Electronic Supporting Information (ESI) for

Highly efficient α -arylation of aryl ketones with aryl chlorides by using bulky imidazolylidene-ligated oxazoline palladacycle

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1. General experimental details

All reagents were commercially available unless otherwise noted. Precatalysts **Cat.1**-**Cat.12** were synthesized according to previously reported literatures.¹ All reactions were carried out under argon atmosphere in dried glassware. Air and moisture sensitive liquids and solutions were transferred *via* syringe. All solvents were dried and distilled by standard procedures. Solutions were concentrated under reduced pressure by rotary evaporation. Chromatographic purification of products was accomplished on silica gel Si 60® (300-400 mesh).

Nuclear magnetic resonance spectra were acquired on a Bruker AMX 400 (400 MHz, and 100 MHz for ¹H, and ¹³C respectively) and a Bruker DRX 600 (500 MHz, and 150 MHz for ¹H, and ¹³C respectively). All ¹H NMR spectra are reported in parts per million (ppm) downfield of TMS and were measured relative to the signals at 7.26 ppm (CDCl₃). All ¹³C NMR spectra were reported in ppm relative to CDCl₃ (77.16 ppm) were obtained with ¹H -decoupling. Data for ¹H-NMR are reported as follows: chemical shift (δ in ppm), multiplicity (s = singlet; brs = broad singlet; vbs = vary broad singlet; d = doublet; t= triplet; q = quartet; quint = quintet; sext = sextet; m = multiplet), coupling constant (Hz), integration. Data for ¹³C-NMR are reported in terms of chemical shift (δ in ppm), multiplicity, coupling constant (Hz). High-resolution mass spectra were obtained on a Finnigan MAT 8200 instrument.

2. Optimization of reaction conditions

2.1 α-Arylation of *p*-methylpropiophenone 2a with chlorobenzene 1a



Scheme S1 α -Arylation of chlorobenzene 1a and *p*-methylpropiophenone 2a

Entry	$H_2O/\mu L$	Yield/%
1	0	26
2	5	30
3	8	58
4	10	73
5	12	76
6	13	87
7	14	72
8	15	71
9	20	70
10	40	26
11	60	trace

Table S1^a Screening the amount of H₂O

^a Standard condition: 1a (1.0 mmol), 2a (1.2 mmol), dioxane (3 mL), Cat.1 (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (x μL), 12 h, 100 °C. Isolated yield.

Entry	Solvent	Yield/%
1	dioxane	87
2	DME	83
3	THF	72
4	MTBE	77

Table S2^a Screening solvents

5	toluene	trace
6	ACN	51
7	EtOH	NR
8	CPME	62
9	DMF	32
10	DMSO	8

^{*a*} Standard condition: **1a** (1.0 mmol), **2a** (1.2 mmol), solvent (3 mL), **Cat.1** (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (13 μ L), 12 h, 100 °C. Isolated yield. THF= terahydrofuran; MTBE =methyl *tert*-butyl ether; ACN = acetonitrile; DMF = dimethyl formamide; DMSO = dimethyl sulfoxide.

Entry	Base	Yield/%
1	tBuONa	87
2	tBuOK	28
3	<i>t</i> BuOLi	9
4	Cs_2CO_3	NR
5	Na ₂ CO ₃	NR
6	K ₃ PO ₄	NR
7	КОН	55
8	Et ₃ N	NR

 Table S3^a Screening bases

^{*a*} Standard condition: **1a** (1.0 mmol), **2a** (1.2 mmol), dioxane (3 mL), **Cat.1** (0.5 mol%), base (2.0 mmol), H₂O (13 μL), 12 h, 100 °C. Isolated yield.

Entry	n equiv.	Yield/%
1	1.0	34
2	1.2	31
3	1.5	66
4	1.8	64

Table S4^a Screening equiv. of tBuONa

5	2.0	87
6	2.2	65
7	2.5	18

^a Standard condition: 1a (1.0 mmol), 2a (1.2 mmol), dioxane (3 mL), Cat.1 (0.5 mol%), *t*BuONa (x mmol), H₂O (13 μL), 12 h, 100 °C. Isolated yield.

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Entry	Cat./mol%	Yield/%
1	0.25	63
2	0.5	87

^a Standard condition: 1a (1.0 mmol), 2a (1.2 mmol), dioxane (3 mL), Cat.1 (x mol%), *t*BuONa (2.0 mmol), H₂O (13 μL), 12 h, 100 °C. Isolated yield.

