

## Supporting Information

### Iron (III) Oxide anchored conductive polymer-graphene ternary nanocomposite decorated disposable paper electrodes for non-enzymatic detection of serotonin

*Sharmila Prashanth<sup>a</sup>, Raifa Abdul Aziz<sup>b</sup>, Shamprasad Varija Raghu<sup>b,c</sup>, Yoon-Bo Shim<sup>d</sup>, K. Sudhakara Prasad<sup>a,\*</sup>,  
Airody Vasudeva Adhikari<sup>a,e,\*</sup>*

<sup>a</sup> Nanomaterial Research Laboratory (NMRL), Nano Division, Yenepoya Research Centre and Centre for Nutrition Studies, Yenepoya (Deemed to be University), Deralakatte, Mangalore 575018, India

<sup>b</sup> Neurogenetics Lab, Department of Applied Zoology, Mangalore University, Mangalagangothri, Mangalore 574199, Karnataka, India

<sup>c</sup> Division of Neuroscience, Yenepoya Research Centre, Yenepoya (Deemed to be University), Deralakatte, Mangalore 575018, India

<sup>d</sup> Department of Chemistry and Institute of Biophysio Sensor Technology, Pusan National University, Busan 46241, Republic of Korea

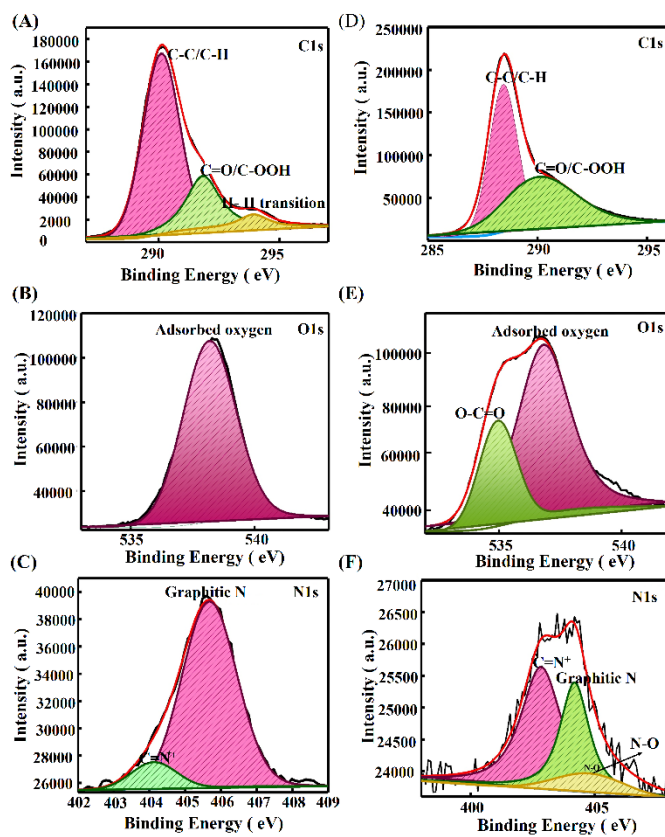
<sup>e</sup> Department of Chemistry, National Institute of Technology Karnataka, Surathkal, Mangalore 575025, India

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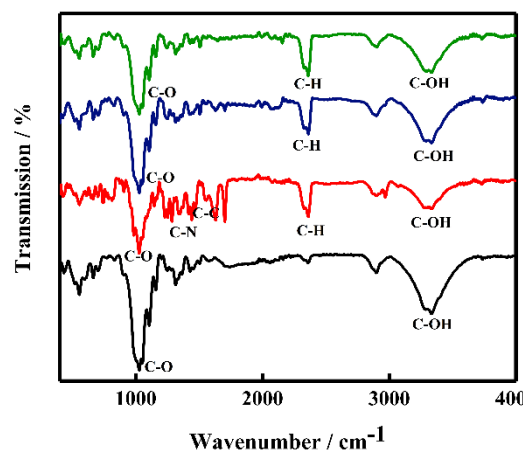
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## RESULTS AND DISCUSSIONS

