

Targeted Development of Sustainable Green Catalysts for Regioselective Acylation of Aromatic Ethers with Carboxylic Acids *via* Chlorosulfonic Acid Coated on Poly(guanidine-triazine-sulfonamide) Grafted γ -Fe₂O₃@quartz

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Characterization of γ -Fe₂O₃@quartz@PGTS-SO₃H nanocomposite

The FT-IR spectra of PGTSA, quartz, γ -Fe₂O₃@quartz, γ -Fe₂O₃@quartz@PGTSA, γ -Fe₂O₃@quartz@PGTS-SO₃H composite systems are shown in Fig. S1. In FT-IR spectrum of PGTSA, the broadband at 3200-3700 cm⁻¹ indicates the presence of NH and NH₂. The vibration bands at 1338 and 1160 cm⁻¹ (because of the O=S=O stretching) demonstrate the presence of sulfone bridges in the PGTSA. Also, absorption bands at 1664 and 1682 cm⁻¹ indicate the presence of guanidine and melamine (Fig. S1a). In the quartz curve, the Si-O stretching vibration appeared at 1099 cm⁻¹ (Fig. S1b). Fig. S1c illustrates the FT-IR spectrum of γ -Fe₂O₃@quartz. FT-IR analysis of γ -Fe₂O₃@quartz displayed a broad absorption band at 3000-3500 cm⁻¹, which is indexed to -SiOH groups' OH symmetric stretching vibrations. Two adsorption bands at 1087 cm⁻¹ are indexed to the symmetric and asymmetric stretching vibrations of the Si-O-Si bond on the quartz surface. Also, the γ -Fe₂O₃ in the quartz structure can be detected through the band at 605 cm⁻¹ (indexed to Fe-O bonds' stretching vibration). It is noteworthy that this band is covered by Si-O bonds' stretching vibrations. These results conform with those in the literature. After PGTSA immobilization on the γ -Fe₂O₃@quartz surface, the Fe-O bonds shifted to a higher wavelength (620 cm⁻¹) in the γ -Fe₂O₃@quartz@PGTSA spectrum. The broad peak at 3100-3400 cm⁻¹ is attributed to the polymeric structure's NH and NH₂ groups (Fig. S1d). In the case of the γ -Fe₂O₃@quartz@PGTSA-SO₃H catalyst, some additional peaks appeared at 629 cm⁻¹, suggesting the successful anchoring of the chlorosulfonic acid to the γ -Fe₂O₃@quartz@PGTSA nanohybrid (Fig. S1e).

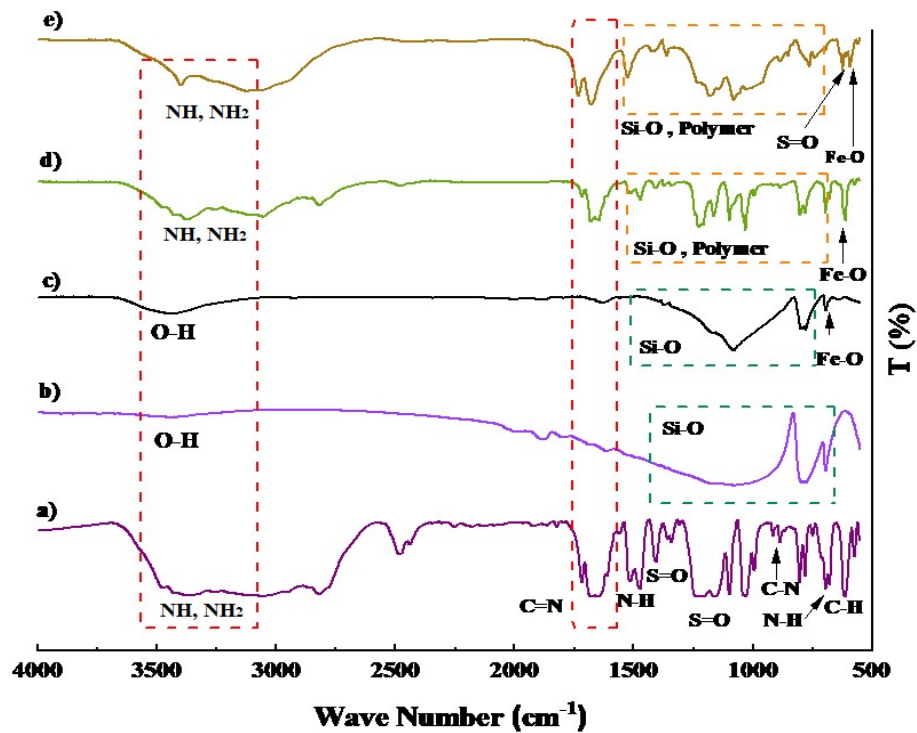


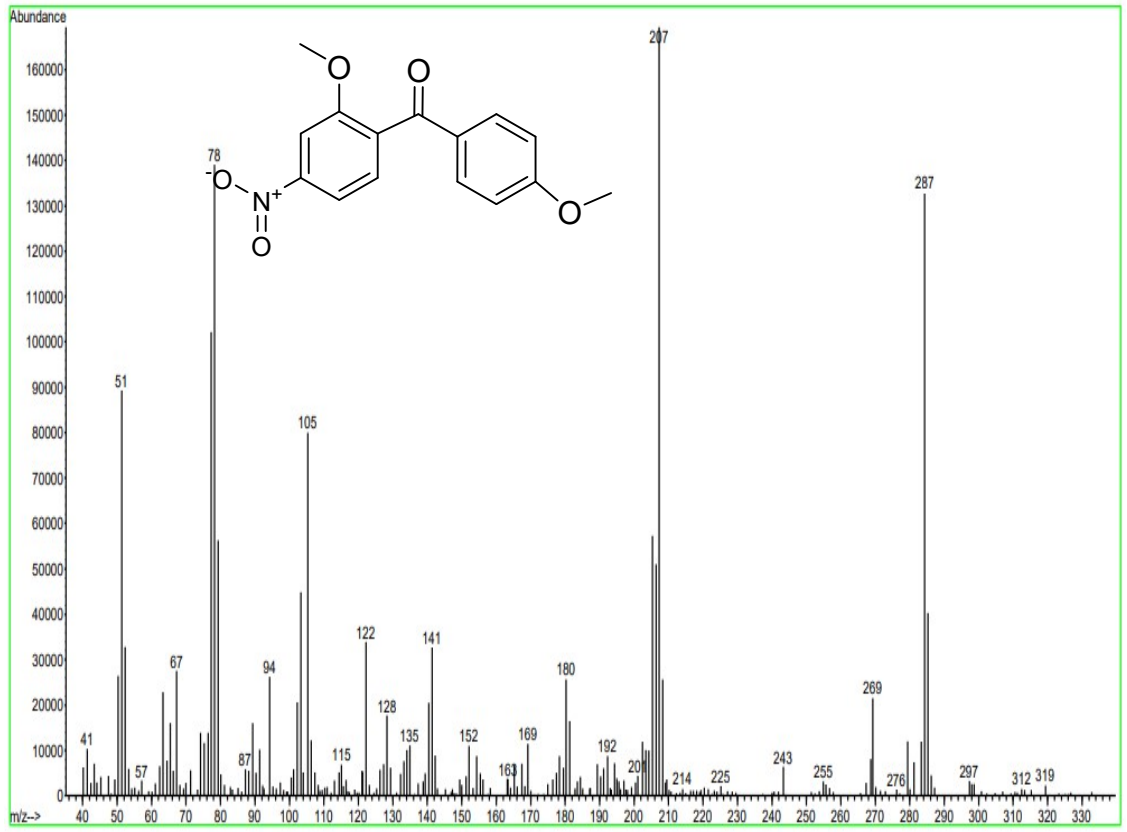
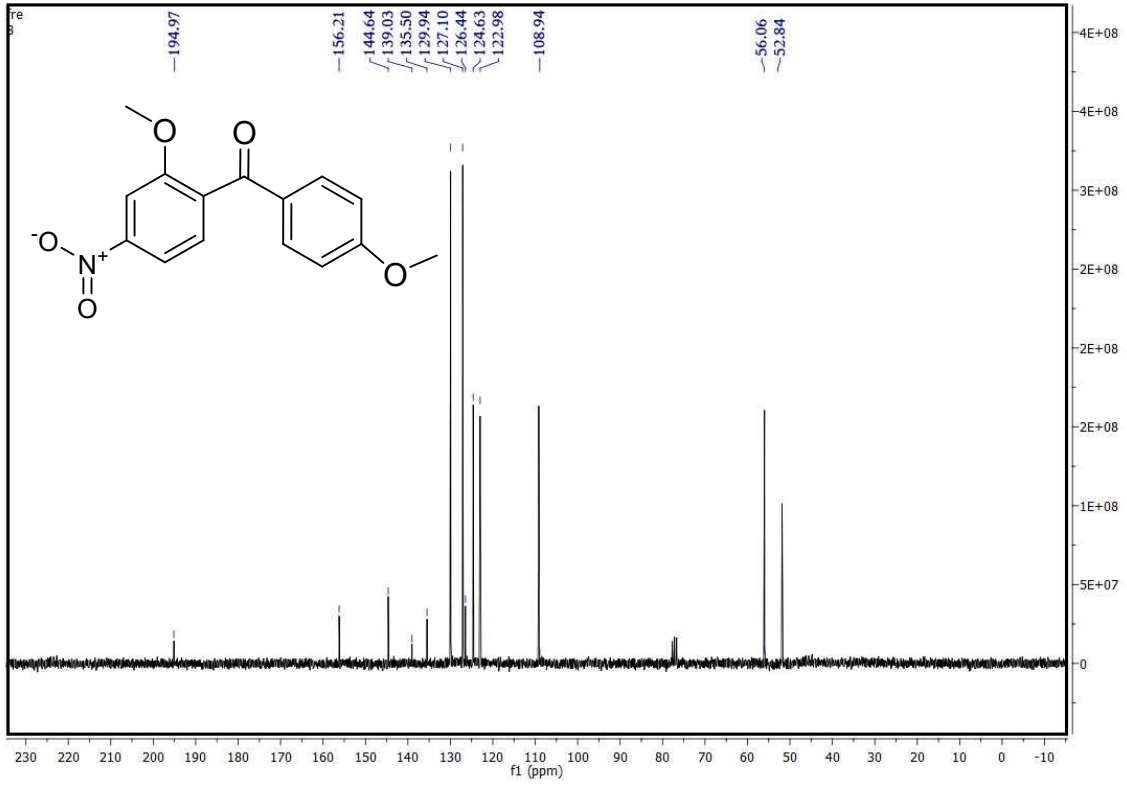
Fig. S1. FT-IR spectra of PGTSA (a), quartz (b), γ -Fe₂O₃@quartz (c), γ -Fe₂O₃@quartz@PGTSA (d), γ -Fe₂O₃@quartz@PGTS-SO₃H (e).



