

Supporting Information

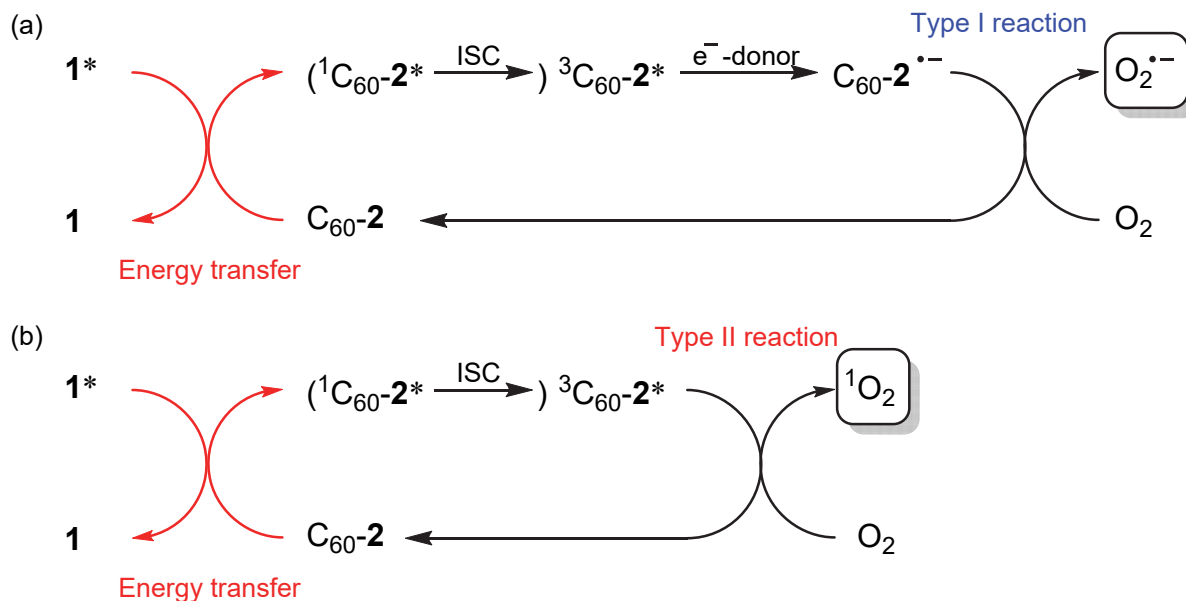
Improvement in photodynamic activity by porphyrin–fullerene composite system in lipid membrane

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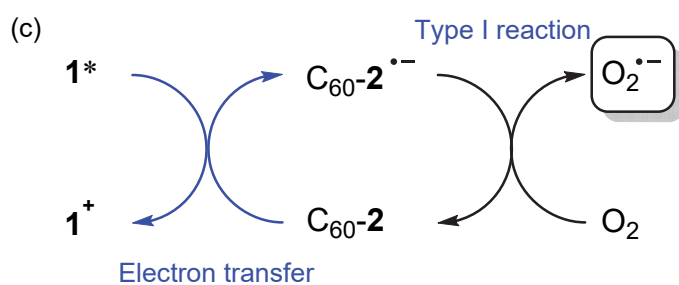
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Scheme S1. Energy or electron transfer mechanism from photoactivated **1** to C₆₀-**2**. If the triplet excited state of C₆₀-**2** (³C₆₀-**2**^{*}) is generated via energy transfer from photoactivated **1** to C₆₀-**2**, either O₂^{•-} or ¹O₂ can be produced by a Type I or Type II reaction between C₆₀-**2** and dissolved oxygen. Conversely, if the radical anion of C₆₀-**2** (C₆₀-**2**^{•-}) is generated via electron transfer from photoactivated **1** to C₆₀-**2**, only O₂^{•-} can be produced by a Type I reaction between C₆₀-**2**^{•-} and dissolved oxygen.

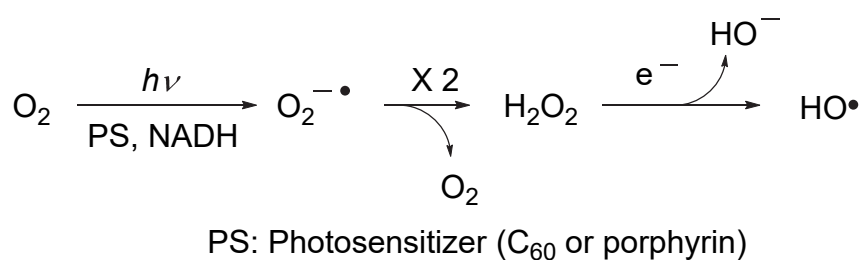
Energy transfer from porphyrin **1** to C₆₀-**2**