Entry	T/°C	Yield/%
1	60	78
2	70	84
3	80	81
4	90	88
5	100	87
6	110	56

 Table S6^a Screening temperature

^a Standard condition: 1a (1.0 mmol), 2a (1.2 mmol), dioxane (3 mL), Cat.1 (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (13 μL), 12 h, T °C. Isolated yield.

Table S7^{*a*} Screening equiv. of *p*-methylphenylacetone 2a

Entry	n equiv.	Yield/%
1	1.0	83
2	1.2	88
3	1.3	88
4	1.4	91

5	1.5	93
6	1.6	83

^{*a*} Standard condition: **1a** (1.0 mmol), **2a** (x mmol), dioxane (3 mL), **Cat.1** (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (13 μ L), 12 h, 90 °C. Isolated yield.

 Table S8^a Screening the volume of solvent

Entry	Volume/mL	Yield/%
1	2	87
2	3	93
3	4	86

^{*a*} Standard condition: **1a** (1.0 mmol), **2a** (1.5 mmol), dioxane (x mL), **Cat.1** (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (13 μL), 12 h, 90 °C. Isolated yield.

Entry	Time/h	Yield/%
1	6	90
2	12	93
3	18	91
4	24	90

 Table S9^a Screening reaction time

^a Standard condition: 1a (1.0 mmol), 2a (1.5 mmol), dioxane (3 mL), Cat.1 (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (13 μL), x h, 90 °C. Isolated yield.

Entry	Cat.	Yield/%
1	Cat.1	93
2	Cat.2	85
3	Cat.3	87
4	Cat.4	91
5	Cat.5	89
6	Cat.6	91

 Table S10^a Screening catalysts

7	Cat.7	92
8	Cat.8	85
9	Cat.9	92

^a Standard condition: 1a (1.0 mmol), 2a (1.5 mmol), dioxane (3 mL), Cat. (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (13 μL), 12 h, 90 °C. Isolated yield.

2.2 α-Arylation of *p*-methylpropiophenone 2a with *p*-chloroanisole 1e



Scheme S2 α -Arylation of *p*-chloroanisole 1e and *p*-methylpropiophenone 2a

Entry	$H_2O/\mu L$	Yield/%
1	0	NR
2	8	63
3	10	64
4	13	61
5	15	44
6	18	44
7	20	NR

Table S11^a Screening the amount of H₂O

^{*a*} Standard condition: **1e** (1.0 mmol), **2a** (1.5 mmol), dioxane (3 mL), **Cat.1** (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (x μL), 12 h, 90 °C. Isolated yield.

Entry	Cat.	Yield/%
1	Cat.1	64
2	Cat.2	81

 Table S12^a Screening catalysts

3	Cat.3	15
4	Cat.4	94/93 ^b
5	Cat.5	89
6	Cat.6	55
7	Cat.7	68
8	Cat.8	67
9	Cat.9	58
10	Cat.10	79
11	Cat.11	93
12	Cat.12	82

^a Standard condition: 1e (1.0 mmol), 2a (1.5 mmol), dioxane (3 mL), Cat. (0.5 mol%),
tBuONa (2.0 mmol), H₂O (10 μL), 12 h, 90 °C. Isolated yield. ^b D₂O (10 μL).



3. Substrate scopes



Scheme S3 Substrate scopes catalyzed by using Cat.1 according to standard condition A: Standard condition (Method A): 1 (1.0 mmol), 2 (1.5 mmol), Cat.1 (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (13 μ L), dioxane (3 mL), 12 h, 90 °C. Isolated yield.



Scheme S4 Substrates tested with poor reactivity by using Cat.4 according to standard condition (Method B): 1 (1.0 mmol), 2 (1.5 mmol), Cat.4 (0.5 mol%), *t*BuONa (2.0 mmol), H₂O (10 μ L), dioxane (3 mL), 12 h, 90 °C. Isolated yield.

4. Control experiments and proposal mechanism



Scheme S5 α -Arylation of *p*-chloroanisole 1e and *p*-methylpropiophenone 2a in the presence of D₂O



Fig. S1 ¹H NMR (600 MHz, CDCl₃) spectrum of 3e' in the presence of D₂O



Fig. S2 ¹H NMR (600 MHz, CDCl₃) spectrum of 3e in the presence of H₂O



Scheme S6 Proposed reaction mechanism of α -arylation catalyzed by Cat.4

At the current stage, we suggested the presence of H_2O effected the keto-enol tautomerism equilibrium, however, its role in the activation of the precatalyst² could be excluded. We will carry out the DFT calculations with other groups to get a deep insight.

