

Supporting Information

A Phosphine-based fluorescent probe for fluorescent imaging of
hypochlorous acid in living cells and zebrafish

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Experimental section

Reagents, materials, and apparatus:

All the solvents used in the experiment were of analytic grade. The reaction progress was monitored by thin-layer chromatography (TLC) on silica gel plates (GF₂₅₄) visualized by UV light. 200-300 mesh silica gel was used for column chromatography. NMR experiments were carried out on a Bruker AV-400 NMR spectrometer with chemical shifts reported in ppm (in CDCl₃, DMSO-*d*₆, or TMS as an internal standard). Mass spectra were measured on an Agilent 1290 LC-MS spectrometer. All pH measurements were made with a Sartorius basic pH-Meter PB-10. Fluorescence spectra were determined on a PerkinElmer LS55 Fluorescence spectrophotometer. Absorption spectra were collected on a Shimadzu UV 2501(PC)S UV-Visible spectrophotometer. The excitation and emission widths for **BBP** were all 1.5.

Preparation of various ROS and RNS species:^[1]

HOCl: Take an appropriate amount of commercially available hypochlorous acid solution and prepare about 10⁻² M hypochlorous acid stock solution with deionized water. Dilute the hypochlorous acid solution, and calibrate its concentration ($\epsilon = 350 \text{ M}^{-1}\text{cm}^{-1}$) through the ultraviolet spectrum absorption value at 292 nm.

ONOO⁻: To a vigorously stirred solution of NaNO₂ (0.6 M, 10 mL) and H₂O₂ (0.7 M, 10 mL) in deionized H₂O at 0 °C was added HCl (0.6 M, 10 mL), immediately followed by the rapid addition of NaOH (1.5 M, 20 mL). Excess hydrogen peroxide was removed by passing the solution through a short column of MnO₂. The concentration of ONOO⁻ was determined by UV analysis with the extinction coefficient at 302 nm ($\epsilon = 1670 \text{ M}^{-1} \text{ cm}^{-1}$). The solution were stored at -20 °C for use.

NO: A solution of the H_2SO_4 (3.6 M) was added dropwise into a stirred solution of NaNO_2 (7.3 M). The emitted gas was allowed to pass through a solution of NaOH (2 M) first and then deionized H_2O to make a saturated NO solution of 2.0 mM.

$^1\text{O}_2$: NaMoO_4 (10 mM) and H_2O_2 (10 mM) was prepared in PBS (10 mM, pH 7.4). Equal aliquots of these solutions were then mixed to yield $^1\text{O}_2$ of 5 mM.

H_2O_2 : Take an appropriate amount of commercially available H_2O_2 solution and prepare about 10^{-2} M H_2O_2 stock solution with deionized water. And its concentration is calibrated by the ultraviolet absorption value at 240 nm ($\epsilon = 43.6\text{M}^{-1}\text{cm}^{-1}$).

$\cdot\text{OH}$: $\cdot\text{OH}$ was generated by Fenton reaction. To a solution of H_2O_2 (1.0 mM, 1.0 mL) in PBS (10 mM, pH 7.4) was added FeSO_4 solution (1.0 mM, 100 μL) at ambient temperature (stock solution 0.1 mM).

$\text{ROO}\cdot$: $\text{ROO}\cdot$ was generated from 2, 2'-azobis(2-amidinopropane)dihydrochloride, which was dissolved in PBS (10 mM, pH 7.4) 1 h before use to make a stock solution of 10 mM.

