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Electronic Supporting Information

Cobalt Catalysts (Co-N-C) for C-O Bond Cleavage in Lignin-Derived Aryl Ethers and Lignin

Daniel Bautista-García, ^a David Macias-José, ^a Paola Aguillón-Rodríguez, ^a Obed Pérez-Reyes ^b and Carmen Ortiz-Cervantes, *^a

^a Instituto de Química, Universidad Nacional Autónoma de México, CU, Coyoacán, 04510, Ciudad de México, México.

^b Instituto Politécnico Nacional, Centro de Investigación en Ciencia Aplicada y Tecnología Avanzada, Unidad Legaria, Ciudad México, México.

carmen.ortiz@iquimica.unam.mx

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1. Characterization Methods

For cobalt materials as well as for organic compounds, various identification and characterization techniques were used, sample preparation was performed according to each specific protocol described below.

Nuclear Magnetic Resonance (NMR) Studies

¹H, ¹³C{¹H} and HSQC NMR spectra were recorded at room temperature on a 400 MHz Bruker Avance III spectrometer in CDCl₃, unless otherwise stated. ¹H and ¹³C{¹H} chemical shifts (δ , ppm) are reported relative to the residual proton resonance in the corresponding deuterated solvent.

Mass Spectrometry

A Jeol mass spectrometer SX 102 A was used with the direct analysis in real time (DART).

GC-MS

Conversion and selectivity of catalytic reactions were determined via GC-MS analysis preformed on Agilent 5975C system equipped with a 30 m DB-5MS capillary (0.32 mm ID) column), He 99.999%.

Electron Paramagnetic Resonance (EPR)

Solid samples were placed in quartz tubes of 1.34 cm diameter and 0.15 cm thickness. Loaded tubes were then introduced in a Jeol JES-TE300 spectrometer operating at X band frequency (9.4 GHz) at 100 kHz field modulation, with a cylindrical cavity (TE011 mode). Samples were run at 77 K.

Fourier Transform Infrared (FTIR) spectra

The ATR-IR spectra were determinate on a FTIR/FIR spectrum FT-IR NICOLET IS-50, Thermo Fisher Scientific, measuring 32 scans per sample, from 4000 to 400 cm⁻¹ with a resolution of 2 cm⁻¹, the sample did not require previous preparation.

Powder X-ray Diffraction (PXRD)

Powder X-Ray diffraction measurements were performed on a Rigaku Ultima IV diffractometer with Cu K_{α} radiation (I=1.54056 Å) at 40 kV and 44 mA. The instrument was operating in a Bragg Brentano geometry with a step increment of 0.02° and an acquisition time of one second per step.

Transmission Electron Microscopy

Transmission electron microscopy studies were carried out in a Jeol ARM-200F operated at 200 kV using copper grids (mesh size of 300 covered with a lacey carbon film) equipped with on a holey carbon. The samples were dispersed in anhydrous toluene.

X-ray photoelectron spectroscopy (XPS)

XPS spectra were recorded at room temperature using a K-alpha+ spectrometer Thermo Fisher Scientific Co. Equipped with an Al Ka (1486.6 eV) monochromatic X-ray source, a dual-beam flood gun for charge neutralization, and 180 double focusing hemispherical analyzer operating in a constant analyzer energy mode (CAE). The measurement spot size was 400 µm and a base pressure of 1 x 10-9 mbar was held in the analytical chamber. Survey scans were recorded using 400 µm spot size and fixed pass energy of 200 eV, whereas high resolution spectra were recorded at 20 eV of pass energy with a step size of 0.1 eV. Charge corrections for all the spectra were referenced to the position of the C1s adventitious peak at 284.8 eV. All spectra were processed with the Avantage software (v5.9925) provided by Thermo-Fisher Scientific Co. The curve fitting was performed with a Voight function and a Shirley-type background. Transition metals having unpaired electrons in the d orbitals, such as Co(II), exhibit the so-called multiplet splitting feature in their XPS spectra, which is manifested as the appearance of additional peaks as a consequence of the different final states due to the coupling of the unpaired spins in d orbitals with the unpaired spin in the nucleus produced by the photoemission of an electron, in this case from the Co2p level. In addition, there can be present shakeup and plasmon loss structures which also contribute to the final spectrum shape. In a transition metal spectrum with the mentioned features the number of contributions as well as their positions and intensities can be different even for the same atom in the same oxidation state since these structures are very sensitive to the chemical environment, as previously described by Biesinger et al ¹and Yang et al² who developed multiplet structure models of transition metal oxides and hydroxides and the contributions do not necessarily have a single chemical identity but contribute to reproduce the spectral shape of the analyzed species and thus can be used in subsequent curve fitting procedures.³

Inductively coupled plasma spectroscopy (ICP-MS) analysis

ICP-MS analysis for Co materials was measured using Thermo Scientific Element XR High-Resolution ICP-MS after digesting materials in aqua regia.

Elemental Analysis

Elemental analysis of Co/C and Co-1 was carried out in Thermo Scientific/Flash 2000.

Gel permeation chromatography/Size exclusion chromatography

The GPC/HPSEC analysis were carry out UV/Vis detector equipped the molecular weights and the molecular distributions (MWD) of the samples from lignin hydrogenolysis product were determinate by GPC chromatograph in Water Alliance e2695 chromatograph separation module apparatus equipped with detector multi angle light scattering (MALLS) Wyatt Technology, model DAWN EOS, polystyrene was used to calibration. The samples were dissolved in THF.

2. Characterization of Co-1.

PDRX

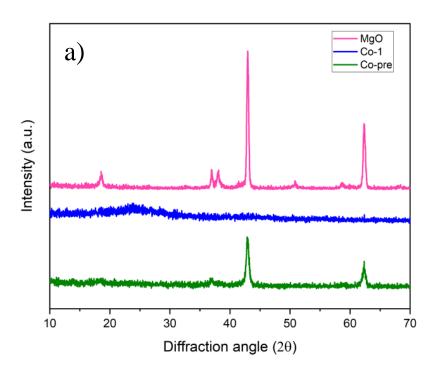


Figure S1. Powder X-ray diffraction (PDRX) patterns of Co-1 before thermal treatment, Co-1 and MgO.

XPS

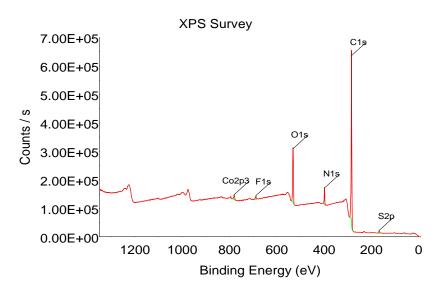


Figure S2. XPS survey spectra of the Co-1.