### XPS characterization of the sensor

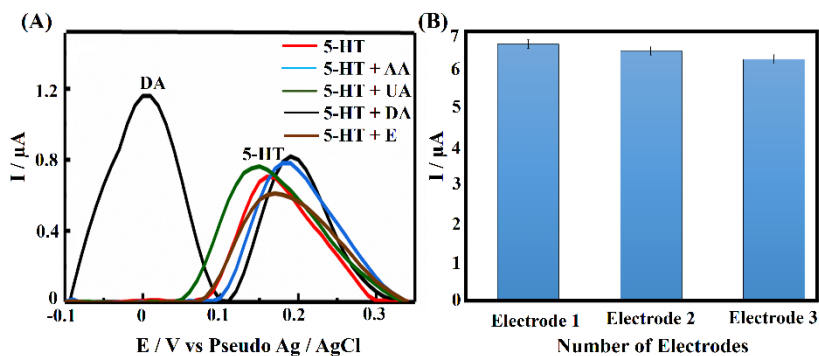


**Figure S1.** Deconvoluted C 1s, O 1s, N 1s XPS spectrum for **(A)** PPE-P (py) (A, B, and C) and **(B)** PPE-P (py) - rGO (D, E, and F).



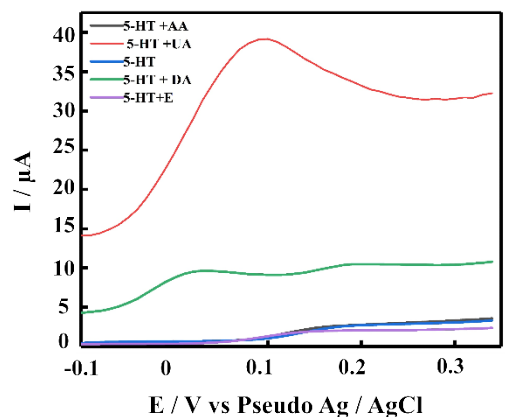
### FTIR characterization of the sensor

**Figure S2** FTIR spectrum obtained for PPE (black), PPE-P(py) (red), PPE-P(py)-rGO (blue), and PPE-P(py)-rGO-Fe<sub>2</sub>O<sub>3</sub> (green).

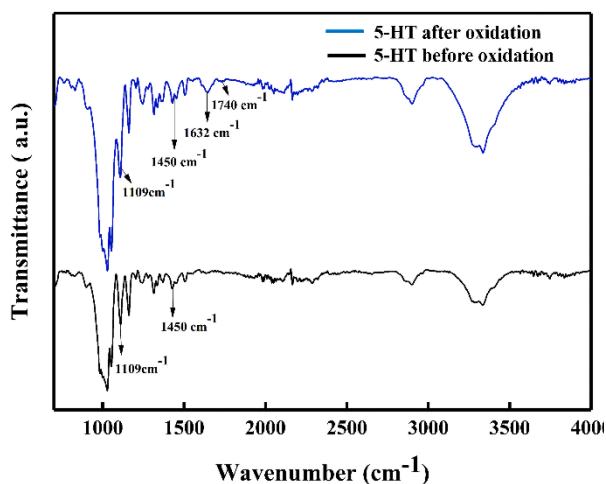


### Selectivity and stability studies

**Figure S3 (A)** Background subtracted DPV response for selectivity studies conducted for 5-HT at a concentration of 75 μM, with 75 μM concentrations of ascorbic acid (AA), uric acid (UA), dopamine (DA), and epinephrine (E) **(B)** Average current response for four electrodes recorded using DPV for 500 μM 5-HT.



**Figure S4** Raw DPV response for selectivity studies conducted for 5-HT at a concentration of 75 μM, with 75 μM concentrations of ascorbic acid (AA), uric acid (UA), dopamine (DA), and



epinephrine (E) without background subtraction

**Figure S5** FTIR spectrum obtained for 5-HT before oxidation (blue) and after oxidation (black)

### Real sample analysis by HPLC-UV method

Chromatographic conditions used to determine the concentration of 5-HT are as follows (1).

