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General Method

All reagents and solvents were commercially available and used without further purification, unless otherwise noted. ¹H, and ¹³C NMR spectra were recorded on Bruker Avance III 400 MHz. Mass spectra were determined on Bruker Daltonics Inc. APEXIII 7.0 TESLA FTMS and Agilent 6520 q-TOF LC-MS. The geometry optimization and energy calculation were performed by using Gaussian09 program with B3LYP-D3/6-31G+(d,p) level. Association constants (K_a) were examined by fluorescence titrations on Agilent CaryEclipse Fluorescence Spectrophotometer, where nonlinear curvefitting method was used to obtain the K_a values through the following equation:

 $\Delta F = 0.5*(\alpha([G]_0 + [H]_0 + 1/K) - (\alpha^2([G]_0 + [H]_0 + 1/K)^2 - 4\alpha^2[H]_0[G]_0)^{-0.5})$

Synthesis

The monomer (2,7-bis(2,4-dimethoxy-phenyl) naphthalene) and macrocycle **NBP-OMe** were prepared according to literature procedures (*Angew. Chem. Int. Ed.*, 2020, **59**, 7214–7218.).



NBP-OH. To the solution of **NBP-OMe** (1.0 g, 1.2 mmol) in DCM (250 mL) was gradually added BBr₃ (4.1 mL, 46 mmol) and stirred at 25 °C for 24 hours. After pouring the reaction solution into iced water, precipitate appeared and then was filtrated. The per-hydroxylated product **NBP-OH** was obtained (0.24 g, 90%) as brown solid. ¹H NMR (400 MHz, CD₃OD) δ (ppm): 7.85 (s, 4H), 7.76 (s, 8H), 7.20 (s, 4H), 6.48 (s, 4H), 3.85 (s, 4H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 155.3, 153.2, 137.9, 134.1, 132.1, 130.0, 128.3, 126.7, 126.1, 119.2, 118.7, 103.1, 27.7. HRMS (m/z): calcd. for [M+H]⁺: 713.2170, found 713.2156.



NBP-COOEt. To the solution of **NBP-OH** (0.20 g, 0.28 mmol) in MeCN (20 mL) was added K₂CO₃ (0.62 g, 4.5 mmol) and refluxed for 6 hours. Then ethyl bromoacetate (0.37 mL, 3.4 mmol) was added and refluxed for another 72 hours. After filtration, the filtrate was evaporated. The residue was purified by column chromatography (eluent: petroleum ether/ethyl acetate, 1/3, v/v) to afford **NBP-COOEt** as white solid (0.19 g, 48%). ¹H NMR (400 MHz, CDCl₃) δ (ppm): 7.80 (s, 8H), 7.74 (s, 4H), 7.09 (s, 4H), 6.45 (s, 4H), 4.62 (s, 8H), 4.44 (s, 8H), 4.25 (t, *J* = 7.2 Hz, 8H), 4.16 (t, *J* = 7.2 Hz, 8H), 4.11 (s, 4H), 1.25 (t, *J* = 7.2 Hz, 12H), 1.20 (t, *J* = 7.2 Hz, 12H); ¹³C NMR (100 MHz, CDCl₃) δ : 169.1, 156.3, 154.4, 136.6, 134.0, 132.9, 131.4, 128.6, 127.7, 126.7, 126.4, 125.3, 123.7, 100.1, 67.0, 66.0, 61.4, 61.3, 28.1, 14.2, 14.2. HRMS (m/z): calcd. for [M+H]⁺: 1401.5112, found 1401.5119.



NBP-COOH. To the solution of **NBP-COOEt** (80 mg, 0.057 mmol) in EtOH (4.0 mL) was added NaOH aqueous solution (1.25 ml, 30%) and refluxed for 12 hours. After evaporating the solvent and adding H₂O (8 mL), HCl was added to acidify the solution and adjust the pH to 2-3. The residue was obtained by filtration to afford **NBP-COOH** as dark brown solid (31 mg, 47%). ¹H NMR (400 MHz, DMSO-*d*₆) δ (ppm): 7.88 (s, 4H), 7.80 (m, 8H), 6.96 (s, 4H), 6.65 (s, 4H), 4.72 (s, 8H), 4.65 (s, 8H), 3.97 (s, 4H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ : 170.3, 156.1, 153.9, 137.0, 133.8, 132.1, 130.6, 128.6, 126.9, 126.2, 122.8, 121.3, 98.3, 65.4, 65.2, 40.0, 27.6. HRMS (m/z): calcd. for [M+Na]⁺: 1199.2433, found 1199.2428; calcd. for [M+NH₄]⁺: 1194.2879, found 1194.281.