Elemental Analysis and ICP

Table S1. wt% Co and N content.

Sample	Co (mg/Kg)/%wt.ª	N (%) ^b	Co(atomic %)/%wt. ^c
Co-1	6040.07/0.60	5.90	0.44/1.98
Co/C	37954.63/3.79	2.91	-
Co-2	13331.42/1.32	-	-

^a Obtained by IPC-MS, ^b elemental analysis, and ^c XPS.

Elemental Analysis

No. de registro	Clave de la muestra	Valor	N [%]	C [%]	H [%]	S [%]	Fecha de análisis
225 Co-C	Co. C	Teórico	2.91	90.98	1.59		24-08-2022
	C0-C	Exp	1.29	71.33	1.25		24-00-2022

No. de registro	Clave de la muestra	Valor	N [%]	C [%]	H [%]	S [%]	Fecha de análisis
224	DM-1-DM-2	Teórico	5.90	74.19	1.15		24-08-2022
224	DIVI-T-DIVI-2	Exp	9.14	59.28	3.21		24-00-2022

TEM

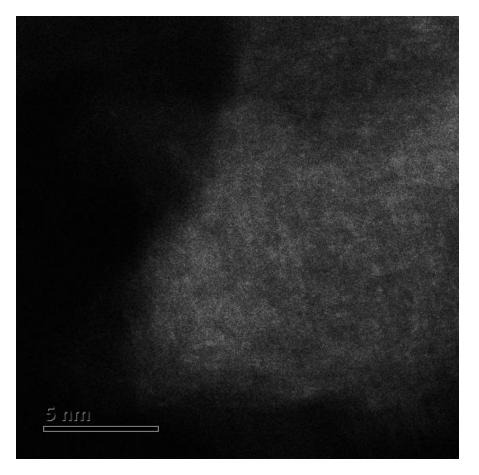


Figure S3. TEM images from Co-1 (scale = 5 nm).

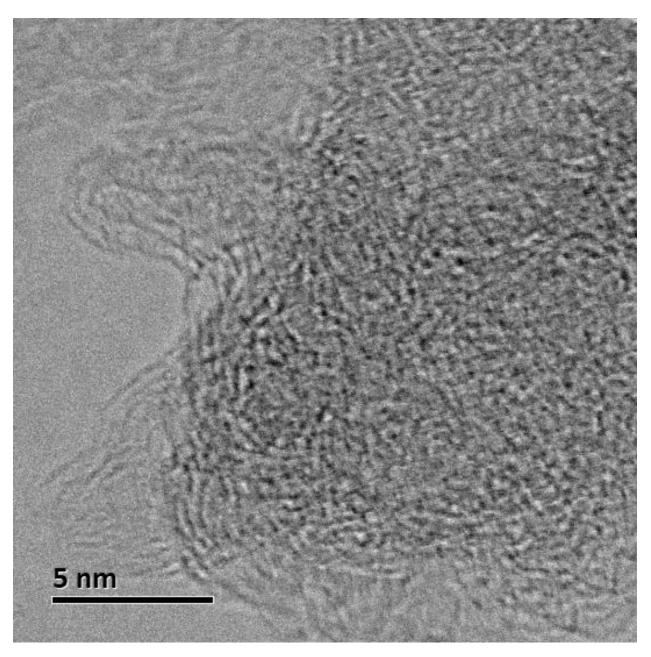


Figure S4. TEM images from Co-1 (scale = 5 nm).

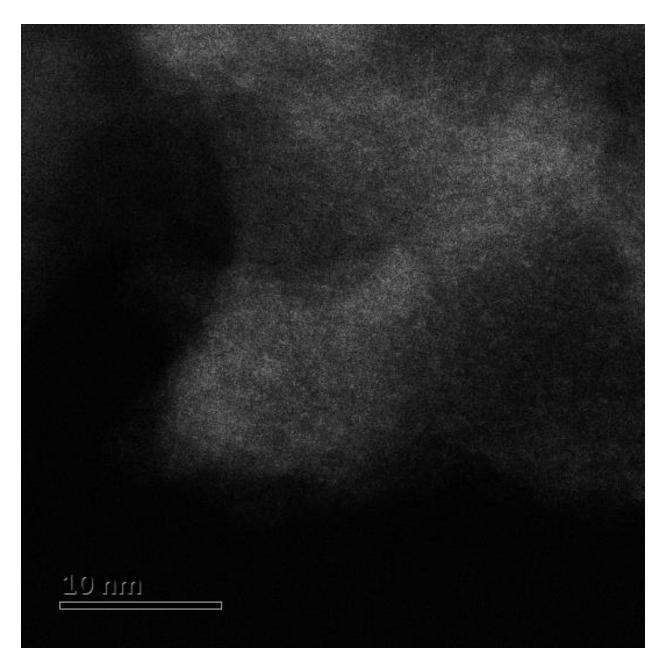


Figure S5. TEM images from Co-1 (scale = 10 nm).

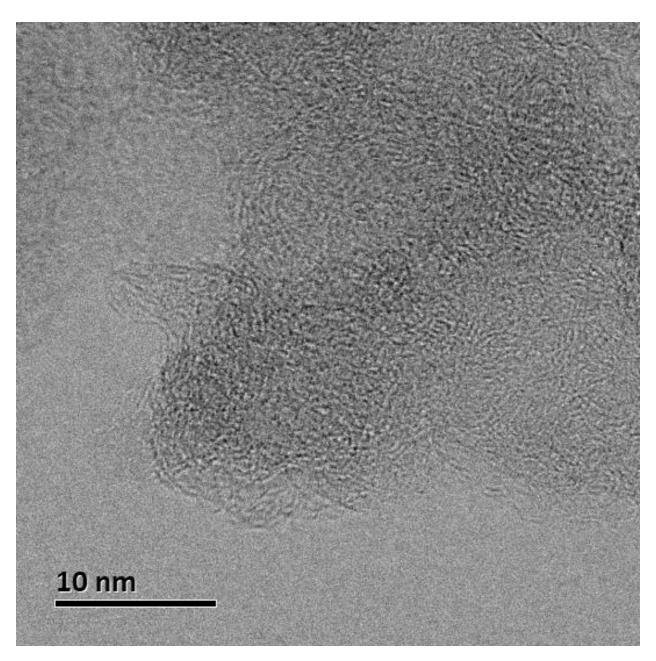


Figure S6. TEM images from Co-1 (scale = 10 nm).