5. General procedure for synthesis of substrates

To a flame dried round bottom flask equipped with a magnetic stir bar, diisopropylamine (0.92 mL, 6.6 mmol) in dry THF (15 mL) was added at -78 °C. Then, *n*BuLi (2.64 mL, 6.6 mmol, 2.5 M in Hexane) was added dropwise, and the reaction mixture was stirred for 30 minutes. Then, acetophenone (0.70 mL, 6 mmol) was added and reacted for 30 minutes. Then, methyl iodide (0.45 mL, 7.2 mmol) in dry THF (15 mL) was added. After 4 h, the reaction was quenched with saturated ammonium chloride. The organic phase was separated, and the aqueous phase was washed with hexane (1×5 mL). The combined organic phases were washed with 1 M HCl (1×10 mL), dried over anhydrous sodium sulfate, filtered, and evaporated under reduced pressure. The crude products were further purified by column chromatography leading to desired products.



2q: Following the **General procedure**, **2q** was obtained as a yellow oil, 645.7 mg, yield = 27%.

¹H NMR (600 MHz, CDCl₃): δ = 7.98-7.95 (m, 2H), 7.57-7.54 (m, 1H), 7.48-7.44 (m, 2H), 3.01 (q, *J* = 7.2 Hz, 2H), 1.23 (t, *J* = 7.2 Hz, 3H). The data was the same to previous literature.³



2u: Following the General procedure, 2u was obtained as a white solid, 359.3 mg, yield = 28%.
¹H NMR (600 MHz, CDCl₃): δ = 8.06-8.03 (m, 2H), 7.70-7.67 (m, 2H), 7.64-7.62 (m, 2H), 7.49-7.46 (m, 2H), 7.46-7.38 (m, 1H), 3.04 (q, J = 7.2 Hz, 2H), 1.26 (t, J = 7.2 Hz, 2H)

3H). The data was the same to previous literature.⁴



2y: Following the **General procedure**, **2y** was obtained as a yellow oil, 325.8 mg, yield = 15%.

¹H NMR (400 MHz, CDCl₃): $\delta = 8.56$ (J = 8.5 Hz, 1H), 7.97 (d, J = 8.0 Hz, 1H), 7.98 (d, J = 8.0 Hz, 1H), 7.90-7.83 (dd, J = 7.5, 1.0 Hz, 2H), 7.62-7.47 (m, 3H), 3.08 (q, J = 7.0 Hz, 2H), 1.29 (t, J = 7.0 Hz, 3H). The data was the same to previous literature.⁵

6. Date for arylation products



3a: Pale yellow oil, 203.1 mg, yield: 91%.

¹H NMR (600 MHz, CDCl₃): δ = 7.86 (d, *J* = 8.2 Hz, 2H), 7.28-7.29 (d, *J* = 4.4 Hz, 4H), 7.22-7.16 (m, 3H), 4.67 (q, *J* = 6.8 Hz, 1H), 2.35 (s, 3H), 1.53 (d, *J* = 6.9 Hz, 3H). Data is consistent with that reported in the literature.⁶

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3b: Pale yellow solid, 206.0 mg, yield: 86%.

¹H NMR (600 MHz, CDCl₃): δ = 7.74 (d, *J* = 8.2 Hz, 2H), 7.20 (d, *J* = 7.1 Hz, 1H), 7.15 (d, *J* = 8.0 Hz, 2H),7.12-7.06 (m, 2H), 7.04-7.02 (d, *J* = 7.4 Hz, 1H), 4.74 (q, *J* = 6.8 Hz, 1H), 2.49 (s, 3H), 2.34 (s, 3H), 1.47 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃): δ = 200.6, 143.4, 140.4, 134.5, 134.1, 130.9, 129.2,

128.6, 127.0, 126.8, 126.7, 44.5, 21.5, 19.6, 18.0.

HR-MS (ESI, m/z): calcd for C₁₇H₁₈O [M+H] ⁺: 239.1430. found: 239.1426. m.p.: 63.4-66.7 °C.



3c: Pale yellow oil, 215.8 mg, yield: 91%.

¹H NMR (600 MHz, CDCl₃): δ = 7.86 (d, *J* = 8.2 Hz, 2H), 7.20-7.15 (m, 3H), 7.08 (d, *J* = 1.8 Hz, 2H), 6.98 (d, *J* = 7.5 Hz, 1H), 4.62 (q, *J* = 6.8 Hz, 1H), 2.35 (s, 3H), 2.30 (s, 3H), 1.50 (d, *J* = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁷



3d: Pale yellow oil, 219.0 mg, yield: 92%.

