Supporting Information

Matrix metalloproteinase responsive hydrogel microplates for programmed killing of invasive tumour cells

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Template: 1cm x 1cm x 0.5cm

Figure S1. Macroscale hydrogel template utilized in this study.



Figure S2. SEM images of lyophilized macroscopic hydrogels from different gel precursor initial concentrations (scale bars left panels: 100 µm, scale bars right panels: 10 µm).



Figure S3. Fabrication and characterization of hydrogel microplates using soft-lithography method, i) Silicon master template with electron beam etched features, ii) PDMS inverse template, iii) Sacrificial PVA replica template, iv) PVA template filled with hydrogel precursor solution and crosslinked with 365 nm UV light, v) dissolution of PVA template in water and purification of hydrogel microplates



Figure S4. Equilibrium swelling ratios (mass) of macroscopic polyethylene glycol peptide thiol-ene hydrogels of varying stiffness (error bar represents S.D. of three measurements).



Figure S5. Size distribution of microplates in solution from optical microscopy, and associated 10 wt% microgel swelling characterization, in comparison to PVA template idealized particle sizes $(20 \times 20 \times 10 \ \mu\text{m})$. Scale bar: 100 μm .



Figure S6. Macroscale hydrogel mass degradation (varying stiffness from different hydrogel precursor initial concentrations) in 50 nM solutions of MMP-2/9 (collagenase IV) at 37 °C over time.



Figure S7. Optical microscopy images of hydrogel microplates and their degradation in either presence (bottom '+ line') or absence (top '- line') of 50 nM concentrations of MMP-2/9 (collagenase IV) at 37 °C over time (scale bars: 100 μ m).



Figure S8. a) DLS size distribution of DTXL containing PLGA SPNs in water, b) Zeta potential distribution of DTXL containing PLGA SPNs in water.