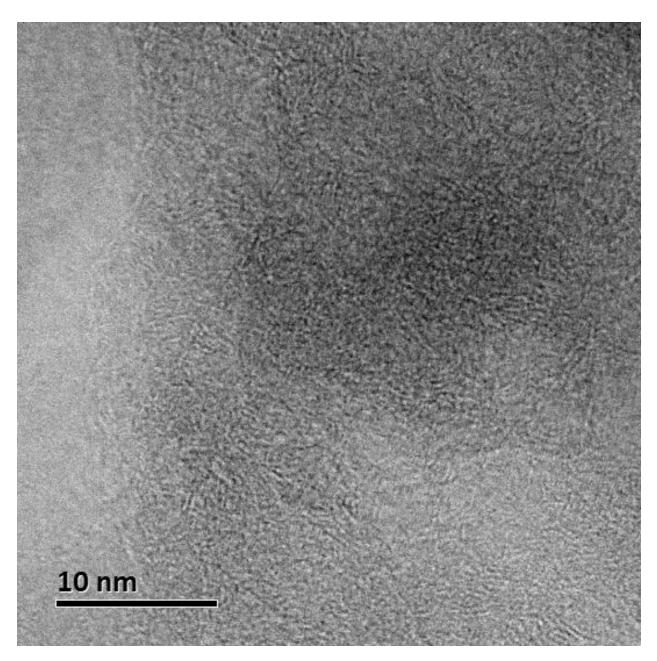


Figure S7. TEM images from Co-1 (scale = 10 nm).

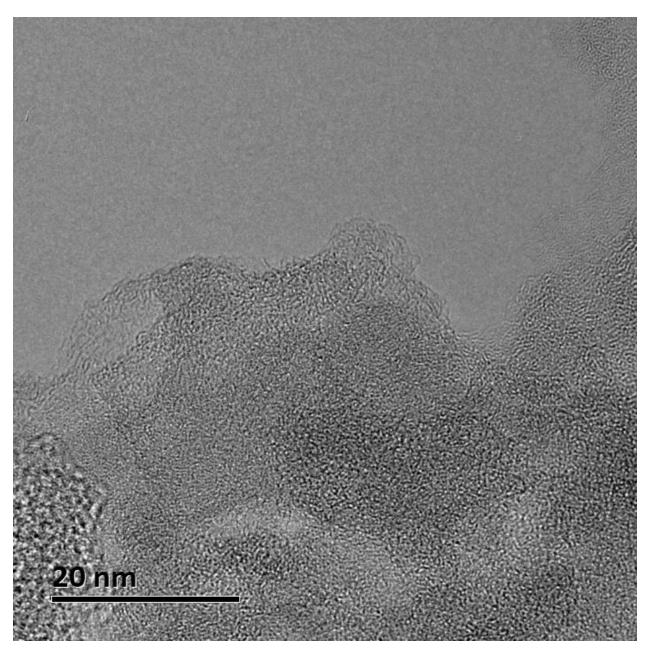


Figure S8. TEM images from Co-1 (scale = 20 nm).

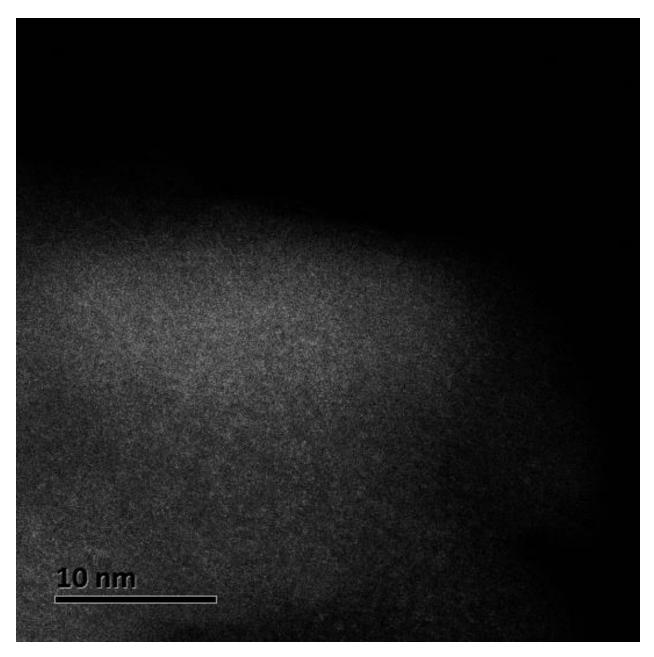


Figure S9. TEM images from Co-1 (scale = 10 nm).

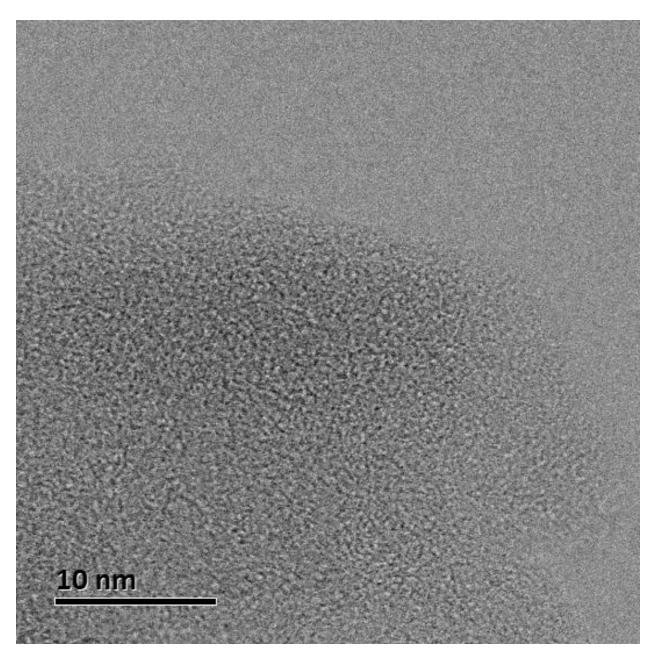


Figure S10. TEM images from Co-1 (scale = 10 nm).

