Electronic Supplementary Information (ESI)

Photocytotoxic Kinetically Stable Ruthenium(II)-N,N-donor Polypyridyl

Complexes of Oxalate for Anticancer Activity Against HepG2 Liver Cancer Cells

Juhi Sayala^a, Ekta Srivastava^b, Priyaranjan Kumar^a, Nitin Shukla^a, Ashok Kumar^b, Ashis K. Patra^{a*}

Authors address: ^aDepartment of Chemistry, Indian Institute of Technology Kanpur, Kanpur 208016, Uttar Pradesh, India, Email: <u>akpatra@iitk.ac.in</u>

^bDepartment of Biological Science & Bioengineering Indian Institute of Technology Kanpur, Kanpur 208016, Uttar Pradesh, India

^cCentre for Environmental Science and Engineering, Indian Institute of Technology Kanpur, Kanpur 208016, India

^dCenter for Nanosciences, Indian Institute of Technology Kanpur, Kanpur 208016, India

^eThe Mehta Family Centre for Engineering in Medicine, Indian Institute of Technology Kanpur, Kanpur 208016, India

Contents	Page No.
Figure S1. Solid-state FTIR overlay (in KBr pellets) for complexes 1–3.	3
Figure S2. ¹ H NMR of complex 1.	4
Figure S3. ¹ H NMR of complex 2.	5
Figure S4. ¹ H NMR of complex 3.	5
Figure S5. Hydrolysis of complexes 2 and 3 in DMF-Tris buffer medium (pH 7.2).	5
Figure S6. Dark stability of complexes 2 and 3 in DMF using absorption spectroscopy.	6
Figure S7. Time-dependent ¹ H-NMR spectra of the complex 1.	6
Figure S8. Time-dependent ¹ H-NMR spectra of the complex 2.	7
Figure S9. Time-dependent ¹ H-NMR spectra of the complex 3.	7
Figure S10. Photoreactivity of the complex 3 in green light.	8
Figure S11 . Green light irradiated time-dependent ¹ H-NMR spectra of complex 1 .	9
Figure S12 . Green light irradiated time-dependent ¹ H-NMR spectra of complex 2 .	10
Figure S13 . Green light irradiated time-dependent ¹ H-NMR spectra of complex 3 .	11
Figure S14. Measurement of singlet oxygen generation by complex 1 using absorption	12
spectroscopy.	
Figure S15. Measurement of singlet oxygen generation by complex 2 using absorption	13
spectroscopy.	
Figure S16. Absorption spectral Singlet oxygen generation of the complex 3.	14
Figure S17. Measurement of singlet oxygen generation of complexes 1, 2, and 3 using	15
emission spectroscopy.	
Figure S18. DNA binding studies by ethidium bromide (EB) displacement assay for	16
complexes 2 and 3.	
Figure S19. Fluorescence emission spectra of BSA upon addition of complex 2.	17
Figure S20. Fluorescence emission spectra of BSA upon addition of complex 3.	18



Figure S1. An overlay of the solid-state FT-IR spectra of the complexes **1**, **2**, **3** and oxalic acid in KBr pellet.



Figure S2. ¹H NMR spectrum of complex [$Ru^{II}(phen)_2(ox)$] (1) (400 MHz, DMSO-d₆).



Figure S3. ¹H NMR spectrum of complex $[Ru^{II}(dpq)_2(ox)]$ (**2**) (400 MHz, DMSO-d₆).



Figure S4. ¹H NMR spectrum of complex [Ru^{II}(dppz)₂(ox)] (**3**) (400 MHz, DMSO-d₆).



Figure S5. The electronic absorption changes of the complex **2** (a) and **3** (b) (48 μ M) upon solvation for 240 min in the dark in 5% (v/v) DMF-5 mM Tris-HCl/NaCl buffer (pH = 7.2) mixture. Inset: Changes in $A_{272 \text{ nm}}$ and $A_{363 \text{ nm}}$ for complexes **2** and **3**.



Figure S6. The electronic absorption changes of the complexes (48 μ M) **2** (a) and **3** (b) upon solvation for 0–240 min in the dark in DMF. Inset: Changes in A_{λ} for complexes **2** and **3**.



Figure S7. Time-dependent ¹H-NMR spectra of the complex **1** (DMSO-d₆, 500 MHz). (a) Overlay of the region 7.50-8.50 ppm.



Figure S8. Time-dependent ¹H-NMR spectra of the complex **2** (DMSO-d6, 500 MHz). (a) Overlay of the region 7.50-9.50 ppm.



Figure S9. Time-dependent ¹H-NMR spectra of the complex **3** (DMSO-d₆, 500 MHz). (a) Overlay of the region 7.50-9.60 ppm.



