

## SUPPLEMENTARY MATERIAL

### **Metabolome biomarkers linking dietary fibre intake with its cardiometabolic effects: Results from the Danish Diet, Cancer and Health - Next Generations MAX study**

Andrea Unión Caballero<sup>1,2#</sup>, Tomás Meroño<sup>1,2#</sup>, Raúl Zamora-Ros<sup>3</sup>, Agnetha Linn Rostgaard-Hansen<sup>4</sup>, Antonio Miñarro<sup>2,5</sup>, Alex Sánchez-Pla<sup>2,5</sup>, Nuria Estanyol-Torres<sup>1</sup>, Miriam Martínez-Huelamo<sup>1,2</sup>, Marta Cubedo<sup>2,5</sup>, Raúl González-Domínguez<sup>1</sup>, Anne Tjønneland<sup>4</sup>, Gabrielle Riccardi<sup>5</sup>, Rikard Landberg<sup>6</sup>, Jytte Halkjær<sup>4\*</sup>, Cristina Andres-Lacueva<sup>1,2\*</sup>

### **Index**

#### **Supplementary discussion: p1-5**

#### **Supplementary tables: p6-7**

Supplementary Table 1. Dietary characteristics of the DCH-NG MAX study population according to tertiles of dietary fibre intake.

#### **Supplementary figures: p8-11**

Supplementary Figure 1. Flowchart of participants and observations included in the study

Supplementary Figure 2. Changes in dietary fibre intake and selected metabolites during the one-year study period.

Supplementary Figure 3. MUVR-PLS analysis showing metabolites associated with dietary fibre intake.

Supplementary Figure 4. Heatmap showing the association between metabolites and food groups in the DCH-NG MAX study.

## Supplementary discussion

The phenolic lipid 2,6-DHBA is one the main metabolites that we found associated with dietary fibre. However, 2,6-DHBA has barely been reported as a biomarker related with dietary fibre consumption, and its specific sources are yet unknown. It has previously been described to be a marker related to the intake of oat and sugar beet fibre (1), and it has also been shown to be a heat stable phenolic compound present in white grape pomace (2). It is thought to be a microbial metabolite primarily derived from alkylresorcinols or lignans fermentation, although the specific microbial enzymes have not been identified in humans yet (3). In a US diet validation study 2,6-DHBA was found to be a reproducible biomarker associated with wholegrain with a reported ICC of 0.62 in evaluations 6 months apart (4). Moreover, following an untargeted plasma metabolomics approach, Johansson-Persson et al. (3) observed an increase in 2,6-DHBA, as well as in 2-aminophenol-sulfate, after a 5-week crossover dietary intervention with a high fibre diet consisting of oat bran, rye bran, and sugar beet fibre in 25 healthy adults. On the other hand, 2-aminophenol-sulfate was also described to be increased in urine in a randomized, controlled crossover trial with an 8-week dietary intervention comparing a wholegrain diet vs. a refined grain diet (5). Nonetheless, in our study, plasma 2,6-DHBA and 2-aminophenol levels were associated between them and with the consumption of wholegrain cereals. Therefore, we may suggest that 2,6-DHBA and 2-aminophenol may be somehow associated with dietary fibre coming from wholegrain cereals; although, we cannot out rule other dietary, host or gut microbial sources.

Indolepropionic acid, a tryptophan metabolite generated by the gut microbiome, has been reported to be associated with the intake of fruits (6), total dietary fibre (7), and higher adherence to different healthy dietary patterns, such as the Healthy Nordic Diet (8), and the French dietary recommendations (9). Indeed, it has been hypothesized that a high intake of dietary fibre could cause a higher production of indolepropionic acid through modifications of gut microbiota composition or activity (10) (11). Importantly, indolepropionic acid levels have been associated with lower likelihood of developing type 2 diabetes, better insulin secretion and low-grade inflammation (10). Given the close relationship between dietary fibre and intake of fruits and vegetables, our results on the association between indolepropionic and dietary fibre intake were mostly confirmatory(12). Nonetheless, herein we report for the first time a moderate reliability and consistency over time of plasma indolepropionic acid levels, with an ICC of 0.52 during a one-year period, underscoring its potential use in more and larger epidemiological studies.

