# **Electronic Supplementary Information**

# Cu-catalyzed carboxylation of

# organoboronic acid pinacol esters with CO<sub>2</sub>

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### [A] Instrumentation and Materials

<sup>1</sup>H and <sup>13</sup>C NMR spectra were taken on a JEOL ECS400 or ECZ600 spectrometer, and chemical shifts are reported as the delta scale in ppm using an internal reference ( $\delta = 7.26$  for <sup>1</sup>H NMR, 77.16 for <sup>13</sup>C NMR, for CDCl<sub>3</sub>,  $\delta = 2.05$ for <sup>1</sup>H NMR, 29.84 for <sup>13</sup>C NMR, for acetone- $d_6$ , and  $\delta = 2.50$  for <sup>1</sup>H NMR, 39.52 for <sup>13</sup>C NMR, for DMSO- $d_6$ ). 1-(4,4,5,5-Tetramethyl-1,3,2-dioxaborolan-2-yl)-9*H*-carbazole (**1j**),<sup>1</sup> 1,4-naphthalenediboronic acid bis(pinacol) ester (**1n**),<sup>2</sup> 2,7-pyrenediboronic acid bis(pinacol) ester (**1n**),<sup>3</sup> (*E*)-styrylboronic acid pinacol ester (**3a**),<sup>4</sup> (*E*)-(4methoxystyryl)boronic acid pinacol ester (**3b**),<sup>4</sup> (phenylethynyl)boronic acid pinacol ester (**3g**),<sup>5</sup> 1,4-bis((*E*)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)vinyl)benzene (**3h**),<sup>4</sup> 1,3-bis((*E*)-2-(4,4,5,5-tetramethyl-1,3,2dioxaborolan-2-yl)vinyl)benzene (**3i**),<sup>4</sup> 1,3,5-tris((*E*)-2-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)vinyl)benzene (**3j**),<sup>4</sup> 3,4-methylenedioxyphenylboronic acid pinacol ester<sup>7</sup> were prepared according to literature procedures.

### **[B] Experimental Procedures and Compound Data**

**General procedure for the copper-catalyzed carboxylation of organoboronic acid pinacol esters with CO<sub>2</sub>.** A 30 mL stainless autoclave containing organoboronic acid pinacol ester (0.5 mmol) and (IPr)CuCl (12.2 mg, 0.025 mmol, 5 mol%) was dried in vacuo and placed in a glove box (purge type) under N<sub>2</sub> atmosphere. 'BuOK (2.81 mg, 0.025 mmol, 5 mol%), CsF (152 mg, 1.00 mmol, dried at 80 °C for 12 h in vacuo and cooled to room temperature in advance) and dry 1,4-dioxane (1.0 mL) were added, and the autoclave was closed. The sealed autoclave was taken out from the glovebox and pressurized with 2.0 MPa of CO<sub>2</sub>. The mixture was stirred at 150 °C for 24 h, and then the autoclave was cooled to room temperature, and the remaining CO<sub>2</sub> was vented slowly. The reaction mixture was acidified with 1 M HCl (aq), and the aqueous layer was extracted with EtOAc twice. The combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and evaporated under reduced pressure. Purification by silica gel column chromatography gave the corresponding carboxylic acid.

