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Supporting Information

Cholesterol appended cyanostyryl thiophene positional isomers with multistimuli responsive emission switching and liquid crystalline properties

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Section 3: References

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Synthesis and characterization

Materials. Solvents and chemicals for the synthesis and photophysical studies were purchased from Sigma Aldrich, Alfa Aeser and TCI and used without further purification.

Characterization. The ¹H NMR and ¹³C NMR spectra were obtained from a Bruker Avance 400 MHz spectrometer operating at room temperature. Photoluminescence spectra and lifetime experiments were performed using Fluorolog-3 with TCSPC spectroflurometer (model FL 3C-221). The mass spectra of the compounds were recorded on a Waters Xevo G2 XS QToF mass spectrometer. The single crystal X-ray diffraction data of CS-1 was collected on Bruker D8 Venture diffractometer attached with PHOTON II detector with CMOS-sensor. The data collection was conducted at room temperature using Mo Ka radiation operated at 50 kV and 40 mA. The powder X-ray diffraction data was collected using PANalytical X'Pert3 Powder X-Ray Diffractometer. X-ray diffraction (XRD) measurements were carried out on powder samples in Lindemann capillaries with CuK α ($\lambda = 0.15418$ nm) radiation using either an Image plate (IP) detector (GeniX3D, Xenocs) from a source operating at 50 kV and 0.6 mA in conjunction with a multilayer mirror was used to illuminate the sample or PANalytical X'Pert PRO MP machine consisting of a focusing elliptical mirror and a fast high-resolution detector (PIXCEL).



Scheme S1: Synthetic route to CS-1 and CS-2

Synthesis of 2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-10,13-dimethyl-17-(6-methylheptan-2-yl)-1Hcyclopenta[a]phenanthren-3-yl 4- (cyanomethyl)phe nylcarbamate, 1

The compound **1** was synthesized following the reported procedure.¹ 2-(4-aminophenyl) acetonitrile (5 mmol) is dissolved in minimum quantity of dichloromethane and cooled to 0 $^{\circ}$ C in an ice bath. The cholesterol chloroformate (4.8 mmol) was added to the reaction mixture followed 0.5 ml pyridine and stirring was continued for 6 hours. The reaction mixture was quenched with water and extracted with dichloromethane. Combined organic layers were washed with brine, and dried over Na₂SO₄. Solvent was evaporated and the residue was purified by column chromatography on silica gel using hexane ethyl acetate (4/1) as eluent to afford compound **1** as a colourless crystalline solid on re-crystallization form ethyl acetate. The product was characterized using ¹H NMR and ¹³C NMR spectroscopy.

Yield: 87 %, ¹H NMR (400 MHz, CDCl3) δ (ppm): 0.67 (s, 3H), 0.87 (m, 6H), 0.91 (t, *J* = 6 Hz, 3H), 0.98 (m, 6H), 1.13 (m, 7H), 1.29 (m, 4H), 1.47 (m, 5H), 1.62 (m, 1H), 1.91 (m, 5H), 2.38 (m, 2H), 3.69 (s, 2H), 4.59 (m, 1H), 5.40 (m, 1H), 6.57 (s, 1H), 7.24 (d, *J* = 8 Hz, 3H), 7.38 (d, *J* = 12 Hz, 2H).

¹³C NMR (400 MHz, CDCl₃) δ (ppm): 11.87, 18.71, 19.34, 21.04, 22.57, 22.83, 23.02, 23.83, 24.29, 28.02, 28.07, 28.24, 31.86. 35.80, 36.18, 36.57, 39.51, 39.72, 42.31, 49.99, 56.12, 56.68, 75.18, 117.94, 119.00, 122.87, 124.38, 128.64, 137.99, 139.49, 152.90.

General method for the synthesis of compounds CS-1 and CS-2

The compounds **CS-1** and **CS-2** were synthesized by adopting reported procedures with suitable modifications.² Compound **1** (545 mg, 1 mmol) and thiophene-2-carbaldehyde or thiophene-3-carbaldehyde (1 mmol) were dissolved in a mixture of ^tBuOH (11 ml) and THF (5

ml) at 50° C. ¹BuOK (0.11 ml of a 1 M solution in THF, 0.11 mmol) and n-Bu₄NOH (1 ml of a 1 M solution in MeOH, 1 mmol) were added. An orange precipitate started to form immediately and was stirred for 15 minutes at 70° C. The reaction mixture was cooled to room temperature and poured into acidified methanol (50 ml containing 1 drop of conc.CH₃COOH). The resulting precipitate was filtered and washed with methanol. The compounds were further re-precipitated 5 times from dichloromethane solutions by adding excess methanol

Compound CS-1: Yield: 87 %, M.P. 188 °C, ¹H NMR (400 MHz, CDCl3) δ (ppm): 0.69 (s, 3H), 0.86 (d, 3H, J=1.84 Hz), 0.88 (d, 3H, J=1.84 Hz), 0.92 (d, 3H, J=6.52 Hz), 0.99 (m, 3H), 1.04 (s, 4H), 1.13 (m, 7H), 1.24 (m, 2H), 1.35 (m, 3 H), 1.49 (m, 5H), 1.66 (m, 1H), 1.86 (m, 2H), 1.99 (m, 3H), 2.42 (m, 2H), 4.62 (m, 1H), 5.42 (m, 1H), 6.66 (s, 1H), 7.41 (m, 1H), 7.47 (d, 2H, J=1.96Hz), 7.59 (m, 2H), 7.76 (dd, 1H, J=5.16Hz and 1.32 Hz), 7.92 (m, 1H).