¹H NMR (600 MHz, CDCl₃): δ = 7.86 (d, *J* = 8.2 Hz, 2H), 7.17 (d, *J* = 8.1 Hz, 4H), 7.08 (d, *J* = 7.9 Hz, 2H), 4.63 (q, *J* = 6.8 Hz, 1H), 2.35 (s, 3H), 2.28 (s, 3H), 1.50 (d, *J* = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁷



3e: Pale yellow oil, 239.6 mg, yield: 94%.

¹H NMR (600 MHz, CDCl₃): δ = 7.85 (d, *J* = 8.2 Hz, 2H), 7.21-7.16 (m, 4H), 6.83-6.81 (dt, *J* = 8.2, 2.6 Hz, 2H), 4.62 (q, *J* = 6.8 Hz, 1H), 3.75 (s, 3H), 2.35 (s, 3H), 1.49 (d, *J* = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁸



new compound

3f: Pale yellow solid, 253.8 mg, yield: 91%.

¹H NMR (600 MHz, CDCl₃): δ = 7.88 (d, *J* = 8.2 Hz, 2H), 7.29 (td, *J* = 8.4, 2.0 Hz, 2H), 7.22-7.18 (m, 4H), 4.66 (q, *J* = 6.8 Hz, 1H), 2.35 (s, 3H), 1.51 (d, *J* = 6.6 Hz, 3H), 1.27 (s, 9H).

¹³C NMR (150 MHz, CDCl₃): δ = 200.2, 149.6, 143.5, 138.5, 134.1, 129.2, 129.0, 127.4, 125.8, 47.1, 34.4, 31.3, 21.6, 19.5.

HR-MS (ESI, m/z): calcd for C₂₀H₂₄O [M+H] ⁺: 281.1900. found: 281.1894. m.p.: 73.8-75.9 °C.



new compound

3g: White solid, 270.4 mg, yield: 90%.

¹H NMR (600 MHz, CDCl₃): δ = 7.90 (dt, *J* = 8.2 Hz, 2H), 7.55-7.51 (dd, *J* = 7.4, 8.2 Hz, 4H), 7.41 (t, *J* = 7.7 Hz, 2H), 7.35 (d, *J* = 8.2 Hz, 2H), 7.33-7.30 (t, *J* = 7.4 Hz, 1H), 7.20 (d, *J* = 8.0 Hz, 2H), 4.72 (q, *J* = 6.8 Hz, 1H), 2.36 (s, 3H), 1.56 (d, *J* = 6.8 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃): δ = 200.0, 143.7, 140.8, 140.7, 139.8, 134.0, 129.3, 129.0, 128.8, 128.2, 127.7, 127.3, 127.0, 47.4, 21.6, 19.5.

HR-MS (ESI, m/z): calcd for C₂₂H₂₀O [M+H] ⁺: 301.1587. found: 301.1583. m.p.: 140.8-143.2 °C.



3h: Pale yellow oil, 234.0 mg, yield: 96%.

¹H NMR (600 MHz, CDCl₃): δ = 7.84 (d, *J* = 8.2 Hz, 2H), 7.25-7.23 (m, 2H), 7.19 (d, *J* = 8.0 Hz, 2H), 6.99-6.95 (tt, *J* = 8.7, 1.83 Hz, 2H), 4.66 (q, *J* = 6.9 Hz, 1H), 2.36 (s, 3H), 1.50 (d, *J* = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁸



3i: Pale yellow oil, 160.8 mg, yield: 55%.

¹H NMR (600 MHz, CDCl₃): δ = 7.85 (d, *J* = 8.2 Hz, 2H), 7.56 (s, 1H), 7.47 (t, *J* = 7.4 Hz, 2H), 7.40 (t, *J* = 7.7 Hz, 1H), 7.21 (d, *J* = 8.0 Hz, 2H), 4.75 (q, *J* = 6.9 Hz, 1H), 2.37 (s, 3H), 1.55 (d, *J* = 6.9 Hz, 3H). Data is consistent with that reported in the literature.⁸



3j: Pale yellow oil, 113.5 mg, yield: 44%.

