Supporting information

ReACT (Redox-Activated Chemical Tagging) chemistry enables direct derivatization and fluorescent detection of *S*-adenosyl-*L*-homocysteine (SAH)

Bohong Lin,[†] Lingling Xiang,[†] Zhijun Yuan, Qi Hou, Yaoping Ruan, Jing Zhang*

Artemisinin Research Center and The First Affiliated Hospital, Guangzhou University of Chinese Medicine, 12 Jichang Road, Guangzhou 510405, China.

Email: jingzhang@gzucm.edu.cn

[†] These authors contributed equally to this work

Table of Contents

Figure S1. ¹ H NMR comparison of probe NapOx, SAH, and their reaction mixture
Figure S2. ¹ H NMR analysis of the reaction mixtures of NapOx with different biomoleculesS2
Figure S3. ¹ H NMR analysis of the reaction mixtures of NapOx with DTT and methionine
under neutral and acidic condition
1. Methods and materials
2. Compound Synthesis
2.1 Synthesis of oxaziridines 1a-1g
2.2 Synthesis of compound NapOx
2.3 Synthesis of compounds 2a-2g
3. Fluorescence measurement
3.1 SAH concentration measurement
3.2 Fluorescence profiling of NapOx reaction with thiol or thioether compound
3.3 NapOx selectivity under neutral or acidic condition
4. HPLC experiment
5. NMR spectra of compounds
6. HRMS spectra of compounds
7. Reference



Figure S1. ¹H NMR comparison of probe **NapOx** (top), SAH (middle), and their reaction mixture (bottom). Reaction condition: **NapOx** (2.0 equiv), SAH (1.0 equiv), D₂O/CD₃CN (5:3), r.t., 20 min.



Figure S2. ¹H NMR analysis of the reaction mixtures of **NapOx** with different biomolecules. Reaction condition: **NapOx** (1.0 equiv), biomolecule (2.0 equiv), CD₃CN/D₂O (3:1), 22 °C, 10 min. For the reaction of methionine, CD₃CN/D₂O (7:3) was applied since methionine showed poor solubility in CD₃CN/D₂O (3:1).



Figure S3. ¹H NMR analysis of the reaction mixtures of **NapOx** with DTT and methionine under neutral and acidic condition. For DTT reaction under neutral condition: **NapOx** (1.0 equiv), DTT (2.0 equiv), CD₃CN/D₂O (3:1), 22 °C, 10 min. For acidic reaction condition, 1 mM deuterated HCl solution was used to replace D₂O. For methionine reaction, CD₃CN/D₂O (7:3) or CD₃CN/1 mM deuterated HCl solution (7:3) were used to replace CD₃CN/D₂O (3:1) or CD₃CN/1 mM deuterated HCl solution (3:1).

1. Methods and materials

All chemicals were used as received unless otherwise stated. ¹H NMR and ¹³C NMR spectra were collected on Bruker AVANCE III HD400 spectrometer. Proton chemical shifts of NMR spectra were calibrated with TMS as internals reference. HR-MS spectral data were recorded on Aglient 7250 and JEOL-JMS-T100LPAccuTOF devices. Analytical thin-layer chromatography (TLC) analysis was performed on TLC silica gel plates (0.2 ± 0.03 mm) and visualized with ultraviolet light (254 nm) to monitor reaction progression. The 96-well plate (black, flat, not treated) was purchased from Corning® (USA). Fluorescence analysis was performed on Thermo ScientificTM VarioskanTM LUX microplate reader. The analytic HPLC was run on Agilent 1260-workstation equipped with a Luna C18 column, 250 x 4.6 mm, particle size 5 µm, pore size 110 Å (Phenomenex) with a flow rate of 1 mL/min. Solvent A: H₂O with 0.1% trifluoroacetic acid; Solvent B: MeCN with 0.1% trifluoroacetic acid.

2. Compound Synthesis

2.1 Synthesis of oxaziridines 1a-1g

The compound **1a** was synthesized following reported procedure.^[1] To a mixture of ethylurea (0.62 g, 5.84 mmol) in anhydrous THF (14 mL) was added benzaldehyde (0.8 mL, 8.16 mmol) and Ti(*Oi*Pr)₄ (2.20 mL, 7.48 mmol) sequentially under N₂. The white suspension was allowed to stir overnight at r.t. The reaction mixture was dried over evaporator, and the obtained crude was dissolved in DCM (5 mL). Then this mixture was added slowly to a suspension of mCPBA (4.60 g, 75% purity, 20 mmol) in saturated K₂CO₃ aqueous solution (20 mL) and DCM (20 mL), and the resulted mixture was stirred at r.t. for 6 h. After that, the mixture was diluted with 20 mL water, and extracted with DCM for three times. The combined organic was washed with brine, and dried over anhydrous Na₂SO₄. The crude mixture was filtered, and the filtrate was concentrated under vacuum. The obtained residue was purified over silica gel chromatography (PE/EA = 4:1) to yield the desired compound **1a** as a white solid (150 mg, 13% yield for two steps). ¹H NMR (400 MHz, CDCl₃) δ 7.46 (m, 5H), 6.04 (br, 1H), 5.00 (s, 1H), 3.33 (m, 2H), 1.22 (t, *J* = 7.2 Hz, 3H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 162.1, 132.4, 131.0, 128.6, 128.0, 79.3, 35.4, 15.2; The data are consistent with those reported in previous literature.^[1]