¹³C NMR (400 MHz, CDCl3) δ (ppm) 11.87, 18.72, 19.34, 21.05, 22.57, 22.83, 23.84, 24.29, 28.02, 28.23, 31.87, 31.91, 35.80, 36.18, 36.58, 36.95, 38.42, 39.52, 39.72, 42.32, 50.00, 56.13, 56.68, 75.31, 109.36, 118.45, 118.67, 122.92, 126.54. 126.63, 127.33, 128.95, 129.02, 134.05, 136.11, 138.87, 139.47, 152.69. HRMS (ESI) m/z calculated 638.3906; Found: 639.4040 [M+H]⁺.

Compound CS-2: Yield: 83 %, M.P. 179 °C, ¹H NMR (400 MHz, CDCl3) δ (ppm) : 0.69 (s, 3H), 0.88 (m, 6H), 0.93 (t, 3H, J=6Hz), 1.01 (m, 6H), 1.12 (m, 7H), 1.30 (m, 5H), 1.48 (m, 5H), 1.63 (m, 2H), 1.92 (m, 5H), 2.41 (m, 2H), 4.62 (m, 1H), 5.41 (d, 1H, J= 4Hz), 6.68 (d, 1H, J= 4Hz), 7.15 (m, 1H) 7.45 (m, 1H), 7.47 (m, 1H), 7.53 (t, 1H, J= 4Hz), 7.58 (t, 2H, J= 8Hz), 7.65 (t, 1H, J= 4Hz).

¹³C NMR (400 MHz, CDCl3) δ (ppm) 11.87, 18.72, 19.34, 21.05, 22.57, 22.83, 23.84, 24.29, 28.02, 28.24, 31.87, 31.91, 35.80, 36.18, 36.58, 36.95, 38.42, 39.52, 39.72, 42.32, 50.00, 56.13, 56.68, 75.31, 107.79, 118.17, 118.70, 122.93, 126.49, 127.83, 128.65, 129.68, 132.00, 132.87, 138.08, 138.91, 139.47, 152.78. HRMS (ESI) m/z calculated 638.3906; Found: 639.3990 [M+H]⁺.



Fig. S1 ¹H NMR spectra of 1in CDCl₃



Fig. S2 ¹³C NMR spectra of 1in CDCl₃



Fig. S3 ¹H NMR spectra of CS-1 in CDCl₃



Fig S4¹³C NMR spectra of CS-1 in CDCl₃



Fig. S5 ¹H NMR spectra of CS-2 in CDCl₃



Fig. S6 ¹³C NMR spectra of CS-2 in CDCl₃



Fig. S7 UV-visible absorption spectra of CS-1 (blue curve) and CS-2 (red curve) in THF (1 µM)



Fig. S8 Emission spectra of CS-2 recorded in THF/water mixtures (50 μ M) by varying the water fraction. ($\lambda_{ex} = 370$ nm)



Fig. S9 Fluorescent intensity in response to the changes of water fractions in THF-H₂O mixtures (a) CS-1 and (b)CS-2 Concentration:

50x10⁻⁶ M; λex: 370nm.



Fig. S10 ORTEP plot of the asymmetric unit present in the single crystals structure of **CS-1** (ellipsoids are drawn at 50 % probability, and hydrogen atoms are removed for clarity)

Identification code	CS-1		
Empirical formula	$C_{41} H_{54} N_2 O_2 S$		
Formula weight	638.92		
Temperature	296(2) K		
Wavelength	0.71073 Å		
Crystal system	Monoclinic		
Space group	P 21		
Unit cell dimensions	a = 18.357(11) Å	$\alpha = 90^{\circ}$.	
	b = 12.475(7) Å	β = 115.78(2) °.	
	c = 18.564(13) Å	$\gamma = 90^{\circ}$.	
Volume	3828(4) Å ³		
Z	4		
Density (calculated)	1.109 mg/m ³		
Absorption coefficient	0.119 mm ⁻¹		
F(000)	1384		
Crystal size	0.396 x 0.189 x 0.114 mm ³		
Theta range for data collection	2.464 to 25.496°.		
Index ranges	-22<=h<=22, -15<=k<=15, -22<=l<=22		
Reflections collected	108710		
Independent reflections	14206 [R(int) = 0.0836]		
Completeness to theta = 25.242°	99.7 %		
Absorption correction	Semi-empirical from equivalents		
Max. and min. transmission	0.7455 and 0.6707		
Refinement method	Full-matrix least-squares on F ²		
Data / restraints / parameters	14206 / 511 / 934		
Goodness-of-fit on F ²	1.107		
Final R indices [I>2sigma(I)]	R1 = 0.0703, $wR2 = 0.1309$		
R indices (all data)	R1 = 0.1327, wR2 = 0.1616		
Absolute structure parameter	0.03(4)		
Extinction coefficient	n/a		
Largest diff. peak and hole	0.354 and -0.201 e Å ⁻³		
CCDC No	2293176		

Table S1. Crystallographic and structure refinement details of CS-1.



Fig. S11 DSC traces of first heating and cooling and second heating and cooling cycles of (a) **CS-1** and (b) **CS-2** recorded at a scan rate of 5 °C/minutes.



Fig. S12 Polarizing optical photomicrograph of **CS-1** showing (a) characteristic oily streak texture of the N* phase at 188 °C and (b) glassy N* phase obtained at room temperature.



Fig. S13 Small angle X-ray diffraction pattern of (a) CS-1 and (b) CS-2 at different temperatures.

References

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