WNBP. The solution of **NBP-COOH** (0.2 g, 0.17 mmol) in NH₃·H₂O (60 mL) was stirred at 25 °C for 6 hours. After evaporating the solvent, the water-soluble macrocycle **WNBP** was obtained as brown solid (0.18 g, 81%). ¹H NMR (400 MHz, D₂O) δ (ppm): 7.98 (s, 4H), 7.94 (d, J = 8.4 Hz, 4H), 7.86 (d, J = 8.0 Hz, 4H), 7.19 (s, 4H), 6.47 (s, 4H), 4.48 (s, 8H), 4.40 (s, 8H), 4.10 (s, 4H); ¹³C NMR (100 MHz, D₂O) δ : 201.7, 189.4, 182.1, 177.0, 171.8, 161.0, 158.2, 154.8, 151.7, 136.3, 134.1, 132.6, 131.4, 128.7, 127.3, 127.1, 122.5, 121.2, 99.8, 70.5, 68.1, 67.2, 65.5, 26.2. HRMS (m/z): calcd. for [M-8NH₄]⁸-: 146.02387, found 146.02362.







Figure S3. HRMS spectrum of NBP-OH.



Figure S4. ¹H NMR spectrum (400 MHz, CDCl₃, 298K) of NBP-COOEt.





Figure S6. HRMS spectrum of NBP-COOEt.



Figure S7. ¹H NMR spectrum (400 MHz, DMSO- d_6 , 298K) of NBP-COOH.





Figure S9. HRMS spectrum of NBP-COOH.



Figure S10. ¹H NMR spectrum (400 MHz, D₂O, 298K) of **WNBP**.



Figure S11. ¹³C NMR spectrum (100 MHz, D₂O, 298K) of WNBP.



Figure S12. HRMS spectrum of WNBP.



Figure S13. Partial ¹H NMR spectra (400 MHz, 0.8 mM, D_2O) of macrocycle WNBP in the presence of G2. (a) G2, (b) WNBP + G2, (c) WNBP.



Figure S14. Partial ¹H NMR spectra (400 MHz, 0.8 mM, D_2O) of macrocycle WNBP in the presence of G4. (a) G4, (b) WNBP + G4, (c) WNBP.



Figure S15. Partial ¹H NMR spectra (400 MHz, 0.8 mM, D_2O) of macrocycle WNBP in the presence of G5. (a) G5, (b) WNBP + G5, (c) WNBP.



Figure S16. Partial ¹H NMR spectra (400 MHz, 0.8 mM, D_2O) of macrocycle WNBP in the presence of G6. (a) G6, (b) WNBP + G6, (c) WNBP.



Figure S17. Fluorescence spectra of WNBP ($1.0 \mu M$) in aqueous solution recorded in the presence of different concentrations of guests at 298 K. (a) G2, (b) G3, (c) G4, (d) G5, (e) G6.



Figure S18. Fluorescence spectra of WNBP (1.0 μ M) in aqueous solution recorded in the presence of different concentrations of (a) G7, (b) G8, (c) G9 and (d) G10.



Figure S19. Job's plot obtained by plotting the fluorescence intensity change (ΔF) of **WNBP** by varying the ratio of the host and guests against the mole fraction of guest. The total concentration of the host and the guest is fixed: [Host] + [Guest] = 2.0 mM.



Figure S20. Partial ¹H NMR spectra (400 MHz, 0.8 mM, D₂O) of **WNBP** in the presence of different concentrations of **G1**.



Figure S21. Titration plots (heat flow versus time and heat flow versus molar ratio) obtained from ITC experiments of **WNPB** with (a) **G1**, (b)**G2**, (c)**G3**, (d)**G4**, (e)**G5** and (f) **G6** in phosphate buffer (10 mM, pH=7.00).

