Supporting Information

A photo-degradable BODIPY modified Ru(II) photosensitizer for safe

and efficient PDT under both normoxic and hypoxic conditions

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1. Experimental section

Synthesis and characterization of [Ru(dip)₂(tpy-BODIPY)]²⁺ (complex 1)¹

Tpy-BODIPY ligand² and [Ru(dip)₂Cl₂]³ were synthesized following the reported methods. ¹H NMR (400 MHz, CDCl₃) for tpy-BODIPY: δ 8.70 (s, 4H), 8.55 (s, 2H), 7.91 (t, *J* = 8 Hz, 2H), 7.38 (s, 2H), 5.99 (s, 2H), 2.58 (s, 6H), 1.53 (s, 6H). HR ESI-MS: Calculated m/z for M + H⁺: 480.2166, found: 480.2154.

Under a N₂ atmosphere, [Ru(dip)₂Cl₂] (100 mg, 0.12 mmol) and 26 mg tpy-BODIPY (58 mg, 0.12 mmol) were dissolved in methanol/H₂O (7:1) and refluxed at 65 °C for 10 h. After the reaction was complete, the solution was filtered and the solvent was evaporated under reduced pressure. The crude product was purified by silica gel column chromatography using CH₃CN/saturated KNO₃ aqueous solution (80:1) as the eluent. The red solid complex 1 was obtained with a yield of 52%. ¹H NMR (400 MHz, CD₃CN) δ 9.81 (s, 1H), 8.83 (s, 1H), 8.72 (d, J = 8.1 Hz, 1H), 8.35–8.26 (m, 2H), 8.15 (m, 4H), 8.03 (m, 2H), 7.79 (d, J = 7.1 Hz, 2H), 7.73 (m, 9H), 7.65 (d, J = 5.7 Hz, 1H), 7.63–7.55 (m, 7H), 7.53 (d, J = 7.4 Hz, 2H), 7.47 (d, J = 7.3 Hz, 2H), 7.43 (s, 1H), 7.35 (dd, J = 15.7, 5.9 Hz, 2H), 7.20 (t, J = 7.4 Hz, 1H), 7.12 (m, 2H), 6.88 (d, J = 5.5 Hz, 1H), 6.41 (s, 1H), 6.20 (s, 1H), 6.08 (s, 1H), 2.49 (d, J = 8.7 Hz, 6H), 1.73 (s, 3H), 0.96 (s, 3H). 13 C NMR (101 MHz, CD₃CN) δ 164.99, 159.65, 157.43, 156.97, 156.60, 156.03, 155.80, 152.23, 151.93, 151.09, 149.02, 148.87, 148.74, 147.99, 147.94, 147.73, 146.79, 146.71, 144.27, 143.23, 142.21, 137.53, 136.55, 135.67, 135.41, 135.23, 135.18, 129.89, 129.70, 129.60, 129.45, 129.37, 129.31, 129.23, 128.96, 128.88, 128.81, 128.73, 128.64, 128.31, 128.04, 127.49, 127.22, 126.00, 125.95, 125.63, 125.59, 125.53, 125.39, 125.31, 125.23, 124.31, 123.14, 122.32, 121.93, 121.75, 18.58, 14.74, 14.07, 13.53. HR ESI-MS: Calculated m/z for M^{2+} : 622.6876, found: 622.6870. HPLC purity > 95%.

Synthesis and characterization of [Ru(dip)₂(tpy)]²⁺ (complex 2)