General procedure for the copper-catalyzed double carboxylation of organodiboronic acid bis(pinacol) esters with CO<sub>2</sub>. A 30 mL stainless autoclave containing organodiboronic acid bis(pinacol) ester (0.5 mmol) and (IPr)CuCl (24.4 mg, 0.050 mmol, 10 mol%) was dried in vacuo and placed in a glove box (purge type) under N<sub>2</sub> atmosphere. 'BuOK (5.61 mg, 0.050 mmol, 10 mol%), CsF (304 mg, 2.00 mmol, dried at 80 °C for 12 h in vacuo and cooled to room temperature in advance) and dry 1,4-dioxane (2.0 mL) were added, and the autoclave was closed. The sealed autoclave was taken out from the glovebox and pressurized with 2.0 MPa of CO<sub>2</sub>. The mixture was stirred at 160 °C or 170 °C for 24 h, and then the autoclave was cooled to room temperature, and the remaining CO<sub>2</sub> was vented slowly. 10% NaOH (aq) was added to the reaction mixture, and the aqueous layer was washed with Et<sub>2</sub>O once. The aqueous layer was acidified with 5% H<sub>2</sub>SO<sub>4</sub> (aq). For **21**, **20**, and **4h**–**j**, a powder precipitated was collected and washed with water and CHCl<sub>3</sub> to give the dicarboxylic acid. For **20**, the crude product was dissolved in DMSO (10 mL), and MeI (1.2 mL, 2.0 mmol) and K<sub>2</sub>CO<sub>3</sub> (276 mg, 2.00 mmol) were added. The mixture was stirred at 50 °C for 12 h. Water was added, and organic products were extracted with CHCl<sub>3</sub>. Purification by silica gel column chromatography (CHCl<sub>3</sub>) gave methyl ester **20'**. For **2m**, **2n**, **2p**, and **2q**, the product was extracted with Et<sub>2</sub>O twice, and the combined organic layer was washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and evaporated under reduced pressure. Suction filtration of the precipitate gave the dicarboxylic acid.

**Table S1** Initial screening of the reaction conditions of the Cu-catalyzed carboxylation of phenylboronic acidpinacol ester (1a) with  $CO_2$ .<sup>a</sup> A photograph of the 30 mL stainless autoclave is shown.



Entry	Cu-catalyst	Solvent	Base	<i>T</i> (°C)	Yield <sup><math>b</math></sup> (%)
$1^c$	IPrCuCl	toluene	KO <sup>t</sup> Bu	110	0
2	IPrCuCl	toluene	KO <sup>t</sup> Bu	150	13
3	IPrCuCl	THF	KO <sup>t</sup> Bu	150	26
4	IPrCuCl	1,4-dioxane	KO <sup>t</sup> Bu	150	48
5	IPrCuCl	1,4-dioxane	KO <sup>t</sup> Bu	130	21
6	IPrCuCl	1,4-dioxane	KO <sup>t</sup> Bu	170	53
7	IMesCuCl	1,4-dioxane	KO <sup>t</sup> Bu	150	5
$8^d$	CuCl + Xantphos	1,4-dioxane	KO <sup>t</sup> Bu	150	2
$9^d$	CuCl + TMEDA	1,4-dioxane	KO <sup>t</sup> Bu	150	0
$10^d$	CuCl + 1,10-phen	1,4-dioxane	KO <sup>t</sup> Bu	150	16
11	IPrCuCl	1,4-dioxane	NaO <sup>t</sup> Bu	150	trace
12	IPrCuCl	1,4-dioxane	$Cs_2CO_3$	150	28
13	IPrCuCl	1,4-dioxane	<sup><i>i</i></sup> Pr <sub>2</sub> NEt	150	0
14	IPrCuCl	1,4-dioxane	$KO^tBu + CsF^e$	150	36
15	IPrCuCl	1,4-dioxane	$\mathrm{CsF}^{e}$	150	98



<sup>*a*</sup> Reaction conditions: **1a** (0.5 mmol), Cu-catalyst (5 mol%), base (1.05 equiv), solvent (1.0 mL), CO<sub>2</sub> (2.0 MPa), 110–170 °C, 24 h, in a 30 mL stainless autoclave. <sup>*b*</sup> Yields were determined by <sup>1</sup>H NMR spectroscopy using 2-methoxynaphthalene as an internal standard. <sup>*c*</sup> 0.1 MPa CO<sub>2</sub> (balloon). <sup>*d*</sup> 5 mol% of ligand was used. <sup>*e*</sup> 2 equiv of CsF was used.

#### benzoic acid (2a)<sup>8</sup>



92% (56.1 mg, 459 μmol) white solid,  $R_f = 0.15$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 7.49 (t, J = 7.6 Hz, 2H), 7.62 (tt, J = 1.2, 7.6 Hz, 1H), 8.13 (dd, J = 1.2, 8.4 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ 128.6, 129.4, 130.4, 134.0, 172.1.