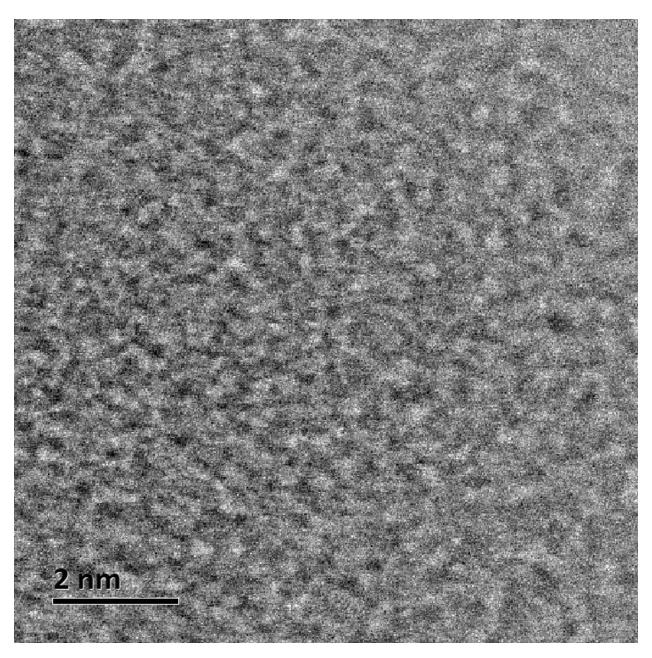


Figure S11. TEM images from Co-1 (scale = 2 nm).

Co/C

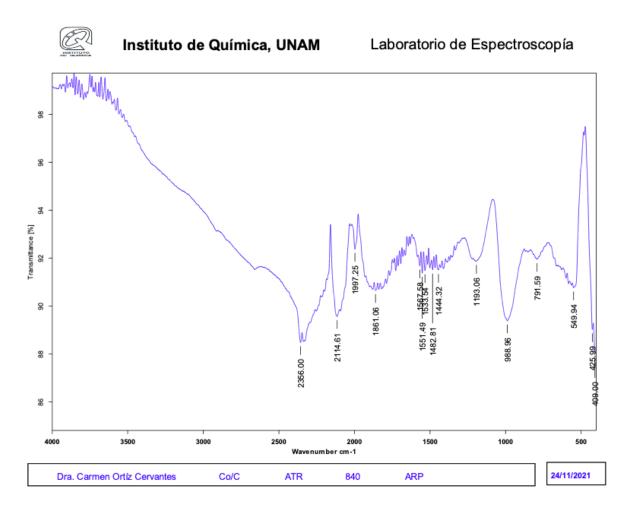


Figure S12. FTIR spectrum of Co/C.

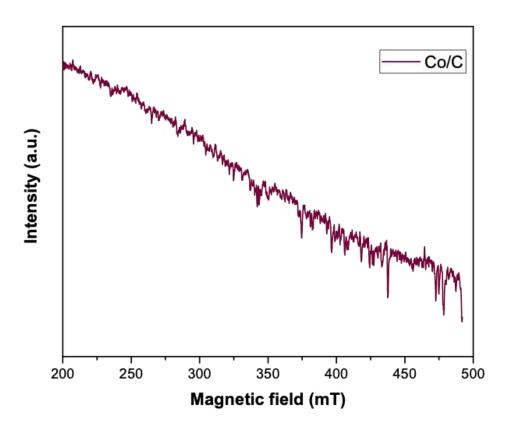
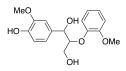


Figure S13. X-band electron paramagnetic resonance (EPR) of the solid for Co/C.

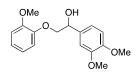
3. Synthesis of substrates

3.1. Synthesis of MM-3



Scheme 1. Lignin model molecule, **MM-3** (guaiacylglycerol-β-guaiacyl ether).

The **MM-3** molecule was prepared according to previously reported procedure without modification.⁴ ¹H NMR (400 MHz, Chloroform-d) δ 7.13 – 6.65 (m, 7H), 5.63 (d, J = 12.5 Hz, 1H), 4.97 – 4.72 (m, 1H), 4.12 – 3.92 (m, 1H), 3.90 – 3.32 (m, 9H), 2.74 (ddd, J = 21.2, 8.0, 5.2 Hz, 1H).



Scheme 2. Lignin model molecule, MM-2.

Compound MM-2 was prepared according to a previously described procedure.⁵

¹H NMR (400 MHz, CDCl₃) d 7.02 – 6.97 (m, 2H), 6.95 – 6.87 (m, 4H), 6.85 (d, J = 8.2 Hz, 1H), 5.05 (dd, J = 9.3, 3.0 Hz, 1H), 4.16 (dd, J = 10.0 Hz, 3.0 Hz), 3.97 (dd, J = 10 Hz, 9.7 Hz, 1H), 3.90 – 3.85 (s, 9H), 3.60 (s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 150.18, 149.20, 148.87, 148.12, 132.34, 122.60, 121.22, 118.73, 115.99, 112.10, 111.16, 109.54, 76.39, 72.22, 56.05, 55.99, 55.94.

3.3 Synthesis of MM-1

Scheme 3. Lignin model molecule, MM-1.

Compound **MM-1** was prepared according to a previously described procedure.⁶ ¹H NMR (400 MHz, CDCl₃) δ 7.06 – 7.02 (m, 1H), 6.97 (m, 1H), 6.91-6.81 (m, 2H), 6.64-6.61 (m, 2H), 4.90 (dd, 2.1 Hz, 8.1 Hz, 1H), 4.54 (s, 1H), 4.40 (dd, *J* = 11.0, 2.6 Hz, 1H), 3.87 (d, *J* = 9.1 Hz, 9H), 3.86 (3, 3H), 3.70 (dd, *J* = 10.9 Hz, 10.1 Hz, 1H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 153.25, 148.98, 148.52, 136.80, 132.00, 124.09, 118.66, 110.93, 109.39, 105.13, 80.15, 72.22, 56.11, 55.92, 55.87.

4. Synthesis of Co/C synthesis

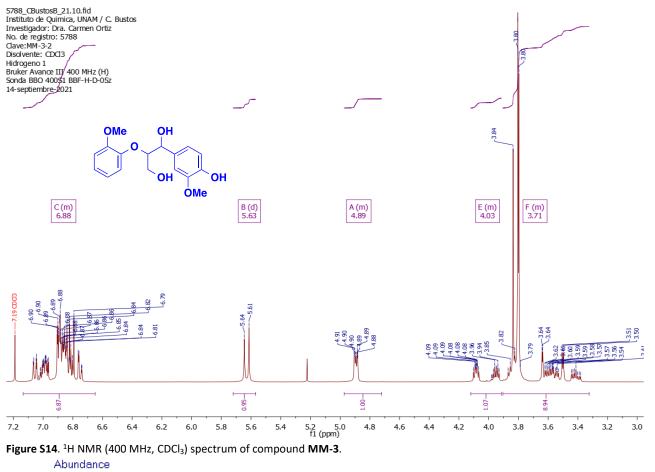
The **Co/C** catalyst were prepared similarly to a procedure reported by Beller group.⁷ Briefly, a 50 mL round bottom flask was charged with Co(acac)₂ (0.5 mmol) and 20 mL of ethanol and stirred for 10 min at room temperature. To this solution, 1,10-phenantroline (0.182 g, 1 mmol) was added and stirred for 120 min at 60 °C. Carbon activated (0.696 g) was added, the resulting suspension was stirred at room temperature for 18 h. The solvent was removed under reduced pressure and dried for 8 h. The resulting solid was ground in an Agate mortar, and the fine powder was transferred to a crucible with a lid and pyrolyzed at 800 °C for 2 h under nitrogen atmosphere.