Complex 2 was synthesized with a procedure similar to that of complex 1. Yield: 86%. ¹H NMR (400 MHz, CD₃CN) δ 8.73 (dd, *J* = 12, 8 Hz, 2H), 8.29 (d, *J* = 4 Hz, 1H), 8.20 (m, 4H), 8.16–8.09 (dd, *J* = 8, 4 Hz, 2H), 8.02 (d, *J* = 8 Hz, 1H), 7.92 (d, *J* = 4 Hz, 1H), 7.77 (d, *J* = 8 Hz, 1H), 7.75–7.55 (m, 20H), 7.53–7.46 (m, 4H), 7.39–7.29 (m, 3H), 7.26 (d, *J* = 8 Hz, 1H), 7.08 (s, 1H), 6.92 (d, *J* = 4 Hz, 2H). ¹³C NMR (101 MHz, CD₃CN) δ 164.99, 159.65, 157.43, 156.97, 156.60, 156.03, 155.80, 152.23, 151.93, 151.09, 149.02, 148.87, 148.74, 147.99, 147.94, 147.73, 146.79, 146.71, 144.27, 143.23, 142.21, 137.53, 136.55, 135.67, 135.41, 135.23, 135.18, 129.89, 129.70, 129.60, 129.45, 129.37, 129.31, 129.23, 128.96, 128.88, 128.81, 128.73, 128.64, 128.31, 128.04, 127.49, 127.22, 126.00, 125.95, 125.63, 125.59, 125.53, 125.39, 125.31, 125.23, 124.31, 123.14, 122.32, 121.93, 121.75, 18.58, 14.74, 14.07, 13.53. HR ESI-MS: Calculated m/z for M²⁺: 499.6306, found: 499.6286. HPLC purity > 95%.

Theoretical calculations

All calculations were carried out with the Gaussian 09 (G09) program package 3, using the density functional theory (DFT) method with Becke's three-parameter hybrid functional and LeeYang-Parr's gradient corrected correlation functional (B3LYP).⁴ The LANL2DZ basis set and effective core potential were used for the Ru atom,⁵ and the 6-31 G** basis set was used for remaining atoms.^{6, 7} The ground-state geometry of the complex was optimized in H₂O using the conductive polarizable continuum model (CPCM), and frequency calculation was performed to verify that the optimized structure was in an energy minimum state. The orbitals were analyzed and plotted using the Gauss View 5.0 program.



Figure S1. ¹H NMR spectrum of tpy-BODIPY in CD₃Cl.

2. ¹H NMR, ¹³C NMR and mass spectra



Figure S2. ¹H NMR spectrum of complex 1 in CD₃CN.



Figure S3. ¹H NMR spectrum of complex 2 in CD₃CN.



170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 Figure S4. 13 C NMR spectrum of complex 1 in CD₃CN.



Figure S5. ¹³C NMR spectrum of complex 2 in CD₃CN.



Figure S6. ESI mass spectrum of tpy-BODIPY in CH_3CN (calculated m/z for M+H⁺: 480.2166, found: 480.2154).



Figure S7. ESI mass spectrum of complex 1 in CH_3CN (calculated m/z for M^{2+} : 622.6876, found: 622.6870).



Figure S8. ESI mass spectrum of complex 2 in CH_3CN (calculated m/z for M^{2+} : 499.6306, found: 499.6286).

3. Photo-stability of complexes 1 and 2



Figure S9. Absorption spectra changes of complex 2 upon irradiation (470 nm, 22.5 mW cm⁻²) in CH_3CN .



Figure S10. ESI mass spectrum of complex 2 after irradiation (470 nm, 22.5 mW cm⁻², 30 min) in CH₃CN. The m/z peak at 424.1090 can be ascribed to the compound that the tpy ligand was replaced by two CH₃CN. Calculated m/z for $[Ru(dip)_2(CH_3CN)_2]^{2+}$: 424.1095, found: 424.1090; Calculated m/z for $[tpy + H^+]$: 234.1031, found: 234.1020.



Figure S11. Absorption spectra changes of complex 1 upon irradiation (470 nm, 22.5 mW cm⁻²) in CH_3CN .



Figure S12. ESI mass spectrum of complex 1 after irradiation (470 nm, 22.5 mW cm⁻², 30 min) in CH₃CN. No free tpy-BODIPY ligand and $[Ru(dip)_2(CH_3CN)_2]^{2+}$ based peaks were detected.



Figure S13. ¹H NMR spectra of complex 2 upon irradiation (470 nm, 22.5 mW cm⁻²) with different times in CD₃COCD₃/D₂O = 2:1. The spectrum of free tpy ligand was also displayed for comparison. ∇ represents the free tpy ligand based peaks.



Figure S14. ¹H NMR spectra of complex 1 upon irradiation (470 nm, 22.5 mW cm⁻²) with different times in CD_3COCD_3/D_2O = 2:1. The spectrum of free tpy-BODIPY ligand was also provided for comparison.

4. Theoretical calculations



Fig. S15. The optimized structure of (a) complex 1 and (b) complex 2 based on Gaussian calculation.