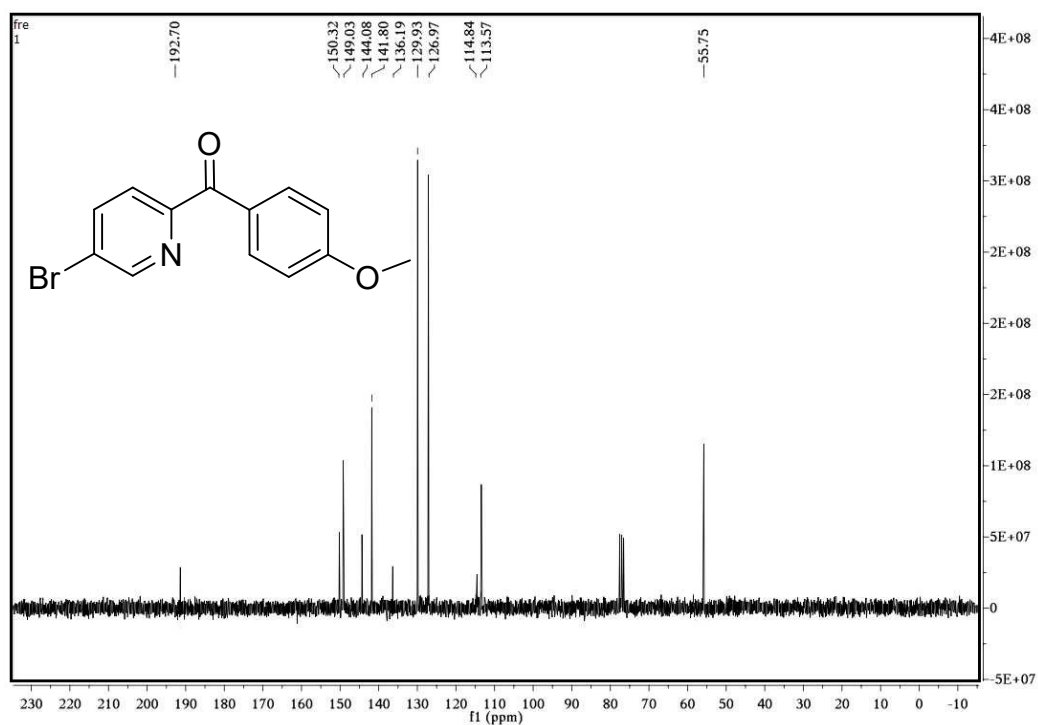
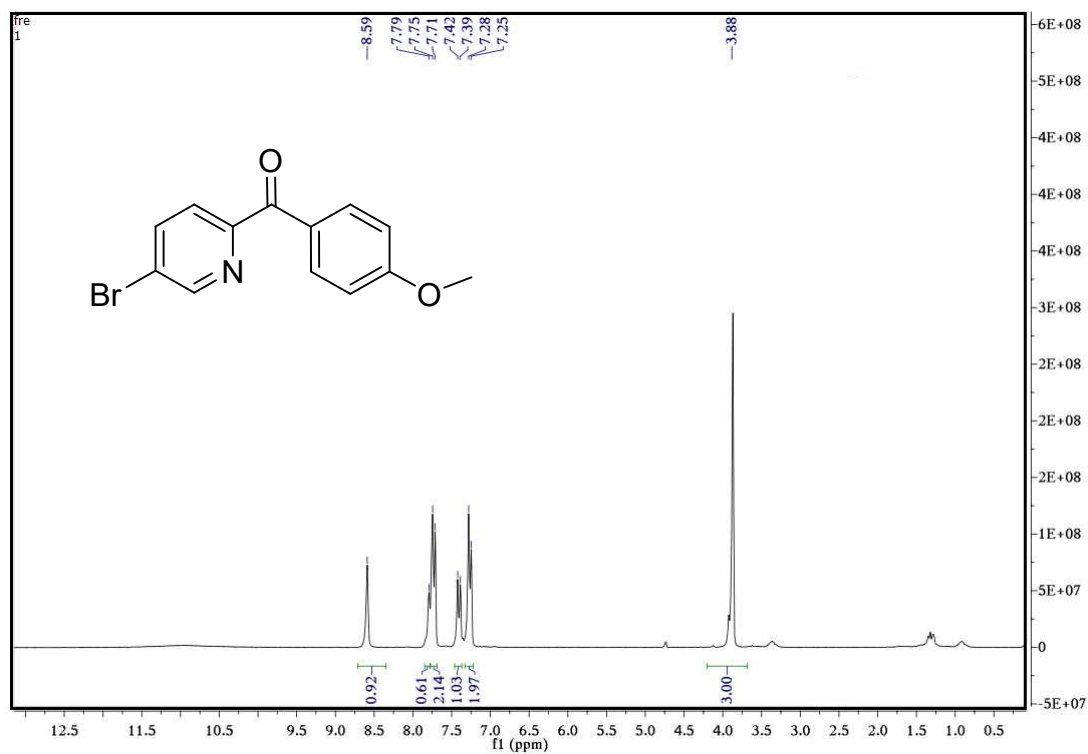
Fig. S2. Images of magnetic recovery A) before the reaction, B) after the reaction.

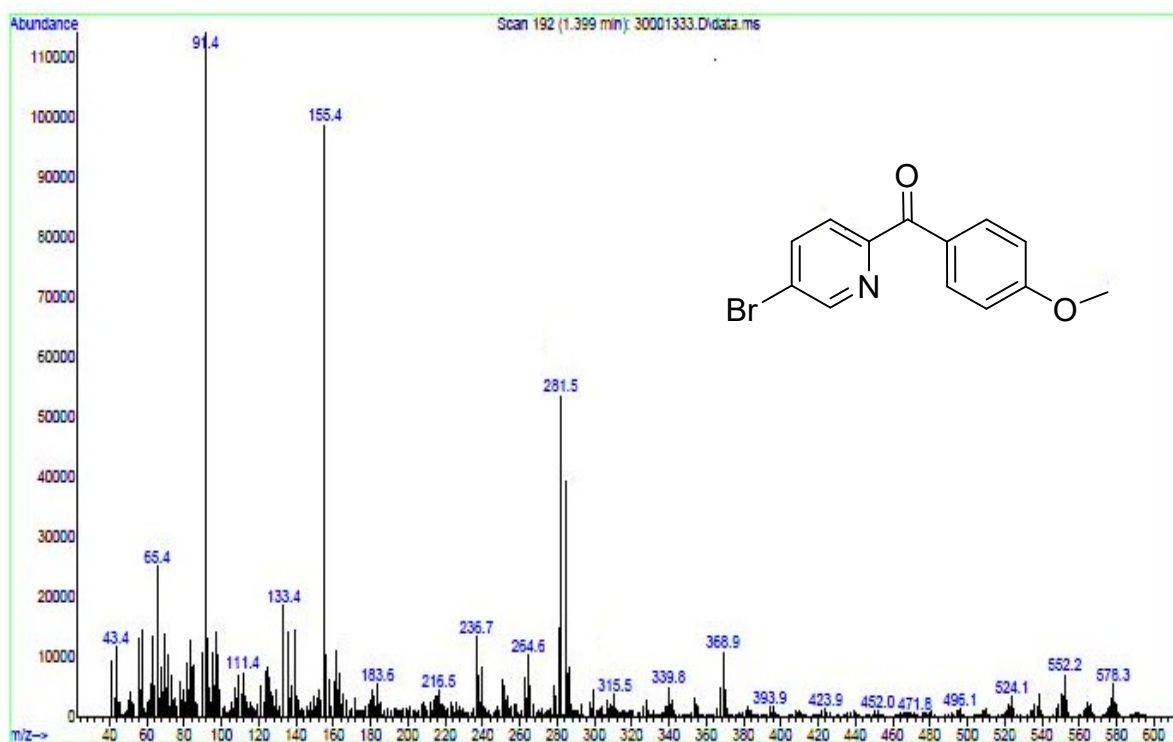
^1H NMR (250 MHz, CDCl_3) δ 9.29 (s, 1H), 7.80 (d, $J = 9.2$ Hz, 1H), 7.62 (d, $J = 8.0$ Hz, 2H), 7.50 (s, 1H), 7.23 (d, $J = 7.7$ Hz, 2H), 3.84 (s, 3H), 3.45 (s, 3H). ^{13}C NMR (63 MHz, CDCl_3) δ 194.97, 156.21, 144.64, 139.03, 135.50, 129.94, 127.10, 126.44, 124.63, 122.98, 108.94, 56.06, 52.84, MS m/z (%): 287.