Additional spectroscopic data

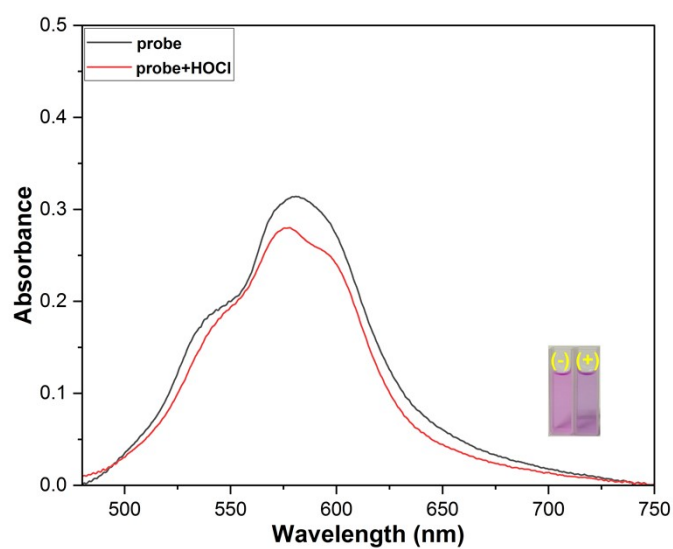


Fig. S1 The UV-vis spectrum of **BBP** (10.0 μM) with and without of HOCl (2.0 equiv.) in PBS buffer solution (10 mM, pH 7.2, containing 10% EtOH). Bottom inset: Cuvette images of the probe **BBP** before and after adding HOCl.

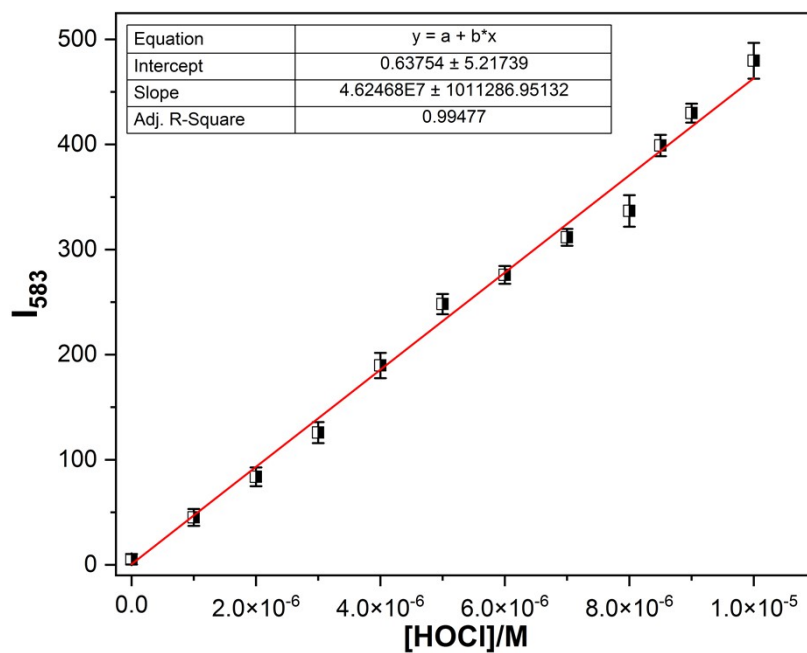


Fig. S2 The fluorescent intensity of **BBP** (10.0 μM) at 583 nm (I_{583}) as a function of HOCl concentration (0-10.0 μM) in PBS buffer solution (10 mM, pH 7.2, containing 10% EtOH, $\lambda_{\text{ex}} = 552$ nm).

The detection limit (DL) of HOCl using **BBP** was determined from the following equation: ^[2]

$$DL = 3 \cdot \sigma / K$$

Where σ is the standard deviation of the blank solution; K is the slope of the calibration curve.