¹H NMR (600 MHz, CDCl₃): δ = 7.83 (d, J = 8.2 Hz, 2H), 7.41-7.39 (m, 1H), 7.18 (d,

J = 8.0 Hz, 2H), 7.15-7.13 (m, 3H), 5.12 (q, *J* = 6.8 Hz, 1H), 2.35 (s, 3H), 1.48 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃): δ = 199.7, 143.8, 139.5, 133.6, 132.9, 129.8, 129.3, 128.7, 128.6, 128.2, 127.5, 44.1, 21.6, 17.8.

HR-MS (ESI, m/z): calcd for C₁₆H₁₅ONaCl [M+Na] ⁺: 287.0704. found: 287.0711.



3k: Pale yellow solid, 194.6 mg, yield: 77%.

¹H NMR (600 MHz, CDCl₃): δ = 7.60 (d, *J* = 8.2 Hz, 2H), 7.07 (d, *J* = 8.0 Hz, 2H),

7.02-6.95 (m, 3H), 4.50 (q, *J* = 6.8 Hz, 1H), 2.30 (s, 9H), 1.50 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃): δ = 201.9, 143.2, 140.1, 135.6, 134.3, 129.5, 129.0, 128.3, 126.6, 46.1, 21.5, 20.6, 14.9.

HR-MS (ESI, m/z): calcd for $C_{18}H_{20}O$ [M+H] ⁺: 253.1587. found: 253.1584.

m.p.: 74.8-78.9 °C.



31: Pale yellow solid, 247.3 mg, yield: 90%.

¹H NMR (600 MHz, CDCl₃): $\delta = 8.25$ (d, J = 8.5 Hz, 1H), 7.90 (d, J = 8.2 Hz, 1H), 7.77 (d, J = 8.2 Hz, 2H), 7.72 (d, J = 8.2 Hz, 1H), 7.64-7.61 (m, 1H), 7.55-7.52 (m, 1H), 7.33 (t, J = 7.9 Hz, 1H), 7.21 (dd, J = 6.6 Hz, 0.6 Hz, 1H), 7.07 (t, J = 8.1 Hz, 2H), 5.36 (q, J = 6.8 Hz, 1H), 2.29 (s, 3H), 1.63 (d, J = 6.8 Hz, 3H). ¹³C NMR (150 MHz, CDCl₃): $\delta = 200.4$, 143.5, 138.3, 134.4, 133.9, 130.7, 129.3, 129.2, 128.7, 127.5, 126.7, 125.9, 125.8, 125.0, 122.6, 43.6, 21.5, 18.6. HR-MS (ESI, m/z): calcd for C₂₀H₁₈O [M+H] ⁺: 275.1430. found: 275.1427. m.p.: 116.8-120.8 °C.



3m: Pale yellow oil, 257.7 mg, yield: 94%.

¹H NMR (600 MHz, CDCl₃): δ = 7.90 (d, *J* = 8.3 Hz, 2H), 7.78 (t, *J* = 8.2 Hz, 3H), 7.72 (t, *J* = 1.2 Hz, 1H), 7.46-7.40 (m, 3H), 7.16 (d, *J* = 8.0 Hz, 2H), 4.83 (q, *J* = 6.8 Hz, 1H), 2.33 (s, 3H), 1.61 (d, *J* = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁸



new compound

3n: Pale yellow solid, 287.6 mg, yield: 81%.

¹H NMR (600 MHz, CDCl₃): δ = 7.82 (d, *J* = 8.3 Hz, 2H), 7.56 (d, *J* = 8.1 Hz, 1H), 7.28-7.26 (m, 3H), 7.16 (s, 1H), 7.10 (d, *J* = 8.1 Hz, 2H), 7.08-7.05 (m, 4H), 6.47 (d, *J* = 3.0 Hz, 1H), 5.26 (d, *J* = 3.4 Hz, 2H), 4.71 (q, *J* = 6.8 Hz, 1H), 2.32 (s, 3H), 1.54 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃): δ = 200.0, 143.6, 140.2, 138.3, 138.0, 127.0, 124.4, 123.8, 123.0, 122.6, 47.6, 29.8, 21.6, 19.9.

HR-MS (ESI, m/z): calcd for C₂₅H₂₃NO [M+H] ⁺: 354.1852. found: 354.1845. m.p.: 97.2-99.3 °C.



30: Pale yellow solid, 266.9 mg, yield: 95%.