*p*-toluic acid (2b)<sup>9</sup>



77% (52.2 mg, 383 µmol) white solid,  $R_f = 0.13$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  2.44 (s, 3H), 7.28 (d, J = 7.6 Hz, 2H), 8.00 (d, J = 8.0 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  21.9, 126.7, 129.4, 130.4, 144.8, 172.3.

*p*-anisic acid (2c)<sup>10</sup>



89% (67.6 mg, 444 µmol) white solid,  $R_f = 0.18$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  3.88 (s, 3H), 6.95 (d, J = 9.2 Hz, 2H), 8.07 (d, J = 9.2 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  55.6, 114.0, 122.0, 132.5, 164.3, 171.6.

#### 4-methoxycarbonylbenzoic acid (2d)<sup>11</sup>



94% (84.3 mg, 468 µmol) white solid,  $R_f = 0.10$  (CHCl<sub>3</sub>/MeOH = 20/1), and  $R_f = 0.13$  (CHCl<sub>3</sub>/MeOH = 9/1); <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz)  $\delta$  3.88 (s, 3H), 8.06 (s, 4H), 13.37 (s, 1H); <sup>13</sup>C NMR (DMSO- $d_6$ , 100 MHz)  $\delta$  52.5, 129.4, 129.7, 133.2, 135.0, 165.6, 166.7.

## 4-trifluoromethylbenzoic acid (2e)<sup>10</sup>



61% (58.3 mg, 307 μmol) white solid,  $R_f = 0.07$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (DMSOd<sub>6</sub>, 400 MHz) δ 7.88 (d, J = 8.0 Hz, 2H), 8.13 (d, J = 8.0 Hz, 2H), 13.49 (s, 1H); <sup>13</sup>C NMR (DMSO-d<sub>6</sub>, 100 MHz) δ 123.8 (q, J = 271.5 Hz), 125.7 (q, J = 2.9 Hz), 130.3, 132.5 (q, J =31.8 Hz), 135.1, 165.9.

## *m*-anisic acid $(2f)^{10}$



76% (58.1 mg, 382 µmol) white solid,  $R_f = 0.13$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  3.87 (s, 3H), 7.16 (ddd, J = 1.2, 2.8, 8.4 Hz, 1H), 7.39 (t, J = 8.4 Hz, 1H), 7.62–7.63 (m, 1H), 7.72 (d, J = 7.6 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  55.6, 114.5, 120.6, 122.8, 129.7, 130.8, 159.7, 172.3.

## o-anisic acid $(2g)^{10}$



33% (25.0 mg, 164 µmol) white solid,  $R_f = 0.13$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  4.09 (s, 3H), 7.07 (d, J = 8.0 Hz, 1H), 7.16 (t, J = 7.2 Hz, 1H), 7.59 (dt, J = 1.6, 7.2 Hz, 1H), 8.20 (dd, J = 2.0, 8.0 Hz, 1H), 10.74 (s, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  56.8, 111.8, 117.7, 122.3, 133.9, 135.2, 158.2, 165.7.

#### 1-naphthoic acid (2h)<sup>10</sup>



68% (58.4 mg, 339 μmol) white solid,  $R_f = 0.13$  (CHCl<sub>3</sub>/MeOH = 20/1), and  $R_f = 0.05$  (hexane/EtOAc = 5/1); <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz) δ 7.57–7.67 (m, 3H), 8.02 (d, J = 7.2 Hz, 1H), 8.14–8.17 (m, 2H), 8.86 (d, J = 8.8 Hz, 1H), 13.14 (s, 1H); <sup>13</sup>C NMR (DMSO- $d_6$ , 100 MHz) δ 124.9, 125.5, 126.2, 127.6, 127.7, 128.6, 129.9, 130.7, 133.0, 133.5, 168.7.