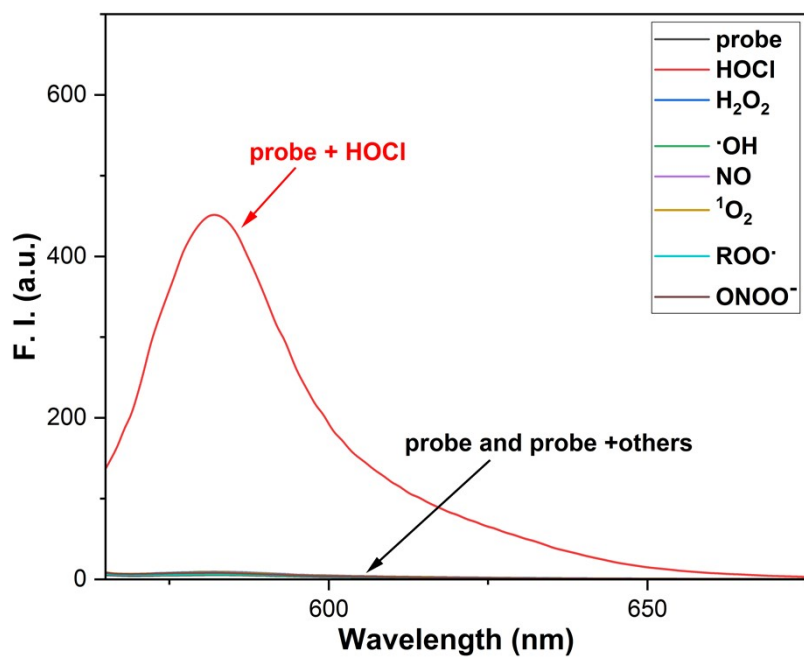


Fig. S3 The fluorescent spectra of **BBP** (10 μM) before and after the addition of various analytes (100 μM each, including $^1\text{O}_2$, H_2O_2 , NO , $\cdot\text{OH}$, $\text{ROO}\cdot$, and ONOO^-), and HOCl (10 μM), (in PBS buffer solution, 10 mM, pH 7.2, containing 10% EtOH, $\lambda_{\text{ex}} = 552$ nm).

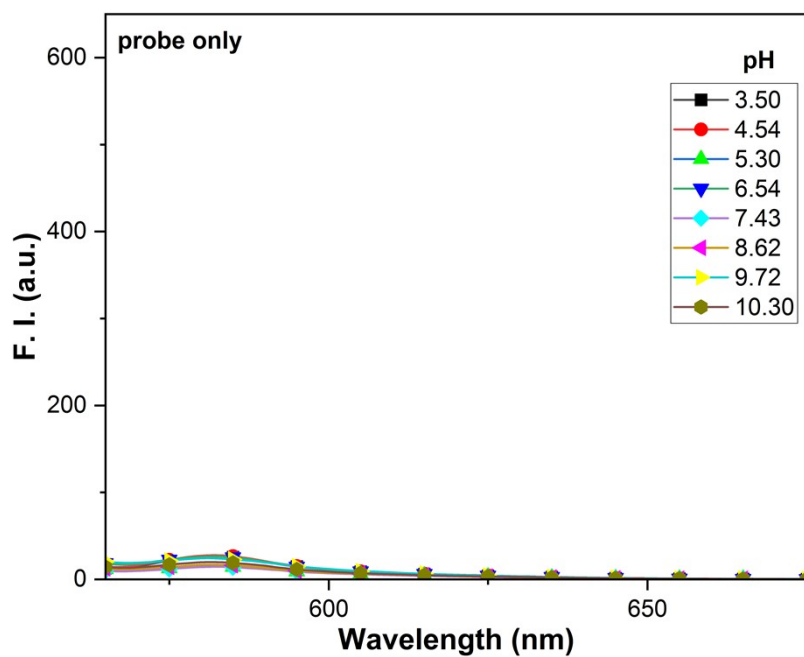


Fig. S4. The fluorescence spectra of the probe **BBP** (10 μM) alone at different pH values ($\lambda_{\text{ex}} = 552$ nm).

