CH_{pyr}), 7.56 (d, ${}^{3}J_{HH}$ = 6.6 Hz, 1H, CH_{PYA}), 7.51 (d, ${}^{3}J_{HH}$ = 7.1 Hz, 1H, CH_{PYA)}, 7.25 (dd, ${}^{3}J_{HH}$ = 7.4, 1.1 Hz, 1H, CH_{pyr}), 7.17 (dd, ${}^{3}J_{HH}$ = 8.3, 6.7 Hz, 1H, CH_{xyl}),

7.08 (d, ${}^{3}J_{HH}$ = 7.5 Hz, 2H, CH_{xyl}), 6.64 (dd, ${}^{3}J_{HH}$ = 7.1, 6.6 Hz, 1H, CH_{PYA}), 3.83 (s, 3H, NCH₃), 2.20 (s, 3H, PYA-CCH₃), 2.02 (s, 6H, xyl-CH₃) ppm. ¹³C{¹H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 166.90 (*C*O), 161.03 (*C*PYA), 158.73 (*C*pyr-xyl), 156.07 (*C*pyrCO), 141.19 (Cxyl-pyr), 140.11 (*C*HPYA), 137.57 (*C*HPYA), 136.91 (*C*Hpyr), 136.29 (xyl-*C*CH3, 2C) 132.40 (PYA-*C*CH3), 127.98 (*C*Hxyl), 127.71 (2C, *C*H_{xyl}), 125.48 (*C*H_{pyr}), 121.91 (*C*H_{pyr}), 112.85 (*C*H_{PYA}), 42.71 (N*C*H₃), 20.40 (2C, xyl-*CH*₃) 19.34 (PYA-CCH₃) ppm. HR-MS (m/z): calculated for C₂₁H₂₂N₃O [M+H]⁺ = 332.1763; found: 332.1751.

Figure S17. ¹H NMR spectrum (CD₂Cl₂, 298 K, 400 MHz) of L₅.

Figure S18. ¹³C $\{^1H\}$ NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of **L**₅.

S.1.3.6 Synthesis of neutral ligand L8

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prec4. To a solution of 2-chloro-3-methylpyridine $(3.0 \text{ mL}, 27.5 \text{ mmol})$ in CH_2Cl_2 (30mL) was added methyl trifluoromethanesulfonate (4.7 mL, 41.2 mmol) dropwise and the resulting solution was stirred for 2 h at 23 °C. The solution was then reduced to 10

mL and precipitated with Et₂O (50mL). The precipitate was filtered, washed with the same solvent and dried to yield the pure title compound as a white solid $(7.88 \text{ g}, 98\%)$. ¹H NMR (400 MHz, DMSO- d_6 , 298 K): δ = 9.07 (d, ${}^{3}J_{HH}$ = 6.2 Hz, 1H, CH_{pyr}), 8.57 (d, ${}^{3}J_{HH}$ = 7.8 Hz, 1H, CH_{pyr}), 8.01 (dd, ${}^{3}J_{HH}$ = 7.8, 6.2 Hz, 1H, CH_{pyr}), 4.34 (s, 3H, NCH₃), 2.54 (s, 3H, CCH₃) ppm. ¹³C{¹H} NMR (101 MHz, DMSO- d_6 , 298 K): δ = 147.02 (*C*Hpyr), 146.83 (*C*Cl), 145.99 (*C*Hpyr), 138.18 (*C*CH3), 125.06 (*C*Hpyr), 120.66 (q, $^{1}J_{CF}$ = 322.2 Hz, *C*F₃), 48.02 (N*C*H₃), 19.28 (C*C*H₃) ppm. ¹⁹F{¹H} NMR (376 MHz, DMSO-*d*₆, 298 K): δ = -79.34 ppm. HR-MS (m/z): calculated for C₇H₉ClN [M-OTf]⁺ = 142.0418; found: 142.0413.

Figure S19. ¹ H NMR spectrum (DMSO-*d*6, 298 K, 300 MHz) of **prec4**.

Figure S20. ¹³C{¹H} NMR spectrum (DMSO- d_6 , 298 K, 75 MHz) of prec4.

 L_8 To a suspension of **prec4** (2.83 g, 9.71 mmol) and K_2CO_3 (1.48 g, 10.68 mmol) in MeCN (8 mL) was added pyridin-2-ylmethanamine (1 mL, 9.71 mmol) and the reaction mixture was stirred 16 h at 80 °C. The resulting suspension was filtered and

washed with MeCN. The solvent was then evaporated under vacuum and the residue was dissolved in aq. KOH (2M, 50 mL, 100 mmol) and stirred 5 min before being extracted in CH_2Cl_2 (5 x 20 mL). Organics were dried over MgSO₄, filtered and evaporated to yield L₈ as a brownish oil (2.03 g, 98%). ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ = 8.46 (dd, *J*_{HH} = 4.8, 1.6 Hz, 1H, C*H*_{pyr}), 7.71 (d, ³*J*_{HH} = 7.4 Hz, 1H, CH_{pyr}), 7.67 (td, *J*_{HH} = 7.4, 1.6 Hz, 1H, CH_{pyr}), 7.11 (d, *J*_{HH} = 7.4, 4.8, 1.6 Hz, 1H, CH_{pyr}) 6.97 $(dd, J_{HH} = 6.7, 1.8 \text{ Hz}, 1H, CH_{PYE}$, $6.63 \text{ (dd, } ^3J_{HH} = 6.7, 1.8 \text{ Hz}, 1H, CH_{PYE})$, $5.55 \text{ (t, } ^3J_{HH} = 6.7 \text{ Hz}, 1H,$ C*H*PYE), 5.15 (s, 2H, C*H*2), 3.40 (s, 3H, PYE-NC*H*3), 2.41 (s, 3H, PYE-CC*H*3) ppm. 13C{1 H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 164.88 (C_{pyr}), 152.69 (C_{PYE}), 148.80 (CH_{pyr}), 137.90 (CH_{PYE}), 136.60 (CH_{pyr}), 135.98 (*C*HPYE), 125.72 (*C*PYA–Me), 121.48 (*C*Hpyr) 121.32 (*C*Hpyr), 100.68 (*C*HPYE), 55.15 (*C*H2), 41.32 (PYE-NCH₃), 22.90 (PYE-CCH₃) ppm. HR-MS (m/z): calculated for C₁₃H₁₆N₃ [M+H]⁺ = 214.1344; found: 214.1336.

Figure S21. ¹H NMR spectrum (CD₂Cl₂, 298 K, 400 MHz) of L_8 .

Figure S22. ¹³C{¹H} NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of **8**.

S.1.4 General procedure for the preparation of PYA palladium complexes 1-8

The neutral PYA ligand $(0.80 \text{ to } 0.84 \text{ mmol}, 1.00 \text{ to } 1.05 \text{ eq.})$ and $\text{[Pd(cod)Cl}_2\text{]}$ $(212 \text{ mg}, 0.8 \text{ mmol}, 1.00 \text{ to } 1.05 \text{ eq.})$ eq.) were dissolved in dry CH₂Cl₂ (5 mL). The resulting yellow mixture was stirred for 2 h at 23 °C and then concentrated to 2 mL. Upon addition of $Et₂O$ (6 mL) a precipitate formed, which was filtered and washed with more Et₂O. The precipitation was repeated if necessary until no palladium precursor was detected. Then the precipitate was dried under vacuo to afford the complex as a bright yellow solid.

