

Supporting information

Table A.1. MZmine parameters used in pre-processing MS data analysis.

Method	Parameters	Value
Mass detection MS¹	Mass detector	Centroid
	Noise level	1E ⁷
Mass detection MS²	Noise level	1E ³
	Min. group size in # of scans	4
Chromatogram building (ADAP Chromatogram builder)	Group intensity threshold	3E ⁷
	Min. highest intensity	3E ⁷
Chromatogram deconvolution (Local minimum search algorithm)	<i>m/z</i> tolerance	10 ppm
	Chromatographic threshold	10%
	Search min. in RT range	0.01 min
	Min. relative height	10%
	Min. absolute height	3E ⁷
	Min. ratio of peak top/edge	2
	Peak duration range	0.05 - 2.00 min
	<i>m/z</i> range for MS2 scan pairing	0.01 Da
	RT range for MS2 scan pairing	0.02 min
	<i>m/z</i> tolerance	10 ppm
Isotope grouping (Isotopic peaks grouper)	Retention time tolerance	0.02 min
	Maximum charge	2
Feature alignment (Join aligner)	<i>m/z</i> tolerance	10 ppm
	Weight for <i>m/z</i>	75
	Retention time tolerance	0.05 min
	Weight for RT	25
Isotope filter and others (Feature list rows filter)	Min. peaks in a row	2
	Keep only peaks with MS2 scan	Selected
Gap filling (Peak finder)	Intensity tolerance	10%
	<i>m/z</i> tolerance	10 ppm
	Retention time tolerance	0.05 min

Table A.2. Features detected in Vorax™ data using MZmine 2.53 according to parameters shown in Table A.1.

Sample Label	V1_25.mzXML Vorax 25 min	V2_25.mzXML Vorax 25 min	V1_30.mzXML Vorax 30 min	V2_30.mzXML Vorax 30 min
1/111.0204mz/18.81min	4.64E+08	4.53E+08	287630.3998	215932.7659
2/154.9902mz/18.92min	6.82E+08	7.05E+08	8425.767673	6454.069045
3/110.0202mz/18.90min	7.26E+08	7.24E+08	20964.2658	9543.340144
4/113.9637mz/18.92min	5.03E+08	4.84E+08	3402.150537	1822.245117
5/139.9880mz/18.81min	2.12E+08	2.06E+08	112757.5261	66297.10008
6/102.0340mz/18.84min	2.03E+08	2.08E+08	NA	NA
7/112.0181mz/18.80min	1.60E+08	1.63E+08	86959.79605	91130.41563
8/149.0234mz/18.80min	5.76E+08	5.98E+08	2058551.385	1563215.175
9/279.1590mz/18.79min	5.53E+08	5.89E+08	3872478.106	6534015.295
10/111.0204mz/22.35min	51679.53852	30781.27165	7.87E+08	8.39E+08
11/110.0202mz/22.50min	163931.3796	147936.5471	1.13E+09	1.14E+09
12/154.9903mz/22.50min	318126.9205	482037.643	1.16E+09	1.16E+09
13/139.9881mz/22.36min	8169.156328	18785.8942	3.60E+08	3.69E+08
14/113.9637mz/22.50min	1824060.092	574990.6511	7.11E+08	7.25E+08
15/102.0340mz/22.43min	110795.6055	191195.5768	3.40E+08	3.61E+08
16/112.0181mz/22.35min	9182.209292	33141.90933	2.85E+08	2.93E+08
17/123.0405mz/22.36min	144313.1498	225371.2682	2.33E+08	2.41E+08
18/149.0234mz/22.40min	2.69E+07	2.76E+07	4.70E+08	1.83E+08
19/279.1591mz/22.22min	7.97E+07	1.77E+07	3.56E+08	1.28E+09
20/149.0234mz/22.27min	4.63E+07	3.53E+07	2.41E+08	2.96E+08
21/279.1590mz/22.27min	5.73E+07	1.45E+07	2.32E+08	3.92E+08
22/536.1649mz/16.53min	61532.91271	41737.55105	38567.2023	108063.4168
23/148.0606mz/0.80min	4.11E+08	3.86E+08	3.81E+08	3.35E+08
24/118.0863mz/0.84min	1.03E+08	3.61E+08	2.42E+08	2.09E+08
25/104.1071mz/0.77min	2.63E+08	2.84E+08	2.25E+08	2.20E+08
26/199.1078mz/1.45min	1.17E+08	1.23E+08	1.53E+08	1.14E+08
27/247.1289mz/1.50min	1.64E+08	8.94E+07	1.22E+08	9.22E+07
28/213.1232mz/2.16min	1.48E+08	1.19E+08	1.33E+08	1.39E+08
29/149.0234mz/22.37min	2.69E+07	4.19E+07	1.28E+09	1.89E+08
30/279.1590mz/22.36min	1.69E+07	1.45E+07	1.25E+09	2.23E+08
31/132.1020mz/1.15min	3.67E+08	3.26E+08	2.98E+08	4.04E+08
32/130.0500mz/0.98min	1.99E+08	2.13E+08	3.82E+07	1.18E+08
33/118.0863mz/0.89min	1.79E+08	1.39E+08	8.41E+07	1.11E+08
34/116.0707mz/0.80min	2.74E+08	2.07E+08	2.57E+08	2.40E+08
35/148.0605mz/0.89min	1.14E+08	9.49E+07	5.64E+07	4.28E+07
36/130.0501mz/0.90min	2.91E+08	2.84E+08	9.32E+07	3.06E+08
37/149.0234mz/22.25min	5.83E+07	3.18E+07	3.48E+08	1.35E+09
38/279.1591mz/6.14min	2.45E+07	1.65E+07	5.25E+07	5.78E+08
39/156.9907mz/22.36min	8447.341162	14201.04281	1.89E+08	1.62E+08
40/149.0235mz/6.08min	4.27E+07	6.62E+07	2.17E+07	3.46E+08
41/149.0234mz/4.72min	3.10E+07	2.37E+07	2.14E+07	6.83E+07
42/149.0235mz/5.34min	1.07E+07	3.39E+07	4.69E+07	3.30E+08

Fig S1. PLS-DA scores plot of leaf extracts from non-treated (C) and VoraxTM-treated (T) samples with 35% of the total variance (28.6% for Component 1 and 6.4% for Component 2). The data were acquired by UHPLC-MS/MS, positive mode, with n = 5 (three repetitions).