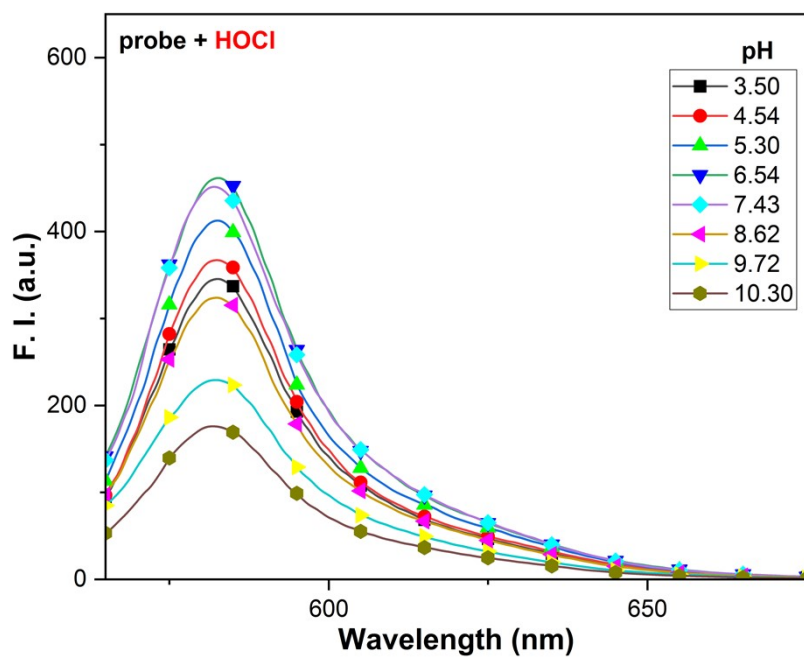


Fig. S5. The fluorescence spectra of the probe **BBP** (10 μM) in the present of HOCl (10 μM) at different pH values ($\lambda_{\text{ex}} = 552 \text{ nm}$).

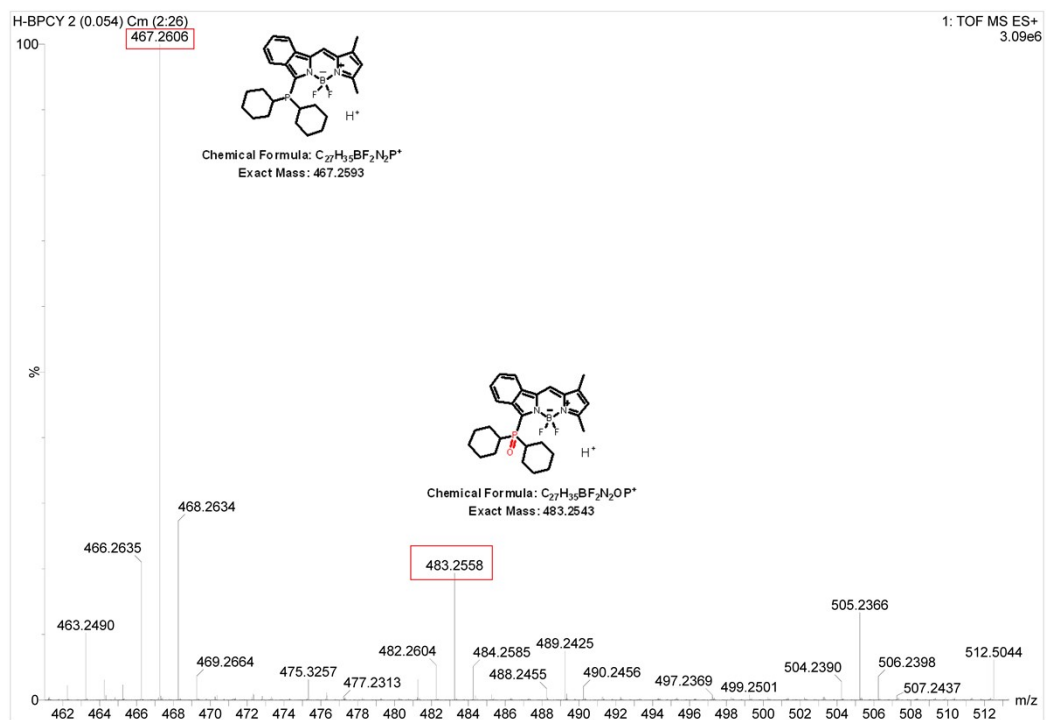


Fig. S6. The HR ESI-MS spectrum of **BBP** and HOCl mixture.