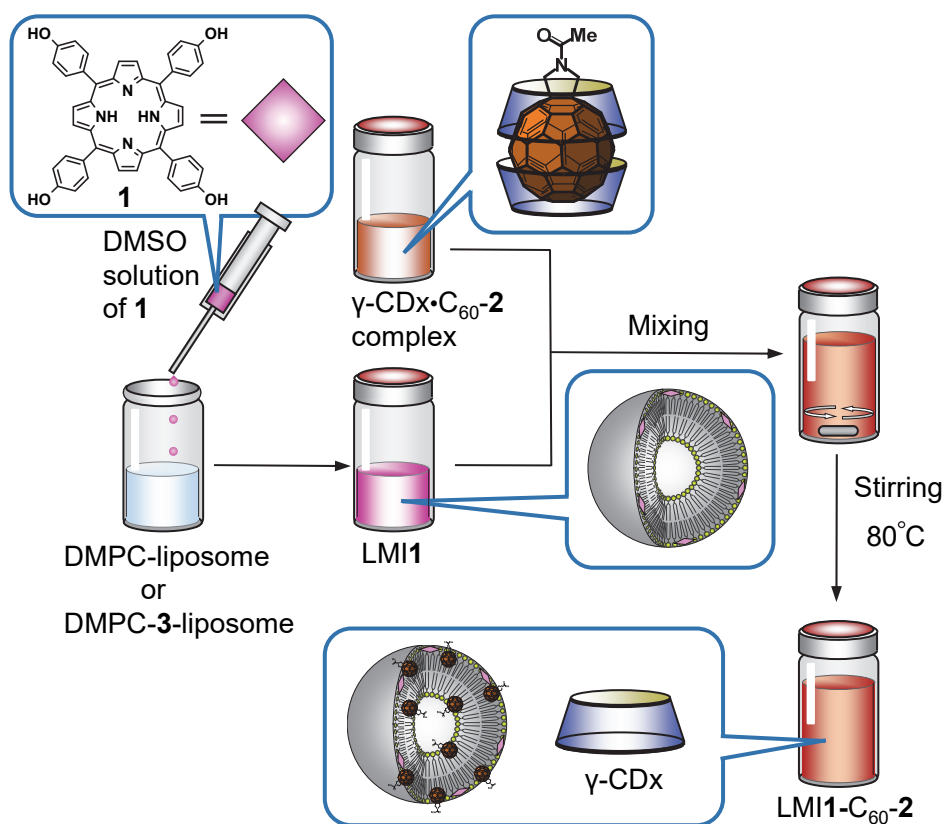
Electron transfer from porphyrin **1** to C₆₀-**2**



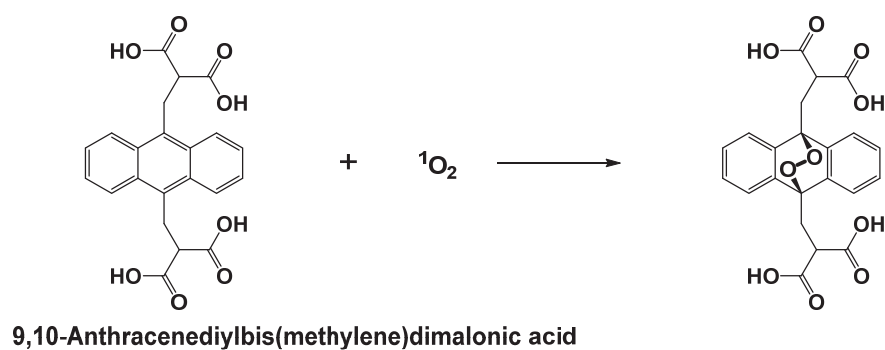
Scheme S2. Generation O₂^{•-} and then OH[•] from dissolved oxygen by photosensitizer²³