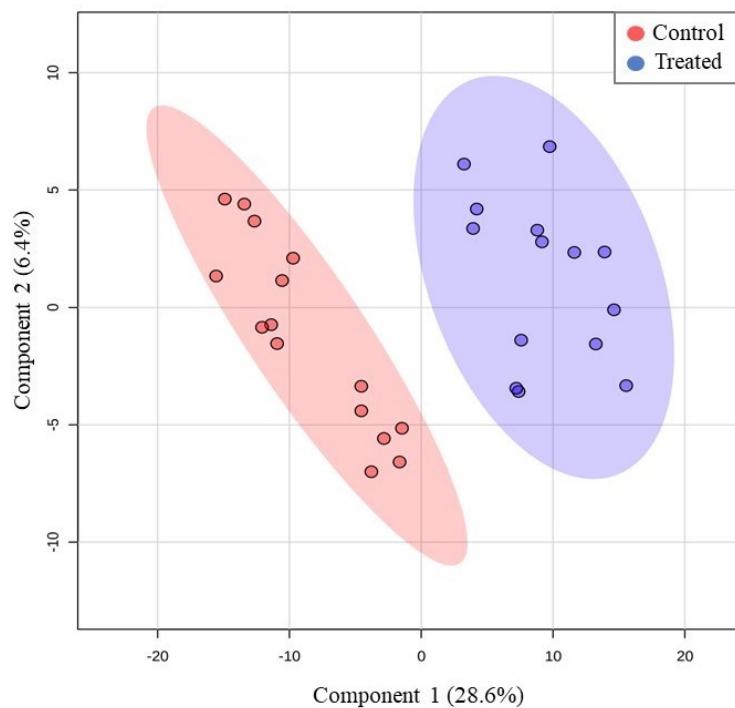


Fig S2. MS/MS spectra of m/z 148.0605 [M+H]⁺ annotated as Glutamic acid.

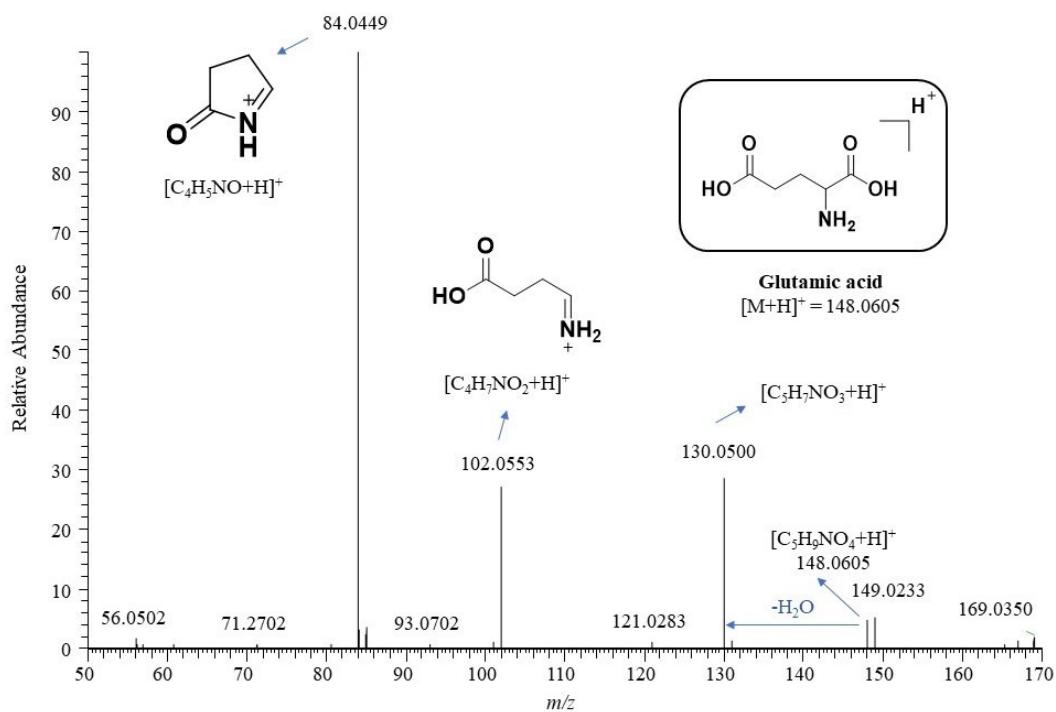


Fig S3. MS/MS spectra of m/z 595.1658 $[M+H]^+$ annotated as Nicotiflorin.