5. Catalytic hydrogenolysis

Catalyst	conditions	Products	Ref.
Co-1	H ₂ O/EtOH, 2 h, 150 °C, Et ₃ N, HCOOH	OMe OH MeO HO	this work
		99 99	
Ni-N-C/Ni-NPs	H ₂ O/EtOH, 2 h, 150 °C, Et ₃ N, HCOOH	$\begin{array}{c} OMe \\ HO \\ H$	8
5wt% Zn/Pd/C	300 psi H ₂ , MeOH, 150 °C, 2 h. Conv. 100%	ОМе	9
CuO/C	435 psi H ₂ , MeOH, 200 °C, 4 h. Conv. 50.2 %	OMe MeO OMe OH HO HO OMe 70 35 9	10
Ni SAC@N-C	iPrOH, 200 °C, 5 h. Conv. >99%	OMe OH HO	11
		80 53 OMe	
Ni@ZIF-8	435 psi H ₂ , MeOH, 260 °C, 8 h.	HO HO HO	12
Cp ₂ TiCl ₂ ,	THF, 22 °C, 2h, In ³⁺ (4 equiv.) TMSCI (4 equiv.)	OMe HO HO	13
DUNZ=0/C	425	60 36 OMe	14
RuNZnO/C	435 psi H ₂ , MeOH, 220 °C, 4 h.	HO HO HO HO	14
		61 23 43	

Table S2. Overview of catalysts for C-O bond cleavage guaiacil-glycerol- β -guiacyl.

6. Spectra and chromatograms



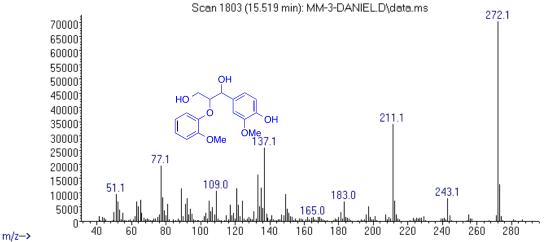


Figure S15. Mass spectrum of compound MM-3.

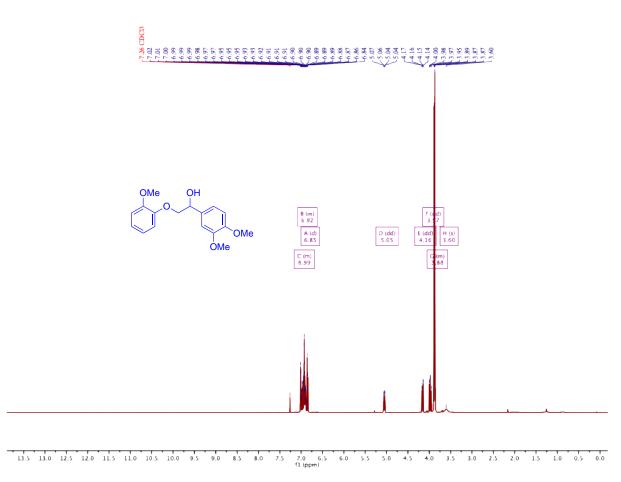


Figure S16. ¹H NMR (400 MHz, CDCl₃) of compound MM-2.

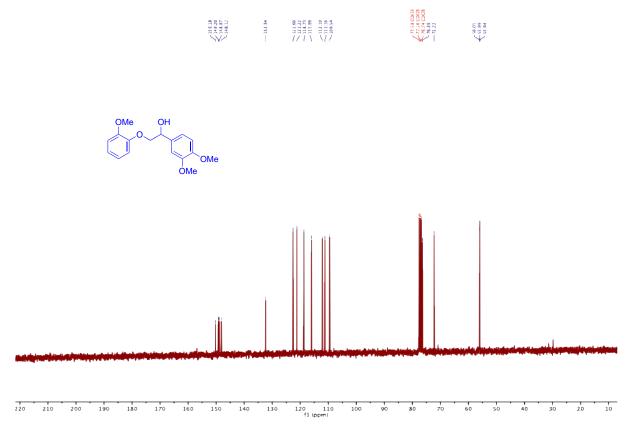


Figure S17. ${}^{13}C{}^{1}H$ NMR (101 MHz, CDCl₃) of compound MM-2.

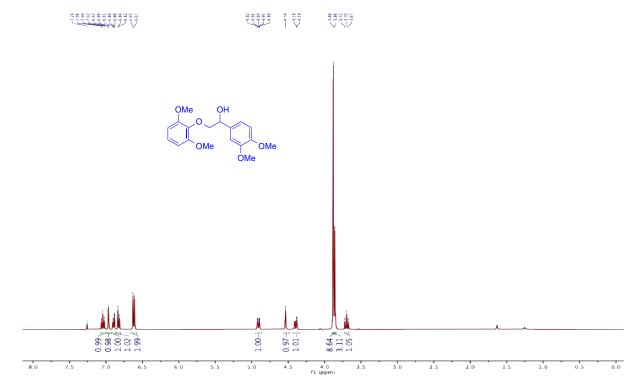


Figure S18. ¹H NMR (400 MHz, CDCl₃) of compound MM-1.