Other metabolites associated with fibre intake in the present study were 3,4-DHBA, linoleoyl-carnitine, ethyl-glucuronide, and proline betaine. To our knowledge, 3,4-DHBA has not been reported as a biomarker associated specifically with dietary fibre. 3,4-DHBA is a phenolic compound widely distributed in many food plants such as olives, white grapes, blackberries and strawberries, but also in beans flours (13) and wholegrain buckwheat (14,15) Accordingly, in our

analysis, 3,4-DHBA was significantly associated with total fruits and legumes. 3,4-DHBA is considered a major metabolite of complex polyphenols, especially anthocyanins (16), and it is thought to be generated by colonic microbiota by B-ring fission of cyanidin-O-glycoside (17). It has been reported to have a strong in vitro and in vivo antioxidant and anti-inflammatory activity (18), to decrease the expression of lipogenic enzymes (19), and to increase glucose uptake in adipocytes (20) among others, but the link of this compound with dietary fibre intake and its metabolic effect is still unclear.

Proline betaine has been consistently proposed as a candidate biomarker for citrus intake (21). Indeed, it has shown to accurately predict orange juice intake measured in 24 h and fasting urine samples, and to provide predictions of citrus intake in good agreement with self-reported intakes (22). Unlike other betaines, proline betaine cannot be synthesized endogenously, and diet is the only source (23). Similar to indolepropionic acid, an observational study showed a positive association between plasma proline betaine levels and higher adherence to the Healthy Nordic Diet (8). In our study, the association between proline betaine and dietary fibre was not consistent across the different time point evaluations. One possible explanation is that in the present study consumption of citrus fruits had a mean contribution of 13% to total fruits intake. However, disregarding its association with dietary fibre, proline betaine levels may reflect a particular role of citrus fruits in host cardiometabolic health that deserves further studies. Linoleoyl-carnitine is another compound significantly associated with dietary fibre in this study. Little is known about its relationship with overall dietary fibre intake. Linoleoyl-carnitine is an acyl-carnitine derived from linoleic acid. Acyl-carnitines play an important role in the mitochondrial energy metabolism, but circulating acyl-carnitines have been speculated to be a marker of detoxification and removal of excess fatty acids (24). Their circulating concentration have also found to be influenced by diet and specific nutrients. Specifically, linoleoyl-carnitine have been associated with the intake of the omega-6 fatty acid linoleic acid, probably coming from vegetable oils used as salad dressings (25). However, in our analysis it was specifically associated with wholegrain cereals, legumes, and vegetables. Last, ethyl-glucuronide has been proven as a biomarker of alcohol consumption (26,27). This negative association could be explained by the fact that plant-foods rich in fibre are not usually linked with alcohol consumption, but rather foods of animal origin as observed in a French cohort (28).

## References:

1. Persson AJ. EFFECTS OF DIETARY FIBRE ON THE HUMAN METABOLISM AND METABOLOME.
2. Mildner-Szkudlarz S, Bajerska J, Zawirska-Wojtasiak R, Górecka D. White grape pomace as a source of dietary fibre and polyphenols and its effect on physical and nutraceutical characteristics of wheat biscuits. *J Sci Food Agric.* 2013 Jan;93(2):389–95.