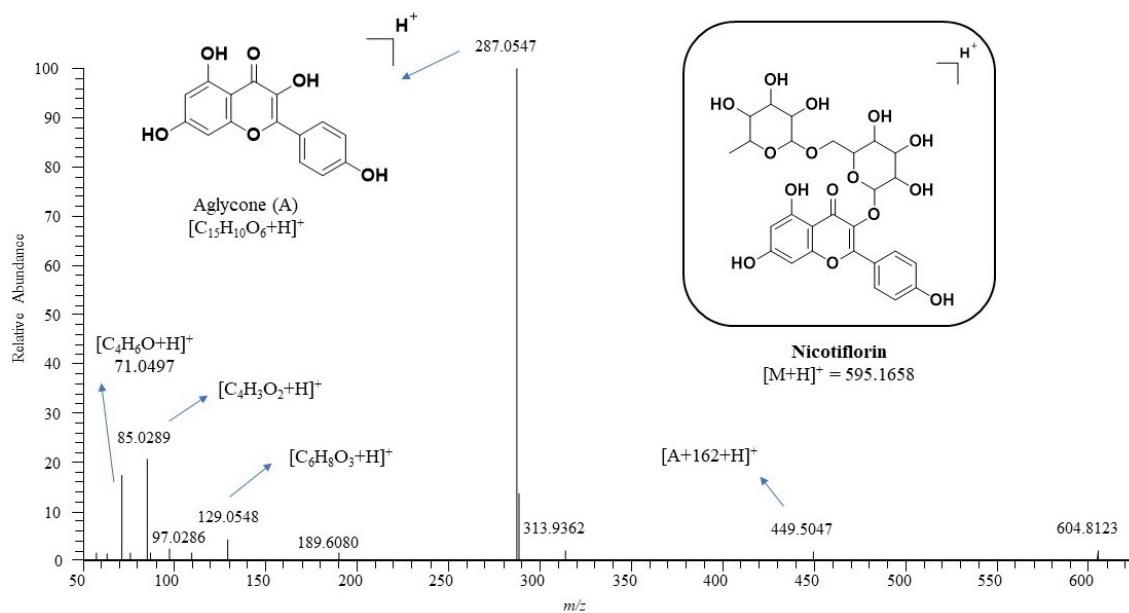


Fig S4. MS/MS spectra of m/z 353.2687 $[M+H]^+$ annotated as 1-linolenoylglycerol.

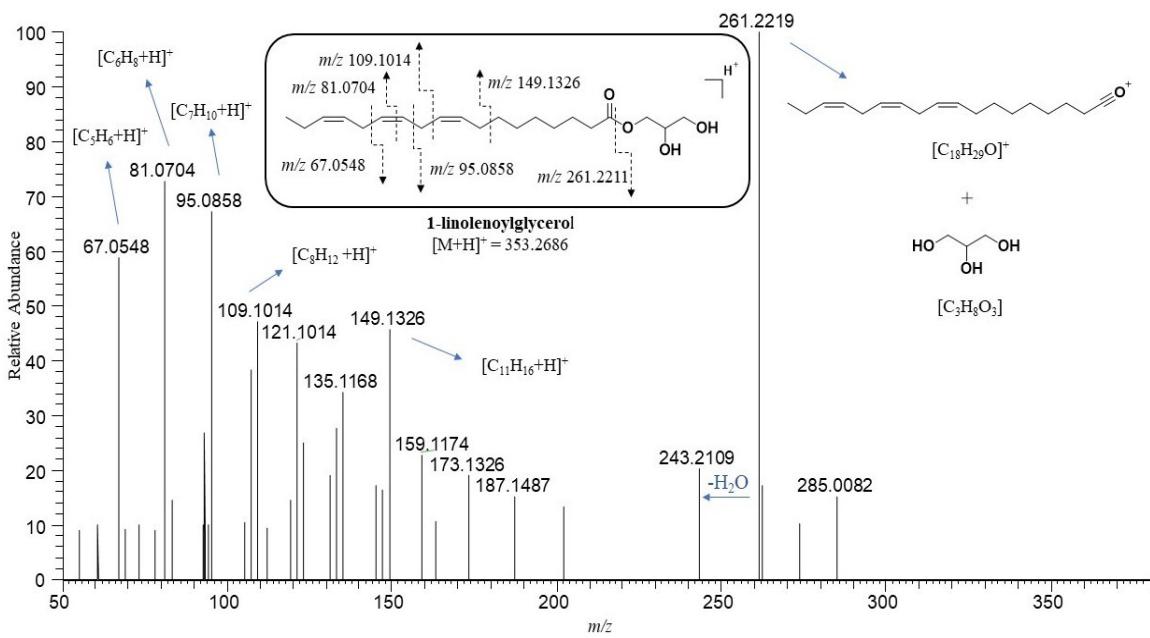


Fig S5. MS/MS spectra of m/z 532.3482 $[M+\text{NH}_4]^+$ annotated as 1-linolenoyl-3-galactopyranosyl-glycerol.

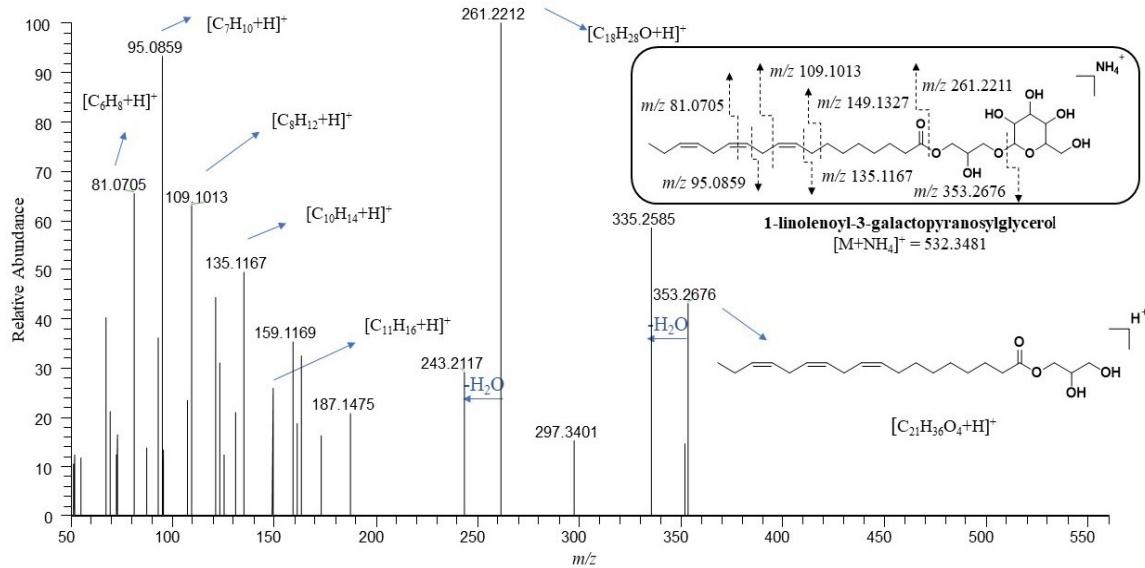


Fig S6. MS/MS spectra of m/z 694.4003 $[M+\text{NH}_4]^+$ annotated as gingerglycolipid A.

