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Electronic Supplementary Information

Using a dual-emission Sm(III)-macrocycle as the perceptive lab-on-a-

molecule chemosensor toward selective and discriminative detection

of nitroaromatic explosives

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Crystal structure determination and refinement

A high-quality single-crystal sample of complex Sm-2₁ was covered with glue and mounted on glass fiber for data collection. Crystallographic data were collected at 150(2) K on a Bruker SMART 1K CCD diffractometer, using graphite mono-chromated MoK α radiation (λ = 0.71073 Å). Absorption corrections were performed to all data and the structure was solved by direct method and refined by full-matrix least-squares method on F_{obs}² by using the SHELXTL-PC software package [1]. All non-H atoms were anisotropically refined and all hydrogen atoms were inserted in the calculated positions assigned fixed isotropic thermal parameters and allowed to ride on their respective parent atoms. A summary of the crystal data, experimental details and refinement results for Sm-2₁ was listed in Table S1. Selected bond lengths and bond angles of Sm-2₁ were tabulated in Table S2, and hydrogen bond parameters were shown in Table S3.

References

1 Sheldrick, G. M. SHELXTL (Version 6.10). Software Reference Manual; Madison, Wisconsin (USA): Bruker AXS, Inc.: **2000**.

Tables

Table S1 Crystal data and structural refinements for Sm-2₁.

Complex	Sm-2 ₁	
Empirical formula	$C_{30}H_{31}BrCl_2N_5O_{10}Sm$	
Formula weight	922.76	
Temperature / K	293(2)	
Wavelength / Å	0.71073	
Crystal Size (mm)	0.14×0.21×0.26	
Crystal system	Monoclinic	
Space group	C2/c	
<i>a</i> / Å	43.493(3)	
b / Å	8.4563(6)	
<i>c</i> / Å	18.8102(15)	
α / °	90	
β / °	93.291(2)	
γ/°	90	
$V/\text{\AA}^3$	6906.8(9)	
$Z / D_{\text{calcd}} (\text{g} / \text{cm}^3)$	8/1.775	
<i>F</i> (000)	3656	
μ / mm ⁻¹	3.074	
h_{\min} / h_{\max}	-52/56	
k_{\min} / k_{\max}	-10/11	
l _{min} / l _{max}	-24/24	
Data / parameters	8050/442	
$R_1, WR_2 [I > 2\sigma(I)]^a$	$R_1 = 0.0406, \mathrm{w}R_2 = 0.0803$	
R_1 , w R_2 (all data) ^{<i>a</i>}	$R_1 = 0.0730, wR_2 = 0.0906$	
S	1.00	
Max/min $\Delta \rho$ /e Å ⁻³	1.05/-0.99	

 $\frac{1}{a R_1 = \Sigma ||Fo| - |Fc|| / \Sigma |Fo|, wR_2 = [\Sigma [w(Fo^2 - Fc^2)^2] / \Sigma w(Fo^2)^2]^{1/2}}$

Table S2 Selected	bond distances	(Å) and angle	es (°) in Sm-2 I.

Bond distances		Bond angles	
Sm-2 ₁			
Sm1–O1	2.319(3)	O1–Sm1–O2	120.74(9)
Sm1–O2	2.723(3)	O1–Sm1–O3	174.83(10)
Sm1–O3	2.675(3)	O1–Sm1–O4	68.92(9)
Sm1–O4	2.355(3)	O1–Sm1–O5	116.98(10)
Sm1–O5	2.581(3)	O1-Sm1-O6	73.81(10)
Sm1–O6	2.529(3)	O1–Sm1–O8	70.53(10)

