Supporting Information

Ultrasound-triggered in situ gelation with ROS-controlled drug

release for cartilage repair

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Figure S1. Synthesis and characterization of ROS-responsive precursor. (a-c) The synthesis process and ¹H NMR spectra of the TK linker, TK-mPEG and DSPE-TK-mPEG. (d) The ¹H NMR spectra of the TK broken by H_2O_2 .



Figure S2. The size of a series of nanoparticles measured by DLS. (a-d) The DLS of Lip, LP, LPT and LPK.



Figure S3. The rupture of LPKT NPs triggered by ROS under US treatment for 3 min. (a) TEM image of Lip NPs with US treatment, (b) TEM image of LPKT NPs with US treatment.



Figure S4. Swelling ratio of nanocomposite hydrogels from initial gelation state to swelling state on the basis of wet hydrogels.



Figure S5. The degradation effect of *in situ* nanocomposite hydrogels in the presence of MMP-13. (a) the overall degradation profile of FT, FLPT+US, and FLPKT+US hydrogels in 25 ng/mL MMP-13 solution for 28 days. (b) The weight rate of in situ nanocomposite hydrogels with or without MMP-13 enzyme at day 28.



Figure S6. Average sizes of large and small pores in the FT, FLPT+US and FLPKT+US hydrogels. (*p < 0.05).



Figure S7. The cell viability of FLPKT+US hydrogels after being cultured for 24 h.



Figure S8. Ultrasonic penetration and gelation capabilities through the pig tissues. (a) The thickness of pig tissues. (b) The effect of surrounding tissues on the capability of gelation under the ultrasonic treatment with 1 MHz, 1 W/cm² for 3 min.



Figure S9. The toxicity of *in situ* nanocomposite hydrogel at 8 weeks. (Scale bar: 500 μ m).

Table S1. Primer sequences for qRT-PCR.

Gene symbol	5'-3'
GAPDH F	GGTTGTCTCCTGCGACTTCA
GAPDH R	TGGTCCAGGGTTTCTTACTCC
ACAN F	AACTCAGTGGCCAAACATCC
ACAN R	TCAGGAATCCCAGATGTTCC
Col2 F	CTCAAGTCGCTGAACAACCA
Col2 R	GTCTCCGCTCTTCCACTCTG
Sox9 F	TGGCAGAGGGTGGCAGACAG
Sox9 R	CGTTGGGCGGCAGGTATTGG
Sox9 R	CGTTGGGCGGCAGGTATTGG

Western Blot Raw Data and Grouping







COL II









p-mTOR