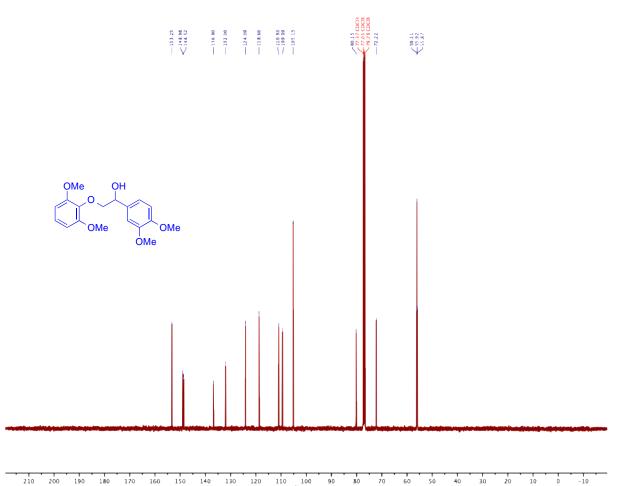
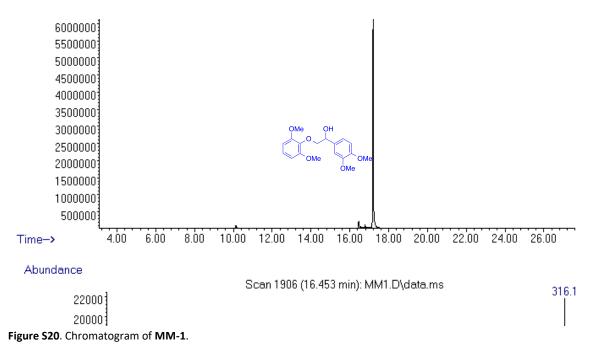




Figure S19. ¹³C{¹H} NMR (101 MHz, CDCl₃) of compound MM-1.



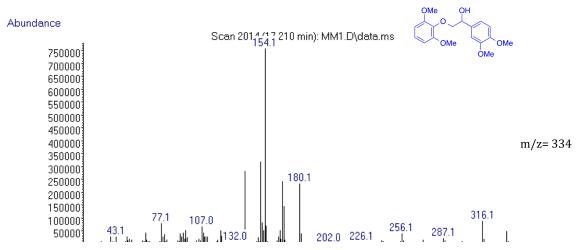
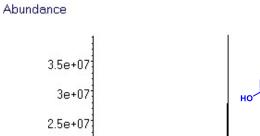


Figure S21. Electronic impact mass spectrum for compound MM-1.



2e+07

1.5e+07

1e+07

5000000

Time->

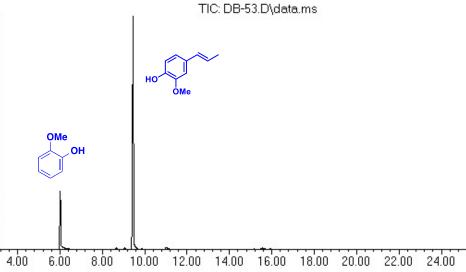


Figure S22. Chromatogram of MM-3 hydrogenolysis with Co-1.

26.00

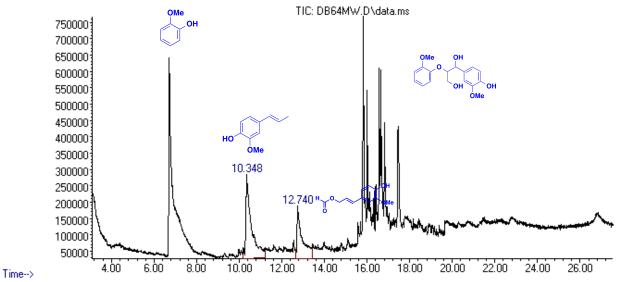
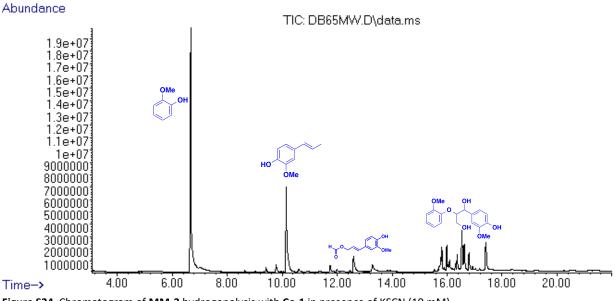
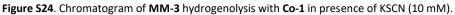


Figure S23. Chromatogram of MM-3 hydrogenolysis with Co/C.





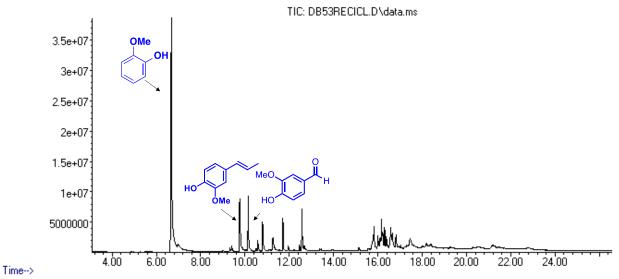


Figure S25. Chromatogram of MM-3 hydrogenolysis with recycled Co-1.

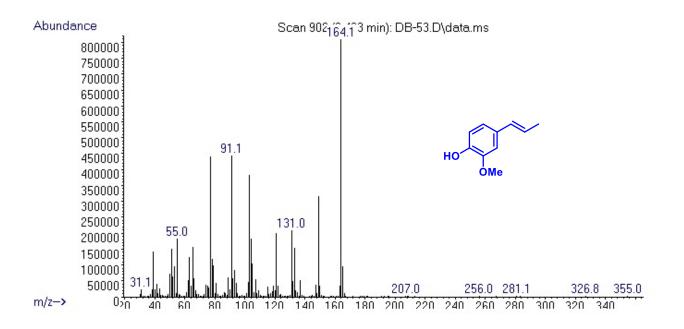
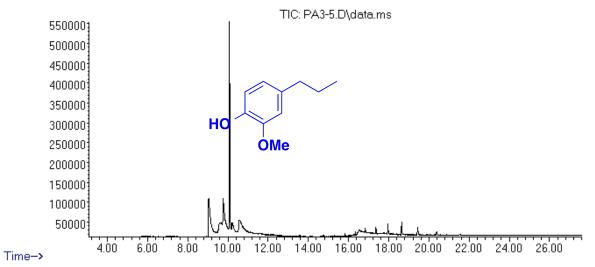
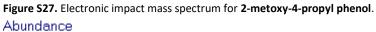


Figure S26. Electronic impact mass spectrum for compound isoeugenol (2-metoxy-4-[(E)-1-prophenyl] phenol).