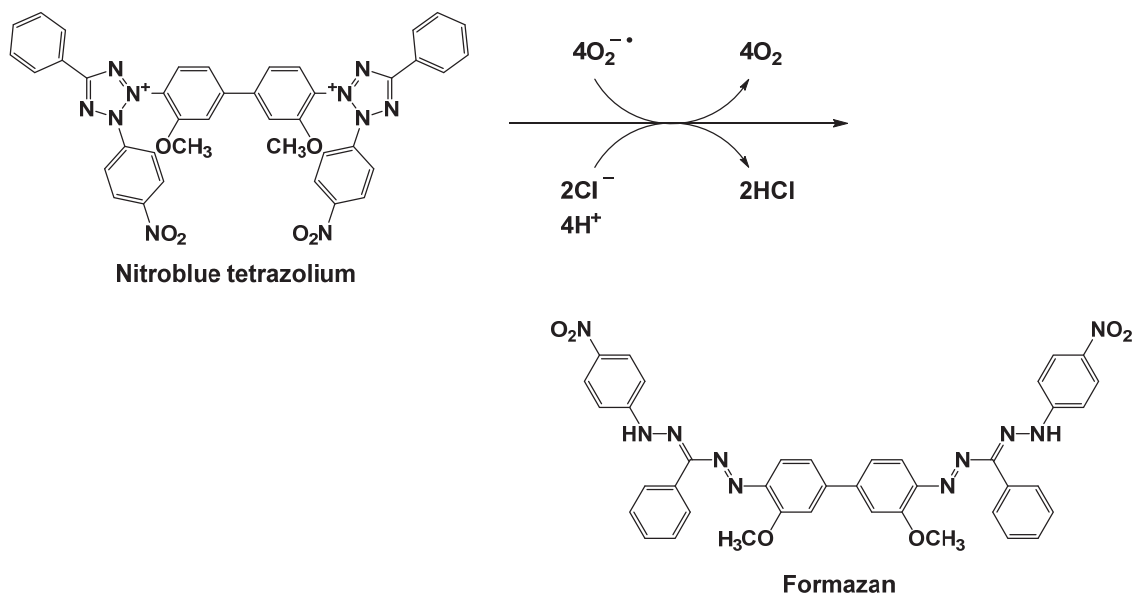
Scheme S3. Representation of the exchange method implemented for the preparation of LMI1–C₆₀-2 (γ -CDx = γ -cyclodextrin; LMI = lipid membrane–incorporated).



Scheme S4. Conversion of 9,10-anthracenediyl-bis(methylene)dimalonic acid to the corresponding endoperoxide as a result of reaction with singlet oxygen (¹O₂).



Scheme S5. Conversion of nitroblue tetrazolium to formazan as a result of reaction with superoxygen ($O_2^{\cdot-}$).