¹H NMR (600 MHz, CDCl₃): δ = 7.88 (d, *J* = 8.3 Hz, 2H), 7.80 (d, *J* = 8.3 Hz, 1H), 7.72 (d, *J* = 1.5 Hz, 1H), 7.41-7.40 (d, *J* = 5.4 Hz, 1H), 7.30-7.28 (dd, *J* = 1.6, 6.7 Hz, 1H), 7.27 (s, 1H), 7.16 (d, *J* = 8.1 Hz, 2H), 4.79 (d, *J* = 6.8 Hz, 1H), 2.33 (s, 3H), 1.58 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃): δ = 200.0, 143.6, 140.7, 137.7, 136.8, 133.9, 129.2, 128.8, 127.0, 124.8, 123.0, 121.3, 121.2, 46.0, 21.6, 18.5.
HR-MS (ESI, m/z): calcd for C₁₈H₁₆SO [M+H] ⁺: 281.0995. found: 281.0985.
m.p.: 71.4-74.8 °C.



3p: Pale yellow solid, 257.8.9 mg, yield: 92%.

¹H NMR (600 MHz, CDCl₃): δ = 7.80 (d, *J* = 8.3 Hz, 2H), 7.74 (d, *J* = 8.0 Hz, 1H), 7.60 (d, *J* = 5.5 Hz, 1H), 7.53 (d, *J* = 5.5 Hz, 1H), 7.25-7.22 (t, *J* = 7.7 Hz, 1H), 7.15 (d, *J* = 7.3 Hz, 1H), 7.11 (d, *J* = 8.0 Hz, 2H), 5.10 (d, *J* = 6.8 Hz, 1H), 2.30 (s, 3H), 1.62 (d, *J* = 6.8 Hz, 3H).

¹³C NMR (150 MHz, CDCl₃): δ = 200.4, 143.5, 138.3, 134.4, 133.8, 130.6, 129.3, 129.2, 128.7, 127.5, 126.7, 125.9, 125.8, 125.0, 122.6, 43.6, 21.6, 18.6.

HR-MS (ESI, m/z): calcd for C₁₈H₁₆SO [M+H]⁺: 281.0995. found: 281.0984. m.p.: 84.0-86.4 °C.



3q: Pale yellow oil, 196.6 mg, yield: 93%.

¹H NMR (600 MHz, CDCl₃): δ = 7.95 (d, *J* = 7.3 Hz, 2H), 7.47 (t, *J* = 6.8 Hz, 1H), 7.38 (t, *J* = 7.4 Hz, 2H), 7.30-7.25 (m, 4H), 7.20 (m, 1H), 4.69 (q, *J* = 6.7 Hz, 1H), 1.53 (d, *J* = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁶

3r: Pale yellow oil, 215.6 mg, yield: 96%.

¹H NMR (600 MHz, CDCl₃): δ = 7.97-7.95 (m, 2H), 7.49-7.46 (m, 1H), 7.40-7.37 (m,

2H), 7.32-7.27 (m, 4H), 7.22-7.19 (m, 1H), 4.45 (t, J = 7.3 Hz, 1H), 2.24-2.17 (m, 1H), 1.90-1.83 (m, 1H), 0.91 (t, J = 7.4 Hz, 3H). Data is consistent with that reported in the literature.⁶



3s: Pale yellow oil, 217.0mg, yield: 91%.

¹H NMR (600 MHz, CDCl₃): δ = 7.97-7.95 (m, 2H), 7.49-7.46 (m, 1H), 7.40-7.38 (t, *J* = 7.7 Hz, 2H), 7.32-7.27 (m, 4H), 7.21-7.18 (m, 1H), 4.56 (t, *J* = 7.3 Hz, 1H), 2.19-2.13 (m, 1H), 1.85-1.79 (m, 1H), 1.38-1.25 (m, 2H), 0.92 (t, *J* = 7.5 Hz, 3H). Data is consistent with that reported in the literature.⁹



3t: Pale yellow oil, 237.0 mg, yield: 96%.

¹H NMR (600 MHz, CDCl₃): δ = 7.95 (d, *J* = 8.8 Hz, 2H), 7.30-7.28 (m, 4H), 7.22-7.18 (m, 1H), 6.85 (d, *J* = 8.8 Hz, 2H), 4.64 (q, *J* = 6.8 Hz, 1H), 3.81 (s, 3H), 1.51 (d, *J* = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁸



3u: Pale yellow oil, 218.7 mg, yield: 76%.