## thiophene-2-carboxylic acid (2i)<sup>10</sup>



53% (34.1 mg, 266 µmol) white solid,  $R_f = 0.05$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.15 (dd, J = 3.8, 4.4 Hz, 1H), 7.65 (dd, J = 1.2, 5.2 Hz, 1H), 7.90 (dd, J = 1.2, 3.6 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  128.3, 133.1, 134.2, 135.3, 168.0.

## carbazole-1-carboxylic acid (2j)<sup>12</sup>



51% (53.8 mg, 255 μmol) white solid,  $R_f = 0.30$  (CHCl<sub>3</sub>/MeOH = 9/1); <sup>1</sup>H NMR (DMSOd<sub>6</sub>, 400 MHz) δ 7.18–7.27 (m, 2H), 7.42 (t, J = 6.8 Hz, 1H), 7.74 (d, J = 8.8 Hz, 1H), 8.00 (d, J = 7.2 Hz, 1H), 8.16 (d, J = 8.0 Hz, 1H), 8.39 (d, J = 7.2 Hz, 1H), 11.32 (s, 1H), 13.17 (s, 1H); <sup>13</sup>C NMR (DMSO-d<sub>6</sub>, 100 MHz) δ 112.3, 112.8, 118.0, 119.3, 120.2, 121.7, 124.2, 125.2, 126.1, 127.5, 139.1, 140.3, 168.0.

## carbazole-3-carboxylic acid (2k)<sup>13</sup>



45%, (47.8 mg, 226 μmol) white solid,  $R_f$  = 0.25 (CHCl<sub>3</sub>/MeOH = 9/1); <sup>1</sup>H NMR (DMSOd<sub>6</sub>, 400 MHz) δ 7.22 (t, *J* = 7.6 Hz, 1H), 7.44 (t, *J* = 7.2 Hz, 1H), 7.54 (d, *J* = 9.2 Hz, 2H), 8.02 (dd, *J* = 1.6, 8.4 Hz, 1H), 8.24 (d, *J* = 8.0 Hz, 1H), 8.78 (s, 1H), 11.66 (s, 1H), 12.58 (s, 1H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 150 MHz) δ 110.6, 111.4, 119.5, 120.6, 121.0, 122.2, 122.49, 122.50, 126.3, 127.0, 140.3, 142.4, 168.1.

terephthalic acid (2l)<sup>11</sup>



65% (54.1 mg, 326 μmol) white solid; <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz) δ 8.04 (s, 4H), 13.29 (s, 2H); <sup>13</sup>C NMR (DMSO- $d_6$ , 100 MHz) δ 129.5, 134.5, 166.7.

isophthalic acid (2m)<sup>14</sup>



51% (42.7 mg, 257 μmol) white solid; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz) δ 7.64 (t, J = 8.0 Hz, 1H), 8.16 (dd, J = 2.0, 8.0 Hz, 2H), 8.48 (s, 1H), 13.25 (s, 2H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz) δ 129.2, 130.1, 131.3, 133.5, 166.7.

## 1,4-naphthalenedicarboxylic acid (2n)<sup>15</sup>



45% (48.7 mg, 225 μmol) white solid; <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz) δ 7.70 (dd, J = 3.6, 6.4 Hz, 2H), 8.10 (s, 2H), 8.78 (dd, J = 3.6, 6.8 Hz, 2H), 13.51 (s, 2H); <sup>13</sup>C NMR (DMSO- $d_6$ , 100 MHz) δ 125.9, 127.6, 127.9, 130.8, 132.3, 168.5.

## dimethyl 2,7-pyrenedicarboxylate (20')<sup>16</sup>



42% (66.6 mg, 209 µmol) light yellow solid; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz)  $\delta$  4.09 (s, 6H), 8.13 (s, 4H), 8.82 (s, 4H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  52.6, 126.3, 128.6, 131.7, 167.5.