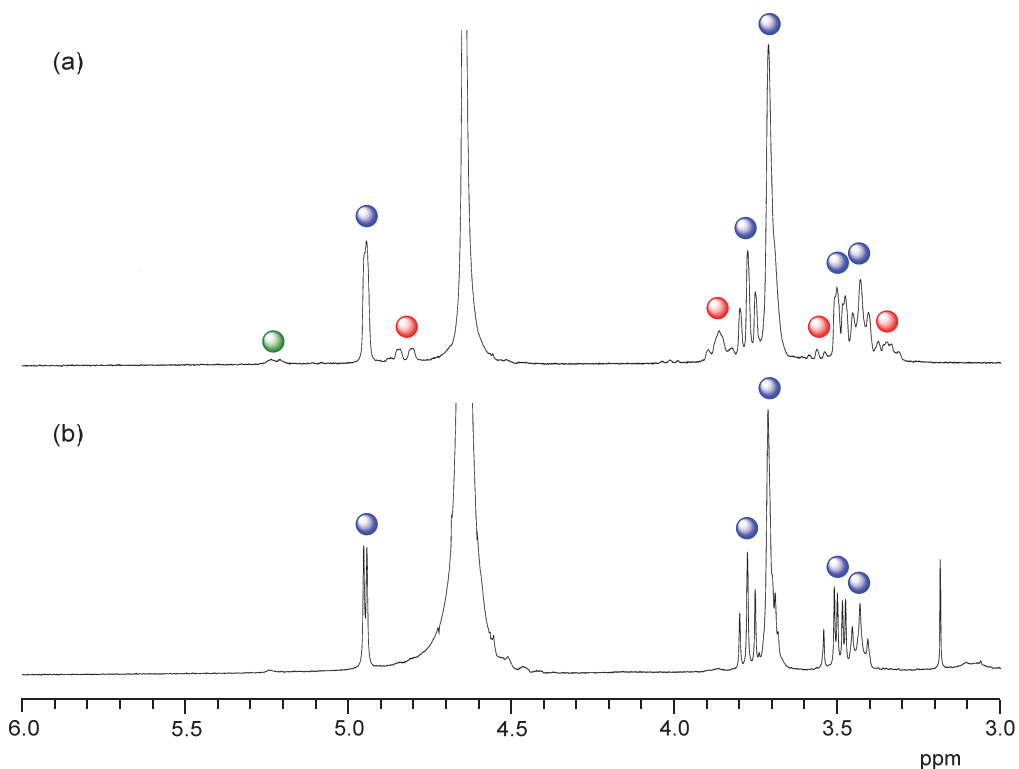


Fig. S1 Sections of the ^1H NMR spectra of the γ -cyclodextrin- $\text{C}_{60}\text{-2}$ complex (a) before and (b) after the addition of the DMPC liposomes in D_2O . The blue circles indicate peaks due to free γ -cyclodextrin; the red circles indicate peaks due to γ -cyclodextrin in the γ -cyclodextrin- $\text{C}_{60}\text{-2}$ complex; the green circle indicates the peak due to $\text{C}_{60}\text{-2}$ in the γ -cyclodextrin- $\text{C}_{60}\text{-2}$ complex. (a) $[\text{C}_{60}\text{-2}] = 0.05 \text{ mM}$; $[\gamma\text{-cyclodextrin}] = 0.7 \text{ mM}$; (b) $[\text{DMPC}] = 1.0 \text{ mM}$; $[\text{C}_{60}\text{-2}]/[\text{DMPC}] = 5.0 \text{ mol\%}$; $[\text{DiD}]/[\text{DMPC}] = 2.5 \text{ mol\%}$.

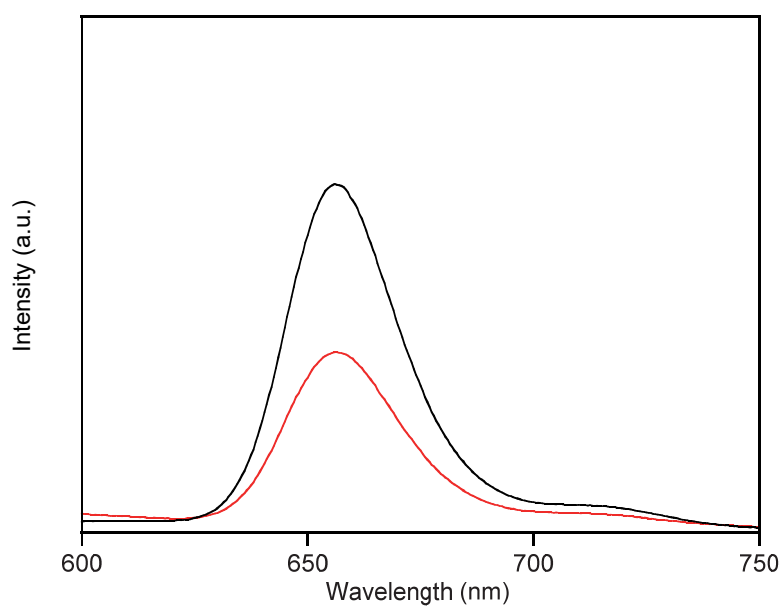


Fig. S2 Fluorescence spectra (λ_{ex} : 570 nm) of LMI1 (black line) and LMI1- $\text{C}_{60}\text{-2}$ (red line) ($[\text{DMPC}] = 0.1 \text{ mM}$; $[\text{1}]/[\text{DMPC}] = 2.5 \text{ mol\%}$; $[\text{C}_{60}\text{-2}]/[\text{DMPC}] = 5.0 \text{ mol\%}$) in water.