3. Johansson-Persson A, Barri T, Ulmius M, Önning G, Dragsted LO. LC-QTOF/MS metabolomic profiles in human plasma after a 5-week high dietary fibre intake. *Anal Bioanal Chem* 2013 40514. 2013 Mar;405(14):4799–809.
4. Wang Y, Hodge RA, Stevens VL, Hartman TJ, McCullough ML. Identification and Reproducibility of Plasma Metabolomic Biomarkers of Habitual Food Intake in a US Diet Validation Study. *Metabolites*. 2020 Oct;10(10):1–20.
5. Munch Roager H, Vogt JK, Kristensen M, Hansen LBS, Ibrügger S, Maerkedahl RB, et al. Whole grain-rich diet reduces body weight and systemic low-grade inflammation without inducing major changes of the gut microbiome: A randomised cross-over trial. *Gut*. 2019 Jan 1;68(1):83–93.
6. Pallister T, Jennings A, Mohny RP, Yarand D, Mangino M, Cassidy A, et al. Characterizing Blood Metabolomics Profiles Associated with Self-Reported Food Intakes in Female Twins. *PLoS One*. 2016 Jun;11(6):e0158568.
7. Tuomainen M, Lindström J, Lehtonen M, Auriola S, Pihlajamäki J, Peltonen M, et al. Associations of serum indolepropionic acid, a gut microbiota metabolite, with type 2 diabetes and low-grade inflammation in high-risk individuals. *Nutr Diabetes* 2018 81. 2018 May;8(1):1–5.
8. Noerman S, Kokla M, Koistinen VM, Lehtonen M, Tuomainen TP, Brunius C, et al. Associations of the serum metabolite profile with a healthy Nordic diet and risk of coronary artery disease. *Clin Nutr*. 2021 May;40(5):3250–62.
9. Lécuyer L, Dalle C, Micheau P, Pétéra M, Centeno D, Lyan B, et al. Untargeted plasma metabolomic profiles associated with overall diet in women from the SU.VI.MAX cohort. *Eur J Nutr* 2020 598. 2020 Jan;59(8):3425–39.
10. De Mello VD, Paananen J, Lindström J, Lankinen MA, Shi L, Kuusisto J, et al. Indolepropionic acid and novel lipid metabolites are associated with a lower risk of type 2 diabetes in the Finnish Diabetes Prevention Study. *Sci Reports* 2017 71. 2017 Apr;7(1):1–12.
11. Zhu C, Sawrey-Kubicek L, Beals E, Rhodes CH, Houts HE, Sacchi R, et al. Human gut microbiome composition and tryptophan metabolites were changed differently by fast food and Mediterranean diet in 4 days: a pilot study. *Nutr Res*. 2020 May;77:62–72.
12. Menni C, Hernandez MM, Vital M, Mohny RP, Spector TD, Valdes AM. Circulating levels of the anti-oxidant indolepropionic acid are associated with higher gut microbiome diversity. *Gut Microbes*. 2019 Nov;10(6):688–95.
13. Pérez-Ramírez IF, Becerril-Ocampo LJ, Reynoso-Camacho R, Herrera MD, Guzmán-Maldonado SH, Cruz-Bravo RK. Cookies elaborated with oat and common bean flours improved serum markers in diabetic rats. *J Sci Food Agric*. 2018 Feb;98(3):998–1007.
14. Sedej I, Sakač M, Mandić A, Mišan A, Tumbas V, Čanadanović-Brunet J. Buckwheat (*Fagopyrum esculentum* Moench) Grain and Fractions: Antioxidant Compounds and Activities. *J Food Sci*. 2012 Sep;77(9):C954–9.
15. Juurlink BHJ, Azouz HJ, Aldalati AMZ, Altinawi BMH, Ganguly P. Hydroxybenzoic acid isomers and the cardiovascular system. *Nutr J*. 2014 Jun;13(1):1–10.
16. Vitaglione P, Donnarumma G, Napolitano A, Galvano F, Gallo A, Scalfi L, et al. Protocatechuic Acid Is the Major Human Metabolite of Cyanidin-Glucosides. *J Nutr*. 2007 Sep;137(9):2043–8.
17. Macià A, Romero MP, Yuste S, Ludwig I, Pedret A, Valls RM, et al. Phenol metabolic fingerprint and selection of intake biomarkers after acute and sustained consumption of