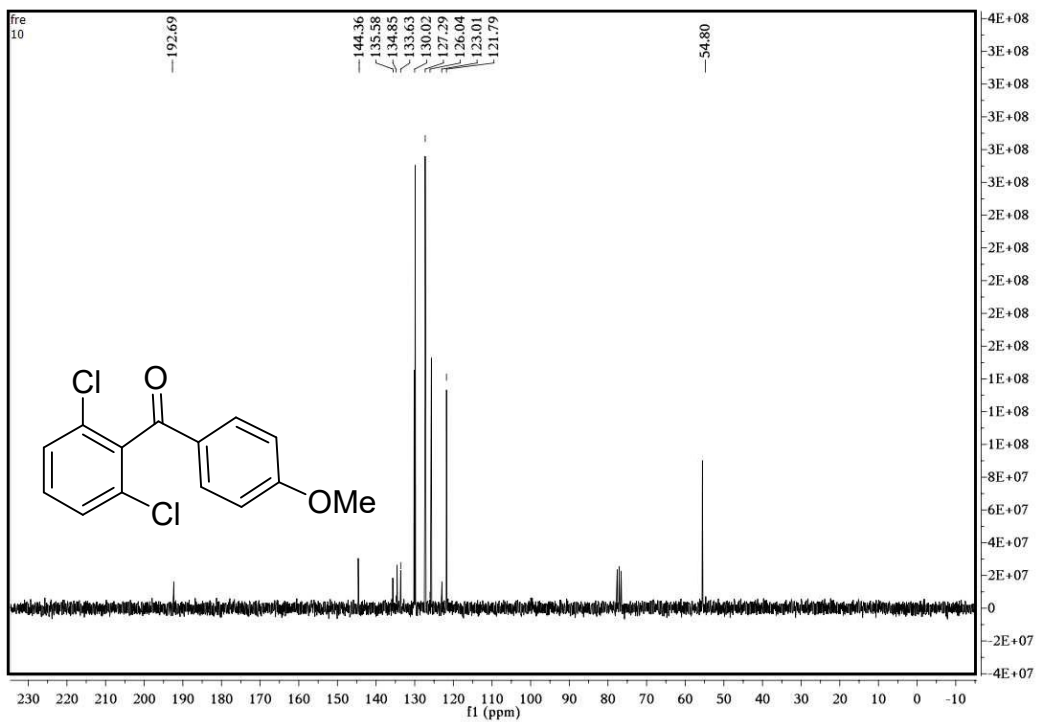
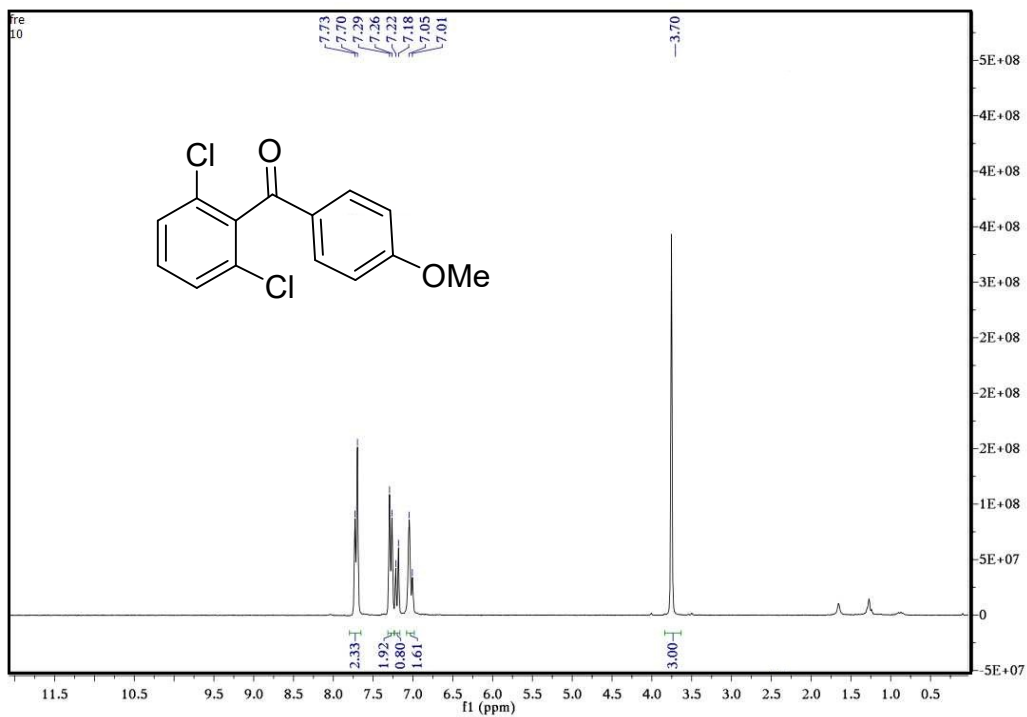


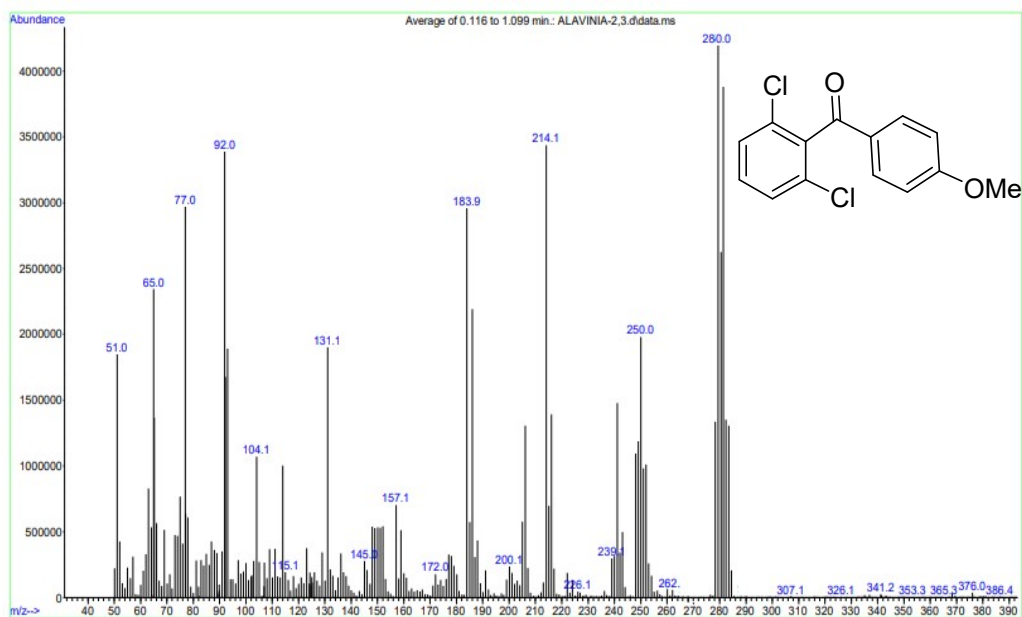
^1H NMR (250 MHz, CDCl_3) δ 8.59 (s, 1H), 7.79 (s, 1H), 7.73 (d, $J = 8.3$ Hz, 2H), 7.40 (d, $J = 8.7$ Hz, 1H), 7.26 (d, $J = 8.1$ Hz, 2H), 3.88 (s, 3H). ^{13}C NMR (63 MHz, CDCl_3) δ 192.70, 150.32, 149.03, 144.08, 141.80, 136.19, 129.93, 126.97, 114.84, 113.57, 55.75. MS m/z (%): 290.