¹H NMR (600 MHz, CDCl₃): δ = 8.02 (d, *J* = 7.8 Hz, 2H), 7.60 (dd, *J* = 11.7, 8.0 Hz, 4H), 7.44 (d, *J* = 7.3 Hz, 2H), 7.38 (d, *J* = 6.9 Hz, 1H), 7.32-7.31 (m, 4H), 7.24-7.20 (m, 1H), 4.72 (q, *J* = 6.8 Hz, 1H), 1.56 (d, *J* = 6.9 Hz, 3H). Data is consistent with that reported in the literature.⁶



3v: Pale yellow oil, 225.2 mg, yield: 80%. ¹H NMR (600 MHz, CDCl₃): $\delta = 8.02$ (d, J = 8.1 Hz, 2H), 7.63 (d, J = 7.2, 8.2 Hz, 2H), 7.32-7.29 (m, 2H), 7.26-7.24 (m, 2H), 7.23-7.20 (m, 1H), 4.65 (q, J = 6.8 Hz, 1H), 1.55 (d, J = 6.9 Hz, 3H). Data is consistent with that reported in the literature.⁸



3w: Pale yellow oil, 190.4 mg, yield: 83%.

¹H NMR (400 MHz, CDCl₃): δ = 7.98-7.95 (m, 2H), 7.32-7.26 (m, 4H), 7.22-7.20(m, 1H), 7.06-7.02 (m, 2H), 4.62 (q, J = 6.8 Hz, 1H), 1.52 (d, J = 6.9 Hz, 3H). Data is consistent with that reported in the literature.¹⁰



3w: Pale yellow oil, 164.7 mg, yield: 72%. ¹H NMR (400 MHz, CDCl₃): δ = 7.71 (d, J = 7.8 Hz, 1H), 7.62 (dt, J = 9.6, 4.2 Hz, 1H), 7.37-7.26(m, 5H), 7.24-7.14 (m, 2H), 4.62 (q, J = 6.8 Hz, 1H), 1.53 (d, J = 6.8 Hz, 3H). Data is consistent with that reported in the literature.⁷



3y: Pale yellow oil, 85.0 mg, yield: 32%.

¹H NMR (600 MHz, CDCl₃): δ = 7.90 (d, J = 8.2 Hz, 1H), 7.82 (d, J = 7.9 Hz, 1H), 7.75 (d, J = 7.1 Hz, 1H), 7.70 (d, J = 7.7 Hz, 1H), 7.54-7.48 (m, 2H), 7.43-7.40 (m, 1H), 7.31-7.29 (m, 2H), 7.27-7.24 (m, 2H), 7.17 (t, *J* = 7.1 Hz, 1H), 4.70 (q, *J* = 6.8 Hz, 1H), 1.64 (d, *J* = 6.9 Hz, 3H). Data is consistent with that reported in the literature.¹¹



3z: Pale yellow oil, 39.3 mg, yield: 18%.

¹H NMR (600 MHz, CDCl₃): δ = 8.10 (dd, *J* = 0.9, 7.8 Hz, 1H), 7.50 (td, *J* = 1.4, 7.5 Hz, 1H), 7.36-7.33 (m, 3H), 7.30-7.27 (m, 2H), 7.21-7.19 (m, 2H), 3.81 (dd, *J* = 2.8, 6.5 Hz, 1H), 3.15-3.04 (m, 2H), 2.46-2.42 (m, 2H). Data is consistent with that reported in the literature.¹²

References

(a) K. Wang, H. Yang, F. Bauer, B. Breit and W. Fang, *Chem. Eur. J.*, 2023, e202300719; (b) T. Tu,
 W. Fang and J. Jiang, *Chem. Commun.*, 2011, **47**, 12358-12360; (c) S. Ghorai, S. U. Rehman, W. Xu,
 W. Huang and C. Li, *Org. Lett.*, 2020, **22**, 3519-3523; (d) Q. Deng, Y. Zhang, H. Zhu and T. Tu, *Chem. Asian J.*, 2017, **12**, 2364-2368; (e) W. Fang, Q. Deng, M. Xu and T. Tu, *Org. Lett.*, 2013, **15**, 3678-3681.
 O. Navarro, N. Marion, Y. Oonishi, R. A. Kelly, 3rd and S. P. Nolan, *J. Org. Chem.*, 2006, **71**, 685-692.