- red-fleshed apple versus common apple in humans. The AppleCOR study. *Food Chem.* 2022 Aug;384:132612.
18. Semaming Y, Pannengpetch P, Chattipakorn SC, Chattipakorn N. Pharmacological properties of protocatechuic acid and its potential roles as complementary medicine. *Evidence-based Complement Altern Med.* 2015;2015.
  19. Liu WH, Lin CC, Wang ZH, Mong MC, Yin MC. Effects of Protocatechuic Acid on Trans Fat Induced Hepatic Steatosis in Mice. *J Agric Food Chem.* 2010 Sep;58(18):10247–52.
  20. Scazzocchio B, Vari R, Filesi C, D'Archivio M, Santangelo C, Giovannini C, et al. Cyanidin-3-O- $\beta$ -glucoside and protocatechuic acid exert insulin-like effects by upregulating PPAR $\gamma$  activity in human omental adipocytes. *Diabetes.* 2011 Sep;60(9):2234–44.
  21. Heinzmann SS, Brown IJ, Chan Q, Bictash M, Dumas ME, Kochhar S, et al. Metabolic profiling strategy for discovery of nutritional biomarkers: proline betaine as a marker of citrus consumption. *Am J Clin Nutr.* 2010 Aug;92(2):436–43.
  22. Gibbons H, Michielsen CJR, Rundle M, Frost G, McNulty BA, Nugent AP, et al. Demonstration of the utility of biomarkers for dietary intake assessment; proline betaine as an example. *Mol Nutr Food Res.* 2017 Oct;61(10):1700037.
  23. Servillo L, D'Onofrio N, Giovane A, Casale R, Cautela D, Ferrari G, et al. The betaine profile of cereal flours unveils new and uncommon betaines. *Food Chem.* 2018 Jan;239:234–41.
  24. Reuter SE, Evans AM. Carnitine and Acylcarnitines. *Clin Pharmacokinet* 2012 519. 2012 Dec;51(9):553–72.
  25. Wedekind R, Rothwell JA, Viallon V, Keski-Rahkonen P, Schmidt JA, Chajes V, et al. Determinants of blood acylcarnitine concentrations in healthy individuals of the European Prospective Investigation into Cancer and Nutrition. *Clin Nutr.* 2022;41:1735–45.
  26. Müller A, Iwersen-bergmann S, Brandy F, Brandy F. Ethyl Glucuronide in Alcoholic Beverages. 2018;53(May):532–8.
  27. Kharbouche H, Sporkert F, Staub C, Mangin P, Augsburg M. L ' éthylglucuronide : un marqueur de la consommation d ' alcool. 2009;1299–306.
  28. Kesse E, Clavel-Chapelon F, Slimani N, Van Liere M. Do eating habits differ according to alcohol consumption? Results of a study of the French cohort of the European Prospective Investigation into Cancer and Nutrition (E3N-EPIC). *Am J Clin Nutr.* 2001;74(3):322–7.

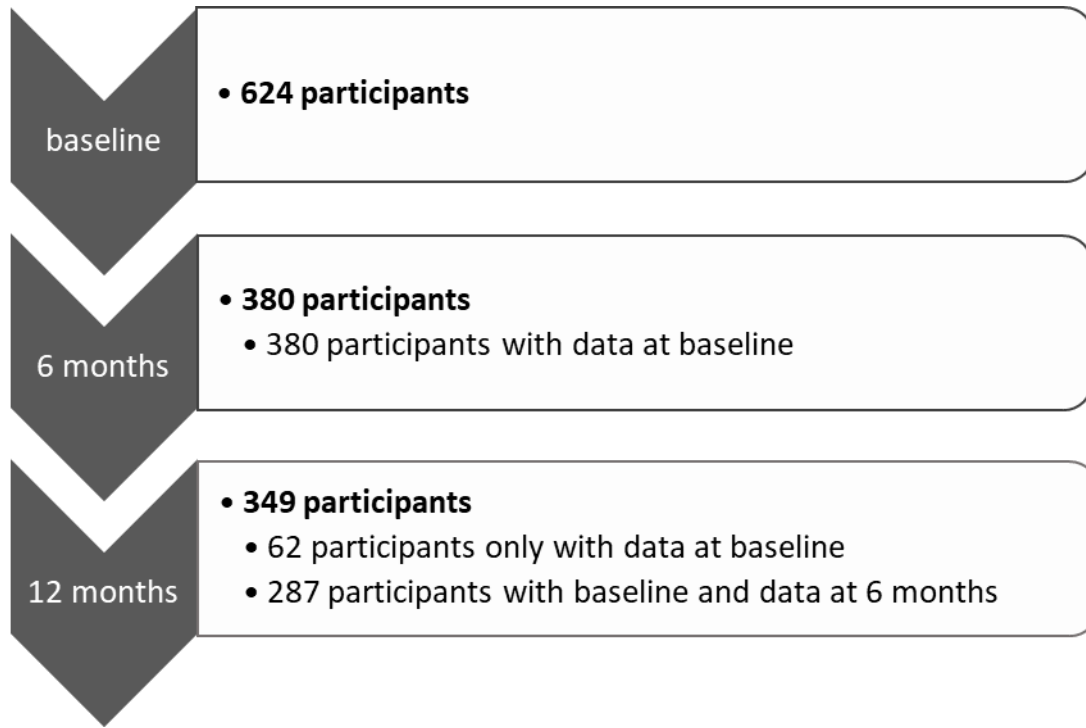
**Supplementary Table 1. Dietary characteristics of the DCH-NG MAX study population according to tertiles of dietary fibre intake.**