S.1.4.1 Synthesis of pyridyl-PYA Pd complex 1

Following the general procedure reaction of **L1** (270 mg, 1.19 mmol) and $[Pd(cod)Cl₂]$ (308 mg, 1.08 mmol) yielded 1 as a bright orange solid (424 mg, 97%). Crystals suitable for Xray diffraction were grown by slow diffusion of $Et₂O$ in a CH₂Cl₂ solution. ¹H NMR (400 MHz, CD₃CN, 298 K): δ = 9.15 (d, *J*_{HH} = 5.6,

1.5 Hz, 1H, CH_{pyr}), 8.38 (d, ³J_{HH} = 6.2, 1H, CH_{PYA}), 8.23 (d, ³J_{HH} = 7.7, 1H, CH_{PYA}), 8.16 (td, J_{HH} = 7.7, 1.5 Hz, 1H, C*H*pyr), 7.88 (dd, *J*HH = 7.7, 1.6 Hz, 1H, C*H*pyr), 7.69 (ddd, *J*HH = 7.7, 5.6, 1.6 Hz, 1H, CH_{pyr}), 7.60 (d, ³ J_{HH} = 7.7, 6.2, 1H, CH_{PYA}), 4.26 (s, 3H, NCH₃), 2.49 (s, 3H, PYA-CCH₃). ¹³C{¹H} NMR (101 MHz, CD₃CN, 298 K): δ = 169.91 (*C*O), 158.25 (*C*_{PYA}), 152.62 (*C*_{pyr}), 149.67 (*C*H_{pyr}), 146.69 (*C*H_{PYA}), 142.77 (*C*H_{PYA}), 141.45 (*C*H_{pyr}), 138.70 (PYA-*CCH*₃), 129.31 (*CH*_{pyr}), 126.93 (*CH*_{pyr}), 123.94 (*C*HPYA), 45.49 (N*C*H3), 18.38 (PYA*-C*H3), ppm. HR-MS (m/z): calculated for $C_{13}H_{13}Cl_2N_3NaOPd$ $[M+Na]^+=$ 425.9368; found: 425.9370. Elemental analysis calculated for $C_{13}H_{13}Cl_2N_3OPd \cdot 0.25 \text{ CH}_2Cl_2$ (%): C 37.37, H 3.20, N 9.87; found: C 37.69, H 3.30, N 9.79.

Figure S23. ¹ H NMR spectrum (CD3CN, 298 K, 400 MHz) of **1**.

Figure S24. ¹³C $\{^1H\}$ NMR spectrum (CD₃CN, 298 K, 101 MHz) of 1.

S.1.4.2 Synthesis of pyridyl-PYA Pd complex 2

Following the general procedure from L_2 (40 mg, 0.188 mmol) and $[Pd(cod)Cl_2]$ (54 mg, 0.188 mmol). The product was further purified by microfiltration of a MeCN solution of the crude solid over Celite and subsequent precipitation upon Et2O addition, yielding complex **2** as an orange solid (67 mg, 91%). Crystals

suitable for X-ray diffraction were grown by slow diffusion of toluene in a DMSO solution of 2. ¹H NMR (400 MHz, DMSO-*d*₆, 298 K): δ = 9.04 (dd, *J*_{HH} = 5.7, 1.5 Hz, 1H, C*H*_{pyr}), 8.88 (dd, *J*_{HH} = 6.3, 1.7 Hz, 1H, CH_{PYA}), 8.45 (td, ³J_{HH} = 7.9, 1.5 Hz 1H, CH_{PYA}), 8.28 (td, ³J_{HH} = 7.7, 1.5 Hz, 1H, CH_{pyr}), 7.94 – 7.87 (m, 2H, C*H*pyr + C*H*PYA), 8.16 (td, *J*HH = 7.7, 1.5 Hz, 1H, C*H*pyr), 7.86 – 7.78 (m, 2H, C*H*pyr + CH_{PYA}), 4.24 (s, 3H, NCH₃) ppm. ¹³C{¹H} NMR (101 MHz, , DMSO-d₆, 298 K): δ = 169.46 (CO), 157.40 (*C*PYA), 151.24 (*C*pyr), 148.45 (*C*Hpyr), 145.24 (*C*HPYA), 144.36 (*C*HPYA), 140.98 (*C*Hpyr), 128.68 (*C*Hpyr or *C*HPYA), 128.48 (*C*Hpyr or *C*HPYA), 126.10 (*C*Hpyr), 123.22 (*C*HPYA), 43.78 (N*C*H3) ppm. HR-MS (m/z): calculated for $C_{16}H_{19}CIN_3OPd$ [M-Cl+CH₃CN]⁺ = 394.9891; found: 394.9878. Despite several attempts no satisfactory elemental analysis for **2** could be obtained. The closest match being $C_{12}H_{11}C_{2}N_{3}OPd \cdot 0.5 C_{4}H_{10}O$ (%): C 39.32, H 3.77, N 9.83; found: C 39.79, H 3.43, N 9.17.

Figure S25. ¹ H NMR spectrum (DMSO-*d*6, 298 K, 400 MHz) of **2**.

Figure S26. ¹³C $\{^1H\}$ NMR spectrum (DMSO- d_6 , 298 K, 101 MHz) of 2.

S.1.4.3 Synthesis of pyridyl-PYA Pd complex 3

Following the general procedure, reaction of **L3** (43 mg, 160 µmol) and [Pd(cod)Cl2] (45 mg, 0.167 mmol) yielded complex **3** as a bright orange solid (60 mg, 80%). Crystals suitable for X-ray diffraction were grown by slow diffusion of Et₂O in a CH₂Cl₂ solution of **3**. ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ = 9.24 (d,

*J*_{HH} = 5.7, 1.5 Hz, 1H, C*H*_{pyr}), 8.18 (dd, *J*_{HH} = 6.3, 1.6 Hz, 1H, C*H*_{PYA}), 8.14 (d, ³*J*_{HH} = 7.6, 1H, C*H*_{PYA}), 8.08 (td, *J*HH = 7.7, 1.5 Hz, 1H, C*H*pyr), 7.93 (dd, *J*HH = 7.7, 1.6 Hz, 1H, C*H*pyr), 7.62 (ddd, *J*HH = 7.7, 5.6, 1.6 Hz, 1H, CH_{pyr}), 7.55 (dd, ³J_{HH} = 7.6, 6.3, 1H, CH_{PYA}), 5.40 (ddd, J_{HH} = 13.4, 9.1, 5.9 Hz, 1H, NCH₂), 4.20 (ddd, *J*HH = 13.4, 9.1, 6.8 Hz, 1H, NC*H*2), 2.56 (s, 3H, PYA-CC*H*3), 2.01 – 1.81 (m, 2H, NCH2C*H*2), 1.41 (sextet, ${}^{3}J_{\text{HH}}$ = 7.4 Hz, 2H, CH₂CH₃), 0.92 (t, ${}^{3}J_{\text{HH}}$ = 7.4 Hz, 3H, CH₂CH₃). ¹³C{¹H} NMR (101 MHz, CD2Cl2, 298 K): δ = 169.77 (*C*O), 157.95 (*C*PYA), 151.83 (*C*pyr), 149.54 (*C*Hpyr), 144.72 (*C*HPYA), 140.26 (*C*Hpyr), 140.11 (*C*HPYA), 139.15 (PYA-*C*CH3), 129.48 (*C*Hpyr), 126.46 (*C*Hpyr), 123.05 (*C*HPYA), 58.02 (N*C*H2), 32.51 (NCH2*C*H3), 20.14 (*C*H2*C*H3), 18.68 (PYA*-C*H3), 13.68 (CH2*C*H3) ppm. HR-MS (m/z): calculated for C₁₆H₁₉ClN₃OPd [M-Cl]⁺ = 410.0251; found: 410.0313. Elemental analysis calculated for $C_{16}H_{19}Cl_2N_3OPd \cdot 0.5 CH_2Cl_2$ (%): C 40.52, H 4.12, N 8.59; found: C 40.26, H 4.05, N 8.51.

