

Magnetically Separable Ag@Fe₃O₄-GO Nanocomposites for SERS Detection, Removal of Organic Pollutants, Oil from Water, and Antibacterial Applications

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Experimental Section

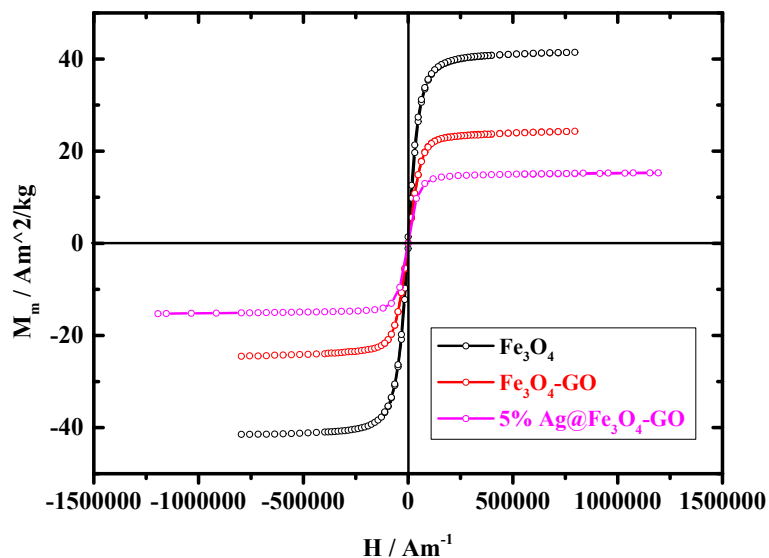
Chemicals: Ferric Chloride hexahydrate (FeCl₃·6H₂O), Sodium Acetate (CH₃COONa), ethylene glycol (C₂H₆O₂), graphite flakes, Hydrogen peroxide (H₂O₂), Silver nitrate (AgNO₃), Potassium carbonate (K₂CO₃), and Ascorbic acid were brought from Merck Specialities Pvt. Ltd., Polyvinyl Pyrrolidone (PVP molecular weight ~ 40000), Phosphoric acid (H₃PO₄), Potassium permanganate (KMnO₄) were purchased from Loba Chemie Pvt. Lmt., Urea (NH₂CONH₂) was brought from Thermo Fisher Scientific India Pvt. Lmt., Sodium nitrite (NaNO₂) was purchased from Samir Tech Chem Pvt. Lmt., concentrated H₂SO₄ was brought from Himedia, Tannic acid was purchased from SRL Chem. All the solutions were prepared in distilled water. All the chemicals were used as provided without any further purification.

Instrumentations/ Characterization techniques: The X-ray diffraction measurements were carried out using XRD Bruker, Model-D8 Advance (Eco) to study the crystal structures of the as-synthesized nanocomposite. The X-ray photoelectron spectroscopy (XPS) measurements were carried out on PHI 5000 Versa Probe III spectrometer (ULVAC-PHI, Inc.) to investigate the elemental composition and oxidation state of metal nanoparticles present in the as synthesized nanocomposite. The surface morphology of the as-prepared Ag@Fe₃O₄-GO composite was observed by a scanning electron microscope (CARL ZEISS Model EVO-18) equipment. Elemental analysis of the Ag@Fe₃O₄-GO composite was done using Energy- Dispersive X-ray (EDX) detector (DS: 51N1000 – EDS System Company: Oxford Instruments Nanoanalysis) which was attached to the SEM machine. The morphology and size of the as-prepared nanocomposite were examined by using Transmission Electron Microscope (Model: Tecnai G2 20 TWIN Company: FEI Company of USA (S.E.A.) PTE, LTD). Magnetic properties of the as-prepared samples were determined using a

Vibrating Sample Magnetometer (MicroSense Easy VSM). Brunauer-Emmett-Teller (BET) experiment was performed on the QuantaChrome iQ2 instrument to evaluate the surface area and pore size. Raman spectra were recorded on a Renishaw inVia Raman microscope to observe structural alterations in the sample during the oxidation process. Fourier transform infrared spectroscopy (FT-IR) studies were conducted using the FTIR-4700 JASCO instrument at a resolution of 2cm⁻¹. Atomic force microscopy (AFM) measurement was conducted using a multi-mode scanning probe microscope by NT-MDT Spectrum Instruments. The dynamic light scattering data was recorded to measure hydrodynamic size of Ag NPs on Malvern Panalytical Zetasizer Ultra (ZSU5700). The UV-vis absorption studies were conducted by a Fluorolog FL-3-11 Horiba Jobin-Yvon spectrophotometer.