1		2	2		
Bond	Length	Bond	Length		
1Ru-17N	2.190	1Ru-17N	2.201		
1Ru-22N	2.100	1Ru-94N	2.104		
1Ru-64N	2.120	1Ru-57N	2.101		
1Ru-75N	2.124	1Ru-46N	2.123		
1Ru-106N	2.099	1Ru-15N	2.118		
1Ru-117N	2.111	1Ru-2N	2.108		

Table S1. Selected bond length (Å) of complexes 1 and 2^a

^a The data in the table were obtained by theoretical calculation.



Figure S16. Other selected frontier molecular orbitals of complex 1

 Table S2. Singlet state transitions of complex 1 based on TD-DFT calculation (H = HOMO, L = LUMO).

Singlet Excited	Energy	Wavelength	Oscillator	Calculated transitions
state	(eV)	(nm)	Strength(f)	and Orbital contributions
S ₁	2.51	494.74	0.0049	H - 1→L + 1 (86%)
S_2	2.60	476.06	0.0059	H→L + 1 (97%)
S_3	2.70	459.63	0.0045	H - 2→L + 1 (65%)
S_4	2.71	457.63	0.0022	H - 1→L + 3 (55%)
S_5	2.73	454.19	0.0071	H - 1→L + 2 (75%)
S_6	2.85	434.62	0.1328	H - 3→L + 1 (75%)
S_7	2.86	433.60	0.0682	H - 1→L (77%) H→L (20%)

Triplet Excited	Energy	Wavelength	Oscillator	Calculated transitions
state	(eV)	(nm)	Strength(f)	and Orbital contributions
T ₁	1.47	842.73	0.0000	H→L (100%) ³ BODIPY*
T ₂	2.33	531.81	0.0000	H - 1→L + 1 (71%) ³ MLCT
T ₃	2.42	510.94	0.0000	H - 1→L + 3 (36%)
T_4	2.45	504.87	0.0000	H - 1→L + 2 (18%)
T ₅	2.49	497.42	0.0000	H - 1→L + 2 (23%)

Table S3. Triplet state transitions of complex 1 based on TD-DFT calculation (H = HOMO, L =LUMO).



Fig. S17. Selected frontier molecular orbitals of tpy-BODIPY.

Singlet Excited	Energy	Wavelength	Oscillator	Calculated transitions
state	(eV)	(nm)	Strength(f)	and Orbital contributions
S_1	2.90	427.72	0.5776	H→L (98%)
S_2	3.31	374.39	0.0332	H - 1→L (92%)
S_3	3.39	365.81	0.0085	H→L + 1 (98%)

Table S4. Three minimum singlet state transitions of tpy-BODIPY based on TD-DFT calculation (H= HOMO, L = LUMO)

Table S5. Three minimum triplet state transitions of tpy-BODIPY based on TD-DFT calculation (H = HOMO, L = LUMO).

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Triplet Excited	Energy	Wavelength	Oscillator	Calculated transitions
state	(eV)	(nm)	Strength(f)	and Orbital contributions
T ₁	1.51	818.66	0.0000	H→L (100%)
T ₂	2.71	457.11	0.0000	H - 2→L (81%)
T ₃	2.89	428.62	0.0000	H→L + 2 (93%)

5. ROS generation



Figure S18. (a) The mechanism of 9,10-anthracenediyl-bis(methylene)-dimalonic acid (ABDA) as the ${}^{1}O_{2}$ probe; Absorption spectra changes of ABDA in acetonitrile upon irradiation (470 nm, 22.5 mW cm⁻²) for different time in the presence of complex 2 (b), tpy-BODIPY (c) and [Ru(bpy)₃]²⁺ (d).



Figure S19. (a) The mechanism of dihydroethidium (DHE) as the $O_2^{-\bullet}$ probe; (b) The quenching mechanism of $O_2^{-\bullet}$ by 1,4-benzoquinone; The fluorescence spectra change of DHE solutions containing 500 µg/mL ctDNA upon irradiation (470 nm, 22.5 mW cm⁻²) for different time in the presence of complex 2 (5 µM) (c), tpy-BODIPY (5 µM) (d), complex 1 (5 µM) and 1,4-benzoquinone (100 µM) (e).