	All n= 624 k=1,353	Fibre <16g/day (T1) k= 452	Fibre: 16-25g/day (T2) k= 450	Fibre >25g/day (T3) k= 451
<b>Dietary characteristics</b>				
Energy (10 <sup>3</sup> kcal/d)	2.1 ± 0.8	1.7 ± 0.6	2.1 ± 0.6	2.6 ± 0.9 *
Total fats (% of Energy)	35 ± 9	35 ± 11	36 ± 8	34 ± 8
SFA (% of Energy)	11 ± 5	12 ± 5	12 ± 4	10 ± 4 ***
MUFA (% of Energy)	12 ± 5	12 ± 5	13 ± 5	12 ± 4
PUFA (% of Energy)	6.0 ± 2.8	5.6 ± 3.1	6.0 ± 2.5	6.5 ± 2.5 ***
Total carbohydrates (% of Energy)	43 ± 10	41 ± 12	43 ± 9	45 ± 9 ***
Total sugars (% of Energy)	14 ± 8	14 ± 9	14 ± 7	14 ± 6
Protein (% of Energy)	17 ± 6	18 ± 6	17 ± 5	16 ± 5 ***
Dietary fibre (g/d)	21 ± 10	11 ± 4	20 ± 2	33 ± 8 ***
[g/1000 kcal]	[11 ± 5]	[7 ± 4]	[11 ± 4]	[14 ± 5] ***
Alcohol (g/d)	0 (0 - 13)	0 (0 - 18)	0 (0 - 14)	0 (0 - 14)
Sodium (g/d)	3.0 ± 1.7	2.4 ± 1.4	3.0 ± 1.4	3.7 ± 2.1 **
<b>Food intake</b>				
Cereal whole grain (g/d)	115 (45-200)	45 (0-90)	122 (70-185)	204 (129-285) ***
Cereal refined (g/d)	45 (0-125)	45 (0-120)	50 (0-142)	40 (0-124)
Fruits (g/d)	150 (0-313)	100 (0-250)	148 (3-270)	210 (54-400) ***
Vegetables (g/d)	140 (30-290)	74 (0-170)	133 (20-265)	255 (130-411) ***
Legumes (g/d)	0 (0-0)	0 (0-0)	0 (0-0)	0 (0-0) *
Nuts and seeds (g/d)	0 (0-10)	0 (0-0)	0 (0-8)	0 (0-30) ***

SFA, saturated fatty acids; MUFA, monounsaturated fatty acids; PUFA, polyunsaturated fatty acids. Mean and standard deviation, or median (Q1-Q3) were used to describe continuous variables following a Gaussian or skewed distribution, respectively. \*p for trend<0.05, \*\* p for trend<0.01, \*\*\* p for trend<0.001 using age- and sex-adjusted generalized linear models.