Figure S27. ¹H NMR spectrum (CD₂Cl₂, 298 K, 400 MHz) of 3.

Figure S28. ¹³C{¹H} NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of 3.

S.1.4.4 Synthesis of pyridyl-PYA Pd complex 4

Following the general procedure, reaction of **L4** (100 mg, 330 µmol) and [Pd(cod)Cl2] (87 mg, 300 mmol) yielded complex **4** as a yellow solid (120 mg, 83%). ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ = 9.24 (d, ³J_{HH} = 5.5 Hz, 1H, CH_{pyr}), 8.15 (d, ${}^{3}J_{\text{HH}} = 7.5$ Hz, 1H, CH_{PYA}), 8.14 (d, ${}^{3}J_{\text{HH}} = 7.6$, 1H, CH_{pyr}), 8.09 – 8.02 (m, 2H, C*H*pyr +C*H*PYA), 7.85 (d, 3 *J*HH = 7.8 Hz, 1H, C*H*pyr), 7.62 (dd, 3 *J*HH = 7.7, 5.5 Hz, 1H, C*H*pyr), 7.50 (t, ${}^{3}J_{\text{HH}}$ = 7.6 Hz, 1H, CH_{PYA}), 7.43 – 7.21 (m, 5H, CH_{benzyl}), 6.21 (d, ²J_{HH} = 14.8 Hz, 1H, CH₂), 5.81 (d, $^{2}J_{\text{HH}}$ = 14.8 Hz, 1H, C*H*₂), 2.58 (s, 3H, PYA-CC*H*₃) ppm.). ¹³C{¹H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 169.40 (*C*O), 158.22 (*C*PYA), 151.73 (*C*pyr), 149.49 (*C*Hpyr), 145.92 (*C*HPYA), 140.24 (*C*Hpyr), 139.87 (*C*HPYA), 138.95 (PYA-*C*CH3), 132.89 (*C*benzyl), 130.06 (2C, *C*Hbenzyl), 129.79 (3C, *C*Hbenzyl), 128.45 (*C*Hpyr), 126.41 (*C*Hpyr), 123.02 (*C*HPYA), 60.14 (*C*H2), 18.71 (PYA*-C*H3) ppm. HR-MS (m/z): calculated for $C_{19}H_{17}CIN_3OPd$ $[M-C1]^+$ = 444.0095; found: 444.0073. Elemental analysis calculated for $C_{19}H_{17}Cl_2N_3OPd \cdot 0.25 \text{ } CH_2Cl_2 (%)$: C 46.07, H 3.51, N 8.37; found: C 46.05, H 3.66, N 8.06.

Figure S29. ¹H NMR spectrum (CD₂Cl₂, 298 K, 400 MHz) of 4.

Figure S30. ¹³C{¹H} NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of 4.

S.1.4.5 Synthesis of pyridyl-PYA Pd complex 5

Following the general procedure, reaction of **L5** (39 mg, 0.117 mmol) and [Pd(cod)Cl2] (33 mg, 0.117 mmol) yielded complex **5** as a dark orange solid (57 mg, 95%). Crystals suitable for X-ray diffraction analysis were grown by diffusion of n-heptane into a CH_2Cl_2 solution of **5**. ¹H NMR (400 MHz, CD_2Cl_2 ,

298 K): $\delta = 8.17 - 8.10$ (m, 2H, 2 CH_{PYA}), $8.09 - 8.01$ (m, 2H, 2 CH_{pyr}), 7.50 (dd, ³J_{HH} = 7.7, 6.2 Hz, 1H, CH_{PYA}), 7.40 (dd, ³J_{HH} = 7.4, 2.4 Hz, 1H, CH_{pyr}), 7.25 (t, ³J_{HH} = 7.5 Hz, 1H, CH_{xyl}), 7.10 (d, ³J_{HH} = 7.5 Hz, 1H, C*H*xyl), 4.35 (s, 3H, NC*H*3), 2.62 (s, 3H, PYA-CC*H*3), 2.33 (d, 6H, xyl-CC*H*3). 13C{1 H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 168.89 (*C*O), 163.65 (*C_{pyr}*), 158.32 (*C_{PYA}*), 153.94 (*C_{pyr}-xyl*), 145.75 (*C*HPYA), 140.76 (*C*HPYA), 139.76 (*C*Hpyr), 139.46 (PYA-*C*CH3), 139.12 (Cxyl-pyr), 136.60 (xyl-*CCH*₃), 136.45 (xyl-*CCH*₃), 132.65 (*CH*_{pyr}), 129.30 (*CH*_{xyl}), 127.76 (*CH*_{xyl}), 127.69 (*CH*_{xyl}), 125.08 (*C*Hpyr), 122.69 (*C*HPYA), 45.59 (N*C*H3), 21.33 (xyl-*C*H3), 21.27(xyl-*C*H3), 19.00 (PYA*-C*H3) ppm. HR-MS (m/z): calculated for $C_{21}H_{21}CIN_3OPd$ [M–Cl]⁺ = 472.0408; found: 472.0410. Elemental analysis calculated for $C_{21}H_{21}C_{2}N_{3}OPd \cdot 0.75 CH_{2}Cl_{2}$ (%): C 45.64, H 3.96, N 7.34; found: C 45.65, H 3.87, N 7.10.

Figure S31. ¹H NMR spectrum (CD₂Cl₂, 298 K, 400 MHz) of 5.

Figure S32. ¹³C{¹H} NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of 5.