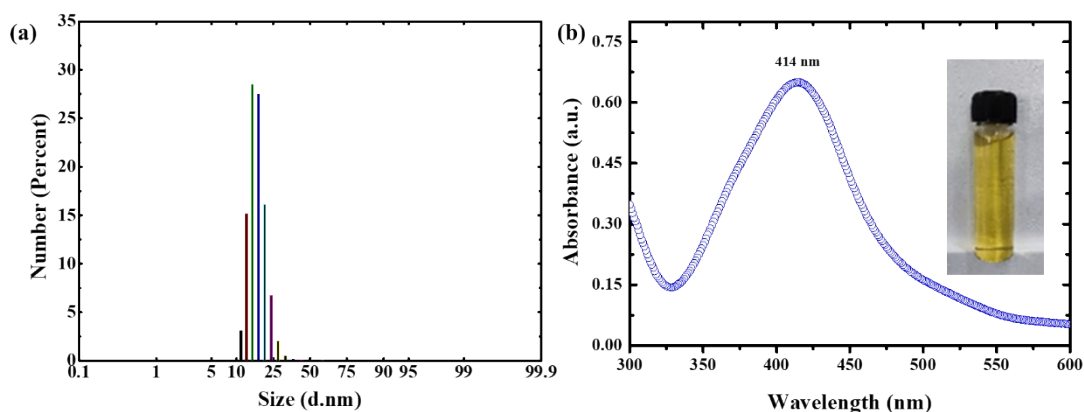
Determination of Antimicrobial activity of Ag@Fe₃O₄-GO nanocomposite: Ag@Fe₃O₄-GO was tested for its ability to inhibit Gram-negative (*Escherichia coli* and *Salmonella enterica*) and Gram-positive (*Listeria ivanovii*) bacteria by modified Kirby-Bauer disk diffusion method (Bauer et al., 1966). Briefly, the pure bacterial cultures were propagated in Müller-Hinton broth at 35°C ± 2°C. A bacterial lawn of each test bacterial strain was prepared by overlaying melted cooled soft agar (8 mL) added with 50 µL fresh test bacterial strain having 10⁶ colony-forming units (CFU)/mL on Müller-Hinton agar plates. As the agar solidified, 8 mm wells were punched into the Müller-Hinton agar plates by a sterile steel borer and sealed by adding 5-10 µL of molten agar (0.75% agar) to prevent leakage from the bottom of the wells. Ag@Fe₃O₄-GO nanocomposite was dissolved @ 50 mg/mL in sterile distilled water (stock suspension) and was serially diluted up to 528-fold to determine the minimum bactericidal concentration (MBC) of the nanocomposite. With help of an autopipette, 100 µL of stock suspension and each dilution of the sample of nanoparticle suspension was poured onto each of wells in agar plates for all bacterial strains mentioned above. Gentamicin was taken as a positive control. Plates were incubated overnight at 35°C ± 2°C, the different levels of zone of inhibition were measured.

Supporting Information Figure 1. VSM of synthesized Fe_3O_4 , $\text{Fe}_3\text{O}_4\text{-GO}$, 2% $\text{Ag}@ \text{Fe}_3\text{O}_4\text{-GO}$, and 5% $\text{Ag}@ \text{Fe}_3\text{O}_4\text{-GO}$ samples