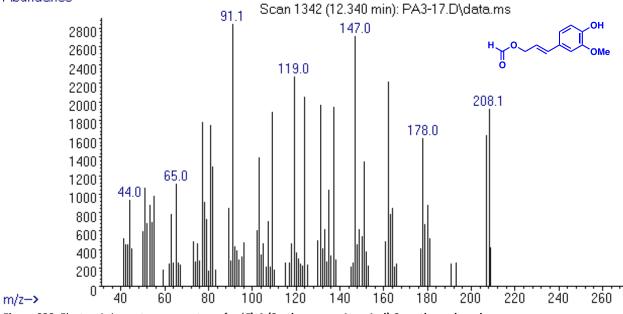


Figure S28. Electronic impact mass spectrum for (E)-4-(3-ethoxyprop-1-en-1-yl)-2-methoxyphenol.

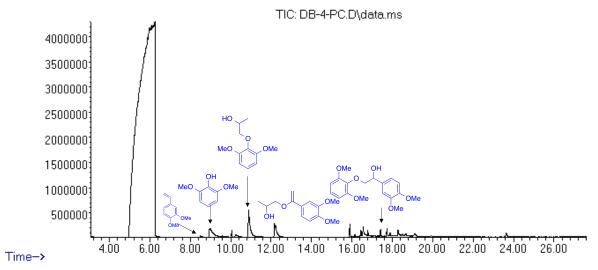
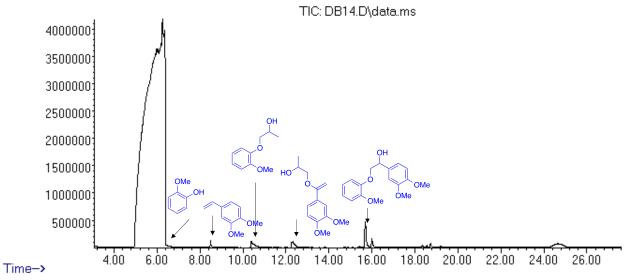
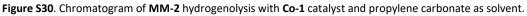


Figure S29. Chromatogram of MM-1 hydrogenolysis with propylene carbonate as solvent.

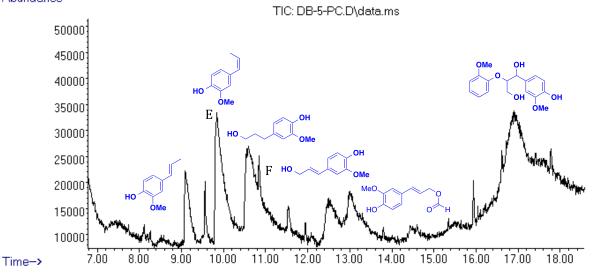


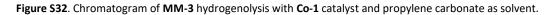


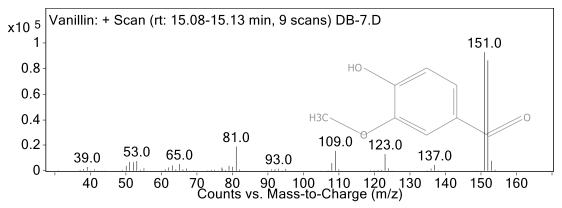
m/z->

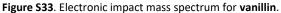
87.0 Scan 1341 (12.333 min): DB14.D\data.ms 360003 34000 32000 30000 28000 26000 24000 22000 20000 18000 59.1 16000 ÓMe 14000 12000 10000 151.1 8000 6000 238.1 41.1 4000 107.0 135.0 2000 179.1 206.9 Ω 70 80 90 100 110 120 130 140 150 160 170 180 190 200 210 220 230 240 40 50 60

Figure S31. Electronic impact mass spectrum for 1-((1-(3,4-dimethoxyphenyl) vinyl)oxy)propan-2-ol. Abundance

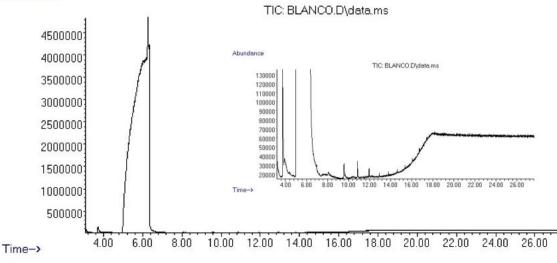


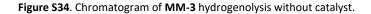












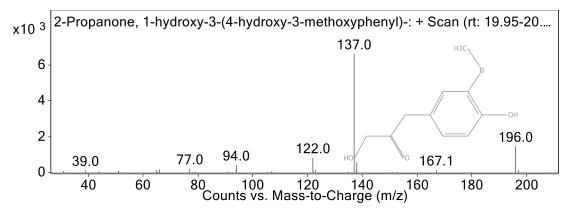


Figure S35. Electronic impact mass spectrum for 1-hydroxy-3-(4-hydroxy-3-methoxyphenyl)-2-propanone.

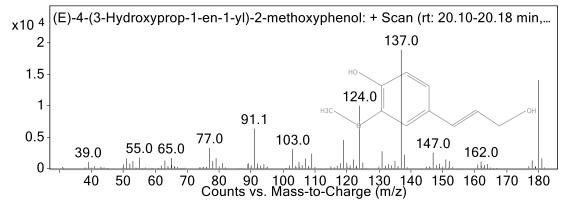


Figure S36. Electronic impact mass spectrum for (E)-4-(3-hidroxyprop-1-en-1-yl)-2-methoxyphenol.

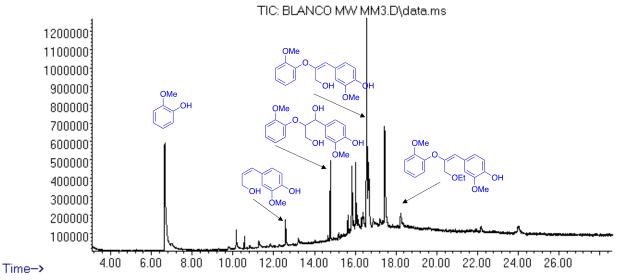


Figure S37. Chromatogram of MM-3 hydrogenolysis in EtOH/water with Et₃N/HCOOH without catalyst at 150 °C for 2h.

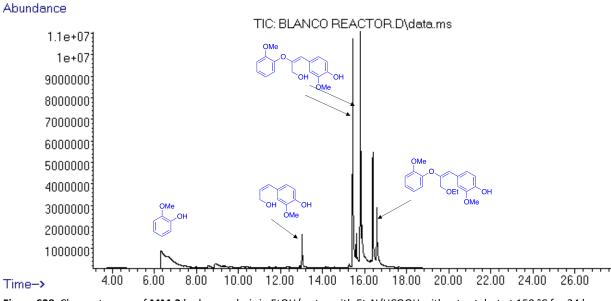


Figure S38. Chromatogram of MM-3 hydrogenolysis in EtOH/water with Et₃N/HCOOH without catalyst at 150 °C for 24 h.