S.1.4.6 Synthesis of pyridyl-PYA Pd complex 7

[L_7 H]I (200 mg, 0.57 mmol) and NH_4 PF₆ (465 mg, 2.85 mmol) were separately dissolved in a minimal amount of H_2O , combined, and the resulting suspension was stirred at 23 °C for 10 min. The suspension was filtered and and the filtrate washed with H₂O and Et₂O. The residue was dried under vacuum to yield $[L_7H]PF_6$ as a

colourless solid (195 mg, 95%). $[L_7H]PF_6$ (180 mg, 0.5 mmol), $[Pd(cod)Cl_2]$ (143 mg, 0.5 mmol) and Cs_2CO_3 (490 mg, 1.5 mmol) were suspended in CH₂Cl₂ (20 mL) and stirred at 23 °C for 16 h. The resulting orange solution was filtered and washed with CH_2Cl_2 until washings were clear. The combined filtrates were concentrated, triturated into $Et₂O$, and the residue was collected and dried under vacuum to yield 7 as an orange solid (146 mg, 75%). ¹H NMR (400 MHz, CD₃CN, 298 K): δ = 9.16 (d, J_{HH} = 5.6, 1.5 Hz, 1H, CH_{pyr}), 8.70 (t, ⁴J_{HH} = 1.5 Hz, 1H, CH_{PYA}), 8.42 (dt, J_{HH} = 8.2, 1.5 Hz, 1H, CH_{PYA}), 8.20 (d, ${}^{3}J_{\text{HH}} = 6.1$ Hz, 1H, CH_{PYA}), 8.10 (td, $J_{\text{HH}} = 7.7$, 1.5 Hz, 1H, CH_{pyr}), 7.88 (dd, $J_{\text{HH}} = 7.7$, 1.6 Hz, 1H, C*H*pyr), 7.78 (dd, *J*HH = 8.2, 6.1 Hz, 1H, C*H*PYA), 7.61 (ddd, *J*HH = 7.7, 5.6, 1.6 Hz, 1H, C*H*pyr), 4.24 $(s, 3H, NCH_3)$ ppm. ¹³C{¹H} NMR (101 MHz, CD₃CN, 298 K): δ = 171.72 (*CO*), 154.37 (*C*_{pyr}), 149.40 (*C*Hpyr), 147.51 (*C*PYA), 145.98 (*C*HPYA), 144.79 (*C*HPYA), 141.21 (*C*Hpyr), 139.35 (*C*HPYA), 128.63 (CH_{pyr}) , 126.77 $(CH_{\text{pyr or}}CH_{\text{PYA}})$, 126.66 $(CH_{\text{pyr or}}CH_{\text{PYA}})$, 48.86 (NCH₃) ppm. HR-MS (m/z): calculated for $C_{12}H_{11}CIN_3OPd$ $[M-Cl]^+$ = 353.9625; found: 353.9619. Elemental analysis calculated for $C_{12}H_{11}C_{2}N_{3}OPd \cdot 0.5 CH_{2}Cl_{2}$ (%): C 34.67, H 2.79, N 9.70; found: C 35.09, H 2.91, N 9.40.

Figure S33. ¹H NMR spectrum (CD₃CN, 298 K, 400 MHz) of 7.

Figure S34. ¹³C $\{^1H\}$ NMR spectrum (CD₃CN, 298 K, 101 MHz) of 7.

S.1.4.8 Synthesis of pyridyl-PYA Pd complex 8

In a N_2 atmosphere, $[Pd(cod)Cl_2]$ (86 mg, 0.3 mmol) was added a solution of L_8 (64 mg, 0.3 mmol) in MeCN (2 mL) and the resulting bright red suspension was stirred for 1 h at 50 °C. The solvent was then evaporated, the residue was dissolved in $CH_2Cl_2(1 \text{ mL})$ and precipitated with Et₂O (5 mL). The precipitate was filtered,

washed with more Et₂O and dried under vacuum to yield **8** as a bright red solid (82 mg, 70%). Crystals suitable for X-ray diffraction analysis were grown by slow diffusion of Et_2O into a 1:1 MeCN/ CH_2Cl_2 solution of **8**. ¹H NMR (400 MHz, CD₂Cl₂, 298 K): δ = 8.94 (d, *J*_{HH} = 5.6, 1.5 Hz, 1H, C*H*_{pyr}), 7.84 (td, *J*HH = 7.7, 1.5 Hz, 1H, C*H*pyr), 7.61 (dd, *J*HH = 6.6, 1.8 Hz 1H, C*H*PYE), 7.50 (d, 3 *J*HH = 7.3 Hz, 1H, C*H*PYE), 7.35 – 7.29 (m, 2H, CH_{pyr}), 6.75 (t, ³J_{HH} = 7.0 Hz 1H, CH_{PYE}), 4.35 (s, 3H, NCH₃), 2.59 (s, 3H, PYE-CCH₃) ppm. Methylene protons not resolved. At 213 K, they appear as a pair of doublets (δ = 5.64 and 4.21 ppm; ${}^{2}J_{\text{HH}} = 15.6 \text{ Hz}$) and at 328 K as a broad singlet ($\delta = 4.92$ ppm; Fig. S36). ¹³C{¹H} NMR (101 MHz, CD₂Cl₂, 298 K): δ = 165.08 (*C*_{PYE}), 154.55 (*C*_{pyr}), 150.71 (*C*H_{pyr}), 142.98 (*C*H_{PYE}), 139.81 (*C*HPYE), 139.18 (*C*Hpyr), 133.73 (PYE-*C*CH3), 123.31 (*C*Hpyr), 119.94 (*C*Hpyr), 113.81 (*C*HPYE), 63.09 (CH_2) , 45.92 (NCH₃), 21.93 (PYE-CH₃) ppm. HR-MS (m/z): calculated for C₁₃H₁₅Cl₂N₃NaPd [M+Na]⁺ = 411.9576; found: 411.9566. Despite several attempts, no satisfactory elemental analysis data could be obtained.

Figure S35. ¹ H NMR spectrum (CD2Cl2, 298 K, 400 MHz) of **8**.

Figure S36. Variable temperature ¹H NMR spectra of complex **8** (CD₂Cl₂, 213 - 328 K, 400 MHz), indicating the coalescence of the AB doublets around 4.2 and 5.6 ppm at 25 °C to a singlet at 5 ppm.

Figure S37. ¹³C $\{^1H\}$ NMR spectrum (CD₂Cl₂, 298 K, 101 MHz) of **8**.

S.2. Crystal structure determinations

Crystals of $[L_1H]PF_6, L_1, 1, 2, 3, 5$ and 8 immersed in parabar oil were mounted at ambient conditions and transferred into the stream of nitrogen (173 K). Measurements for $[L_1H]PF_6, L_1, 1, 2$ and 8 were made on a RIGAKU Synergy S area-detector diffractometer1 using mirror optics monochromated Cu Kα radiation (λ = 1.54184 Å). Measurements for **3** and **5** were made on an *Oxford Diffraction SuperNova* area-detector diffractometer using mirror optics monochromated Mo $K\alpha$ radiation ($\lambda = 0.71073$ Å) and Al filtered.

Data reduction was performed using the *CrysAlisPro*^{S3} program. The intensities were corrected for Lorentz and polarization effects, and an absorption correction based on the multi-scan method using SCALE3 ABSPACK in *CrysAlisPro*^{S3} was applied.

All the structures were solved by intrinsic phasing using *SHELXT*^{S4}, which revealed the positions of all non-hydrogen atoms of the title compound. All non-hydrogen atoms were refined anisotropically. For **[L1H]PF6**, **L1**, **5** H-atoms were located from the difference density map and had their positions and isotropic displacement parameters refined freely. For **1, 3** and **8** H-atoms were assigned in geometrically calculated positions and refined using a riding model where each H-atom was assigned a fixed isotropic displacement parameter with a value equal to 1.2Ueq of its parent atom (1.5Ueq for methyl groups). For **2,** H-atoms were located from the difference density map and had their positions and isotropic displacement parameters refined freely, except for H2 and H5 (CIF labelling), where the atoms were refined using a riding model where each H-atom was assigned a fixed isotropic displacement parameter with a value equal to 1.2Ueq of its parent atom. In addition, anharmonic motion was refined for Pd1.