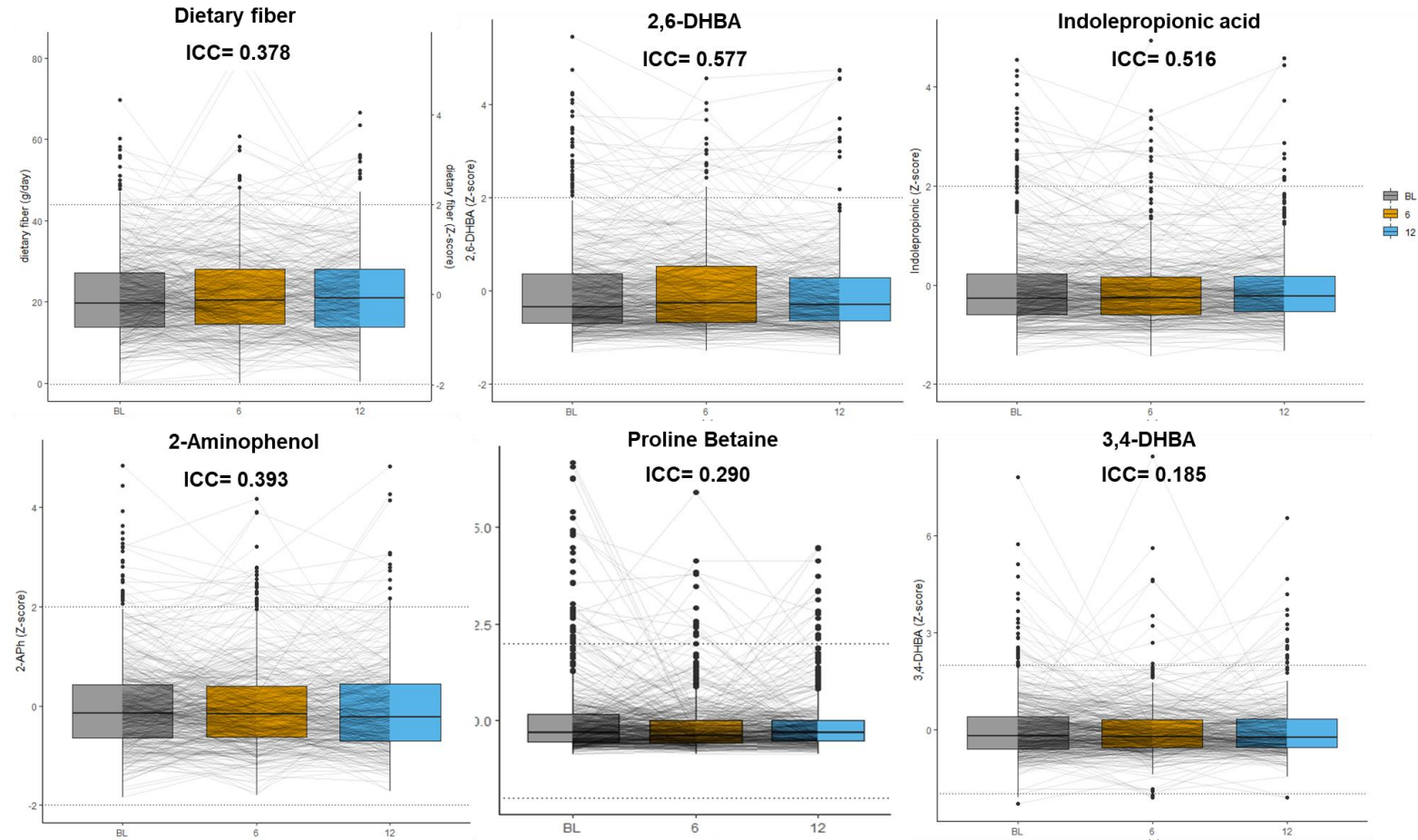
Variables with skewed distribution were log-transformed before entering the analyses.  
n=participants, k=observations.