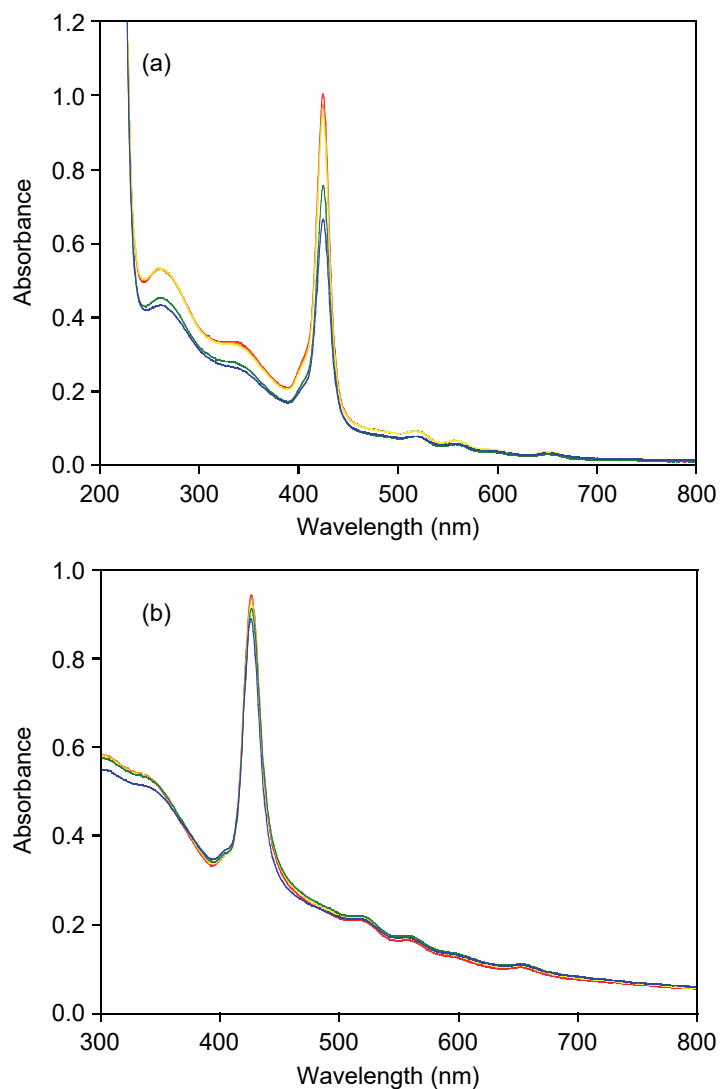


Fig. S3 UV-Vis absorption spectra of LMI1-C₆₀-2 (a) in water after incubation times of 0 (red), 1 (orange), 3 (yellow), 5 (green), and 7 (blue) days and (b) in blood serum after incubation times of 0 (red), 2 (orange), 4 (yellow), 6 (green), and 24 (blue) hours at 37 °C using a 1-cm cell ([DMPC] = 0.1 mM; [1] = 2.5 μM; [C₆₀-2] = 5.0 μM).

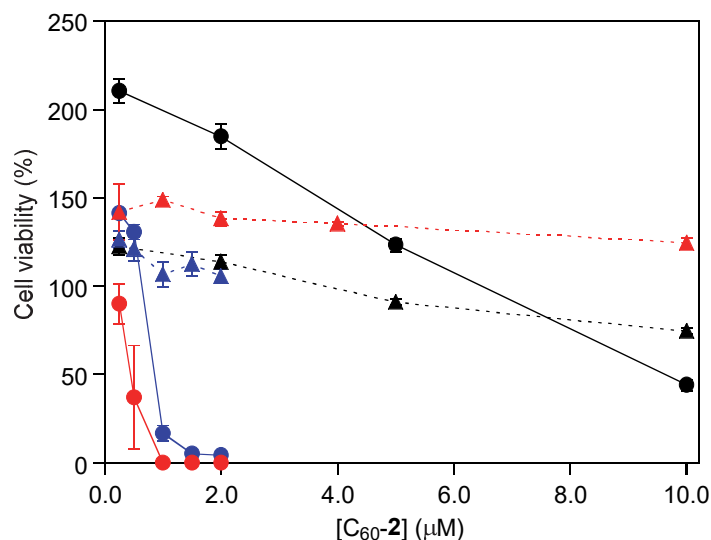


Fig. S4 Cell viability of Colon26 cells treated with LMIC₆₀₋₂ (black), LMI1-C₆₀₋₂ (red), and LMI1+LMIC₆₀₋₂ mixture (blue) in the dark (triangles and dashed lines) and after photoirradiation at 610–740 nm for 30 min (circles and solid lines) at concentrations of C₆₀₋₂ ranging from 0.25 to 10.0 μM. The cell viability was evaluated 24 h after treatment using the WST-8 method. The error bars represent the mean ± standard deviation for $n = 3$.