Refinements were carried out on $F²$ using full-matrix least-squares procedures, which minimized the function Σw(F_o² – F_c²)². The weighting scheme was based on counting statistics and included a factor to downweight the intense reflections. All calculations were performed using the *SHELXL-2014*/7 ^{S5} program in OLEX2.S6

A disorder solvent could not be modeled in **1**, therefore it is not included in the model and instead a mask was used to calculate the electron density present in the void area and its contribution was included into the calculated structure factors (total electron count 63e).

The structure of **3** was twinned by a polymorph (monoclinic crystal system). The multi crystal separation during the data reduction led to an overall 66% completeness of the data only due to reflection overlap. Since the unit cells differ there is no twin law that can be use in the refinement and therefore the refinement had to proceed using the major component polymorph (orthorhombic crystal system) against the reflections containing the overlap, which leads to artificial residual densities around the pseudo translated Pd position on the second polymorph (about 0.25 along the b unit cell axis). In addition, the structure is refined as an inversion twin.

For the structure of **5**, a disorder model was used for part of the structure where the occupancies of each disorder component was refined through the use of a free variable. The sum of equivalent components was constrained to 1 (100%). Final occupancies are listed in the associated crystallographic data.

For **8**, twinning was detected on the frames where the second component corresponded to a twofold rotation around [-0.26 -0.96 0.04] (reciprocal) or [-0.48 -0.88 -0.07] (direct). The refinement proceeded against reflections of the major component only.

Data collection and refinement parameters for $[L_1H]PF_6, L_1, 1, 2, 3, 5$ and 8 are given in Tables S1–S4. Crystallographic data for all structures reported in this paper have been deposited with the Cambridge Crystallographic Data Centre (CCDC) as supplementary publication numbers 2115311–2115316.

| CCDC No. | 2297421 | 2297422 |
|---|---------------------------------------|---|
| Empirical formula | $C_{13}H_{14}F_6N_3OP$ | $C_{13}H_{13}N_3O$ |
| Formula weight | 373.24 | 227.26 |
| Temperature/K | 173.01(10) | 173.01(10) |
| Crystal system | monoclinic | monoclinic |
| Space group | P2 ₁ /n | P2 ₁ /c |
| $a/\text{\AA}$ | 21.02657(6) | 10.25664(8) |
| b/A | 14.43623(4) | 9.94244(8) |
| $c/\text{\AA}$ | 21.17731(6) | 11.26378(9) |
| α /° | 90 | 90 |
| β /° | 90.6664(2) | 91.5286(7) |
| $\gamma/^\circ$ | 90 | 90 |
| Volume/ \AA^3 | 6427.82(3) | 1148.226(15) |
| Ζ | 16 | 4 |
| $\rho_{\rm calc}$ g/cm ³ | 1.543 | 1.315 |
| μ /mm ⁻¹ | 2.208 | 0.697 |
| F(000) | 3040.0 | 480.0 |
| Crystal size/ $mm3$ | $0.432 \times 0.338 \times 0.207$ | $0.155 \times 0.13 \times 0.075$ |
| Radiation | Cu Kα (λ = 1.54184) | Cu Kα (λ = 1.54184) |
| 20 range for data collection/° | 5.89 to 147.988 | 8.624 to 140.468 |
| Index ranges | $-26 \le h \le 26, -18 \le k \le 18,$ | $-12 \le h \le 12$, $-12 \le k \le 12$, |
| | $-26 \le l \le 26$ | $-13 \le l \le 13$ |
| Reflections collected | 246423 | 22145 |
| Independent reflections | 13039 [$R_{int} = 0.0341$] | 2186 [$R_{int} = 0.0303$, |
| | $R_{\text{sigma}} = 0.0102$ | $R_{\text{sigma}} = 0.0123$ |
| Data/restraints/parameters | 13039/0/1090 | 2186/0/207 |
| Goodness-of-fit on F^2 | 1.050 | 1.042 |
| Final R indexes $[I>=2\sigma(I)]$ | $R_1 = 0.0424$, $wR_2 = 0.1161$ | $R_1 = 0.0312$, $wR_2 = 0.0806$ |
| Final R indexes [all data] | $R_1 = 0.0434$, $wR_2 = 0.1168$ | $R_1 = 0.0320$, $wR_2 = 0.0813$ |
| Largest diff. peak/hole / e Å ⁻³ | $0.73/-0.39$ | $0.21/-0.16$ |

Table S1. Crystal data and structure refinement for $[L_1H]PF_6$ and L_1 .

| Identification code | 2297423 | 2297424 |
|--------------------------------------|---------------------------------------|---|
| Empirical formula | $C_{13}H_{13}Cl_2N_3OPd$ | $C_{12}H_{11}Cl_2N_3OPd$ |
| Formula weight | 404.56 | 390.561 |
| Temperature/K | 173.01(10) | 173.00(10) |
| Crystal system | orthorhombic | monoclinic |
| Space group | Pbca | $P2_1/n$ |
| $a/\text{\AA}$ | 13.80951(16) | 10.44149(12) |
| b/A | 14.64287(17) | 11.38265(13) |
| $c/\text{\AA}$ | 20.4888(2) | 11.56249(13) |
| α ^o | 90 | 90 |
| β /° | 90 | 90.8439(10) |
| $\gamma/^\circ$ | 90 | 90 |
| Volume/ \AA^3 | 4143.06(8) | 1374.07(3) |
| Z | 8 | $\overline{4}$ |
| $\rho_{\rm calc}$ g/cm ³ | 1.297 | 1.888 |
| μ /mm ⁻¹ | 9.593 | 14.435 |
| F(000) | 1600.0 | 768.0 |
| Crystal size/ $mm3$ | $0.254 \times 0.091 \times 0.059$ | $0.214 \times 0.18 \times 0.113$ |
| Radiation | Cu Kα (λ = 1.54184) | Cu Kα (λ = 1.54184) |
| 20 range for data collection/° | 9.806 to 136.152 | 10.9 to 152.88 |
| Index ranges | $-16 \le h \le 16, -17 \le k \le 17,$ | $-12 \le h \le 13$, $-14 \le k \le 14$, |
| | $-22 \le l \le 24$ | $-14 \le l \le 14$ |
| Reflections collected | 30476 | 27616 |
| Independent reflections | 3774 [$R_{int} = 0.0572$, | 2890 [$R_{int} = 0.0692$, |
| | $R_{\text{sigma}} = 0.0252$] | $R_{\text{sigma}} = 0.0235$] |
| Data/restraints/parameters | 3774/0/183 | 2890/0/234 |
| Goodness-of-fit on F^2 | 1.095 | 1.045 |
| Final R indexes $[I>=2\sigma(I)]$ | $R_1 = 0.0394$, $wR_2 = 0.1222$ | $R_1 = 0.0291$, $wR_2 = 0.0747$ |
| Final R indexes [all data] | $R_1 = 0.0425$, $wR_2 = 0.1250$ | $R_1 = 0.0292$, $wR_2 = 0.0747$ |
| Largest diff. peak/hole / e A^{-3} | $1.33/-0.77$ | $0.62/-0.80$ |

Table S2. Crystal data and structure refinement for complexes **1** and **2**.