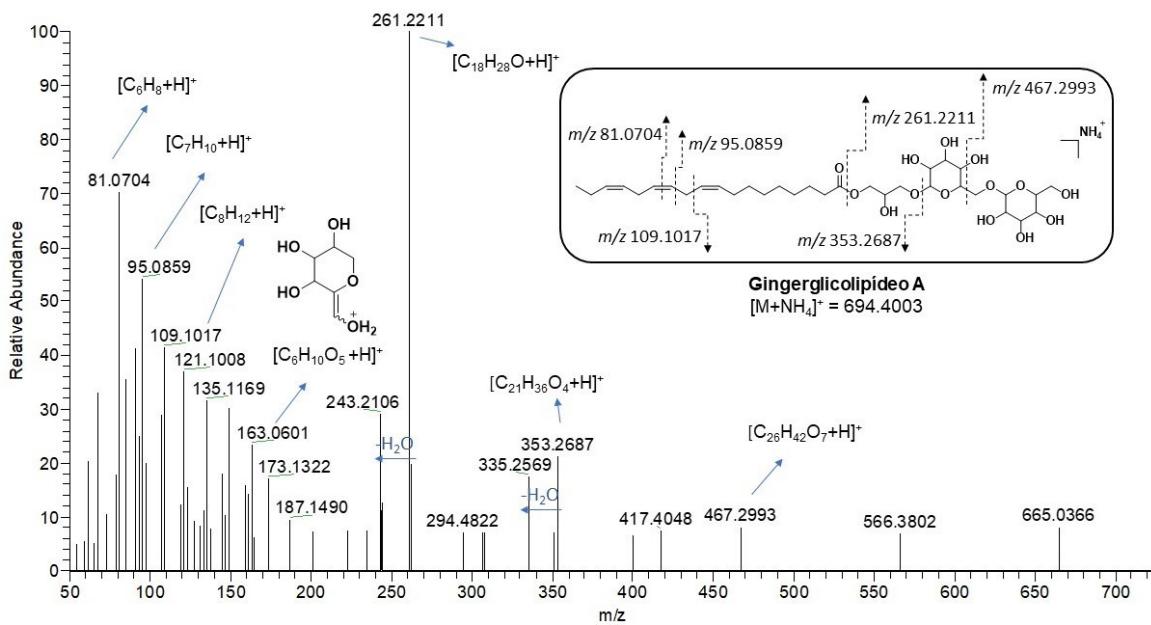


Fig S7. Mirror match obtained from GNPS for astragalin annotation (m/z 449.1078), $\cos = 0.92$.

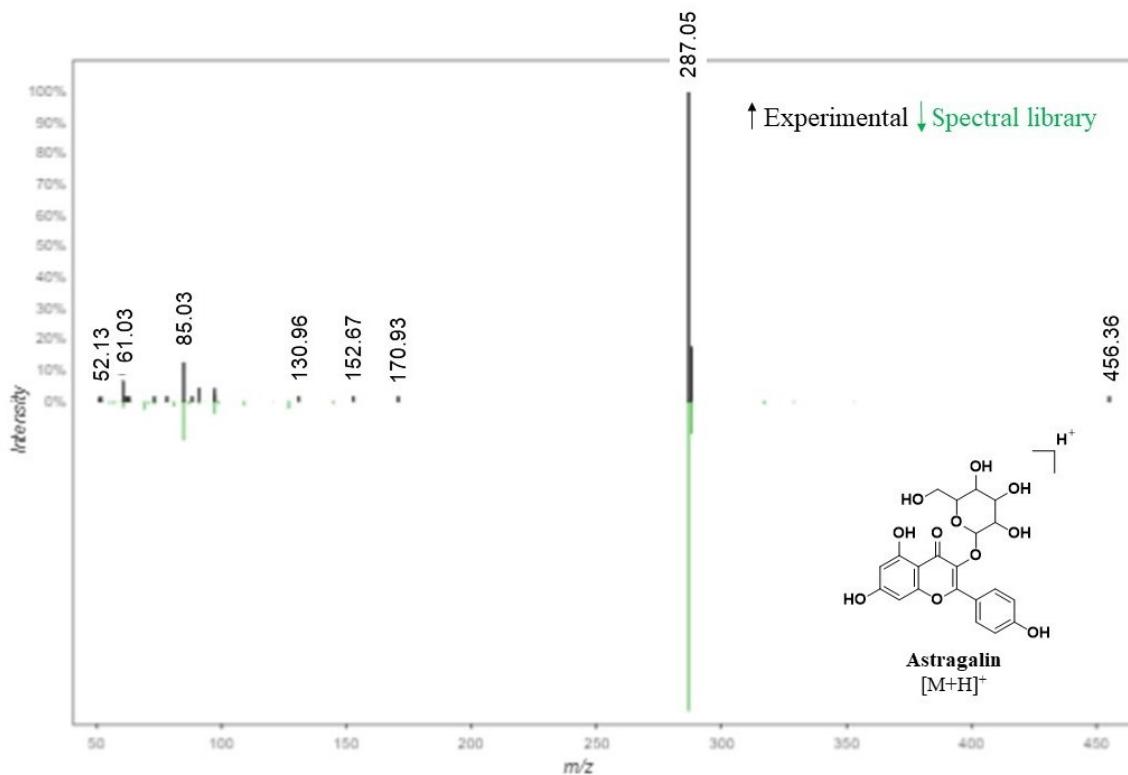


Fig S8. Mirror match obtained from GNPS for rutin annotation (m/z 611.1598), $\cos = 0.90$.