## 2,5-thiophenedicarboxylic acid (2p)<sup>17</sup>



46% (40.0 mg, 232 µmol) white solid; <sup>1</sup>H NMR (DMSO- $d_6$ , 400 MHz)  $\delta$  7.71 (s, 2H), 13.59 (s, 2H); <sup>13</sup>C NMR (DMSO- $d_6$ , 100 MHz)  $\delta$  133.3, 139.8, 162.5.

#### 2,5-pyrroledicarboxylic acid (2q)<sup>18</sup>



6% (5.0 mg, 32.2 μmol) gray solid; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz) δ 6.74 (d, J = 1.2 Hz, 2H), 12.20 (s, 1H) 12.72 (s, 2H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz) δ 115.1, 127.3, 161.3.

## (E)-cinnamic acid $(4a)^{19}$



90% (69.5 mg, 469 µmol) white solid,  $R_f = 0.15$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  6.47 (d, J = 15.6 Hz, 1H), 7.41–7.42 (m, 3H), 7.55–7.57 (m, 2H), 7.79 (d, J = 16.0 Hz, 1H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  117.4, 128.5, 129.1, 130.9, 134.2, 147.3, 172.6.

## (E)-4-methoxycinnamic acid (4b)<sup>20</sup>



84% (75.9 mg, 426 μmol) white solid,  $R_f$  = 0.07 (CHCl<sub>3</sub>/MeOH = 30/1), and  $R_f$  = 0.25 (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 3.85 (s, 3H), 6.32 (d, *J* = 15.6 Hz, 1H), 6.92 (d, *J* = 9.2 Hz, 2H), 7.51 (d, *J* = 9.2 Hz, 2H), 7.74 (d, *J* = 16.0 Hz, 1H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz) δ 55.3, 114.6, 126.8, 130.0, 143.8, 161.0, 167.8.

#### (E)-4-bromocinnamic acid (4c)



54% (61.7 mg, 272 µmol) white solid,  $R_f = 0.05$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (acetone- $d_6$ , 400 MHz)  $\delta$  6.57 (d, J = 16.0 Hz, 1H), 7.61–7.67 (m, 5H), 10.86 (s, 1H); <sup>13</sup>C NMR (acetone- $d_6$ , 100 MHz)  $\delta$  120.2, 124.7, 130.8, 132.9, 134.8, 144.0, 167.4; HRMS (ESI) m/z: [M–H]<sup>–</sup> Calcd for C<sub>9</sub>H<sub>6</sub>O<sub>2</sub>Br 224.9557; Found 224.9565.

(*E*)-4-trifluoromethylcinnamic acid  $(4d)^{21}$ 



61% (36.1 mg, 167 μmol) from **3d** (250 μmol) white solid,  $R_f = 0.07$  (CHCl<sub>3</sub>/MeOH = 20/1), and  $R_f = 0.10$  (EtOAc); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz) δ 6.68 (d, J = 16.0 Hz, 1H), 7.66 (d, J = 16.0 Hz, 1H), 7.76 (d, J = 8.0 Hz, 2H), 7.92 (d, J = 8.4 Hz, 2H), 12.62 (s, 1H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz) δ 122.2, 124.1 (q, J = 271.1 Hz), 125.7 (q, J = 3.8 Hz), 128.9, 129.8 (q, J = 31.8 Hz), 138.3, 142.1, 167.2.

2-phenylacetic acid (4e)<sup>19</sup>

82% (55.8 mg, 410 μmol) white solid,  $R_f$  = 0.20 (CHCl<sub>3</sub>/MeOH = 9/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 3.66 (s, 2H), 7.38–7.28 (m, 5H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ 127.5, 128.7, 129.5, 133.4, 178.1.

octanoic acid (4f)<sup>22</sup>

28% (18.5 mg, 128 μmol) colorless liquid; <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz) δ 0.88 (t, J = 6.8 Hz, 3H), 1.16–1.38 (m, 8H), 1.56–1.70 (m, 2H), 2.35 (t, J = 7.2 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz) δ 14.2, 22.7, 24.8, 29.1, 29.2, 31.8, 34.2, 180.3.