Sm1–O8	2.579(4)	O1–Sm1–O9	109.11(11)
Sm1–O9	2.499(4)	O1-Sm1-N2	66.26(10)
Sm1-N2	2.688(4)	O1-Sm1-N3	117.45(10)
Sm1-N3	2.610(3)	O2–Sm1–O3	59.43(9)
		O2-Sm1-O4	137.50(9)
		O2–Sm1–O5	70.33(10)
		O2-Sm1-O6	72.06(10)
		O2–Sm1–O8	127.18(10)
		O2–Sm1–O9	80.64(10)
		O2-Sm1-N2	62.83(10)
		O2-Sm1-N3	121.70(10)
		O3–Sm1–O4	114.76(9)
		O3–Sm1–O5	68.16(10)
		O3–Sm1–O6	110.53(10)
		O3–Sm1–O8	105.07(10)
		O3–Sm1–O9	65.72(11)
		O3–Sm1–N2	110.91(10)
		O3–Sm1–N3	62.31(10)
		O4–Sm1–O5	69.23(10)
		O4–Sm1–O6	72.17(10)
		O4–Sm1–O8	95.33(10)
		O4–Sm1–O9	138.80(11)
		O4–Sm1–N2	133.23(10)
		O4–Sm1–N3	68.55(10)
		O5–Sm1–O6	49.63(10)
		O5–Sm1–O8	156.31(11)
		O5–Sm1–O9	133.35(11)
		O5–Sm1–N2	121.65(10)
		O5–Sm1–N3	87.10(10)
		O6–Sm1–O8	144.32(11)
		O6–Sm1–O9	148.69(11)
		O6–Sm1–N2	83.17(11)
		O6–Sm1–N3	129.71(10)
		O8–Sm1–O9	49.76(11)
		O8–Sm1–N2	82.04 (11)
		O8–Sm1–N3	70.19(11)
		O9–Sm1–N2	70.68(11)
		O9–Sm1–N3	77.89(11)
		N2-Sm1-N3	147.12(11)

D−H…A D–H $H{\cdots}A$ $D{\cdots}A$ ∠DHA Sm-2₁ $N1\text{--}H1\cdots O1$ 0.98 1.95 2.689(4) 132 N1-H1...04 0.98 1.98 2.769(4) 136

Table S3 Intramolecular hydrogen bond parameters (Å, °) in Sm-2₁.

Table S4 Bond distances $(d_{sm,j})$, bond valences $(v_{sm,j})$ and total samarium atom valence (V_{sm}) in the crystal structure of **Sm-2**₁.

Compound	Atom	Donor type	$d_{ m Sm,j}/ m \AA$	$v_{\mathrm{Sm,j}}$	$V_{\rm Sm}$
N2 N3 O1 O2 Sm-2 ₁ O4 O5 O6 O8 O9	N2	HL2 ₁ -	2.688	3 0.252	
	N3 O1	$HL2_{l}$	2.610	0.311	
		$HL2_{l}$	2.319	0.495	
	O2	$HL2_{l}$	2.723	0.166	
	O3	$HL2_{l}$	2.675	0.189	2.026
	O4	$HL2_{l}$	2.355	0.449	2.936
	05	NO ₃ -	2.581	0.244	
	O6	NO ₃ -	2.529	0.281	
	08	NO ₃ -	2.579	0.245	
	09	NO ₃ -	2.499	0.304	

chemosensor	Sensing mechanism and effect	NACs	Linear range	LOD	Solvent	Reference
GCDs	FL quenching	TNP	0.1-0.15µM	0.091µM	Water	Acta A: Mol. Biomol.Spectrosc. 137 (2015) 1213
sensor 1 sensor 2	FL quenching	TNP	0-100μΜ	70 ppb 300ppb	CH ₃ CN	Anal. Chim. Acta 936 (2016) 216
PI-CONs	FL enhancement	TNP	0.5-10µM	0.25µM	EtOH	Interfaces. 2017, 9,13415
[Bi(L ¹)(NO ₃)] _n [Bi(L ²)(NO ₃) ₃]	FL quenching	2,4- DNP TNP	5-70μM 5-70μM 5-55μM 5-100μM	0.0968μM 0.0924μM	DMF	Sens. Actuators B Chem. 264 (2018) 363
BaAlF ₅ :Eu ²⁺ @PEI BaSiF ₆ :Eu ²⁺ @PEI	FL quenching	TNP	1-5 ng/ml 3-20 ng/ml	0.57ng/ml 2.82ng/ml	EtOH	J. Rare Earth. 39 (2021) 952
L_1 $L_1@C_b$	FL quenching	TNP	NR	29.3±3.7n M 29nM	THF	ACS Omega 5 (2020) 25747
Sensor 3 Sensor 4	FL quenching	TNP	NR	4.1nM 5.9nM	DMF	Spectrochim. Acta A. 226 (2020) 117583
Zn-MOFs 1 Zn-MOFs 2 Zn-MOFs 3 Zn-MOFs 4	FL quenching	2-NP	0-0.125mM 0-0.170 mM 0-0.120 mM 0-0.170 mM	0.79μM 1.34μM 2.98μM 0.73μM	Water	Cryst. Growth Des. 21 (2021) 5558
EY@Zr-MOF	FL quenching	2-NP	0.01-1mM	NR	EtOH	ACS Sustain. Chem. Eng. 7 (2019) 6196
[Nd ₂ CdL ₂ (NO ₃) ₂ (D MF) ₂](OH) ₂	FL quenching	2-NP	0-640µM	14.73µM	CH ₃ CN	Front. Chem. 7 (2019) 139

Table S5 Analytical parameters of fluorescent chemosensors studied herein for detection of nitroaromatic explosives.