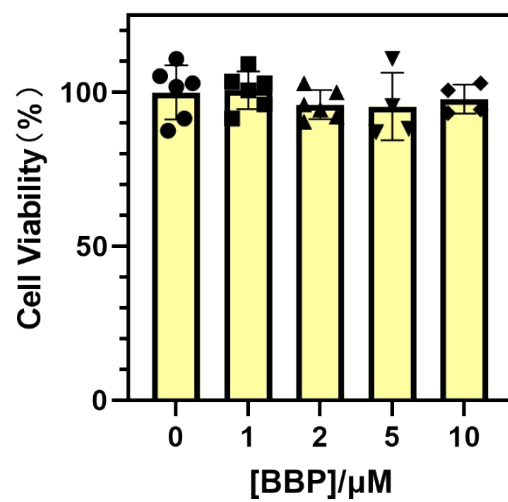
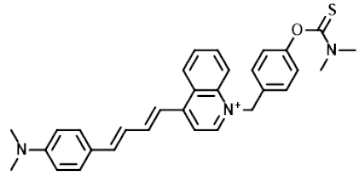
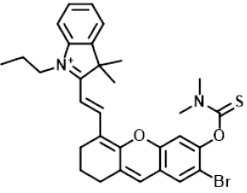
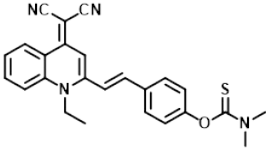
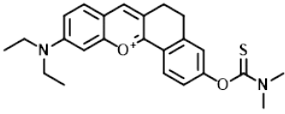
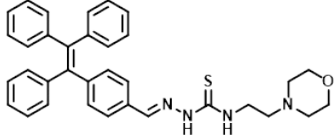
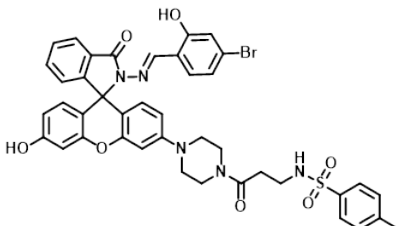
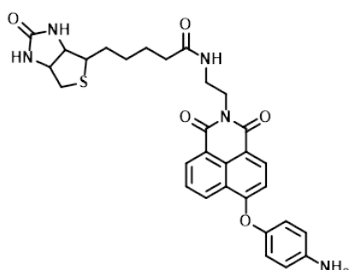
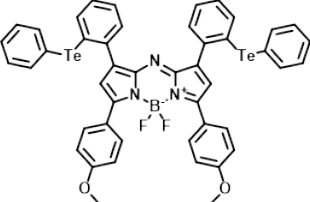
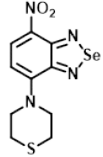
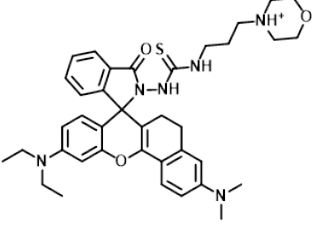
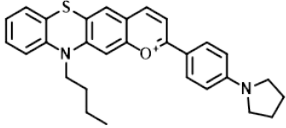
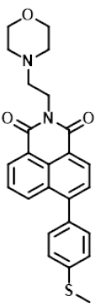
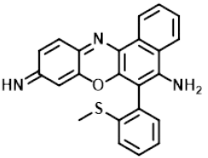
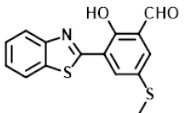
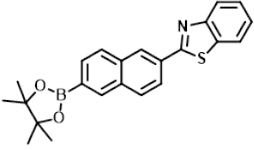
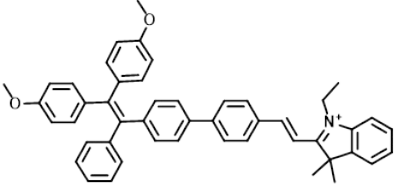
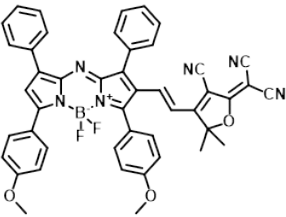
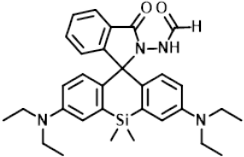
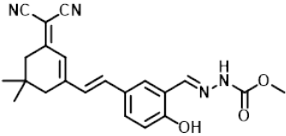
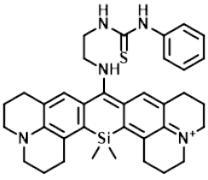
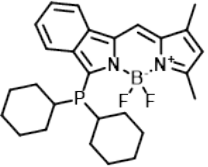