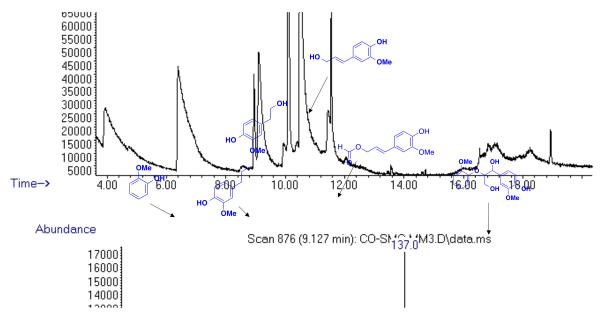
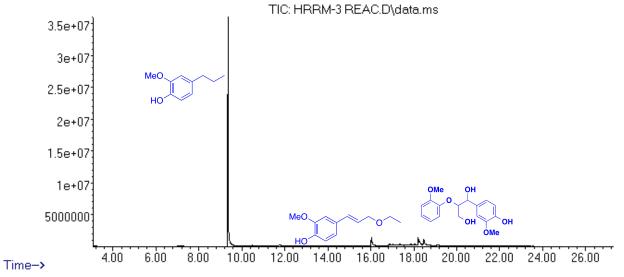
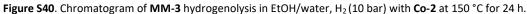


Figure S39. Chromatogram of **MM-3** hydrogenolysis in EtOH/water with HCOOH and without Et₃N with **Co-2** at 150 °C for 24 h. Abundance





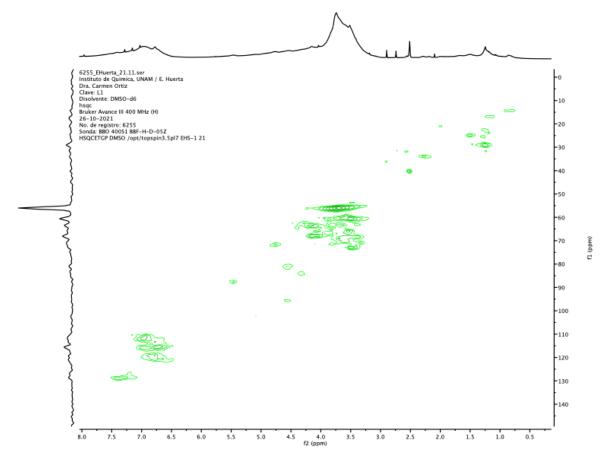


Figure S41. $^1\text{H}\text{-}^{13}\text{C}$ gHSQC NMR (DMSO-d6, 400 MHz) of pine dioxasolv lignin.

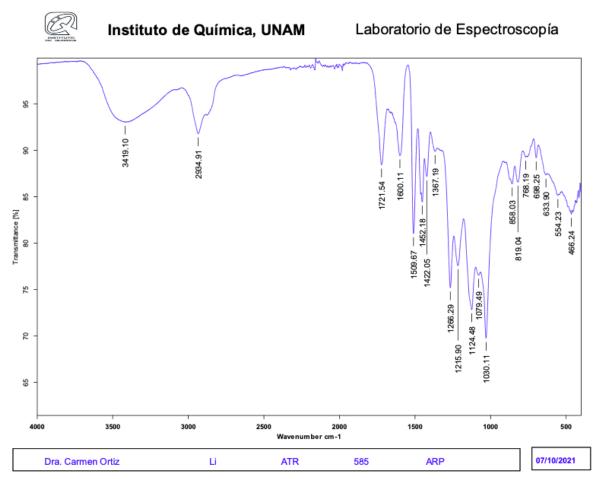


Figure S42. FTIR spectrum of pine dioxasolv lignin.

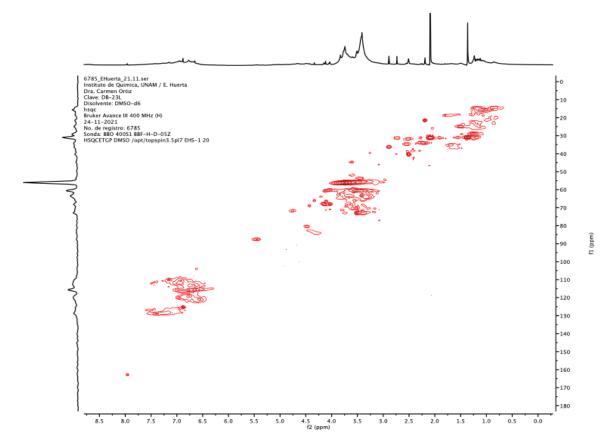


Figure S43. ¹H-¹³C gHSQC NMR (DMSO-d*6*, 400 MHz) of hydrogenolysis of dioxasolv lignin with **Co-1**.

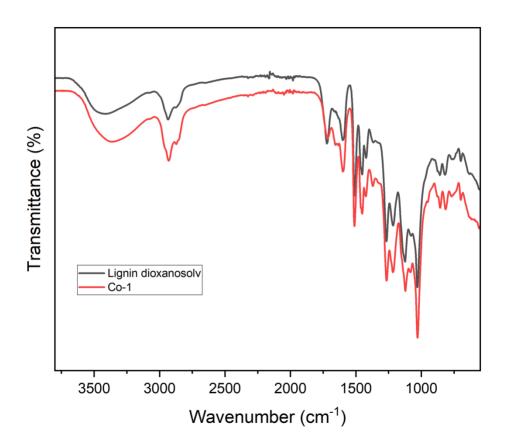


Figure S44. FTIR spectrum of dioxasolv lignin hydrogenolysis with Co-1.

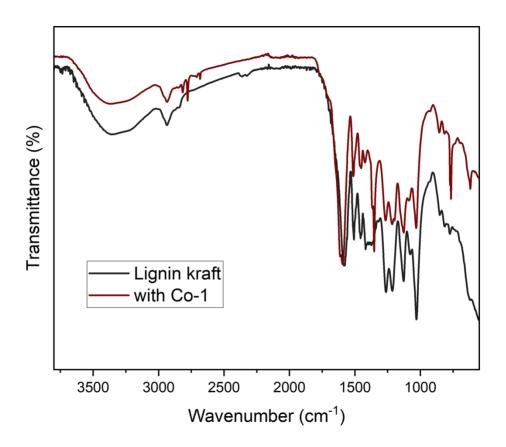


Figure S45. FTIR spectrum of kraft lignin hydrogenolysis with Co-1.

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