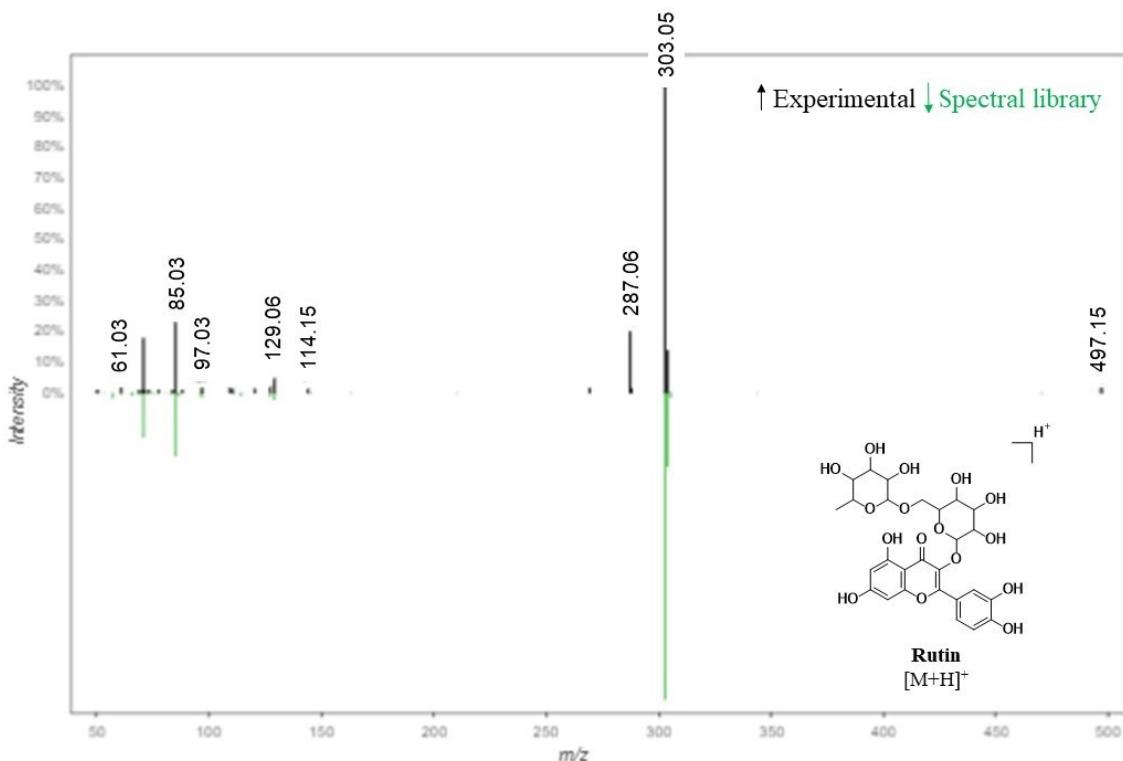


Fig S9. PLS-DA scores plot of leaf extracts from WST, WS and NSNT samples with 42.2% of the overall variance (18.6% for Component 1 and 23.6% for Component 2). The data were acquired by UHPLC-MS/MS, positive mode, with n = 8 (two repetitions).