Fig. S7. Cell viability of the probe in a standard MTT assay in living RAW 264.7 cells for 24 h. The experiment was repeated three times.

Table S1. Comparison of fluorescent probes for HOCl detection.

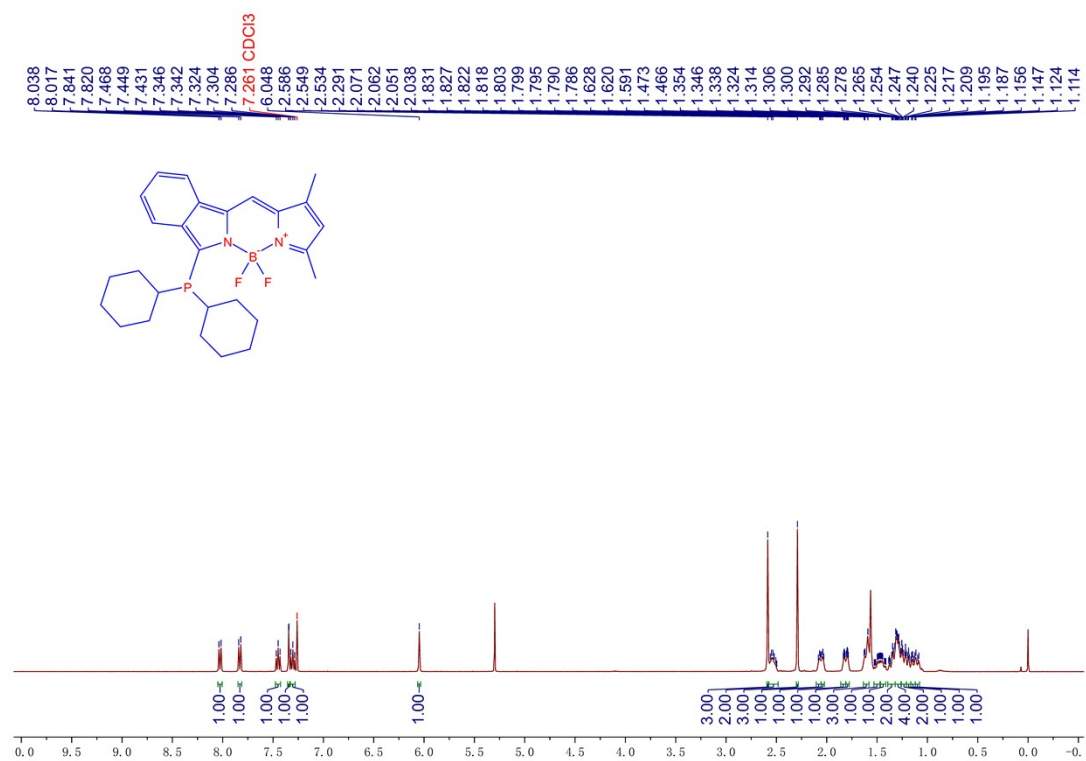
No.	Structures	Abs/Em	Stocks Shift	LOD	Cell Imaging	Mouse/Zebra Fish Imaging	Refs
1		387/590	203	2.4 nM	√	×	[3]
2		694/712	18	89.7 nM	√	√	[4]
3		430/620	190	30.8 nM	√	×	[5]
4		567/623	56	5.8 μM	√	√	[6]
5		350/492	142	250 nM	√	×	[7]
6		513/556	43	785 nM	√	×	[8]
7		468/553	85	17 nM	√	×	[9]
8		692/738	46	90 nM	√	×	[10]