Figure S10. The green light (λ_{irrad} = 530 nm) LED (3 V, 158 lm@700 mA) induced spectral changes observed for the complexes **3** in DMF. Absorption spectral traces of complex **3** (24 µM) for the 0-60 min. Inset: Changes in $A_{273 \text{ nm}}$, $A_{362 \text{ nm}}$ and $A_{380 \text{ nm}}$ of complex **3** with photoirradiation time.



Figure S11. Green light (λ_{irr} =530 nm) irradiated Time-dependent ¹H-NMR spectra of the complex **1** (DMSO-d₆, 500 MHz). (a) Overlay of the region 7.50-9.60 ppm. Maroon trace (0 h) and green trace (4 h).



Figure S12. Green light (λ_{irr} =530 nm) irradiated time-dependent ¹H-NMR spectra of the complex **2** (DMSO-d₆, 500 MHz). (a) Overlay of the region 7.50-9.60 ppm. Maroon trace (0 h) and green trace (4 h).



Figure S13. Greenlight (λ_{irr} =530 nm) irradiated Time-dependent ¹H-NMR spectra of the complex **3** (DMSO-d₆, 500 MHz). (a) Overlay of the region 7.50-9.60 ppm. Maroon trace (0 h) and green trace (4 h).



Figure S14. The absorption spectral profile of DPBF (50 μ M) with **1** (10 μ M) in the dark for 10 min shows the dark stability (a). Overlay of A/A_0 at 414 nm for complex **1** with DPBF in the dark and green-light LED (λ_{irr} = 530 nm, 3V, 158 lm@700mA) for 0–10 min (b). Spectra were recorded at 298 K in DMF solution.



Figure S15. Absorption spectral profile of DPBF (50 μ M) with **2** (10 μ M) upon green light irradiation (λ_{irr} =530 nm) for 10 min shows the ${}^{1}O_{2}$ generation (a) and, in the dark for 10 min shows the dark stability (b). Overlay of A/A_{0} at 414 nm for complex **2** with DPBF in the dark and green-light LED (λ_{irr} = 530 nm, 3V 158 lm@700mA) for 0–10 min (c). Spectra were recorded at 298 K in DMF solution.



Figure S16. Absorption spectral profile of DPBF (50 μ M) with **3** (10 μ M) upon green light irradiation (λ_{irr} = 530 nm) for 10 min shows the ${}^{1}O_{2}$ generation (a) and, in the dark for 10 min shows the dark stability (b). Overlay of A/A_{0} at 414 nm for complex **3** with DPBF in the dark and green-light LED (λ_{irr} = 530 nm, 3V, 158 lm@700 mA) for 0–10 min (c). Spectra were recorded at 298 K in DMF solution.



Figure S17. The emission spectral profile of DPBF (50 μ M) with complexes **1**, **2** and **3** (5 μ M) upon green light irradiation (λ_{irr} = 530 nm) for 10 min shows the ${}^{1}O_{2}$ generation (b, d, f) and, in the dark for 10 min shows the dark stability (a, c, e), Green-light LED (λ_{irr} = 530 nm, 3V, 158 lm@700 mA) for 0–10 min (c). Spectra were recorded at 298 K in DMF solution, λ_{exc} =415 nm, λ_{em} =460 nm.



Figure S18. Emission spectrum of ethidium bromide (EB)-bound to DNA in the presence of complexes **2** (a) and **3** (b). ([EB] = 12.5 μ M, [DNA] = 15 μ M, [**2**] = 0-102 μ M, [**3**] = 0-45 μ M, λ_{ex} = 546 nm, Ex. and Em. slit width = 10 nm. The arrow shows the intensity change upon increasing complex concentration.



Figure S19. (a) The BSA (2 μ M) binding of the complex **2** (0-15 μ M) in 0.7% (v/v) DMF-5 mM Tris-HCl/NaCl buffer (pH = 7.2) at 298 K, $\lambda_{ex/em}$ = 295/345 nm and slit width = 10/5. (b) The Scatchard plot for the determination of the static equilibrium binding constant from the intercept and number of binding sites available (*n*) from the slope of the plot. (c) The tyrosine fluorescence emission quenching of BSA upon increasing concentration of complex **2** using synchronous fluorescence studies. (d) The tryptophan fluorescence emission quenching of BSA upon increasing concentration of complex **2** using synchronous fluorescence studies.



Figure S20. (a) The BSA (2 μ M) binding of the complex **3** (0-15 μ M) in 0.7% (v/v) DMF-5mM Tris-HCl/NaCl buffer (pH=7.2) at 298 K, $\lambda_{ex/em}$ = 295/345 nm and Ex./Em. slit widths= 10/5 nm. (b) The tyrosine fluorescence emission quenching of BSA upon increasing concentration of complex **3** using synchronous fluorescence studies. (d) The tryptophan fluorescence emission quenching of BSA upon increasing concentration of complex **3** using synchronous fluorescence studies.