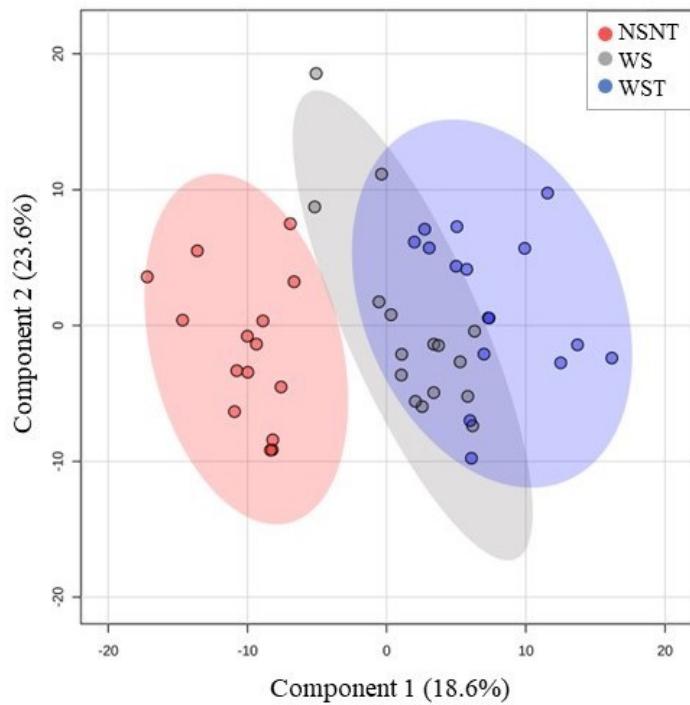
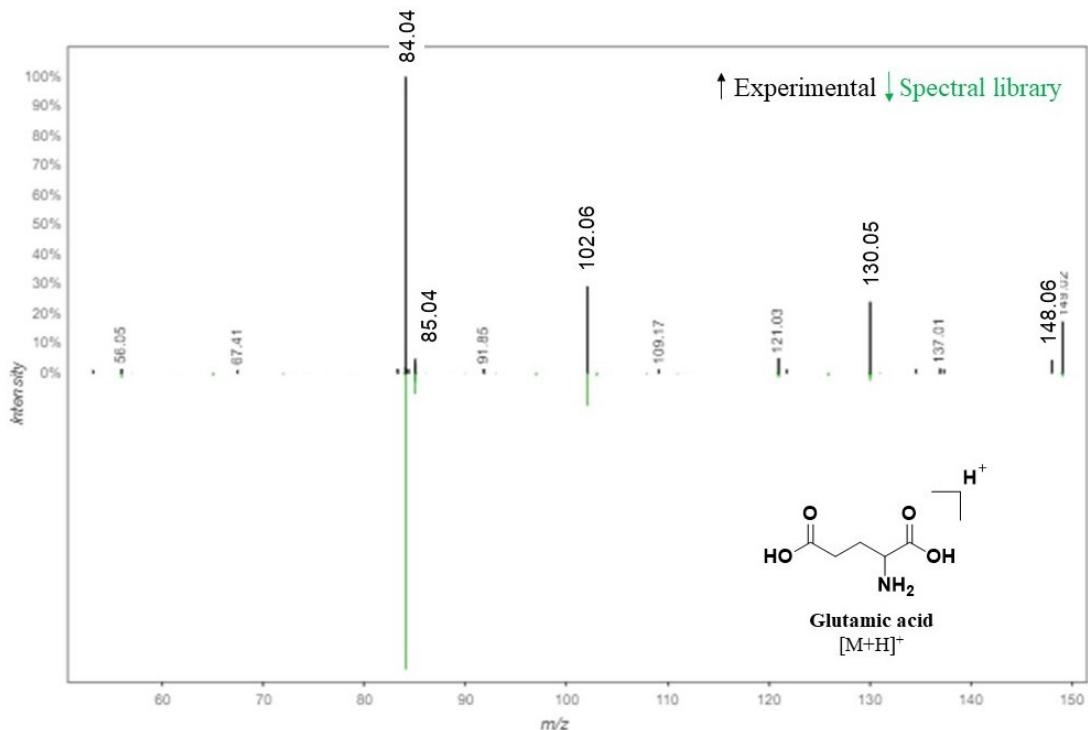
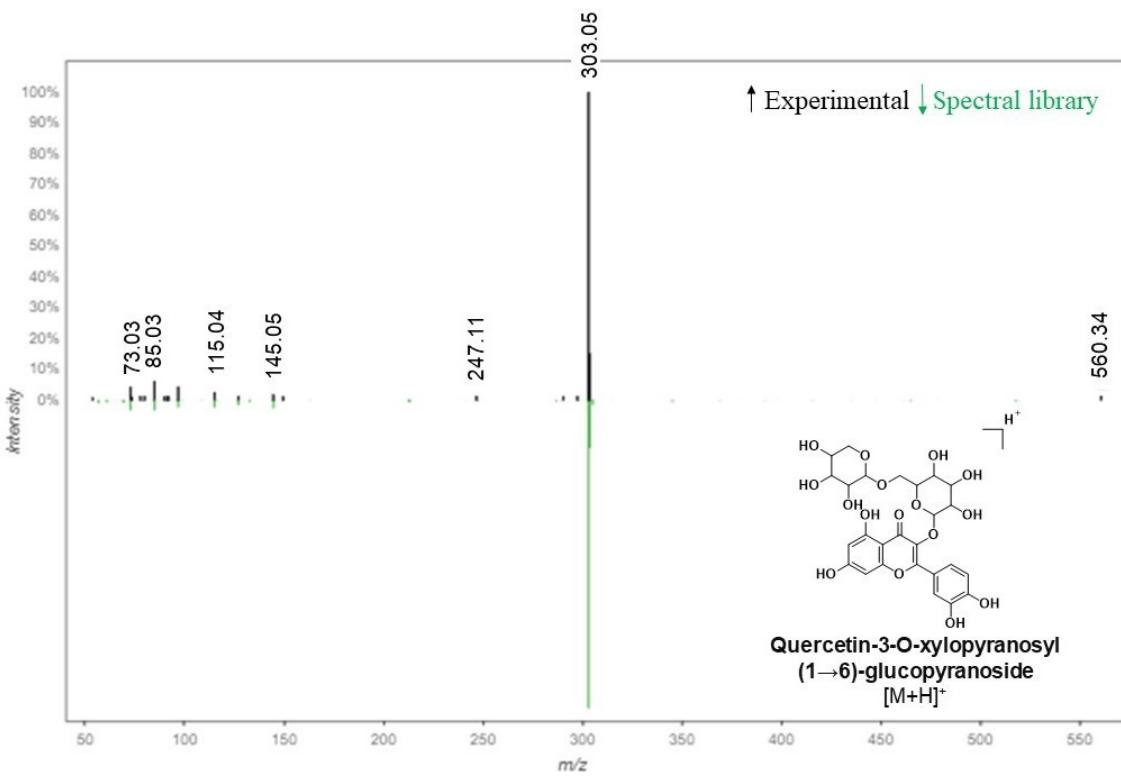
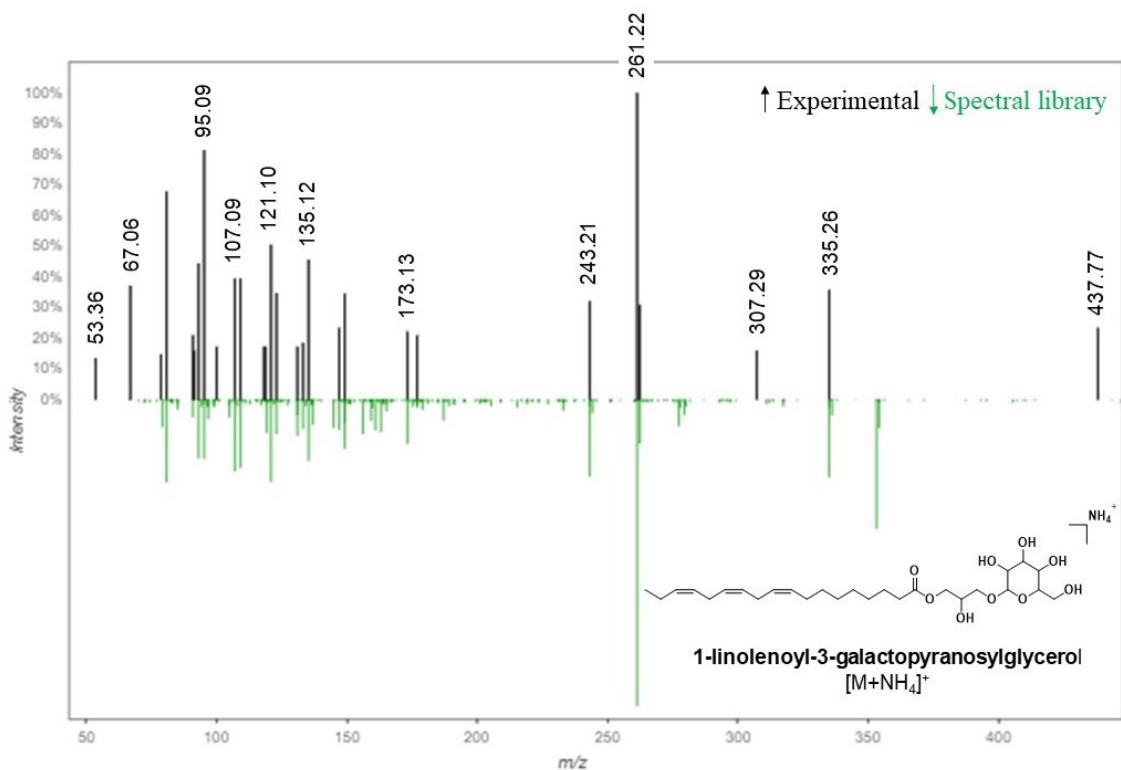


Fig S10. Annotation level 2 of differential metabolites obtained by the analysis of the VIP scores in the water stress assay.