Table S3. Crystal data and structure refinement for **3** and **5**.

| μ /mm ⁻¹ | 1.366 | 1.190 |
|---------------------------------------|---|---------------------------------------|
| F(000) | 2128.0 | 2048.0 |
| Crystal size/ $mm3$ | $0.19 \times 0.142 \times 0.086$ | $0.218 \times 0.132 \times 0.073$ |
| Radiation | Mo Ka $(\lambda = 0.71073)$ | Mo Ka $(\lambda = 0.71073)$ |
| 20 range for data collection/ \circ | 4.094 to 60.82 | 4.516 to 59.148 |
| Index ranges | $-27 \le h \le 27$, $-14 \le k \le 14$, | $-42 \le h \le 42, -13 \le k \le 13,$ |
| | $-31 \le l \le 31$ | $-19 \le l \le 19$ |
| Reflections collected | 62815 | 55339 |
| Independent reflections | 12785 [$R_{int} = 0.0391$, | 5712 [$R_{int} = 0.0342$, |
| | $R_{\text{sigma}} = 0.0305$] | $R_{\text{sigma}} = 0.0165$ |
| Data/restraints/parameters | 12785/1/474 | 5712/43/258 |
| Goodness-of-fit on F^2 | 1.085 | 1.101 |
| Final R indexes $[1>=2\sigma(1)]$ | $R_1 = 0.0892$, w $R_2 = 0.2152$ | $R_1 = 0.0207$, $wR_2 = 0.0533$ |
| Final R indexes [all data] | $R_1 = 0.0925$, $wR_2 = 0.2198$ | $R_1 = 0.0221$, $wR_2 = 0.0540$ |
| Largest diff. peak/hole / e A^{-3} | $11.27/-1.35$ | $0.46/-0.64$ |

Table S4. Crystal data and structure refinement for **8**.

| | $[L_1H]^{b}$ | L_1 | 1 |
|------------------------|--------------|------------|------------|
| $C1-NPYA$ | 1.401(4) | 1.333(1) | 1.395(4) |
| $C2-C3$ | 1.384(3) | 1.370(2) | 1.373(5) |
| $C3-C4$ | 1.368(5) | 1.397(2) | 1.369(5) |
| $C1-N1$ | 1.355(3) | 1.378(1) | 1.344(4) |
| $N1-C5$ | 1.353(3) | 1.3648(14) | 1.355(5) |
| N_{PVA} -C6 | 1.355(2) | 1.343(14) | 1.340(4) |
| $O1-C6$ | 1.216(3) | 1.237(2) | 1.221(4) |
| Pd-Cl1 | | | 2.303(9) |
| Pd –Cl ₂ | | | 2.2935(11) |
| A^c | 89(7) | 39.9(1) | 87.8(4) |
| δ_{NCH3} | 4.32 | 3.86 | 4.34 |
| $\delta_{\text{H}(4)}$ | 7.83 | 6.73 | 7.54 |

Table S5. Extended solid-state X-ray and solution ¹H NMR metrics for $[L_1H]^+$, L_1 and complex 1.^{*a*}

^{*a*} bond lengths in Å, bond angles in deg, chemical shifts in ppm (in CD₂Cl₂); ^{*b*} average value from 4 independent molecules in the asymmetric unit; ^{*c*} torsion angle between amide N–C=O group and PYA heterocycle defined by N1–C1–N_{PYA}–C6, see also Fig. S38.

Note that in L_1 the carbonyl bond length is increased compared to $[L_1H]^+$, suggesting contribution from an imidate resonance structure in **L1**. However, this zwitterionic L-type resonance form contributes less to **1** when considering that the O1-C6 bond length is essentially identical to that of the protonated ligand precursor $[L_1H]^+$.

Figure S38. Graphical representation of the dihedral angle θ in complex 1. Planes were generated from C6-N_{PYA}-C1 (red, amide plane) and NPYA-C1-N1 (blue, pyridinium plane). Analogous atoms were used for the determination of θ in all other structures except 8, where the atoms Pd-N_{PYE}-C1 were used to generate the red plane.

S.3. Buried volume calculations

Buried volume calculations were carried out using the SambVca 2.1 Software.^{S8} The calculations were performed using the following parameters: Bonds radii scaled by 1.17x; Sphere radius of 3.5; Distance of the coordination point from the center of the sphere 0.0; Mesh spacing for numerical integration 0.10; H atoms were not included in the calculations.

Complex **1**:

Complex **2**:

DRAW

Complex **3**:

Complex **5**:

Complex **8**:

| %V Free | %V Buried | | | % V Tot/V Ex | | 3 |
|-----------|-----------|----------------|----------------|--------------|-----------------|----------------|
| 58.4 | 41.6 | | | 99.9 | | \overline{a} |
| | | | | | | ı |
| Quadrant | Vf | V _b | V _t | %V f | %V _b | |
| SW | 28.3 | 16.5 | 44.9 | 63.1 | 36.9 | 0 |
| NW | 30.1 | 14.8 | 44.9 | 67.0 | 33.0 | |
| NE | 25.1 | 19.8 | 44.9 | 55.9 | 44.1 | |
| SE | 21.3 | 23.6 | 44.9 | 47.4 | 52.6 | -2 |
| | | | | | | -3 |

S.4. Catalytic α-arylation of ketones

Typical procedure: NaOtBu (106 mg, 1.1 mmol), complex **1** (4.05 mg, 0.01 mmol) and hexamethylbenzene (32.5 mg, 0.2 mmol) were placed in a 10 mL microwave vial that was closed, evacuated and backfilled with N_2 three times. Dry 1,4-dioxane (1.0 mL), ketone (1.0 mmol) and aryl halide (1.0 mmol) were then added in this order. The resulting suspension was heated at 105^oC and vigorously stirred for the indicated time. The course of the reaction was followed by taking aliquots (20 μ L) and diluting them with either CDCl₃ or wet 1,4-dioxane for NMR or GC-MS analysis, respectively. At the end of the reaction the suspension was poured into H₂O (10 ml), extracted with CH₂Cl₂ (5 x 10) mL), dried over Na2SO4, filtered and concentrated under vacuum. If necessary, the crude product was purified by gradient flash column chromatography (0 to 15% EtOAC in n-hexane; SiO2).

S.4.1. Optimization of reaction conditions

Table S6. Screening of different bases for the α-arylation of propiophenone with bromobenzene. *a*

 α Reaction conditions: ArBr (1.0 mmol), propiophenone (1.0 mmol), [1] (0.01 mmol), 1,4-dioxane (1 mL) in a 10 mL microwave vial, 105 °C for 90 min under N₂; ^{*b*} yields determined by GC analysis with hexamethylbenzene as internal standard; *^c* NaO*t*Bu (2 M in THF, 0.53 mL, 1.1 mmol) was used yielding a dioxane/THF ratio of ~65:35; *^d* THF (53 µL) was added to the reaction mixture to mimic the solvent ratio of entry 9; *^e* NaOtBu solution (2 M in THF, 0.87 mL, 1.8 mmol) was used yielding a dioxane/THF ratio of ~53/47.