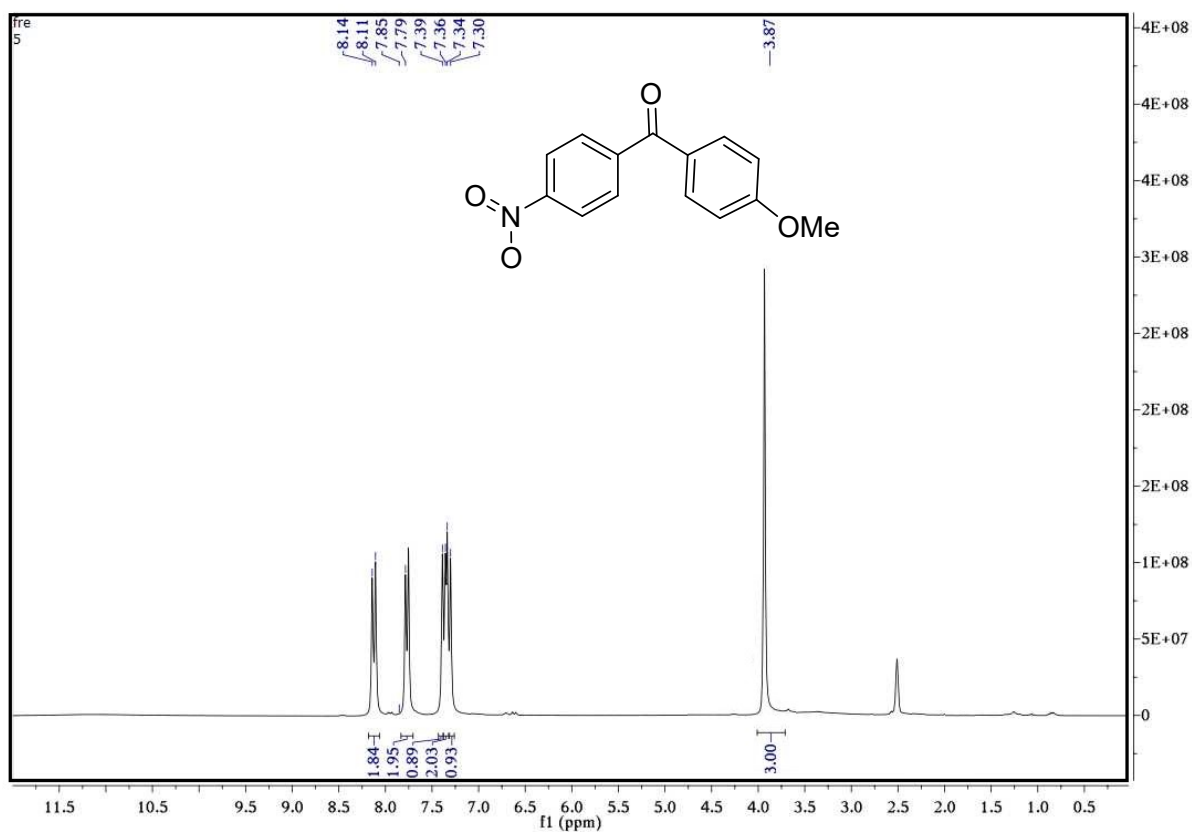


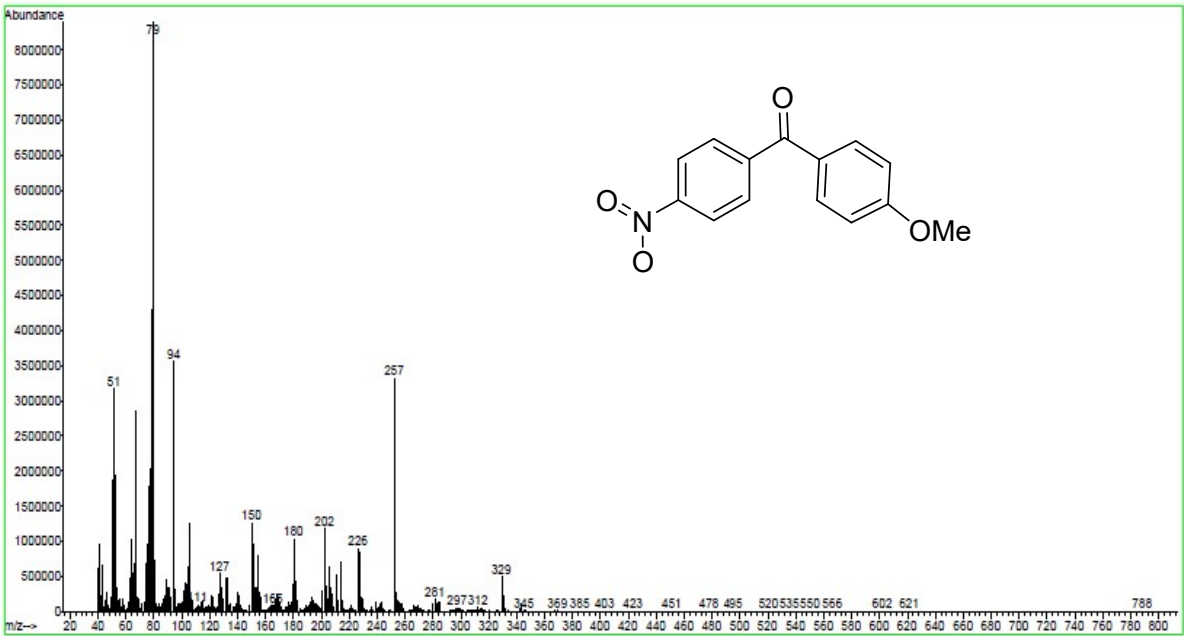
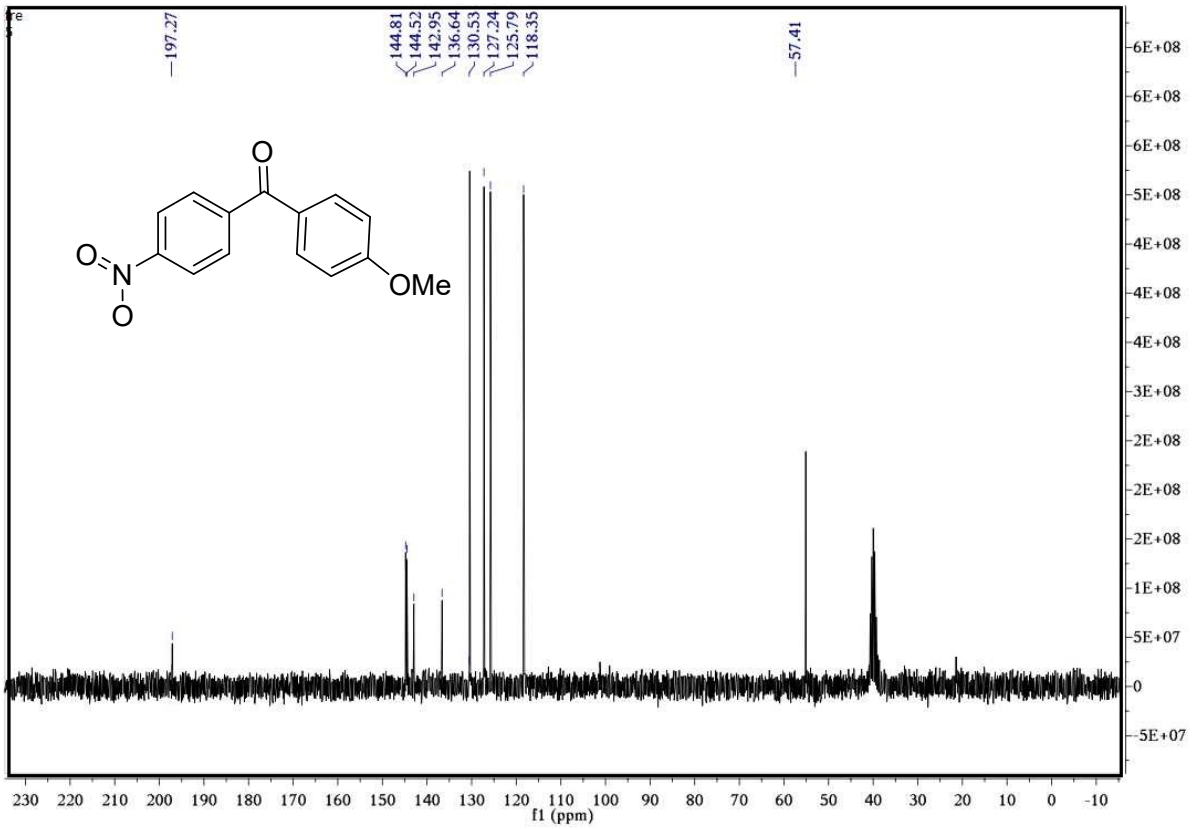
^1H NMR (250 MHz, CDCl_3) δ 7.71 (d, $J = 7.8$ Hz, 2H), 7.28 (d, $J = 7.7$ Hz, 2H), 7.20 (d, $J = 8.6$ Hz, 1H), 7.03 (d, $J = 9.7$ Hz, 2H), 3.70 (s, 3H). ^{13}C NMR (63 MHz, CDCl_3) δ 192.69, 144.36, 135.58, 134.85, 133.63, 130.02, 127.29, 126.04, 123.01, 121.79, 54.80. MS m/z (%): 280.