Solution A: 0.1% formic acid in milli-Q water

Solution B: acetonitrile

Mobile phase: solution A: solution B (90:10%)

Column: shim-pack GIST 5C1g, 4.6\*250 mm

Elution method: Isocratic

Detector wavelength: 280 nm

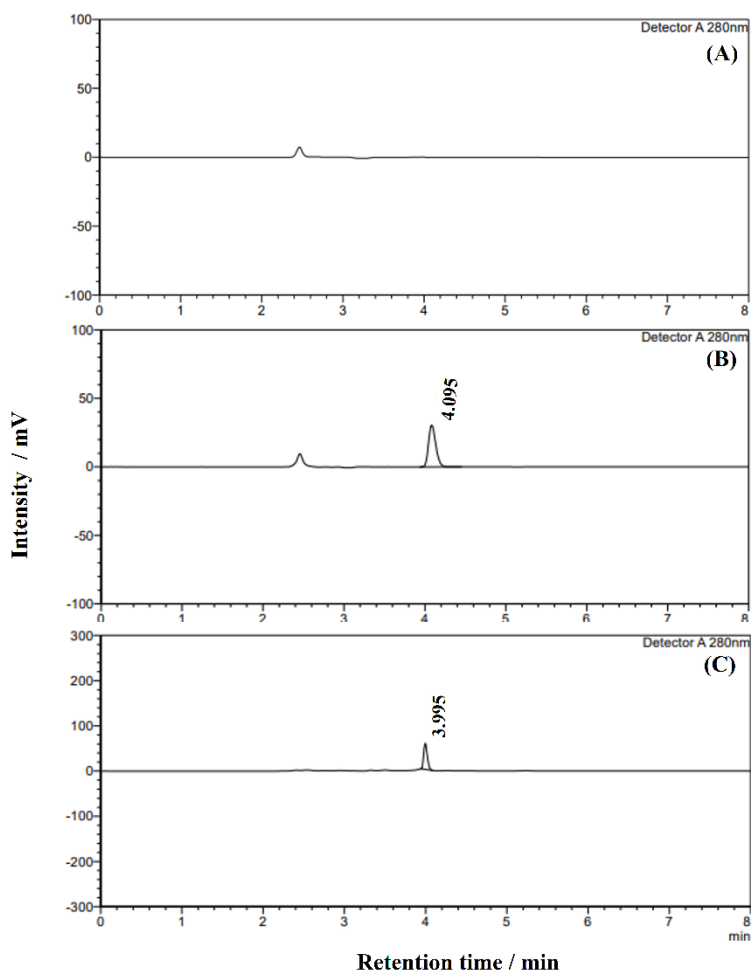
Flow rate: 1.0 mL/min

Column oven temperature: 25°C

Run time: 8 min

Injection volume: 60  $\mu$ L

The concentration of 5-HT in *Drosophila melanogaster* was determined by comparing the peak area obtained from the real sample analysis with the peak area obtained from the standard 5-HT solution of known concentrations (Figure S4)



**Figure S6(A)** Chromatogram of blank solution **(B)** Chromatogram of standard 5-HT solution **(C)** Chromatogram of real sample solution.

**Table S1. Recovery analysis was carried out by standard addition method.**

Sl. No	Actual conc. ( $\mu\text{M}$ )	Calculated conc. ( $\mu\text{M}$ )	% recovery
1.	25	25.0	100
2.	50	47.22	99.4
3.	75	81.38	104.4
4.	100	96.66	96.7

**Table S2. Comparison of composition, detection limit and linear range of different modified electrodes for the determination of 5-HT.**

Electrode type	Modification	Electrode modification	LOD ( $\mu\text{M}$ )	Linear range ( $\mu\text{M}$ )	Real sample	Ref
GCE	Ag/PPy/Cu <sub>2</sub> O	Sonochemical and oxidative polymerisation	0.0124	0.01-250	-	(2)
SPE	MWCNT-AONP	Hydrothermal method	0.0246	0.016-0.16	Tomato	(3)
GCE	rGO-Co <sub>3</sub> O <sub>4</sub>	Hydrothermal	1.128	1 - 10	-	(4)
GCE	MWCNT-NiO, MWCNT-ZnO MWCNT-Fe <sub>3</sub> O <sub>4</sub>	Drop – dry	0.118 0.129 0.166	0.0059– 62.8	Urine	(5)
GCE	Fe <sub>3</sub> O <sub>4</sub> -MWCNT-poly(BCG)	Electropolymerisation followed by drop-dry method	0.080	0.5 - 100	Serum	(6)
Gold mylar substrate	rGO-PEDOT/PSS-nafion	Drop cast method	0.1	1 - 10	-	(7)
PPE	P(py)-rGO-Fe <sub>2</sub> O <sub>3</sub>	Amperometry, CV	0.022	0.01-500	Brain sample of <i>Drosophila melanogaster</i>	This work

## REFERENCES

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