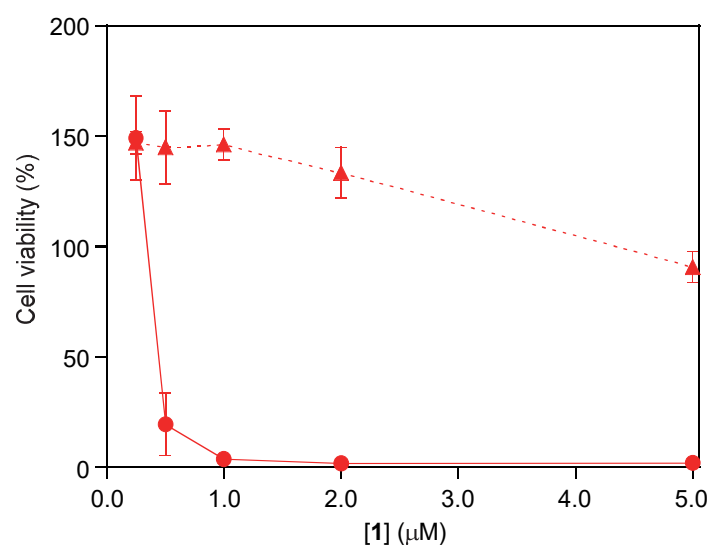


Fig. S5 Cell viability of L929 cells treated with LMI1-C₆₀₋₂ in the dark (triangles and dashed lines) and after photoirradiation at 610–740 nm for 30 min (circles and solid lines) at concentrations of C₆₀₋₂ ranging from 0.25 to 5.0 μM. The cell viability was evaluated 24 h after treatment using the WST-8 method. The error bars represent the mean ± standard deviation for $n = 3$.

