Electronic Supplementary Information For

Rapid Purification and Enrichment of Viral Particles Using Self-

Propelled Micromotors

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Supporting Figures.

Figure S1. Morphological characterization and and EDX analysis of Fe₃O₄@Ag NPs.

Figure S2. Chemical schematic diagram of interface modification of micromotor.

Figure S3. FTIR spectra of CSFA micromotors.

Figure S4. SEM images of anti-SP-CSFA particles before incubation with pseudovirus.

Figure S5. Speed of micromotor with different concentration of surfactant and fuel.

Figure S6. Standard curves for quantification of pseudovirus by RT-PCR.

Figure S7. Enrichment pseudovirus from large-volume samples with and without micromotors.

Figure S8. Scheme of a typical micromotor-based nucleic acid extraction test kit.



Figure S1. Elemental analysis of Fe₃O₄@Ag NPs using TEM and EDX measurements.



Figure S2. Chemical schematic diagram of the connection process of micromotors and 3D-PETx by EDC and NHS.



Figure S3. FTIR spectra of CSFA micromotors.



Figure S4. SEM images of anti-SP-CSFA particles before incubation with pseudovirus.



Figure S5. Speed of micromotor with different concentration of (a) surfactant, and (b) fuel.

All experiments were repeated at least three times and the standard deviation of the threshold time in each experiment is shown by the error bars.



Figure S6. Standard curves for quantification of pseudovirus by RT-PCR. The RT-qPCR assay had efficiency of 99.27% with the slope of the standard curve -3.36.



Figure S7. (a) One hundred microliters of Pseudovirus were spiked into uninfected viral PBS medium to range from 200µL to 2mL, and (b) a total volume of 5 mL or 10 mL.



Figure S8. Scheme of a typical micromotor-based nucleic acid extraction test kit containing a thermostatic heating instrument, handheld syringe, and three buffer solution tubes containing enzyme-free water, micromotor fuel, and dry micromotor powder.