3 T. J. Malinski and D. E. Bergbreiter, Tetrahedron Lett., 2018, 59, 3926-3929.

4 S. Mao, Z. Chen, L. Wang, D. B. Khadka, M. Xin, P. Li and S. Zhang, J. Org. Chem., 2019, 84, 463-471.

5 M. Sai, Eur. J. Org. Chem., 2022, 2022, e202200052.

6 J. Templ and M. Schnürch, J. Org. Chem., 2022, 87, 4305-4315.

7 F. Liu, Y. Hu, D. Li, Q. Zhou and J. Lu, Tetrahedron, 2018, 74, 5683-5690.

8 M. Escudero-Casao, G. Licini and M. Orlandi, J. Am. Chem. Soc., 2021, 143, 3289-3294.

9 R. Kulasekharan, M. V. S. N. Maddipatla, A. Parthasarathy and V. Ramamurthy, *J. Org. Chem.*, 2013, 78, 942-949.

10 J. Templ and M. Schnurch, J. Org. Chem., 2022, 87, 4305-4315.

11 G. M. Badger, J. Chem. Soc., 1941, 535-538.

12 H. Y. Yin, X. L. Lin, S. W. Li and L. X. Shao, Org. Biomol. Chem., 2015, 13, 9012-9021.

7. NMR and HR-MS (ESI) spectra



¹H NMR (600 MHz, CDCl₃) spectrum of **2q**.



¹H NMR (600 MHz, CDCl₃) spectrum of **2u**.



¹H NMR (400 MHz, CDCl₃) spectrum of 2y.



¹H NMR (600 MHz, CDCl₃) spectrum of **3a**.



¹H NMR (600 MHz, CDCl₃) spectrum of $\mathbf{3b}$.



¹³C NMR (CDCl₃, 150 MHz) spectrum of **3b**.



HR-MS (ESI) spectra of 3b.



¹H NMR (600 MHz, CDCl₃) spectrum of **3c**.



¹H NMR (600 MHz, CDCl₃) spectrum of 3d.



¹H NMR (600 MHz, CDCl₃) spectrum of **3e**.



¹H NMR (600 MHz, CDCl₃) spectrum of **3f**.



¹³C NMR (CDCl₃, 150 MHz) spectrum of **3f**.



HR-MS (ESI) spectra of 3f.



¹H NMR (600 MHz, CDCl₃) spectrum of **3g**.



¹³C NMR (CDCl₃, 150 MHz) spectrum of **3g**.



HR-MS (ESI) spectra of 3g.



 1 H NMR (400 MHz, CDCl₃) spectrum of **3h**.



¹H NMR (600 MHz, CDCl₃) spectrum of 3i.



¹H NMR (600 MHz, CDCl₃) spectrum of **3**j.



¹³C NMR (CDCl₃, 150 MHz) spectrum of **3j**.

Single Mass Analysis Tolerance = 20.0 PPM / DBE: min = -1.5, max = 50.0 Element prediction: Off Number of isotope peaks used for i-FIT = 3



1: TOF MS ES+ 3.52e+006

HR-MS (ESI) spectra of 3j.



¹H NMR (600 MHz, CDCl₃) spectrum of **3**k.



 ^{13}C NMR (CDCl₃, 150 MHz) spectrum of **3k**.



HR-MS (ESI) spectra of 3k.



¹H NMR (600 MHz, CDCl₃) spectrum of **3**l.



¹³C NMR (CDCl₃, 150 MHz) spectrum of **3l**.



HR-MS (ESI) spectra of 3l.



¹H NMR (600 MHz, CDCl₃) spectrum of **3m**.



¹H NMR (600 MHz, CDCl₃) spectrum of 3n.



¹³C NMR (CDCl₃, 150 MHz) spectrum of **3n**.



HR-MS (ESI) spectra of **3n**.



¹H NMR (600 MHz, CDCl₃) spectrum of **30**.



 ^{13}C NMR (CDCl₃, 150 MHz) spectrum of **30**.



HR-MS (ESI) spectra of 30.



¹H NMR (600 MHz, CDCl₃) spectrum of **3p**.



¹³C NMR (CDCl₃, 150 MHz) spectrum of **3p**.



HR-MS (ESI) spectra of **3p**.



¹H NMR (400 MHz, CDCl₃) spectrum of **3q**.



¹H NMR (600 MHz, CDCl₃) spectrum of 3r.



¹H NMR (600 MHz, CDCl₃) spectrum of **3s**.



¹H NMR (600 MHz, CDCl₃) spectrum of **3t**.



¹H NMR (600 MHz, CDCl₃) spectrum of 3u.



¹H NMR (600 MHz, CDCl₃) spectrum of 3v.



¹H NMR (600 MHz, CDCl₃) spectrum of **3w**.



¹H NMR (600 MHz, CDCl₃) spectrum of 3x.



¹H NMR (600 MHz, CDCl₃) spectrum of **3y**.



¹H NMR (600 MHz, CDCl₃) spectrum of 3z.