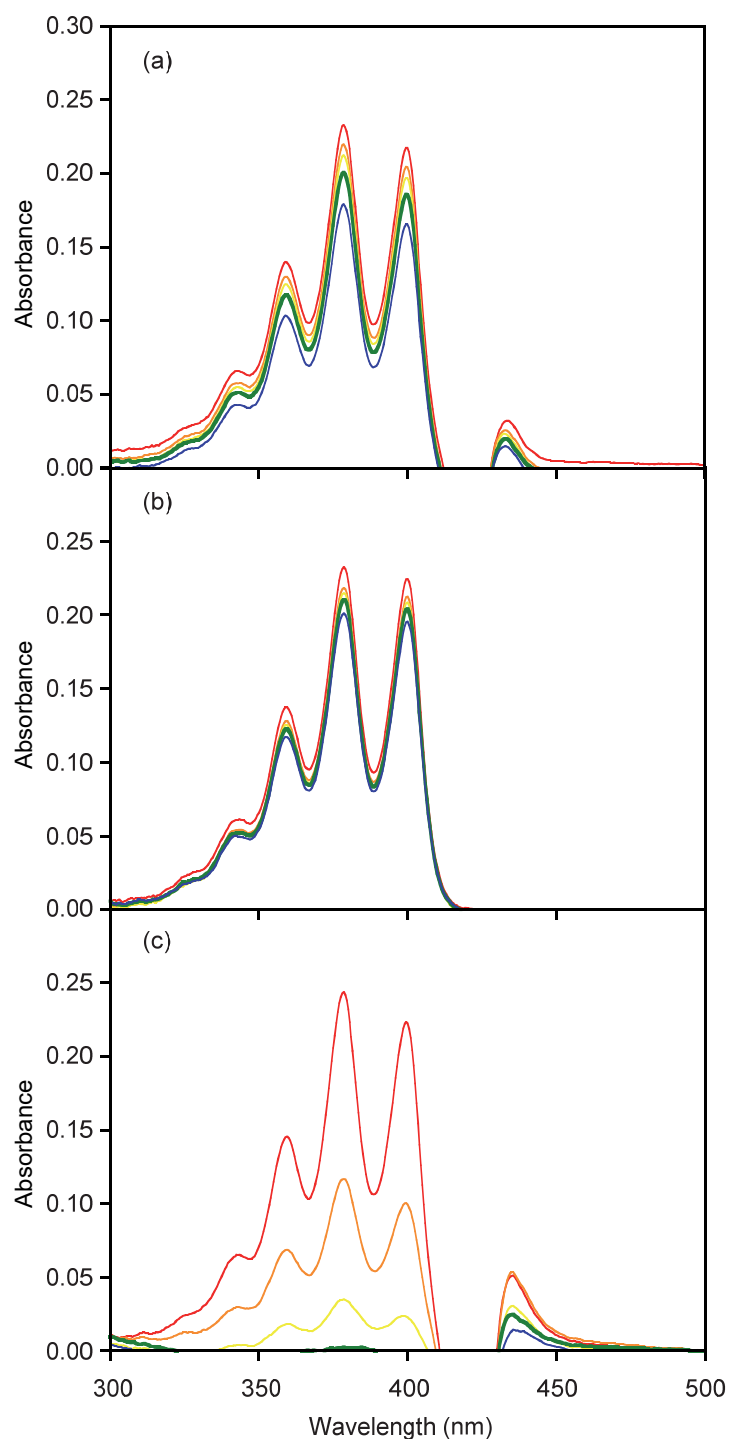


Fig. S6 Time-dependent bleaching of 9,10-anthracenediyl-bis(methylene)dimalonic acid (ABDA) caused by the generation of singlet oxygen by (a) LMI1, (b) LMIC₆₀₋₂, and (c) LMI1-C₆₀₋₂, upon photoirradiation ($\lambda > 620$ nm, 15 mW cm^{-2}) for 0 (red line), 7.5 (orange line), 15 (yellow line), 30 (green line), and 60 (blue line) min. A dimethyl sulfoxide solution of ABDA was injected into aqueous solutions of the liposomes. The absorption spectra were obtained by subtracting the absorption of LMI1, LMIC₆₀₋₂, and LMI1-C₆₀₋₂, respectively. [DMPC] = 0.1 mM, [1] = 0 or 2.5 μM , [C₆₀₋₂] = 0 or 5.0 μM , and [ABDA] = 25 μM ; 1-cm cell. The experiments were conducted under an oxygen atmosphere at 25 °C.

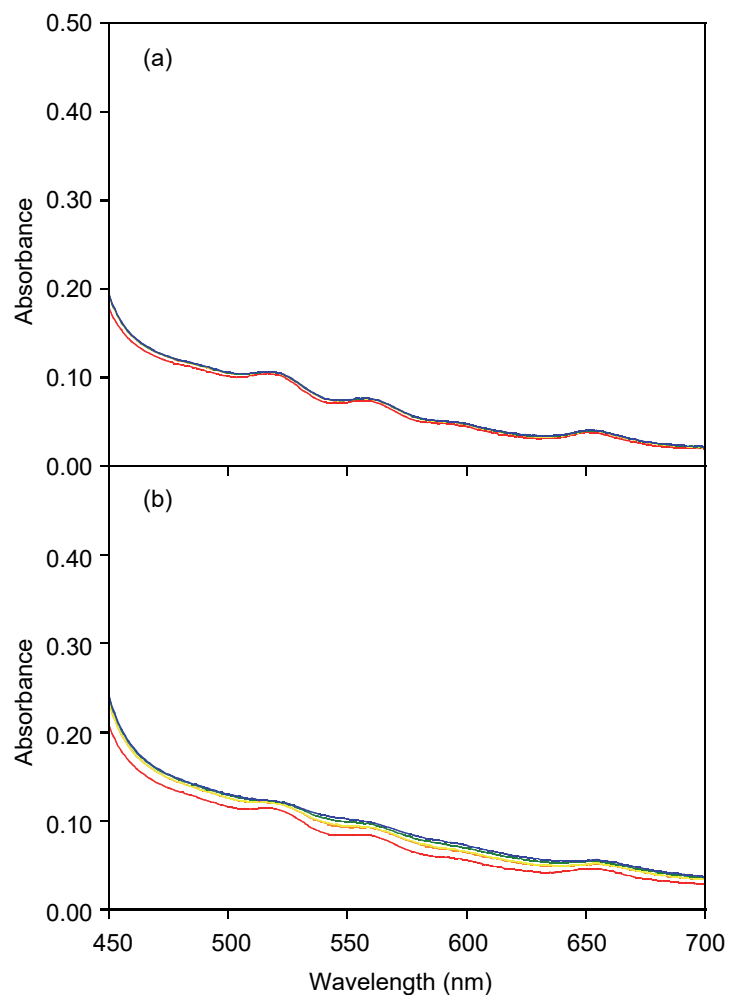


Fig. S7 Changes in the UV–Vis absorption of formazan generated by the reduction of NBT in the presence of LMI1–C₆₀-2 in (a) the absence and (b) the presence of NADH, after photoirradiation at a wavelength greater than 620 nm for 0 (red line), 7.5 (orange line), 15 (yellow line), 30 (green line), and 60 (blue line) min. [DMPC] = 0.1 mM, [1] = 0 or 2.5 μ M, [C₆₀-2] = 0 or 5.0 μ M, [NBT] = 0.20 mM, and [NADH] = 0 or 0.50 mM; 1-cm cell. The experiments were conducted under an oxygen atmosphere at 25 °C.