3-phenylpropiolic acid (4g)<sup>19</sup>



36% (26.7 mg, 183 µmol) white solid,  $R_f = 0.05$  (CHCl<sub>3</sub>/MeOH = 20/1), and  $R_f = 0.08$  (CHCl<sub>3</sub>/MeOH = 9/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta$  7.40 (t, J = 7.6 Hz, 2H), 7.49 (t, J = 7.2 Hz, 1H), 7.62 (d, J = 6.8 Hz, 2H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 100 MHz)  $\delta$  80.4, 89.5, 119.2, 128.8, 131.3, 133.5, 158.5.

1,4-benzenediacrylic acid (4h)<sup>23</sup>



62% (64.1 mg, 294 μmol) light brown solid; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz) δ6.60 (d, J = 16.0 Hz, 2H), 7.60 (d, J = 15.6 Hz, 2H), 7.73 (s, 4H) 12.46 (s, 2H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 150 MHz) δ 120.2, 128.7, 135.9, 143.1, 167.5.

#### 1,3-benzenediacrylic acid (4i)



73% (79.7 mg, 365 μmol) light yellow solid; <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 400 MHz)  $\delta$  6.67 (d, *J* = 16.0 Hz, 2H), 7.46 (t, *J* = 8.0 Hz, 1H), 7.60 (d, *J* = 16.0 Hz, 2H), 7.72 (dd, *J* = 1.6, 8.0 Hz, 2H), 8.08 (s, 1H), 12.44 (s, 2H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 100 MHz)  $\delta$  120.2, 127.7, 129.5, 129.9, 135.0, 143.3, 167.6; HRMS (ESI) *m/z*: [*M*-H]<sup>-</sup> Calcd for C<sub>12</sub>H<sub>10</sub>O<sub>4</sub> 217.0506; Found 217.0515.

#### 1,3,5-benzenetriacrylic acid (4j)



piperonylic acid



 $d_6$ , 400 MHz)  $\delta$  6.78 (d, J = 16.0 Hz, 3H), 7.60 (d, J = 16.4 Hz, 3H), 8.08 (s, 3H), 12.35 (s, 3H); <sup>13</sup>C NMR could not detect peaks due to very low solubility; HRMS (ESI) m/z:  $[M-2H]^{2-}$  Calcd for C<sub>15</sub>H<sub>10</sub>O<sub>6</sub> 143.0244; Found 143.0241.

40% (19.3 mg, 67.0 µmol) from 3j (167 µmol) brown solid; <sup>1</sup>H NMR (DMSO-

80% (66.2 mg, 398 µmol) white solid,  $R_f = 0.11$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (DMSO-*d*<sub>6</sub>, 600 MHz)  $\delta$  6.12 (s, 2H), 7.00 (d, J = 8.4 Hz, 1H), 7.36 (d, J = 1.8 Hz, 1H), 7.54 (dd, J = 1.8, 8.4 Hz, 1H), 12.77 (s, 1H); <sup>13</sup>C NMR (DMSO-*d*<sub>6</sub>, 150 MHz)  $\delta$  102.0, 108.1, 108.8, 124.6, 125.0, 147.5, 151.2, 166.6; HRMS (ESI) *m/z*: [*M*–H]<sup>-</sup> Calcd for C<sub>8</sub>H<sub>5</sub>O<sub>4</sub> 165.0193; Found 165.0187.

felbinac



45% (35.5 mg, 166 μmol) light yellow solid,  $R_f = 0.11$  (CHCl<sub>3</sub>/MeOH = 20/1); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz) δ 3.71 (s, 2H), 7.35–7.38 (m, 3H), 7.42–7.45 (m, 2H), 7.56–7.59 (m, 4H); <sup>13</sup>C NMR (CDCl<sub>3</sub>, 150 MHz) δ 40.6, 127.2, 127.5, 127.6, 128.9, 129.9, 132.4, 140.5, 140.8, 176.5; HRMS (ESI) *m/z*: [*M*–H]<sup>–</sup> Calcd for C<sub>14</sub>H<sub>11</sub>O<sub>2</sub> 211.0765; Found 211.0760.