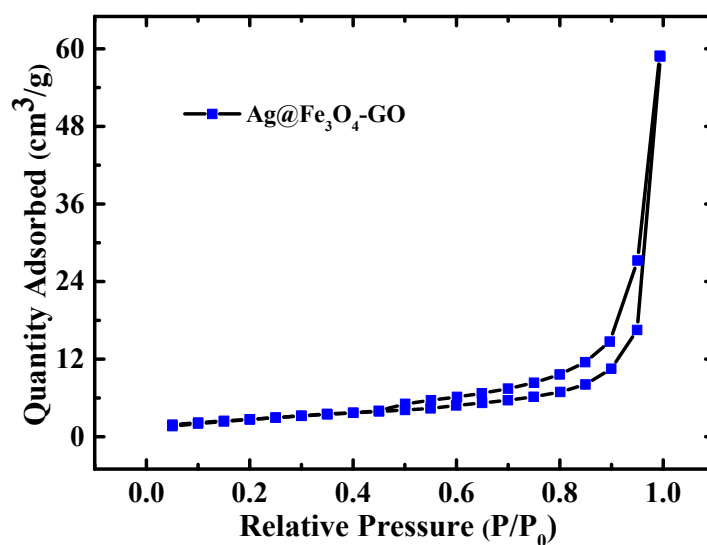
SI Figure 1: VSM of synthesized porous Fe_3O_4 , $\text{Fe}_3\text{O}_4\text{-GO}$, 2% $\text{Ag}@ \text{Fe}_3\text{O}_4\text{-GO}$, and 5% $\text{Ag}@ \text{Fe}_3\text{O}_4\text{-GO}$ samples

Supporting Information Figure 2. (a) and (b) represent the DLS and UV-visible spectra of synthesized Ag nanoparticles respectively. Figure 2 (b) inset visual picture of Ag nanoparticles.



SI Figure 2. (a) and (b) represent the DLS and UV-Visible spectra of synthesized Ag nanoparticles respectively. Figure 2 (b) inset visual picture of Ag nanoparticles.

Supporting Information Figure 3 represents the N₂ adsorption-desorption isotherm of Ag@Fe₃O₄-GO NCs.



SI Figure 3. N₂ adsorption-desorption BET plot of Ag@Fe₃O₄-GO NCs.

Table 1: Concentration of Ag@Fe₃O₄-GO NCs vs clearance diameter (cm) data

Dilution	Concentration (mg/ml)	Diameter of Zone of clearance* (cm)		
		<i>Escherichia coli</i> (Gram-negative)	<i>Salmonella enterica</i> (Gram-negative)	<i>Listeria ivanovii</i> (Gram-positive)
1X	50	2.7±0.03	2.4±0.03	1.7±0.03
2X	25	2.5±0.03	2.2±0.02	1.7±0.03
4X	12.5	2.6±0.01	2.1±0.03	1.5±0.03
8X	6.25	2.7±0.03	2.0±0.03	1.5±0.06
16X	1.5625	2.2±0.03	1.9±0.03	1.4±0.09
32X	0.78125	2.0±0.03	1.7±0.03	1.3±0.07
64X	0.390625	1.4±0.00	1.5±0.03	1.3±0.03
132X	0.1953125	0.9±0.00	1.2±0.03	1.4±0.03
264X	0.09765625	0.9±0.00	1.1±0.03	1.4±0.07
528X	0.048828125	0.9±0.00	0.8±0.00	1.3±0.06
1056X	0.0244140625	0.8±0.00	0.8±0.00	0.8±0.00
	Gentamicin (32 mg/L)	3.2±0.03	3.6±0.03	1.9±0.03

*Zone of clearance more than 2mm is considered positive inhibitory activity. Zone of clearance calculated here includes the agar well diameter of 8mm

Table 2: Performance of the Ag@ Fe₃O₄-GO nanocomposites with existing materials.

S No	Materials	Application	Ref
01	Ag@ Fe ₃ O ₄ -GO Nanocomposites	Gaseous elemental mercury removal from coal-fired flue gas	2
02	Fe ₃ O ₄ /GO/Au-Ag nanocomposite	Organic molecule synthesis (spirooxindole-dihydropyridines)	3
03	Ag/ Fe ₃ O ₄ /RGO nanocomposites	(a) Reduction of organic pollutants (b) SERS detection	4
04	Fe ₃ O ₄ @ GO@ Ag@ PDA Composite	SERS detection of PAH	5
05	Ag-Fe ₃ O ₄ -graphene oxide	Electrochemical sensor	6
05	Ag@Fe ₃ O ₄ nanoparticles decorated GO	(a) Photocatalytic (b) Supercapacitor (c) Antibacterial applications	7
06	Fe ₃ O ₄ /RGO Nanocomposite	Enhanced Photocatalytic Performance for Cr(VI) Reduction, Phenol Degradation, and Antibacterial Activity	8
07	Ag@ Fe₃O₄-GO Nanocomposites	(a) SERS-based detection of organic dyes (methylene blue and methyl red) (b) Removal of oil and methylene blue from water. (c) Absorption and degradation of organic dyes and nitro aromatics. (d) Antibacterial activity.	Our Work

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