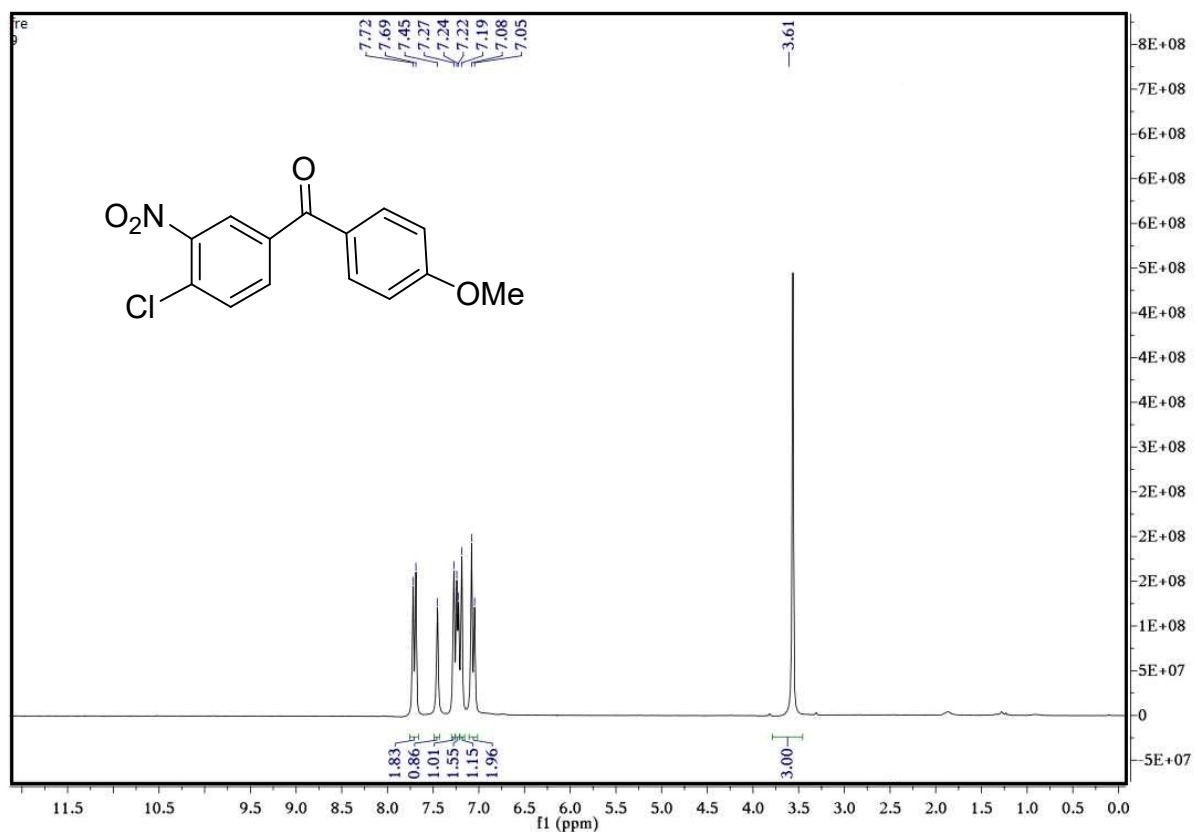


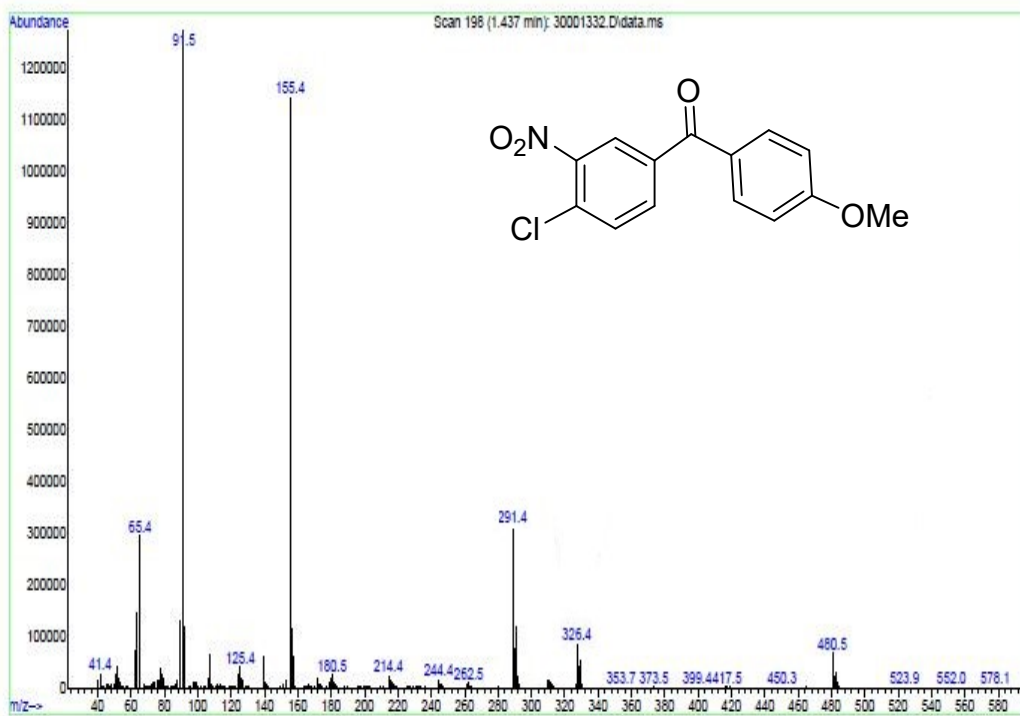
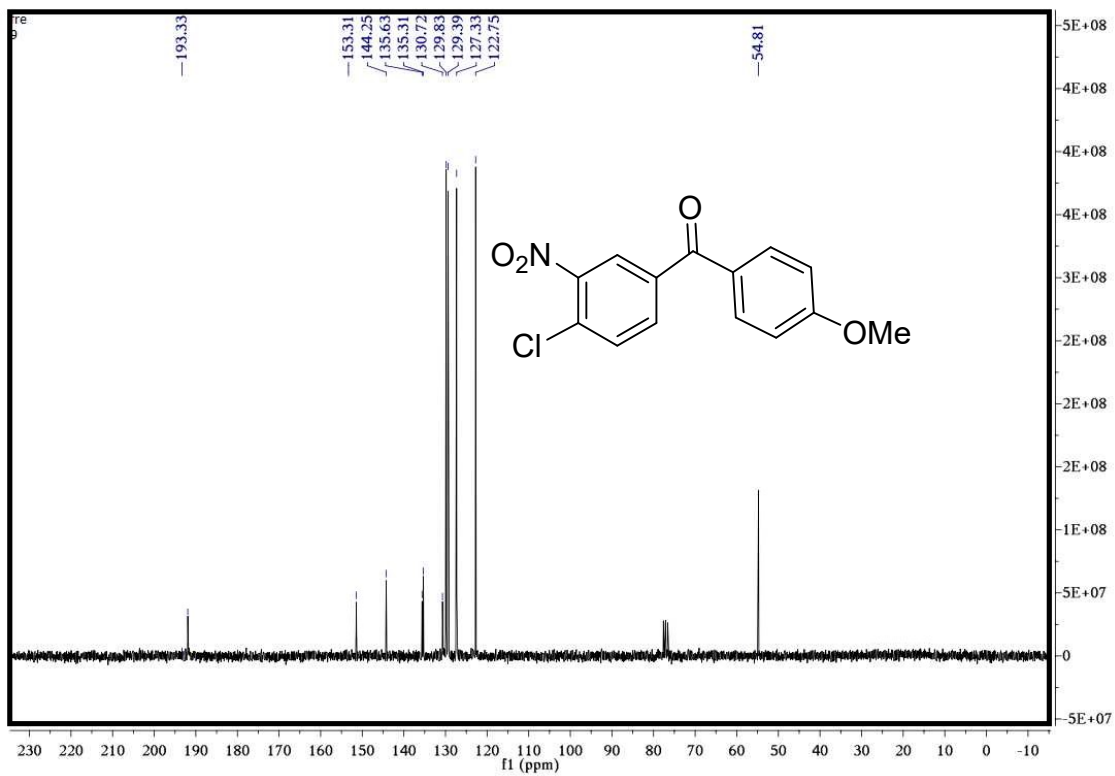
^1H NMR (250 MHz, DMSO-d_6) δ 8.12 (d, $J = 8.9$ Hz, 2H), 7.77 (d, $J = 7.9$ Hz, 2H), 7.39 (s, 1H), 7.35 (d, $J = 4.8$ Hz, 2H), 7.30 (s, 1H), 3.87 (s, 3H). ^{13}C NMR (63 MHz, DMSO) δ 197.27, 144.81, 144.52, 142.95, 136.64, 130.53, 127.24, 125.79, 118.35, 57.41. MS m/z (%): 257.



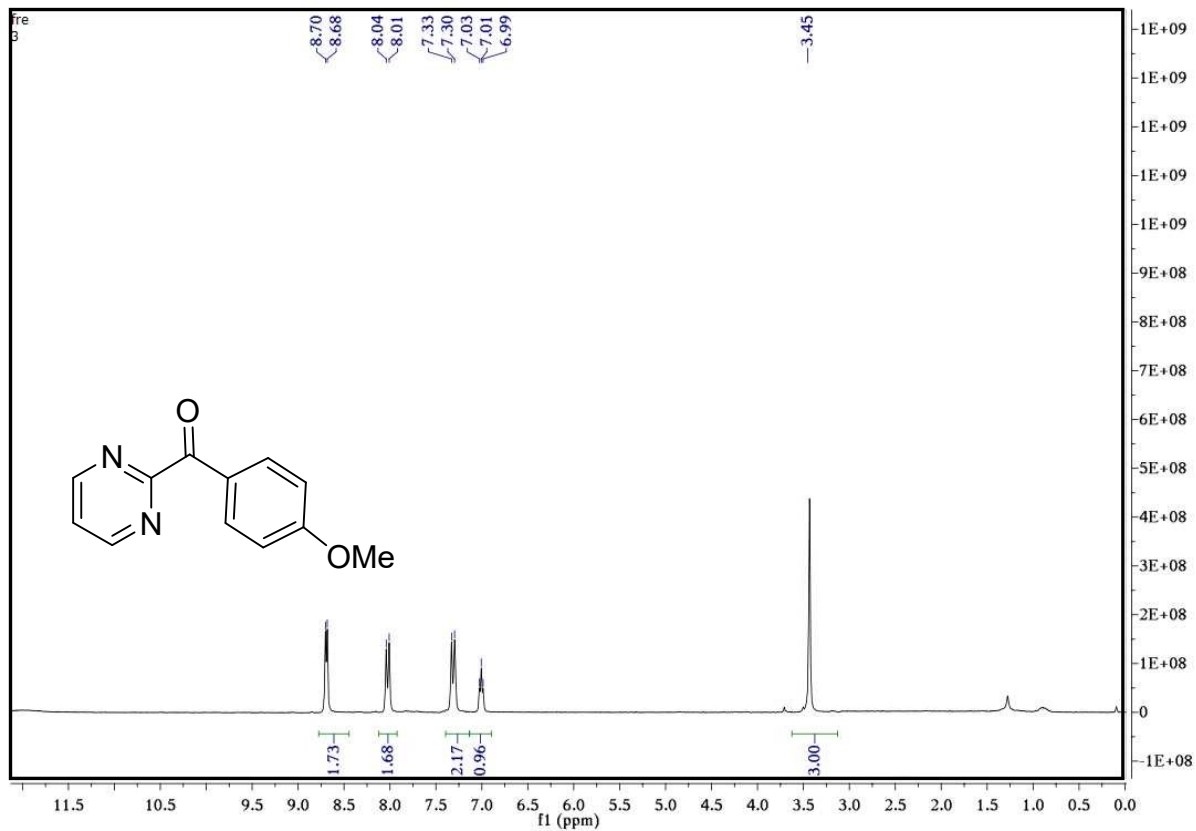


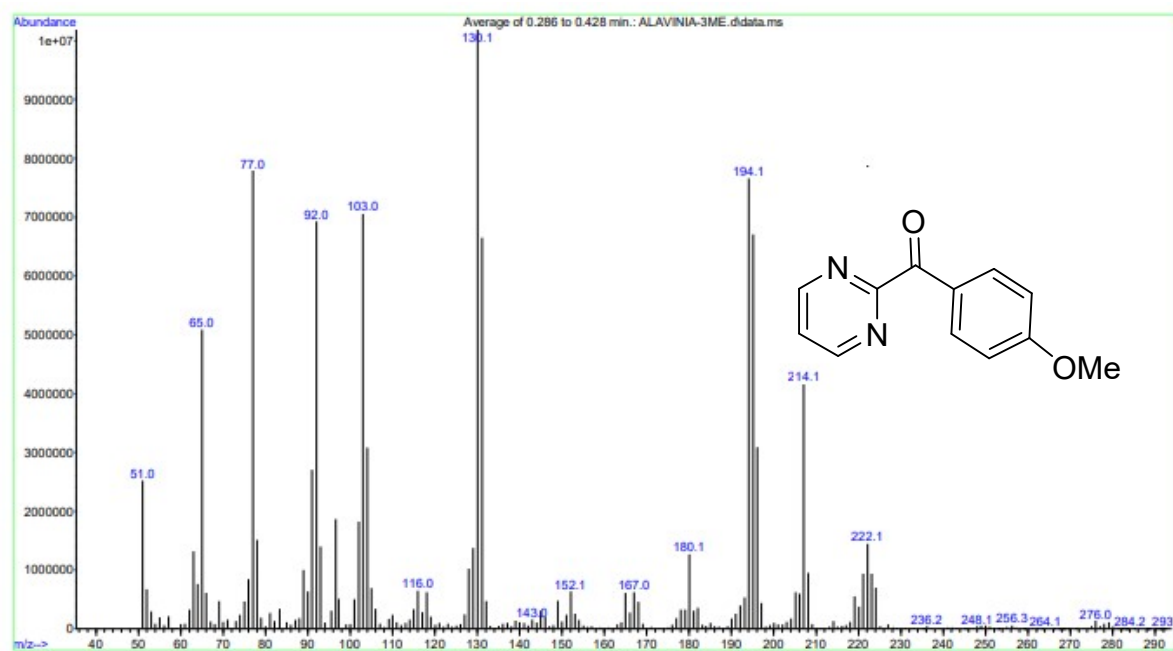
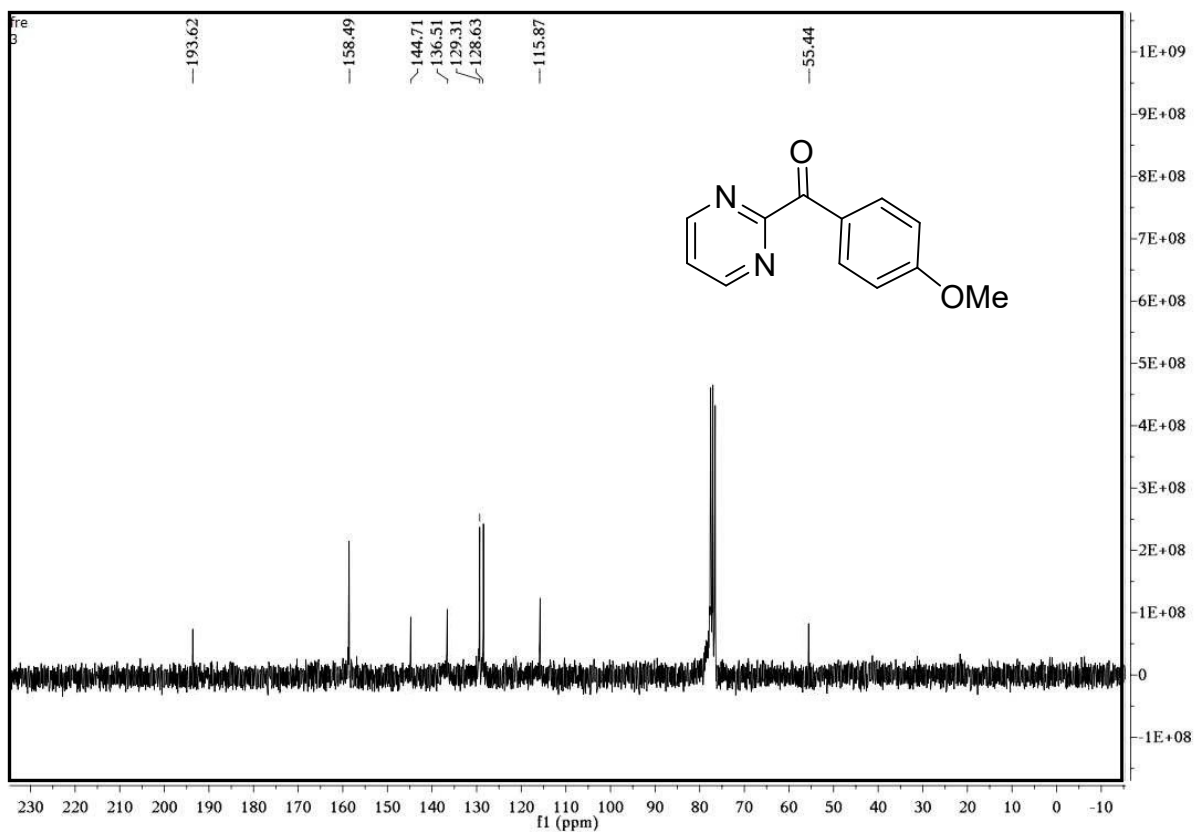
^1H NMR (250 MHz, CDCl_3) δ 7.70 (d, $J = 7.9$ Hz, 2H), 7.45 (s, 1H), 7.27 (s, 1H), 7.23 (d, $J = 5.0$ Hz, 1H), 7.19 (s, 1H), 7.06 (d, $J = 8.4$ Hz, 2H), 3.61 (s, 3H). ^{13}C NMR (63 MHz, CDCl_3) δ 193.33, 153.31, 144.25, 135.63, 135.31, 130.72, 129.83, 129.39, 127.33, 122.75, 54.81. MS m/z (%): 291.