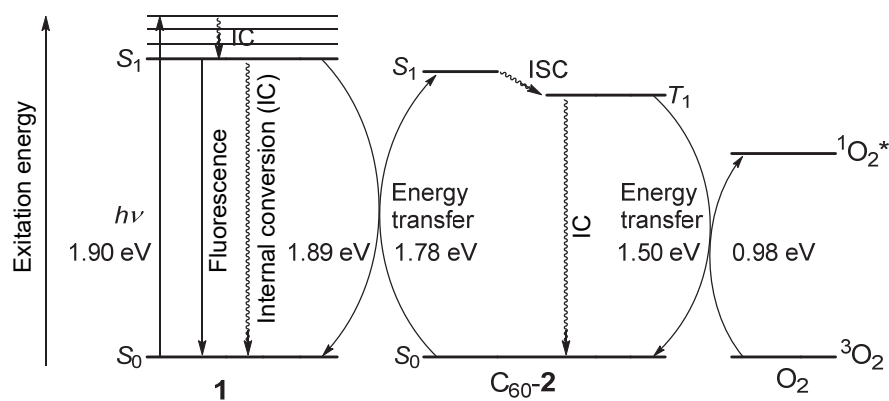


Fig. S8 Schematic illustrations of Jablonski diagrams in LMI1-C₆₀-2 (The energy potential (S₀ → S₁) of C₆₀-2 was determined using the wavelength of maximum fluorescent emission of γ-cyclodextrin-C₆₀-2 complex.²⁸ The energy potential (T₁ → S₀) of C₆₀-2 was determined from wavelength of maximum phosphorescence emission of *N*-methyl fulleropyrrolidine in methylcyclohexane, which was reported in Ref. S1 and S2, because the phosphorescence of C₆₀-2 has not reported.).

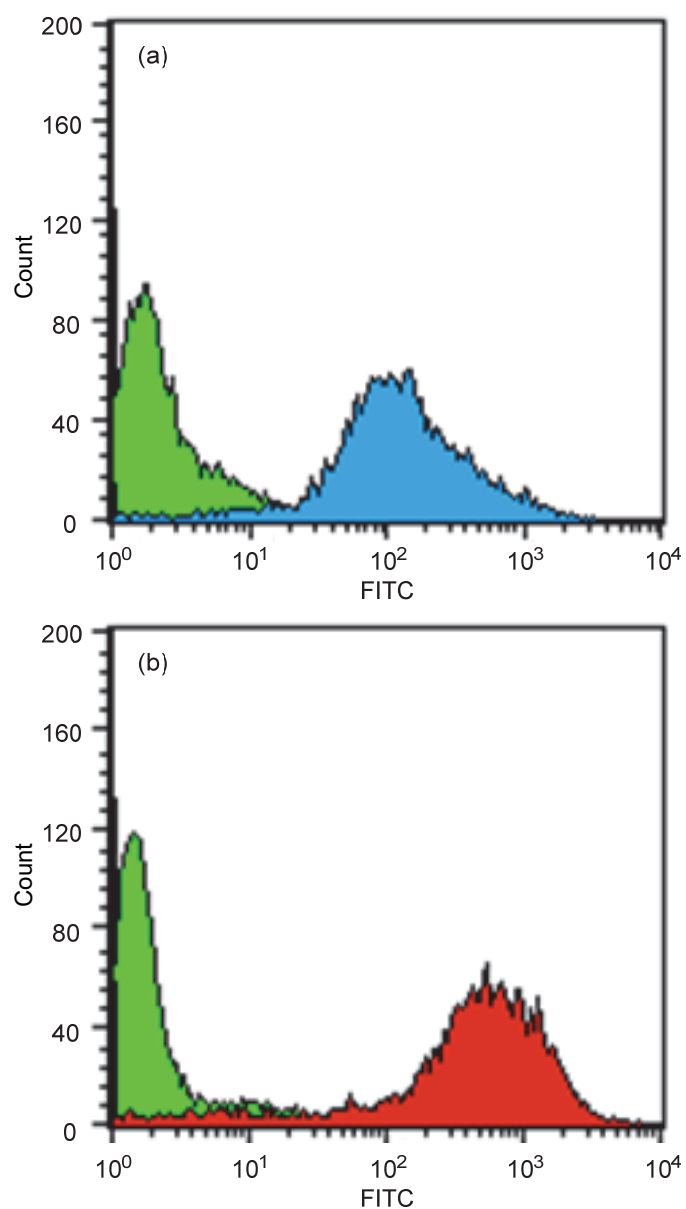


Fig. S9 Flow cytometry analysis for the detection of LMI1–C₆₀-2 in (a) Colon26 (blue) and (b) HeLa (red) cells treated with LMI1–C₆₀-2 ([DMPC] = 0.04 mM, [1] = 1.0 μM, [C₆₀-2] = 2.0 μM) and control (green; [DMPC] = [1] = [C₆₀-2] = 0 mM).

References

- S1 R. M. Williams, J. M. Zwier and J. W. Verhoeven, *J. Am. Chem. Soc.*, 1995, **117**, 4093–4099.
- S2 C. Luo, M. Fujitsuka, A. Watanabe, O. Ito, L. Gan, Y. Huang and C.-H. Huang, *J. Chem. Soc., Faraday Trans.*, 1998, **94**, 527–532.