Figure S20. (a) The mechanism of DHR 123 as the $O_2^{-\bullet}$ probe; The fluorescence spectra of DHR 123 in H₂O upon irradiation (470 nm, 22.5 mW cm⁻²) for different time in the presence of complex 1 (5 μ M) (b), complex 2 (5 μ M) (c) and none (control, d).



Figure S21. (a) Fluorescence spectra of DHE (20 μ M) after irradiation (470 nm, 22.5 mW cm⁻²) for different times in the presence of complex 1 (0.1 μ M) in PBS; (b) Fluorescence spectra of DHE (20 μ M) incubated with increasing concentrations of xanthine (0-50 μ M) in the presence of xanthine oxidase (0.005 U ml⁻¹) in PBS. (c) The calibration curve of DHE fluorescence intensity to actual O₂^{-•} concentrations which was constructed using the xanthine/xanthine oxidase system combined with ferricytochrome c.



Figure S22. ESR spectra for detection of ${}^{1}O_{2}$ (a) and $O_{2}^{-\bullet}$ (b) in the presence of 2 or tpy-BODIPY in the dark or upon irradiation.



6. Photo-degradation of complex 1

Figure S23. Absorption spectral changes of complex 1 upon irradiation (470 nm, 22.5 mW cm⁻²) in solutions with different proportions of water and acetone. (a) 70% H₂O; (b) 80% H₂O; (c) 85% H₂O; (d) 90% H₂O; (e) 95% H₂O; (f) Plots of Δ Abs (A₀-A) at 506 nm of complex 1 in different solutions.



Figure S24. ¹H NMR spectra of complex 1 before (top) and after irradiation (470 nm, 22.5 mW cm⁻²) for 10 min in H_2O (bottom). The spectra were recorded in CD_3CN . After irradiation, BODIPY based peaks at 6.20, 6.08, 1.73 and 0.96 ppm disappeared.



Figure S25. ESI mass spectrum of complex 1 in H_2O after light-irradiation (470 nm, 22.5 mW cm⁻², 10 min).



Figure S26. (a) Absorption spectral changes of complex 1 in degassed H_2O upon light-irradiation; (b) Plots of ΔAbs (A_0 -A) at 506 nm of complex 1 in H_2O upon light-irradiation for different time intervals under normoxic and deoxygenated conditions.



Figure S27. Absorption spectral changes of ABDA in H_2O after irradiation (470 nm, 22.5 mW cm⁻²) for different time in the presence of complex 1 (a) and pre-irradiated (10 min in H_2O) complex 1 (b); (c) The fluorescence spectral changes of DHR 123 in H_2O after irradiation (470 nm, 22.5 mW cm⁻²) for different time in the presence of pre-irradiated complex 1 (10 min in H_2O).

7. Experimental data in vitro



Figure S28. The *n*-octanol/water partition coefficients of complexes 1 and 2.



Figure S29. Detection of intracellular ROS production by DCFH-DA for A549 cells treated with complex 2 or tpy-BODIPY (0.1 μ M) in the dark or upon irradiation (470 nm, 22.5 mW cm⁻² for 10 min) under normoxic conditions. The scale bars represent 50 μ m.



Figure S30. Detection of intracellular $O_2^{-\bullet}$ by DHE for A549 cells treated with complex 2 or tpy-BODIPY (0.1 μ M) in the dark or upon irradiation (470 nm, 22.5 mW cm⁻² for 10 min) under normoxic conditions. The scale bar represents 50 μ m.



Figure S31. Cytotoxicity of complex 1 towards A549 cells in the dark under normoxic or hypoxic conditions.



Figure S32. Cytotoxicity of complex 1 towards HeLa (a, c), B16-F10 (b, d) cells in the dark (a, b) or upon irradiation (470 nm, 22.5 mW cm⁻² for 10 min) (c, d) under normoxic or hypoxic conditions.



Figure S33. Cytotoxicity of complex 2 towards A549 (a, d), HeLa (b, e), B16-F10 (c, f) cells in the dark (a, b, c) or upon light-irradiation (470 nm, 22.5 mW cm⁻² for 10 min) (d, e, f) under normoxic or hypoxic conditions.



Figure S34. Calcein-AM and PI co-stained images of A549 3D MCSs treated with 1 or 2 (0.5 μ M) or PBS without irradiation. Scale bars: 250 μ m.

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