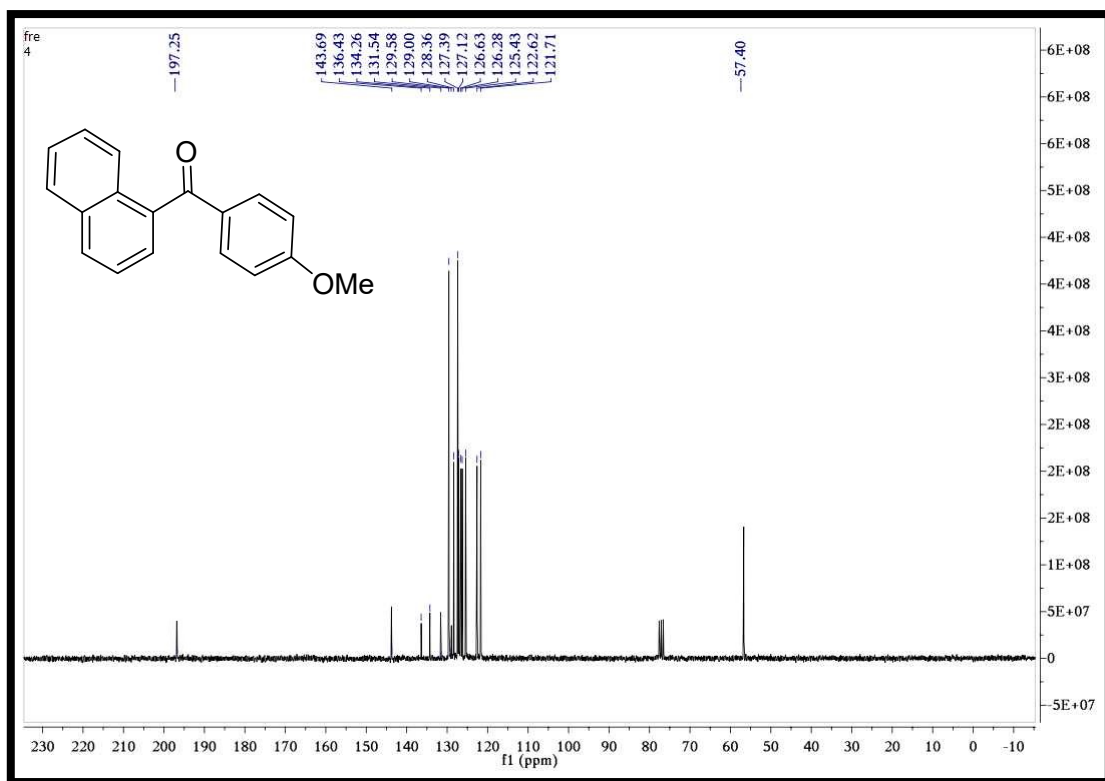
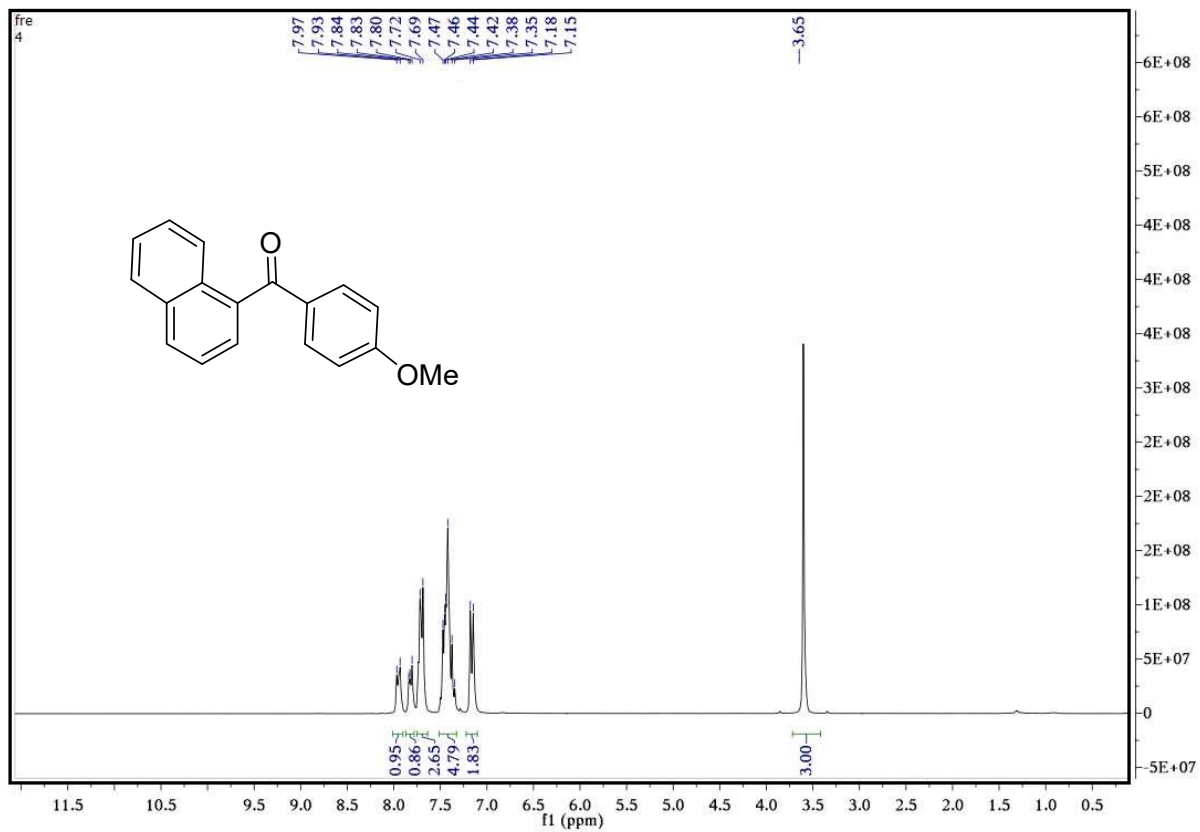


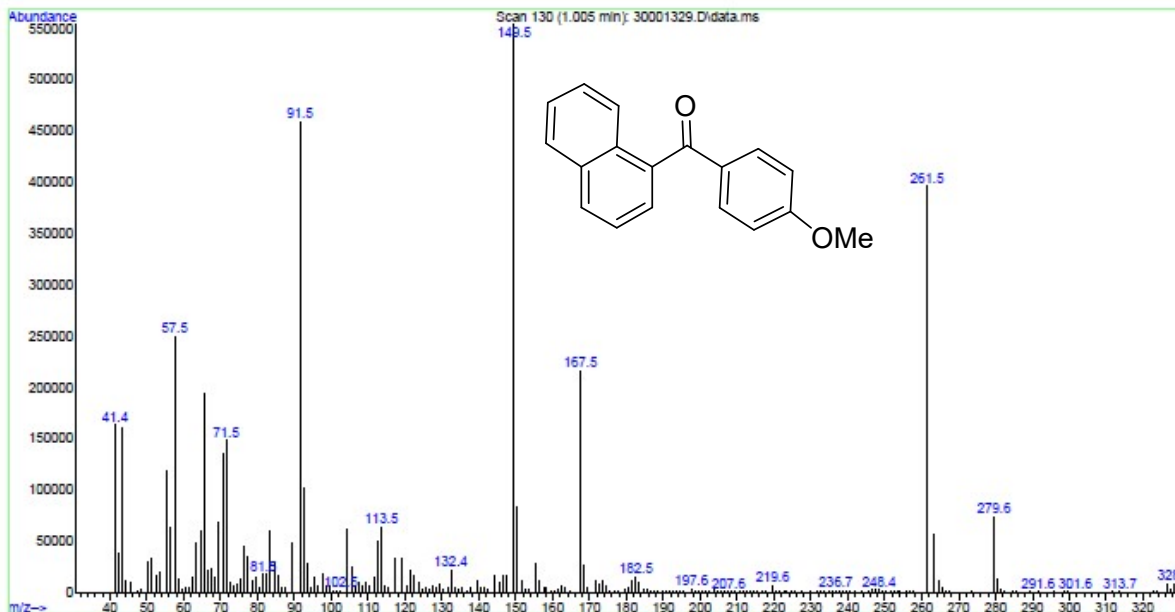
^1H NMR (250 MHz, CDCl_3) δ 8.69 (d, $J = 4.8$ Hz, 2H), 8.02 (d, $J = 8.0$ Hz, 2H), 7.31 (d, $J = 8.2$ Hz, 2H), 7.01 (t, $J = 4.8$ Hz, 1H), 3.45 (s, 3H). ^{13}C NMR (63 MHz, CDCl_3) δ 193.62, 158.49, 144.71, 136.51, 129.51, 128.63, 115.87, 55.44. MS m/z (%): 214.