**Supplementary Figure 1. Flowchart of participants at each time-point.**

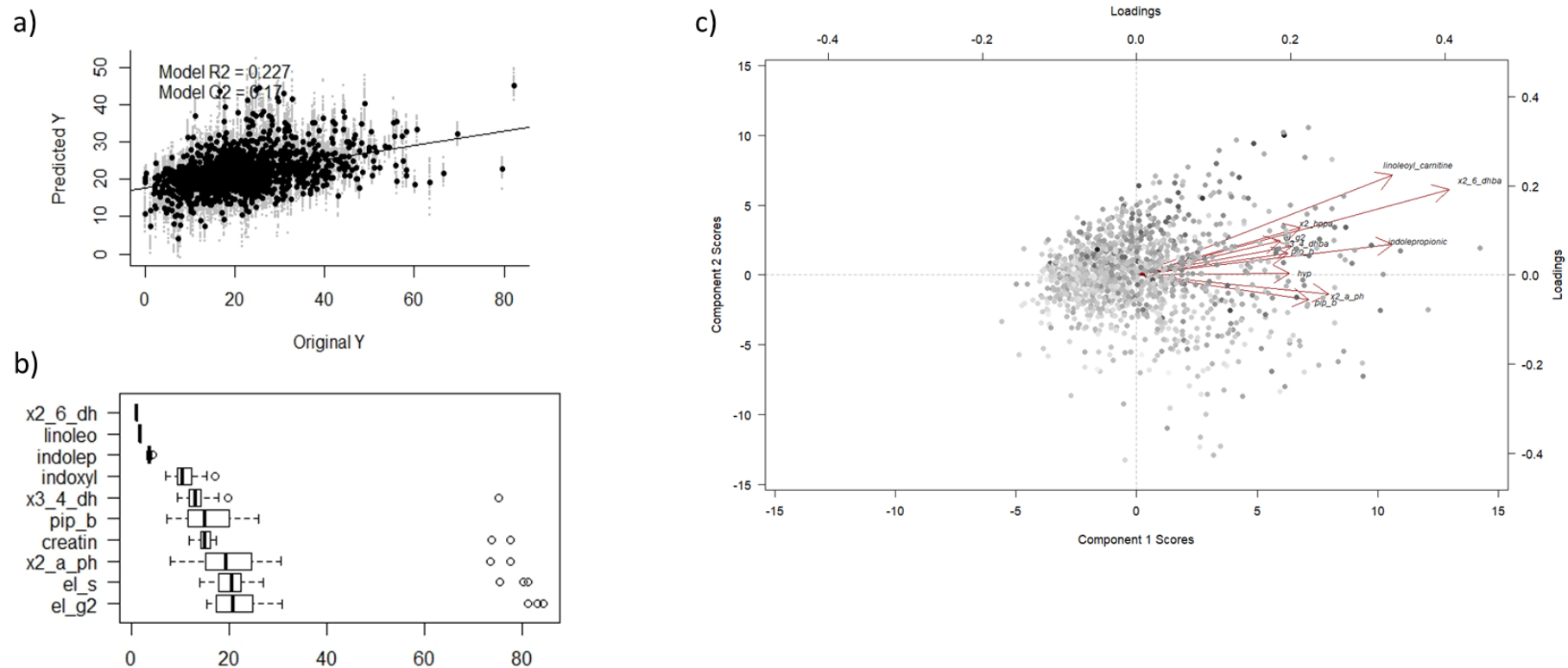




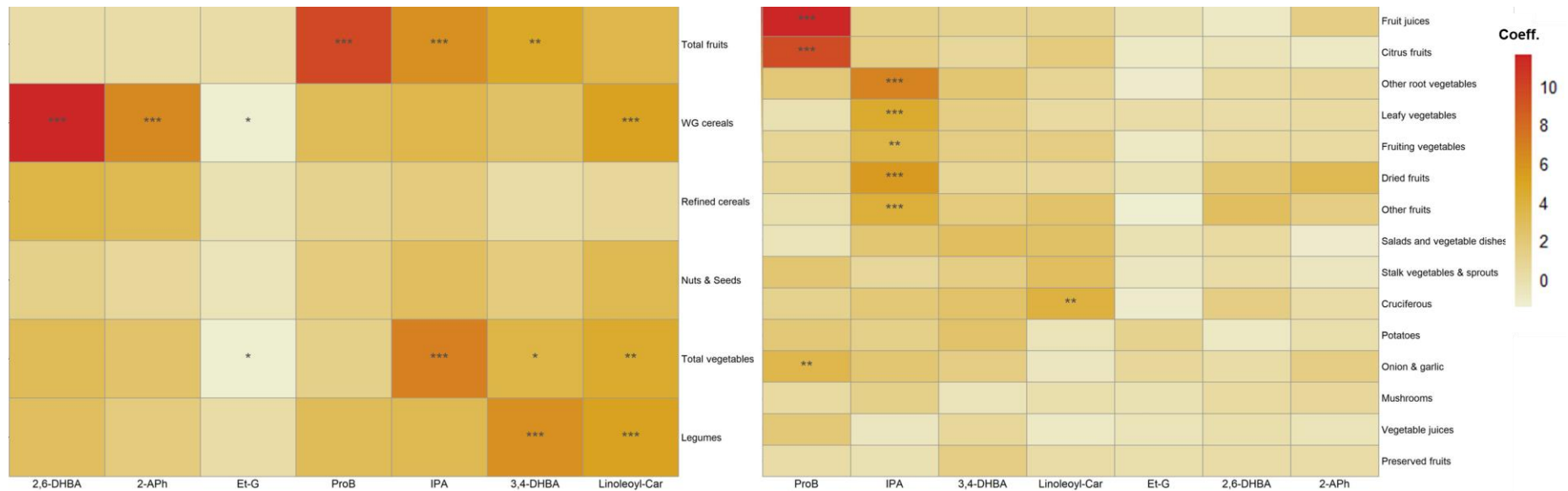
Supplementary Figure 2. Changes in dietary fibre intake and selected metabolites during the one-year study period.



Box plots and spaghetti plots including all the data from all the participants available ( $n=624$ ,  $k=1,353$ ). Intraclass correlation coefficients were calculated from linear mixed models adjusted for age, sex, BMI and time with random intercepts.  $n$ =participants,  $k$ =observations



**Supplementary Figure 3. MUVR-PLS analysis showing metabolites associated with dietary fibre intake (n=624, k=1,353).** a) Predicted fibre intake and self-reported fibre intake according to MUVR-PLS. b) Boxplot of the variable importance rank per-repetition ( $p < 0.0001$ ). Lower rank means higher importance. c) Loading plot of the PLS model. x2\_6\_dh, 2,6-dihydroxybenzoic acid; linoleo, linoleoyl carnitine; indolep, indolepropionic acid; indoxyl, indoxyl sulfate; x3\_4\_dh, 3,4-dihydroxybenzoic acid; pip\_b, pipecolic acid betaine; x2\_a\_ph, 2-aminophenol; el\_s, enterolactone-sulfate; el\_g2, enterolactone-glucuronide.



**Supplementary Figure 4. Heatmap showing the association between metabolites and food groups in the DCH\_NG MAX study.** Coefficients calculated using linear mixed models in the whole study population (n=624, k=1,353). 2,6-DHBA, 2,6-dihydroxybenzoic acid; 2-Aph, 2-Aminophenol; 3,4-DHBA, 3,4-dihydroxybenzoic acid; IPA, indolepropionic acid; ProB, proline betaine; Car, carnitine; Et-G, ethyl-glucuronide. \*FDR-adjusted p-value<0.05, \*\* FDR-adjusted p-value<0.01, \*\*\* FDR-adjusted p-value <0.001 using age-, sex- and time-adjusted linear mixed models with random intercepts. n=participants, k=observations. Contributions to total fibre intake of the major food groups were: Total fruits (12%), WG cereals (41%), refined cereals (12%), Nuts & Seeds (5%), Total vegetables (22%), and Legumes (1%).