9		520/620	100	-	×	×	[11]
10		622/655	33	30 nM	√	√	[12]
11		438/503	65	25.8 nM	√	×	[13]
12		374/595	221	118 nM	√	√	[14]
13		596/638	42	94.7 nM	√	×	[15]
14		435/456	21	0.14 μM	√	×	[16]
15		328/394	66	-	√	×	[17]
16		364/530	166	32 μM	√	×	[18]
17		700/-	-	0.36 μM	√	×	[19]

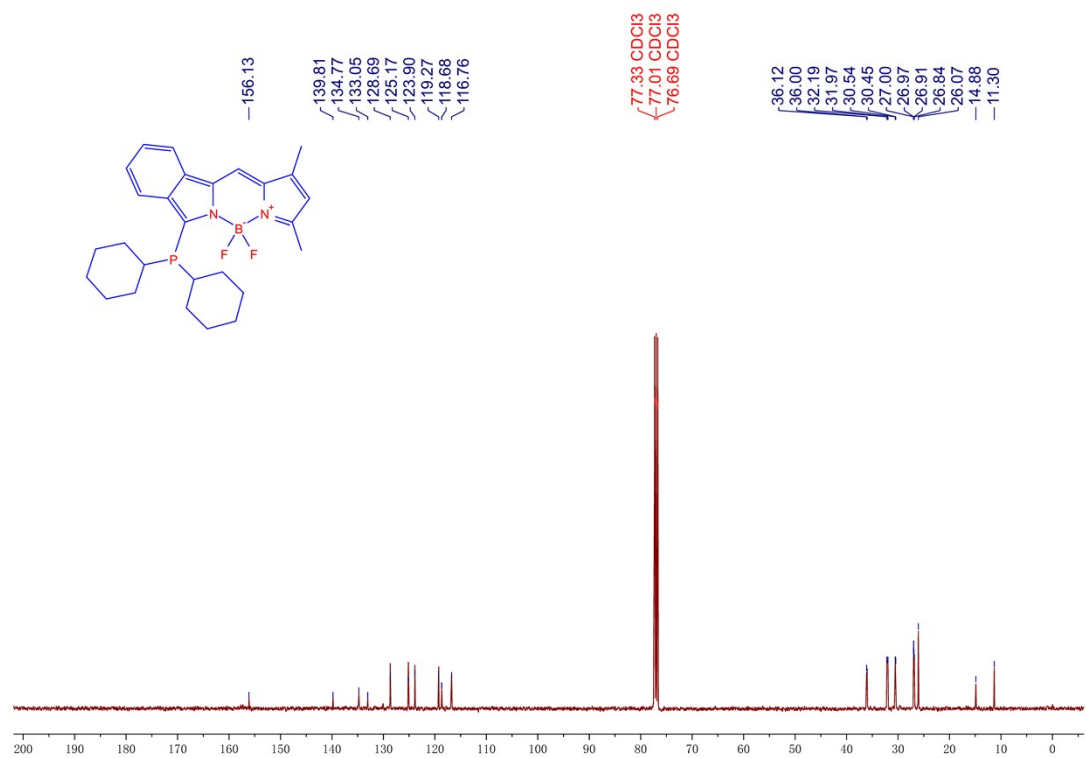
27		653/669	16	0.23 μ M	√	×	[29]
28		425/653	228	15.3 nM	√	×	[30]
29		504/642	142	16.1 nM	√	√	[31]
30		575/583	8	15.3 nM	√	√	This work