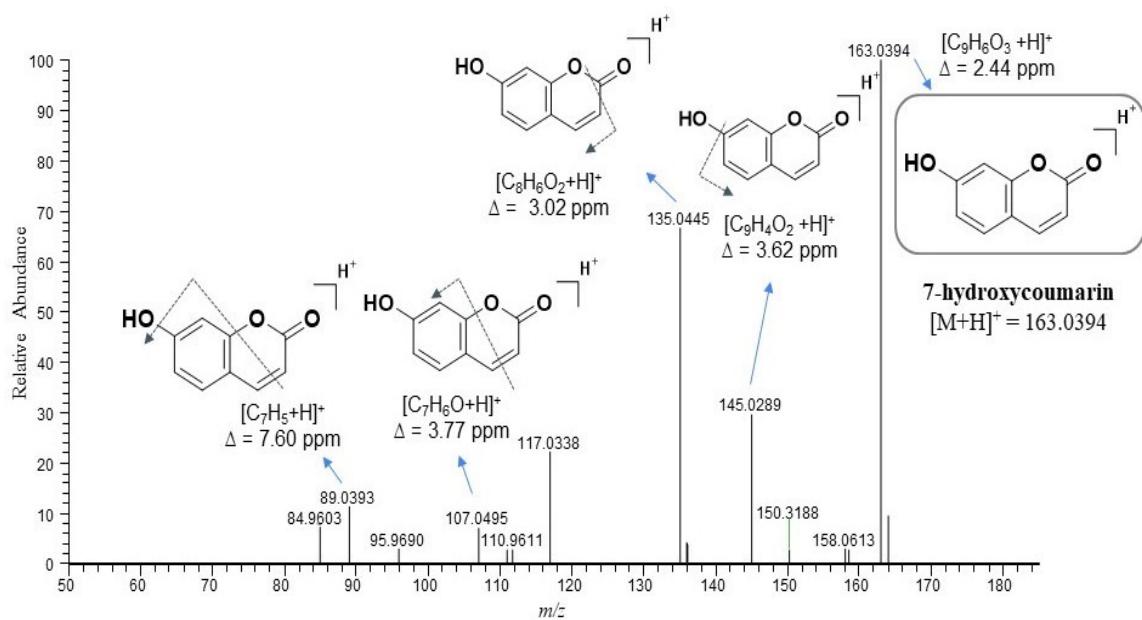
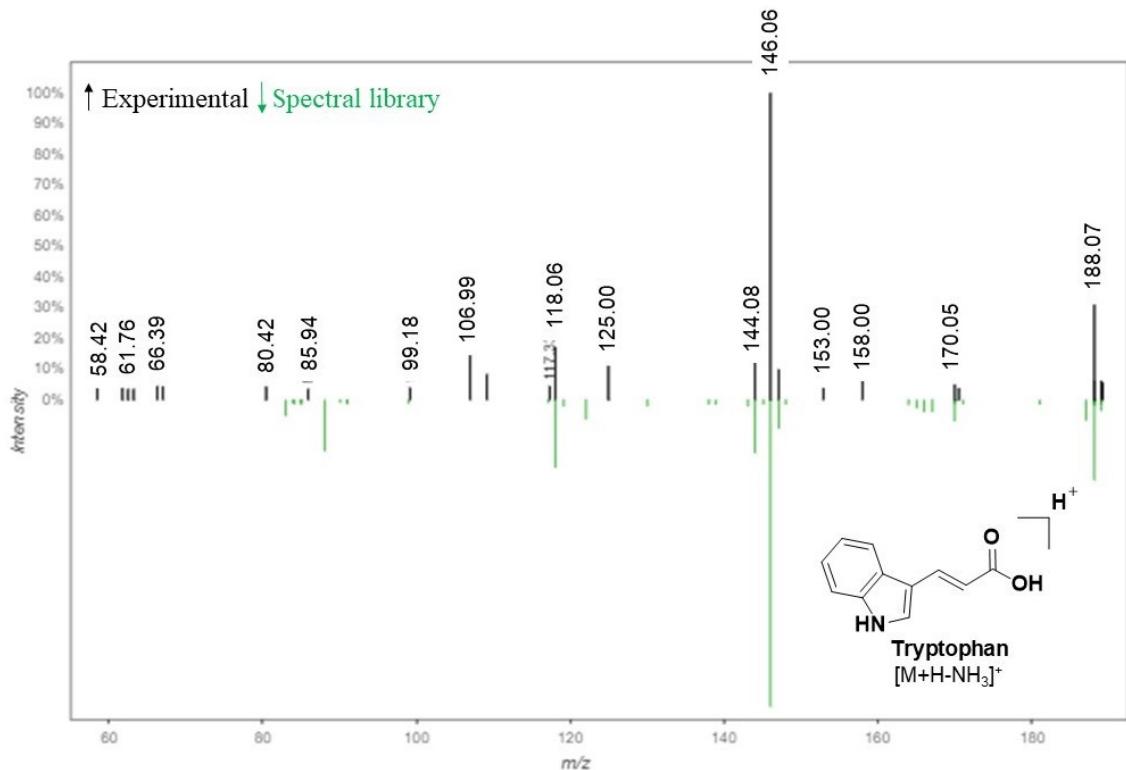


Fig S11. Boxplot chart of the metabolites that exhibited an increase in the concentration in the two trials subjected to water stress but more expressive in WST.