	K_{a} (in H ₂ O)	K_{a} (in phosphate buffer, 10 mM,	K_{a} (in phosphate buffer, 10 mM,	K_{a} (in phosphate buffer, 10 mM,
G1	$(7.08 \pm 0.80) \times 10^{6}$	$(3.73 \pm 0.42) \times 10^6$	$(1.51 \pm 0.19) \times 10^6$	$(1.03 \pm 0.32) \times 10^7$
G2	$(1.06 \pm 0.16) \times 10^{6}$	$(4.28 \pm 1.09) \times 10^{6}$	$(1.80 \pm 0.38) \times 10^{6}$	$(2.43 \pm 0.22) \times 10^{6}$
G3	$(2.29 \pm 0.27) \times 10^{6}$	$(2.91 \pm 0.59) \times 10^{6}$	$(1.67 \pm 0.45) \times 10^{6}$	$(2.77 \pm 0.53) \times 10^{6}$
G4	$(1.30 \pm 0.28) \times 10^7$	$(8.74 \pm 2.09) \times 10^{7}$	$(2.66 \pm 0.69) \times 10^7$	$(7.53 \pm 1.65) \times 10^{6}$
G5	$(1.26 \pm 0.40) \times 10^{6}$	$(7.83 \pm 2.82) \times 10^{6}$	$(2.24 \pm 0.64) \times 10^{6}$	$(3.17 \pm 1.08) \times 10^{6}$
G6	$(3.17 \pm 0.52) \times 10^{6}$	$(4.12 \pm 0.81) \times 10^{6}$	$(3.52 \pm 1.27) \times 10^{6}$	$(1.47 \pm 0.71) \times 10^{6}$

Table S1. Association constants (K_a , M^{-1}) of **WNBP** with the cationic guests in different solution mediums at 298 K.



Figure S22. Fluorescence spectra of **WNBP** (1.00 μ M) in phosphate buffer solutions (1.0 mM) recorded in the presence of different concentrations of **G1** at 298 K, (a) pH=6.00, (b) pH=7.00, (c) pH=8.00. Nonlinear least-squares analyses used to calculate the <u>K</u>_a value, (d) pH=6.00, (e) pH=7.00, (f) pH=8.00.



Figure S23. Fluorescence spectra of **WNBP** (1.00 μ M) in phosphate buffer solutions (1.0 mM) recorded in the presence of different concentrations of **G2** at 298 K, (a) pH=6.00, (b) pH=7.00, (c) pH=8.00. Nonlinear least-squares analyses used to calculate the K_a value, (d) pH=6.00, (e) pH=7.00, (f) pH=8.00.



Figure S24. Fluorescence spectra of **WNBP** (1.00 μ M) in phosphate buffer solutions (1.0 mM) recorded in the presence of different concentrations of **G3** at 298 K, (a) pH=6.00, (b) pH=7.00, (c) pH=8.00. Nonlinear least-squares analyses used to calculate the K_a value, (d) pH=6.00, (e) pH=7.00, (f) pH=8.00.



Figure S25. Fluorescence spectra of **WNBP** (1.00 μ M) in phosphate buffer solutions (1.0 mM) recorded in the presence of different concentrations of **G4** at 298 K, (a) pH=6.00, (b) pH=7.00, (c) pH=8.00. Nonlinear least-squares analyses used to calculate the K_a value, (d) pH=6.00, (e) pH=7.00, (f) pH=8.00.



Figure S26. Fluorescence spectra of **WNBP** (1.00 μ M) in phosphate buffer solutions (1.0 mM) recorded in the presence of different concentrations of **G5** at 298 K, (a) pH=6.00, (b) pH=7.00, (c) pH=8.00. Nonlinear least-squares analyses used to calculate the K_a value, (d) pH=6.00, (e) pH=7.00, (f) pH=8.00.



Figure S27. Fluorescence spectra of **WNBP** (1.00 μ M) in phosphate buffer solutions (1.0 mM) recorded in the presence of different concentrations of **G6** at 298 K, (a) pH=6.00, (b) pH=7.00, (c) pH=8.00. Nonlinear least-squares analyses used to calculate the K_a value, (d) pH=6.00, (e) pH=7.00, (f) pH=8.00.



Figure S28. Rigid potential energy surface (PES) scan of the **WNPB-G1** complex, 1.0 Å/step, 40 steps.



Figure S29. The Change of single point energy in the rigid potential energy surface scan of the **WNPB-G1** complex, 1.0 Å/step, 40 steps.