The characterization data of BBP

^1H NMR of **3** (BBP)

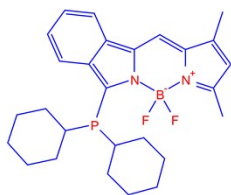


^{13}C NMR of **3** (BBP)

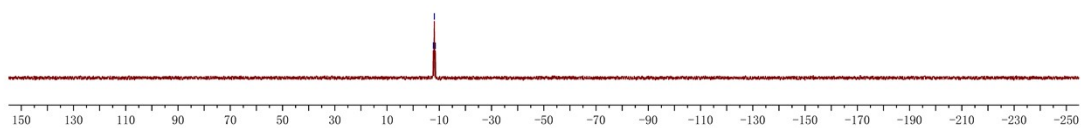


^{31}P NMR of **3** (BBP)

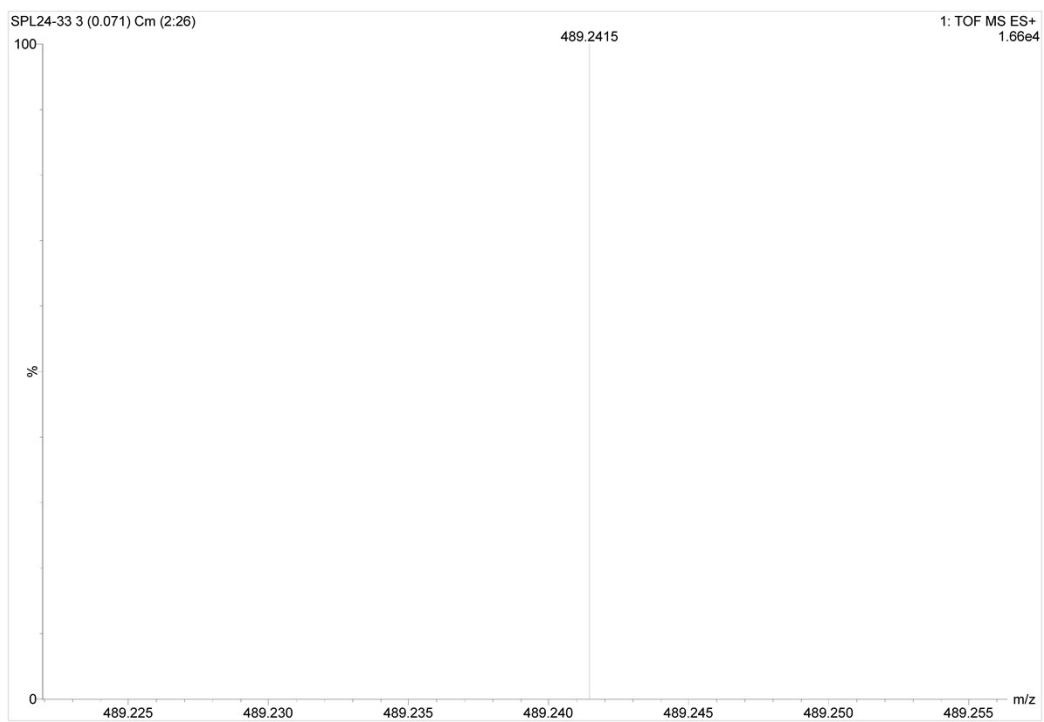
^{31}P NMR (162 MHz, Chloroform-*d*) δ -8.10 (t, $J = 66.6$ Hz).



δ -7.687
 δ -8.102
 δ -8.510



HR-MS of **3** (BBP)



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