Figure S39. Influence of the base cation on the reaction rate. Conditions: ArBr (1.0 mmol), propiophenone (1.0 mmol), **2** (0.01 mmol), base (1.1 mmol), solvent (1.0 mL) in a 10 mL microwave vial, 105 °C for 90 min under N2. Yields determined by GC analysis with hexamethylbenzene as internal standard.

NaO*t*Bu (1.1 eq) **solvent**

Table S7. Screening of solvents for the α-arylation of propiophenone with bromobenzene. *a*

 $\frac{1 \text{ mol\% [1]}}{\text{NaOfBu (1.16)}}$

^a Reaction conditions: ArBr (1.0 mmol), propiophenone (1.0 mmol), [**1**] (0.01 mmol), NaO*t*Bu (1.1 mmol), solvent (1.0 mL) in a 10 mL microwave vial, 105 °C for 90 min under N₂; ^b yields and conversions determined by GC analysis with hexamethylbenzene as internal standard.

Table S8. Influence of the concentration of the reactants on the α-arylation of propiophenone. *a*

| entry | substrate concentration | spectroscopic yield ^b | ketone conv | PhBr conv | TOF_{max} /h ⁻¹ |
|----------------|-------------------------|----------------------------------|-------------|-----------|-------------------------------------|
| 1^c | 0.1 _M | 8% | 36% | 34% | |
| 2 | 0.5 _M | 77% | 91% | 89% | --- |
| | 1.0 M | 91% | 94% | $>99\%$ | 220 |
| 4 | 2.0 _M | 87% | 96% | $>99\%$ | 280 |
| | 4.0 M | 80% | 86% | 85% | 270 |
| 6 ^d | 8.0 M | 85% | 86% | 87% | 360 |

^a Reaction conditions: Bromobenzene (1.0 mmol), propiophenone (1.0 mmol), complex **1** (0.01 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (10 to 0.25 mL) in a 10 mL microwave vial, 105 °C for 1 h under N₂; ^{*b*} yields and conversions determined by GC analysis with hexamethylbenzene as internal standard; *^c* reaction was performed in a 30 mL microwave vial. *^d* all the quantities were doubled and the solvent volume used was 0.25 mL.

Figure S40. Time conversion profile for the a-arylation of propiophenone with bromobenzene catalyzed by **1** at different substrate concentrations (molarity with respect to propiophenone); ratio of ketone/ bromobenzene/ base /catalyst consistently at 1:1:1.1:0.01 as detailed in Table S8.

Table S9. Variation of the substrate stoichiometry in the α-arylation of propiophenone. *a*

^a Reaction conditions: PhBr (1.0 mmol), propiophenone (1.0 mmol), complex **1** (0.01 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial, 105 °C, 60 min, under N₂, yields relative to the limiting substrate; *PhBr (1.2 mmol);* $*c*$ *propiophenone (1.2 mmol).*

Figure S41. Conversion profile for the α -arylation of propiophenone with bromobenzene at different reagent stoichiometries; for conditions see Table S9.

Table S10. Effect of the reaction temperature on the α -arylation of propiophenone catalyzed by complexes **1**. *a*

^a Reaction conditions: ArBr (1.0 mmol), propiophenone (1.0 mmol), complex **1** (0.01 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial, 1 h reaction time under N₂; ^{*b*} spectroscopic yield after 150 min.

Figure S42.Time dependent conversion profiles for the coupling of propiophenone and bromobenzene catalyzed by complex **1** at different temperatures, dotted lines indicate initial rates. Reaction conditions: ArBr (1.0 mmol), propiophenone (1.0 mmol), complex **1** (0.01 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial. Spectroscopic yields measured by GC-FID with hexamethylbenzene used as internal standard.

Figure S43. Eyring plot for the α -arylation of propiophenone catalyzed by complex 1. Least-square regression and integration of standard errors from the least-square method gives a slope of $-12,643$ (± 1199) and an intercept of 13.08 (\pm 3.18). From the Eyring equation (1)

$$
\ln(k/T) = (1/T) \times \Delta H^{\ddagger}/R + \ln(k_B/h) + \Delta S^{\ddagger}/R \tag{1}
$$

 ΔH^{\ddagger} was calculated from the slope,^{S9} as follows:

$$
\Delta H^{\ddagger} = 12,643 \ (\pm 1199) \times 8.31 = 105.1 \ (\pm 9.9) \text{ kJ mol}^{-1}
$$

 ΔS^{\ddagger} was calculated from the intercept as follows:

$$
13.076 \text{ (\pm 3.178)} = \ln(\text{kg/h}) + \Delta S^{\ddagger} / R
$$
\nand hence:

\n
$$
\Delta S^{\ddagger} = [13.08 \text{ (\pm 3.18)} - 23.76] \times 8.31 = -88.8 \text{ (\pm 26.4)} \text{ J (K mol)}^{-1}
$$

S.4.2. Variation of catalyst precursors

Figure S44. Time dependent conversion profile of the a-arylation of propiophenone with bromobenzene catalyzed by different palladium complexes*.* Reaction conditions: ArBr (1.0 mmol), propiophenone (1.0 mmol), [Pd] (0.01 mmol), NaOtBu (1.1 mmol), 1,4-dioxane (1.0 mL) in a 10 mL microwave vial, 105 °C, 90 min, N₂ atmosphere.

Figure S45. Time-conversion profile for the α -arylation of propiophenone upon lowering the catalyst loading. Reaction conditions for 1 mol% catalyst loading: PhBr (1.0 mmol), propiophenone (1.0 mmol), complex **1** (0.01 mmol), NaOtBu (1.1 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial, 90 min at 125°C; for 0.04 mol% catalyst loading: PhBr (3.09 mmol), propiophenone (3.09 mmol), complex **1** (1.24 µmol), NaO*t*Bu (3.40 mmol), 1,4-dioxane (1.55 mL) in a 10 mL microwave vial, 90 min at 125 °C. For 0.01 mol% catalyst loading: PhBr (12.36 mmol), propiophenone (12.36 mmol), complex **1** (1.24 µmol), NaO*t*Bu (13.59 mmol), 1,4-dioxane (6.2 mL) in a 25 mL microwave vial, 90 min at 125 °C.

Figure S46. Time conversion profiles of the α -arylation of propiophenone with complexes 1 and 5 at 105 and 125 °C. Reaction conditions: PhBr (1.0 mmol), propiophenone (1.0 mmol), [Pd] (0.01 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial.

S.4.3. Variation of substrates

Figure S47. Maximum rate determination for the α -arylation of various *para*-substituted propiophenones with bromobenzene. Reaction conditions: PhBr (1.2 mmol), propiophenone (1.0 mmol), complex **1** (0.01 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial at 105°C. Spectroscopic yields determined by GC-FID relative to hexamethylbenzene as internal standard. Dotted lines represent maximum rates.

Figure S48. Initial rate determination (TOF10min) for the α-arylation of propiophenone with various *para*substituted bromobenzenes. Reaction conditions: ArBr (1.0 mmol), propiophenone (1.2 mmol), complex **1** (0.01 mmol), NaO*t*Bu (1.32 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial at 105 °C. Spectroscopic yields determined by GC-FID relative to hexamethylbenzene as internal standard.