NR = Not Reported

References

1 Z. Li, Y. Wang, Y. Ni, S. Kokot, A sensor based on blue luminescent graphene quantum dots for analysis of a common explosive substance and an industrial intermediate, 2,4,6-trinitrophenol, *Acta A: Mol. Biomol. Spectrosc.* 2015, **137**, 1213–1221.

- 2 C. Wu, J.-L. Zhao, X.-K. Jiang, X.-L. Ni, X. Zeng, C. Redshaw, T. Yamato, Clickmodified hexahomotrioxacalix[3]arenes as fluorometric and colorimetric dual-modal chemosensors for 2,4,6-trinitrophenol, *Anal. Chim. Acta* 2016, **936**, 216–221.
- 3 C. L. Zhang, S. M. Zhang, Y H. Yan, F. Xia, A. Huang, Y. Z. Xian, Highly fluorescent polyimide covalent organic nanosheets as sensing probes for the detection of 2,4,6-trinitrophenol, *ACS Appl. Mater. Inter.* 2017, **9**, 13415–13421.
- 4 O. Gungor, M. Kose, Selective detections of nitroaromatic explosives by monomeric and polymeric Bi(III) complexes, *Sensor. Actuat. B-Chem.* 2018, **264**, 363–371.
- 5 X. Zhang, L. T. Liu, W. Zhang, L. Y. Na, R. Hua, Detection of 2,4,6-trinitrophenol based on f-f transition of Eu²⁺, *J. Rare Earth.* 2021, **39**, 952–958.
- 6 A. Narula, M. A. Hussain, A. Upadhyay, C. P. Rao, 1,3-Di-naphthalimide conjugate of Calix[4]arene as a sensitive and selective sensor for trinitrophenol and this turns reversible when hybridized with carrageenan as beads, ACS Omega 2020, 5, 25747–25756.
- 7 F. Qiu, Y.-H. Huang, Q. M. Ge, M. Liu, H. Cong, Z. Tao, The high selective chemosensors for TNP based on the mono- and di-substituted multifarene[2,2] with different fluorescence quenching mechanism, *Spectrochim. Acta A.* 2020, 226, 117583.
- 8 W. B. Liu, N. Li, X. Zhang, Y. Zhao, Z. Zong, R. X. Wu, J. P. Tong, C. F. Bi, F. Shao, Y. h. Fan, Four Zn(II)-MOFs as highly sensitive chemical sensor for the rapid detection of tetracycline, o-nitro phenol, Cr₂O₇²⁻/PO₄³⁻, Fe³⁺/Al³⁺ in water environment, *Cryst. Growth Des.* 2021, **21**, 5558–5572.
- 9 Y. K. Li, Z. H. Wei, Y. Zhang, Z. F. Guo, D. S. Chen, P. Y. Jia, P. Chen, H. Z. Xing, Dual-emitting EY@Zr-MOF composite as self-calibrating luminescent sensor for selective detection of inorganic ions and nitroaromatics, ACS Sustain. Chem. Eng. 2019, 7, 6196–6203.
- 10 H. F. Chen, X. P. Yang, W. Z. Jiang, D. M. Jiang, D. L. Shi, B. C. Yuan, F. Wang, L. J. Zhang, S. M. Huang, Anion dependent self-assembly of polynuclear Cd-Ln Schiff base nanoclusters: NIR luminescent sensing of nitro explosives, *Front. Chem.* 2019, 7, 139.

Figures



Fig. S1 Analogues of nitroaromatic explosives discussed in this work.



Fig. S2 ¹H NMR spectrum of dialdehyde H₂Q₁ in CDCl₃.



Fig. S3 FT-IR spectrum of dialdehyde H₂Q₁ (a) and Sm(III)-macrocycle Sm-2₁ (b).