**Scheme S1** (a) A proposed reaction mechanism for the Cu-catalyzed carboxylation of boronic esters.<sup>24</sup> (b) Stoichiometric transmetalation of (IPr)CuO'Bu with PhBpin.<sup>25</sup> IPr = 1,3-bis(2,6-diisopropylphenyl)imidazol-2-ylidene.

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#### [D] Comparison with the Reported Reaction Conditions

1a (0.73 mmol)

11 (0.37 mmol)

(0.4 MPa)

We conducted the Cu-catalyzed carboxylation under Bayer's reaction condition in a stainless autoclave. Unfortunately, we could not reproduce the high yields they achieved by using a specialized glassware (Scheme S2). In addition, double carboxylation did not take place at all under the reaction condition (Scheme S3).



41%<sup>a</sup> (in a stainless autoclave) 81%<sup>b,c</sup> (in a specialized glassware)

**2l** <1%

2a

<sup>a</sup> NMR yield. <sup>b</sup> Isolated yield. <sup>c</sup> Reference data: Chem. Eur. J., 2020, 26, 6064.

then HCI (aq.)

#### Scheme S2



Scheme S3

then HCI (aq.)

(0.4 MPa)



400 MHz <sup>1</sup>H NMR spectrum of **2a** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2a** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2b** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2b** in CDCl<sub>3</sub>.



400 MHz  $^{1}$ H NMR spectrum of **2c** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2c** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2d** in DMSO-*d*<sub>6</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2d** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2e** in DMSO-*d*<sub>6</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2e** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2f** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2f** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2g** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2g** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2h** in DMSO-*d*<sub>6</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2h** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2i** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2i** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2j** in DMSO-*d*<sub>6</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2j** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of 2k in DMSO- $d_6$ .



150 MHz <sup>13</sup>C NMR spectrum of **2k** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **2l** in DMSO-*d*<sub>6</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **2l** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of 2m in DMSO- $d_6$ .



100 MHz <sup>13</sup>C NMR spectrum of 2m in DMSO- $d_6$ .



400 MHz <sup>1</sup>H NMR spectrum of 2n in DMSO- $d_6$ .



100 MHz <sup>13</sup>C NMR spectrum of **2n** in DMSO-*d*<sub>6</sub>.



600 MHz <sup>1</sup>H NMR spectrum of **20'** in CDCl<sub>3</sub>.



150 MHz <sup>13</sup>C NMR spectrum of **20'** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of 2p in DMSO- $d_6$ .



100 MHz <sup>13</sup>C NMR spectrum of **2p** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of 2q in DMSO- $d_6$ .



100 MHz <sup>13</sup>C NMR spectrum of **2q** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of 4a in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of 4a in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4b** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **4b** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4c** in acetone-*d*<sub>6</sub>.



100 MHz  $^{13}$ C NMR spectrum of **4c** in acetone-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4d** in DMSO-*d*<sub>6</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **4d** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4e** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **4e** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4f** in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **4f** in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of 4g in CDCl<sub>3</sub>.



100 MHz <sup>13</sup>C NMR spectrum of 4g in CDCl<sub>3</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4h** in DMSO-*d*<sub>6</sub>.



150 MHz  $^{13}$ C NMR spectrum of **4h** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4i** in DMSO-*d*<sub>6</sub>.



100 MHz <sup>13</sup>C NMR spectrum of **4i** in DMSO-*d*<sub>6</sub>.



400 MHz <sup>1</sup>H NMR spectrum of **4j** in DMSO-*d*<sub>6</sub>.



600 MHz <sup>1</sup>H NMR spectrum of piperonylic acid in DMSO-*d*<sub>6</sub>.



150 MHz <sup>13</sup>C NMR spectrum of piperonylic acid in DMSO-*d*<sub>6</sub>.



600 MHz <sup>1</sup>H NMR spectrum of felbinac in CDCl<sub>3</sub>.



150 MHz <sup>13</sup>C NMR spectrum of felbinac in CDCl<sub>3</sub>.