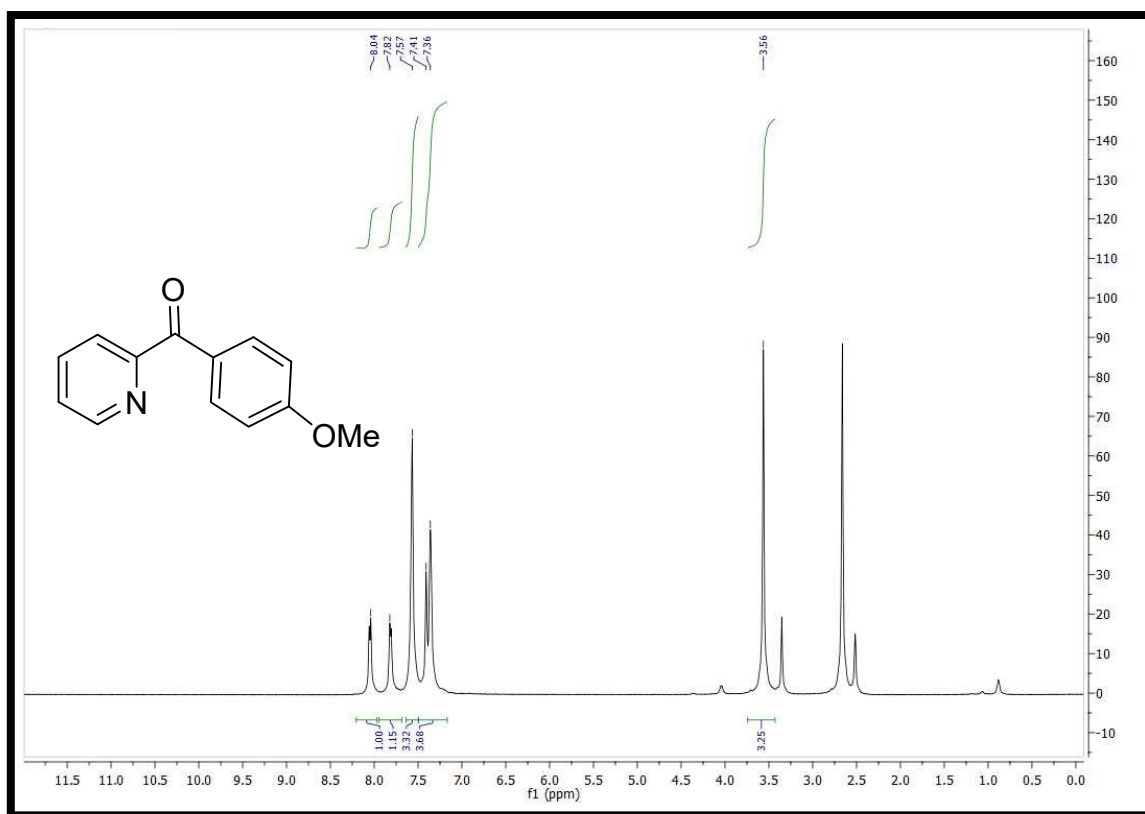


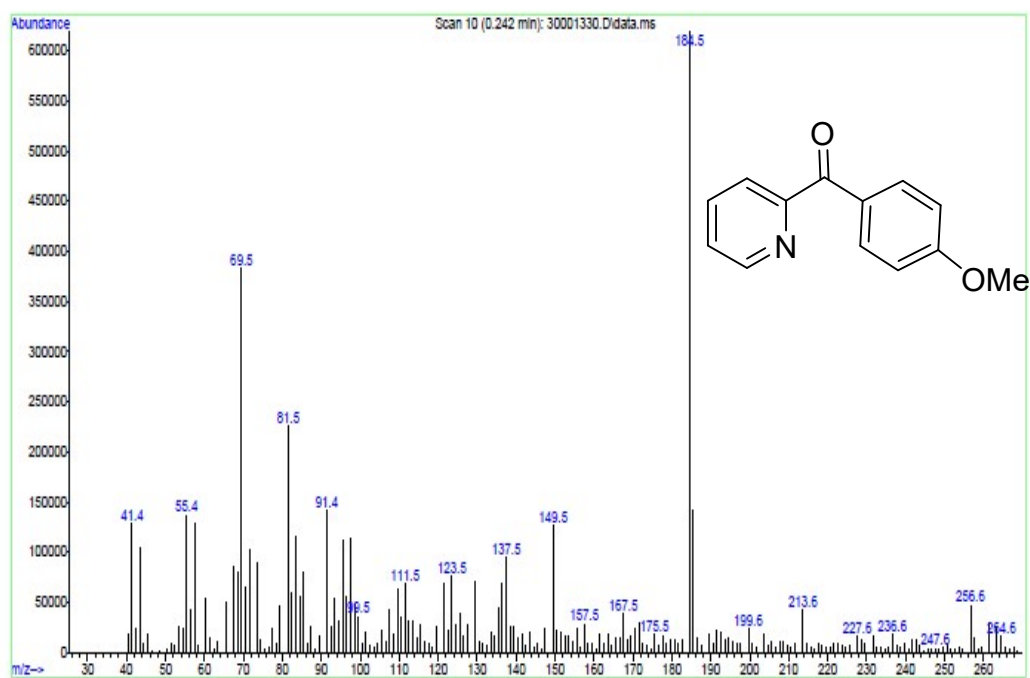
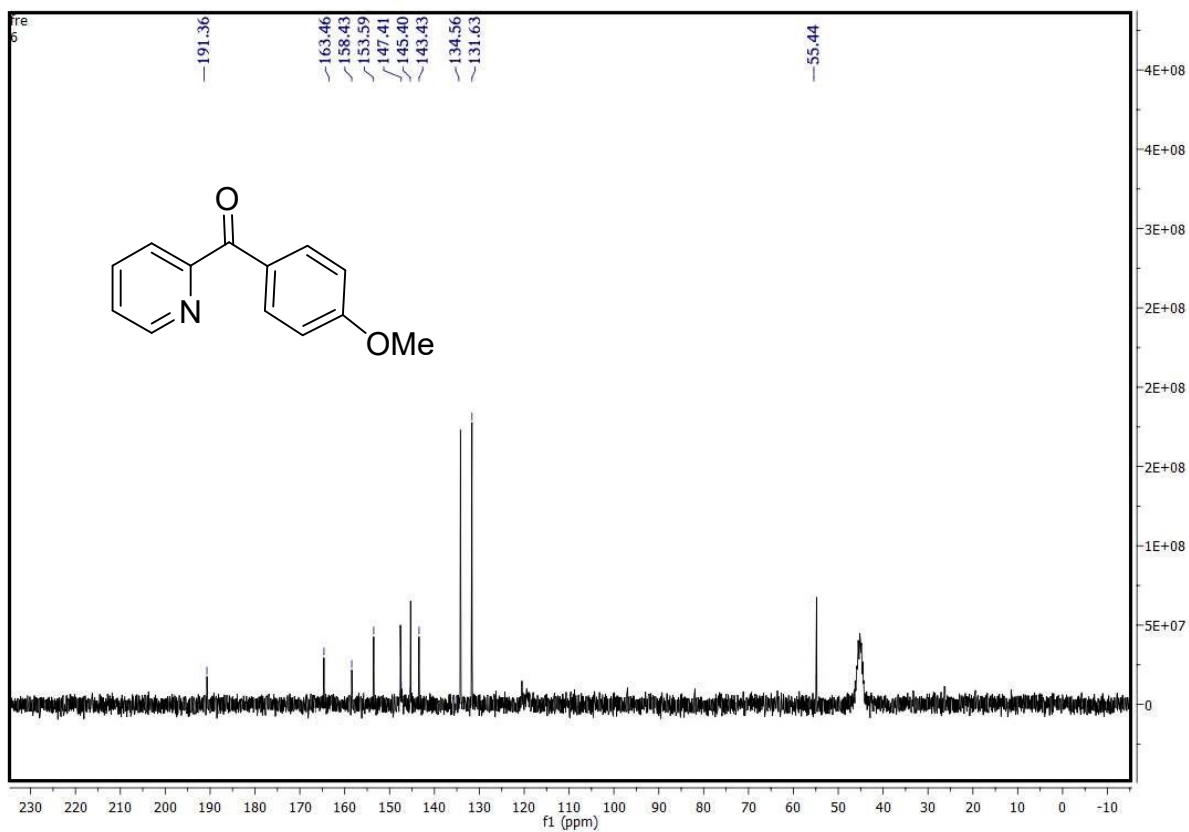
¹H NMR (250 MHz, CDCl₃) δ 7.95 (d, *J* = 8.9 Hz, 1H), 7.87 – 7.78 (m, 1H), 7.75 – 7.64 (m, 3H), 7.51 – 7.33 (m, 5H), 7.16 (d, *J* = 7.9 Hz, 2H), 3.65 (s, 3H). ¹³C NMR (63 MHz, CDCl₃): δ: 57.40, 121.71, 122.62, 125.43, 126.28, 126.63, 127.12, 127.39, 128.36, 128.94, 129.58, 131.62, 134.26, 136.43, 143.78, 197.25. MS m/z (%): 261.