Fig. S4 ESI-MS (positive) of Sm(III)-macrocycle **Sm-2**₁ together with its inserted experimental (a) and simulative (b, calculation for $[C_{34}H_{48}BrCl_2N_5O_{14}Sm]$) peaks of isotopic distribution corresponding to the peak at m/z = 1054.3352.



Fig. S5 UV-vis spectra of H_2Q_1 (a, 40.0 μ M) and Sm-2₁ (b, 45.0 μ M) in DMF.



Fig. S6 (a) Emission spectra ($\lambda_{ex} = 273$, 293, 307 and 363 nm) of **Sm-2**_I (30.0 µM) in DMF-H₂O (19:1, ν/ν). Inset: Excitation spectra of **Sm-2**_I ($\lambda_{em} = 643$ nm, black line). (b) Emission spectra ($\lambda_{ex} = 363$ nm) of **Sm-2**_I (25.0 µM) in 10 common solvents. (c) Emission spectra ($\lambda_{ex} = 363$ nm) of **Sm-2**_I at different concentrations in DMF-H₂O (19:1, ν/ν) and citric acid-sodium citrate buffer (pH = 6.0, 5.00 mM). (d) Time-dependent emission spectra of **Sm-2**_I (30.0 µM) in DMF-H₂O (19:1, ν/ν) at 298 K.



Fig. S7 Absorption (a) and fluorescence (b) spectra of **Sm-2**₁ (25.0 μ M) mixed with explosives (125.0 μ M) in DMF-H₂O (19:1, ν/ν) and citric acid-sodium citrate buffer (pH = 6.0, 5.00 mM).



Fig. S8 Emission spectra (λ_{ex} = 363 nm) of different mixtures contained **Sm-2**₁ (25.0 µM) in DMF-H₂O (19:1, *v*/*v*). (mixA: mixture of 17 explosive analogues without TNP, mixB: mixture of 17 explosive analogues without 2-NP)



Fig. S9 Fluorescence changes of **Sm-2**_I (25.0 μ M) upon increasing TNP (a, 0–125.0 μ M), 2-NP (b, 0–750.0 μ M) (λ_{em} = 643 nm) versus the concentration of NEs in DMF-H₂O (19:1, *v*/*v*) and citric acid-sodium citrate buffer (pH = 6.0, 5.00 mM) at 298 K.



Fig. S10 Absorption changes of **Sm-2**₁ (25.0 μ M) upon increasing contents of TNP (a, 0–125.0 μ M), 2-NP (c, 0–750.0 μ M) together with the inserted linearity of absorbance (360nm) versus the concentration of NEs in DMF-H₂O (19:1, v/v) and citric acid-sodium citrate buffer (pH = 6.0, 5.00 mM) at 298 K.



Fig. S11 ESI-MS (positive) of Sm(III)-macrocycle **Sm-2**₁ (25.0 μ M) mixed with TNP (125.0 μ M) together with its inserted experimental (a) and simulative (b, calculation for [C₃₁H₃₅BrCl₂N₅NaO₁₁Sm]) peaks of isotopic distribution corresponding to the peak at m/z = 979.9790.



Fig. S12 ESI-MS (positive) of Sm(III)-macrocycle **Sm-2**_I (25.0 μ M) mixed with 2-NP (125.0 μ M) together with its inserted experimental (a) and simulative (b, calculation for [C₃₁H₃₅BrCl₂N₅NaO₁₁Sm]) peaks of isotopic distribution corresponding to the peak at m/z = 980.2629.



Fig. S13 Absorption spectra of explosive analogues (125.0 μ M) in DMF-H₂O (19:1, v/v).



Fig. S14 Emission spectra of **Sm-2**₁ (25.0 μ M) under the presence of serial two-component mixtures: 3-NP/4-NP (a), 2,4-DNT/TNT (b), TNT/TNP (c), and RDX/DMNB (d) in DMF-H₂O (19:1, *v*/*v*) and citric acid-sodium citrate buffer (pH = 6.0, 5.00 mM) at 298 K. For mixtures, we set up a series of five contents (0%, 25%, 50%, 75% and 100%) for one component.



Fig. S15 Fluorescence spectra of **Sm-2**₁ mixed with different contents of explosives (a, 125.0 μ M; b, 2.5 μ M) in real water samples.