Figure S49. Hammett plot for the arylation of 4-substituted propiophenones **(**see Fig S48 for reaction conditions and initial rates) showing a very poor correlation.

Table S11. Reaction optimization for the α-arylation of propiophenone with phenyl chloride catalyzed by complex **1**. *a*

^a Reaction conditions: PhCl (1.0–2.0 mmol), propiophenone (1.0 mmol), complex **1** (0.01- 0.05 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (0.5 mL) in a 10 mL microwave vial, 60 min at indicated temperature; yields did not increase upon extension of reaction times.

Figure S50. Initial time conversion profile for the α -arylation of propiophenone with chlorobenzene as substrate, dotted line indicates maximum rate (TOF $_{max}$ = 50 h⁻¹). The yield after 60 min is 9%, barely higher than the yield after 10 min. Reaction conditions: PhCl (1.2 mmol), propiophenone (1.0 mmol), complex **1** (0.01 mmol), NaO*t*Bu (1.1 mmol), 1,4-dioxane (1.0 mL) in a 10 mL microwave vial at 125 °C.

Substrate scope

¹H NMR spectroscopy was typically used to determine spectroscopic yields of α -arylated ketone products. Hexamethylbenzene was used as internal standard. Aliquots (20 µL) were taken from the reaction mixture after purging a syringe three times with nitrogen gas and were directly dissolved in $CDCl₃ (0.3 mL)$ and analysed by ¹H NMR spectroscopy.

Figure S51. Stacked 1 HNMR spectra of a typical catalytic run of bromobenzene (**9d**) and propiophenone (**10c**) in 1,4-dioxane.

Figure S52. Zoom of the Stacked ¹H NMR spectra from Fig. S51 indicating the signals at δ _H = 3.0 and 4.7 used for quantifying conversion and yield, respectively.

S.4.4. Characterization of products

All products were characterized as crude mixtures or isolated products when the ${}^{1}H$ NMR spectroscopic yield was not clear.

Product 11ac

Figure S53. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of 11ac from the crude reaction mixture, in agreement with literature data.^{S10} HR-ESI-MS (m/z): calculated for C₁₇H₂₀NO [M+H]⁺ = 254.1545; found: 254.1536.

Product 10bc

Figure S54. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated **11bc**, in agreement with literature data.^{S11} HR-MS (m/z): calculated for $C_{16}H_{17}O_2$ [M+H]⁺ = 241.1229; found: 241.1222.

Product 11cc

Figure S55. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated 11cc, in agreement with literature data.^{S11} HR-EI-MS (m/z): calculated for C16H18O = 224.1201; found: 224.1196.

Product 11dc (also **11jc** and **11kc**)

Figure S56. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated 11dc, in agreement with literature data.^{S11} HR-MS (m/z): calculated for C15H16O [M+H] ⁺ = 211.1123; found: 211.1127. Note: Products **11jc** and **11kc** had identical analytics (same specie).

Product **11ec**

Figure S57. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated 11ec, in agreement with literature data.^{S12} HR-EI-MS (m/z): calculated for $C_{15}H_{13}FO = 228.0950$; found: 228.0942.

Product 11fc

Figure S58.¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11fc from the reaction mixture, in agreement with literature data.^{S13} Due to low conversion, only the aliphatic protons were integrated. Conversion was calculated from the crude mixture. HR-EI-MS (m/z): calculated for $C_{22}H_{18}O_2 = 314.1307$; found: 314.1304.

Product 11gc

Figure S59. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated 11gc, in agreement with literature data.^{S10} HR-EI-MS (m/z): calculated for $C_{16}H_{13}F_3O = 278.0918$; found: 278.0918.

Product 11hc

Figure S60. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11hc from the reaction mixture, in agreement with literature data.^{S11} Due to low conversion, only the aliphatic protons were integrated. Conversion was calculated from the crude mixture. HR-EI-MS (m/z): calculated for $C_{16}H_{16}O = 224.1201$; found: 224.1195.

Product 11da

Figure S61. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated 11da, in agreement with literature data.^{S12} HR-ESI-MS (m/z): calculated for $C_{16}H_{17}O_2$ [M+H]⁺ = 241.1229; found: 241.1220.

Product 11db

Figure S62. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11db from the reaction mixture, in agreement with literature data.^{S12} HR-ESI-MS (m/z): calculated for $C_{16}H_{17}O$ [M+H] = 225.1279; found: 225.1273.

Product 11dd

Figure S63. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11dd from the reaction mixture, in agreement with literature data.^{S12} HR-EI-MS (m/z): calculated for $C_{15}H_{13}FO = 228.0950$; found: 241.1220.

Product 11de

Figure S64. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11de from the reaction mixture, in agreement with literature data.^{S12} HR-ESI-MS (m/z): calculated for $C_{15}H_{13}ClO = 244.0655$; found: 244.0653.

Figure S65. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of the crude product 11de zoomed in to the two aliphatic resonances of the product showing the appearance of other minor overlapping signals attributed to aryl chloride activation.

Product 11df

Figure S66. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated 11df, in agreement with literature data.^{S14} HR-EI-MS (m/z): calculated for C16H13F3O = 278.0918; found: 278.0918.

Product 11dg

Figure S67. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11dg from the reaction mixture, in agreement with literature data.^{S15} Aromatic protons could not be integrated due to uncomplete conversion. HR-EI-MS (m/z): calculated for $C_{20}H_{16}O = 272.1201$; found: 272.1198.

Product 11dh

Figure S68. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11dh from the reaction mixture, in agreement with literature data.^{S16} Aromatic protons could not be integrated due to incomplete conversion. HR-ESI-MS (m/z): calculated for $C_{16}H_{17}O$ [M+H] = 225.1279; found: 225.1274.

Product 11di

Figure S69. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of isolated 11di, in agreement with literature data.^{S17} HR-ESI-MS (m/z): calculated for C14H13O [M+H] = 197.0966; found: 197.0959.

Product 11dj

Figure S70. ¹ H NMR spectrum (MeOD-*d4*, 298 K, 300 MHz) of crude **11dj** from the reaction mixture, in agreement with literature data.^{S13} Aromatic protons could not be integrated due to incomplete conversion. Conversion and yield were assessed in a MeOD-*d4* solution to avoid potential overlap of the compound geminal methyls signal with H₂O in CDCl₃. HR-EI-MS (m/z): calculated for C₁₆H₁₆O [M+H] = 224.1201; found: 224.1197.

Product 11dk

Figure S71. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11dk from the reaction mixture, in agreement with literature data.^{S18} HR-EI-MS (m/z): calculated for C₁₃H₁₈O = 190.1358; found: 190.1352.

Product 11dl

Figure S72. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11dl from the reaction mixture, in agreement with literature data.^{S19} HR-ESI-MS (m/z): calculated for $C_{12}H_{17}O$ [M+H] = 177.1279; found: 177.1272.

Product 11dm

Figure S73. ¹H NMR spectrum (CDCl₃, 298 K, 300 MHz) of crude 11dm from the reaction mixture, in agreement with literature data.^{S20} HR-EI-MS (m/z): calculated for $C_{13}H_{11}NO = 197.0841$; found: 197.0836.

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