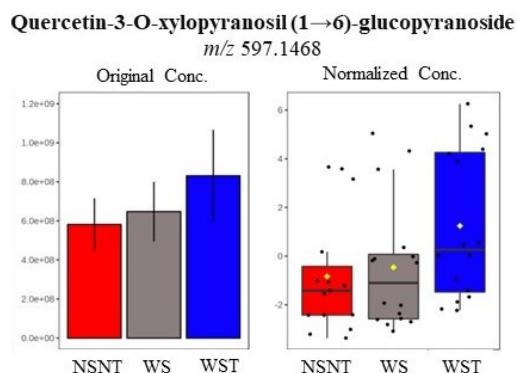
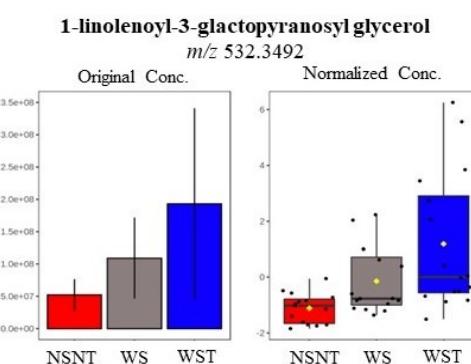
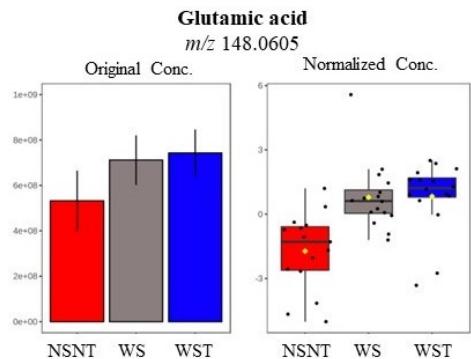


Fig S12. Boxplot chart of the metabolites that exhibited a decrease in the concentration in the two trials subjected to water stress (WS and WST).

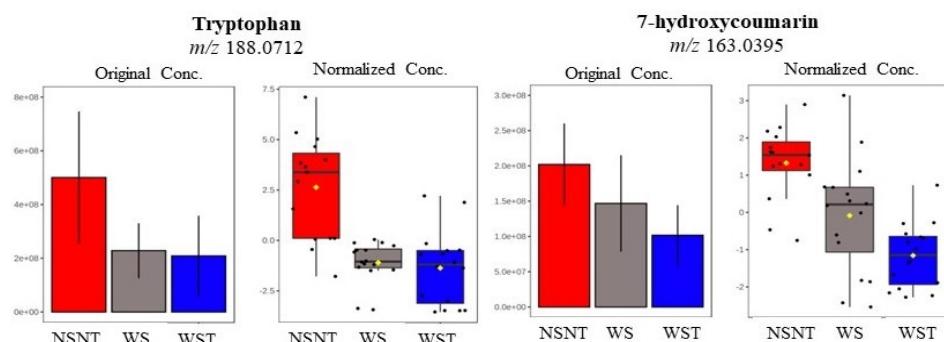


Fig S13. Radicles length for *P. vulgaris* seed germination assays with **A)** Exposure of photoperiod 12h/12h light/dark (+) and **B)** Complete absence of sunlight (-).

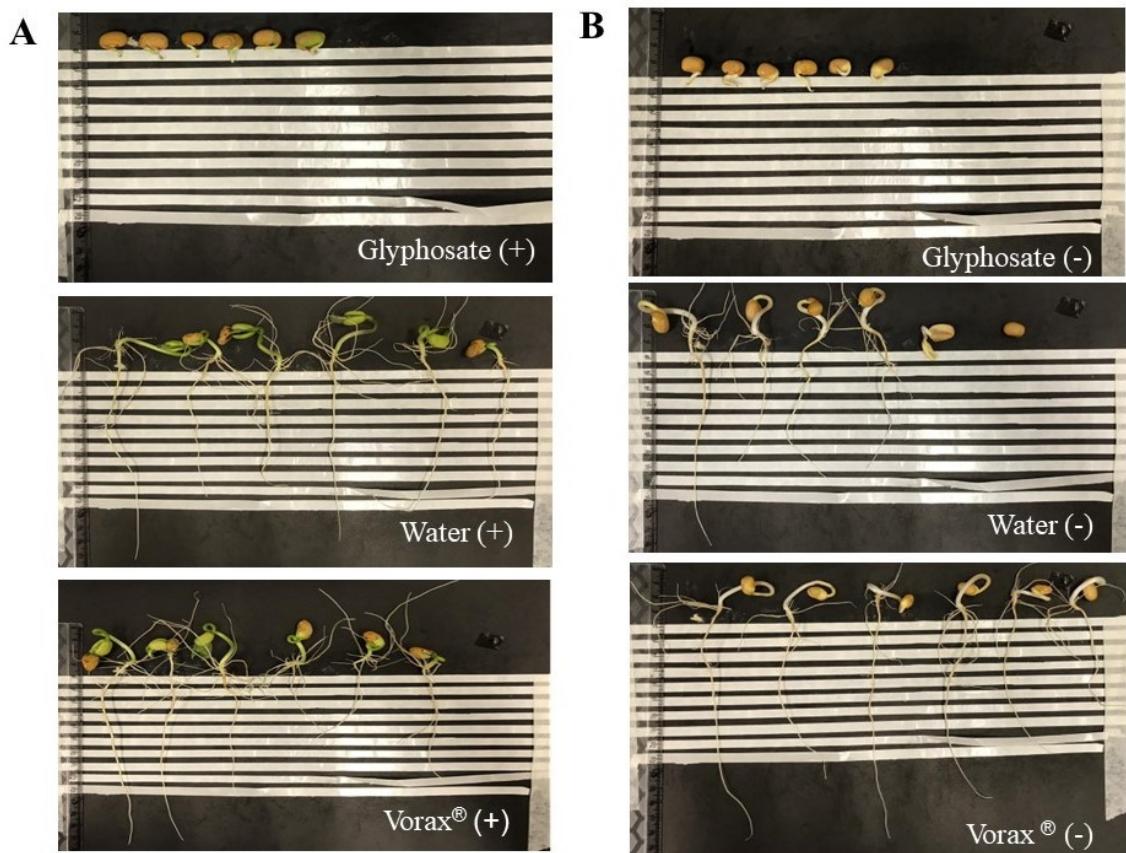


Fig S14. **A)** PLS-DA component 1 (29.7%) x component 2 (15.8%) with data of the two assays (+) and (-), **B)** PLS-DA comp. 1 (15.3%) x comp. 2 (20%) of the (+) data and **C)** PLS-DA comp 1 (21.2%) x comp 2 (21%) of the (-) data. The data were acquired by UHPLC-MS/MS, positive mode, with n = 5 (two repetitions).