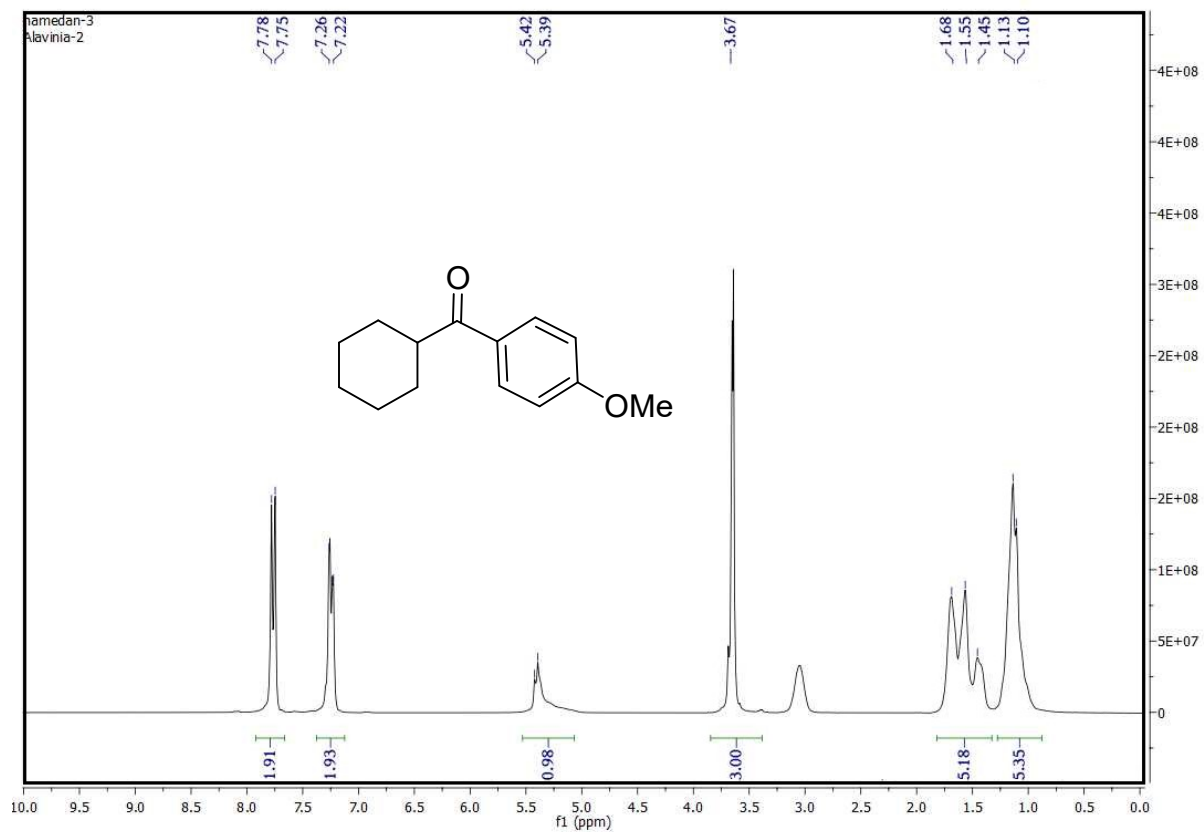
^1H NMR (250 MHz, DMSO-d_6) δ 8.04 (d, $J = 7.9$ Hz, 1H), 7.82 (d, $J = 7.9$ Hz, 1H), 7.37 (s, 3H), 7.39 (d, $J = 8.2$ Hz, 3H), 3.56 (s, 3H). ^{13}C NMR (63 MHz, DMSO-d_6) δ 191.36, 163.46, 158.43, 153.59, 147.41, 145.40, 143.43, 134.56, 131.63, 55.44. MS m/z (%): 213.

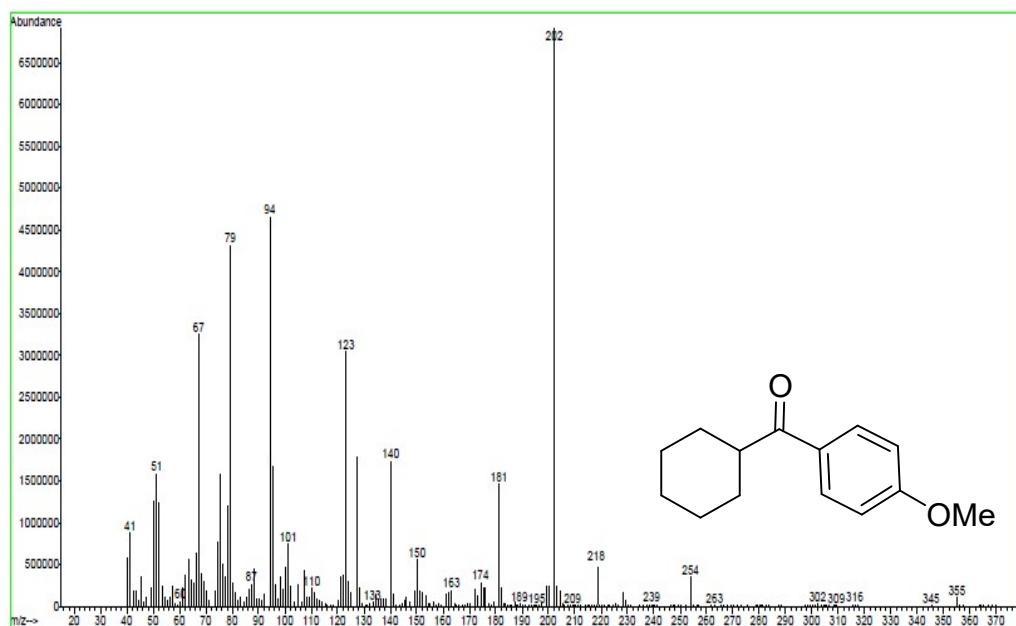
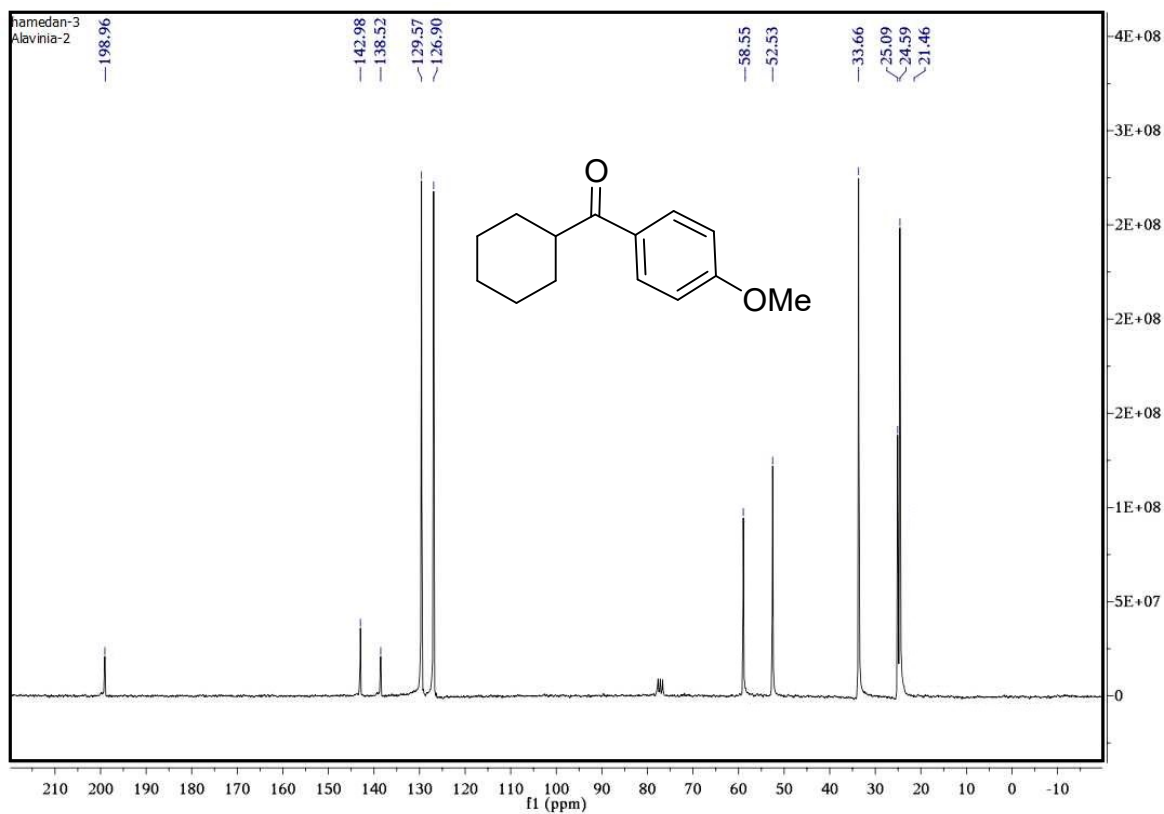




¹H NMR (250 MHz, CDCl₃) δ 7.76 (d, *J* = 8.1 Hz, 2H), 7.24 (d, *J* = 9.7 Hz, 2H), 5.41 (d, *J* = 7.0 Hz, 1H), 3.67 (s, 3H), 1.82 – 1.33 (m, 5H), 1.11 (d, *J* = 7.3 Hz, 5H), ¹³C NMR (63 MHz,

CDCl₃) δ 198.96, 142.98, 138.52, 129.57, 126.90, 58.55, 52.53, 33.66, 25.09, 24.59. MS m/z (%): 218.





^1H NMR (250 MHz, CDCl_3) δ 7.78 (d, $J = 6.6$ Hz, 2H), 7.24 (d, $J = 7.4$ Hz, 2H), 5.39 (d, $J = 11.2$ Hz, 1H), 3.64 (s, 3H), 1.16 (s, 9H). ^{13}C NMR (63 MHz, CDCl_3) δ 196.50, 142.68, 140.63, 129.42, 126.92, 59.15, 54.38, 30.03. MS m/z (%): 192.8.