1b was synthesized with the same procedure as compound **1a**. The obtained residue was purified over Al₂O₃ column chromatography (DCM) to yield the desired compound **1b** as a yellow liquid (100 mg, 17% yield for two steps).¹H NMR (400 MHz, CDCl₃) δ 7.41 (m, 5H), 6.13 (br, 1H), 5.00 (s, 1H), 2.87 (d, *J* = 4.8 Hz, 3H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 162.9, 132.4, 131.0, 128.6, 128.0, 79.4, 27.0; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₉H₁₁N₂O₂ 179.0815; found 179.0815.



1c was synthesized with the same procedure as compound **1a**. The obtained residue was purified over Al₂O₃ column chromatography (PE/ EA = 6:1) to yield the desired compound **1c** as a white solid (102 mg, 15% yield for two steps). ¹H NMR (400 MHz, CDCl₃) δ 7.45 (m, 5H), 6.08 (br, 1H), 5.00 (s, 1H), 3.25 (m, 2H), 1.59 (m, 2H), 0.96 (t, *J* = 7.6 Hz, 3H). Please note that the integration of the hydrogens around δ 1.59 ppm is ca. 3.7 in supplementary NMR figure, which

is higher than 2. This is due to interference of extra H₂O signal at ca. 1.6 ppm. ${}^{13}C{}^{1}H$ NMR (101 MHz, CDCl₃) δ 162.2, 132.4, 131.0, 128.6, 128.0, 79.4, 42.1, 22.7, 11.2; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₁H₁₅N₂O₂ 207.1128; found 207.1124.

Synthesis of 1-(Prop-2-yn-1-yl)urea^[1]

$$\mathbb{NH}_{2} \xrightarrow{1) \text{ KOCN, HCI, 60°C}} \mathbb{NH}_{2}$$
2) silical gel, MeOH, r,t.

This compound was synthesized following reported procedure^[1]. To a solution of propargylamine (1.6 mL, 25 mmol) in aqueous HCl solution (1.0 mol, 25 mL) was added KOCN (8 g, 100 mmol). After being heated at 60 °C for 40 h, the mixture was placed in the -20 °C refrigerator to obtain a white precipitate. After vacuum filtration, the obtained solid was dissolved in MeOH (75 mL) and stirred with silica gel (10 g) for 6 h. The mixture was then filtered, and the residue was washed with MeOH. The combined filtrate was concentrated under vacuum to attain the desired urea as a white solid without further purification (1.9 g, 78%). ¹H NMR (400 MHz, *d*₆-DMSO) δ 6.29 (t, *J* = 6.0 Hz, 1H), 5.60 (s, 2H), 3.75 (dd, *J* = 5.6, 2.4 Hz, 2H), 3.03 (t, *J* = 2.4 Hz, 1H). ¹³C{¹H} NMR (101 MHz, *d*₆-DMSO) δ 158.6, 83.0, 72.9, 29.1; The data are consistent with those reported in previous literature.^[1]



1d was synthesized with the same procedure as compound **1a**. The obtained residue was purified over Al₂O₃ column chromatography (DCM) to yield the desired compound **1d** as a yellow liquid (111 mg, 17% yield for two steps). ¹H NMR (400 MHz, CDCl₃) δ 7.45 (m, 5H), 6.37 (br, 1H), 5.03 (s, 1H), 4.07 (m, 2H), 2.30 (t, *J* = 2.4 Hz, 1H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 162.0, 132.0, 131.1, 128.7, 128.0, 79.5, 78.3, 72.6, 30.2; The data are consistent with those reported in previous literature.^[1]



1e was synthesized with the same procedure as compound **1a**. The obtained residue was purified over Al₂O₃ column chromatography (EA) to yield the desired compound **1e** as a white solid (40 mg, 4% yield for two steps). ¹H NMR (400 MHz, CDCl₃) δ 7.45 (m, 5H), 6.52 (br, 1H), 5.04 (s,

1H), 3.77 (q, J = 5.2 Hz, 2H), 3.46 (m, 2H), 2.19 (t, J = 5.2 Hz, 1H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 163.1, 132.2, 131.1, 128.7, 128.0, 79.5, 61.5, 42.7; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₀H₁₃N₂O₃ 209.0926; found 209.0907.



If was synthesized with the same procedure as compound 1a. The obtained residue was purified over Al₂O₃ column chromatography (PE/ EA = 6:1) to yield the desired compound 1f as a white solid (29 mg, 6% yield for two steps). ¹H NMR (400 MHz, CDCl₃) δ 7.44 (m, 5H), 5.96 (br, 1H), 4.98 (s, 1H), 3.66 (m, 1H), 1.96 (m, 2H), 1.74 (m, 2H), 1.63 (m, 1H), 1.37 (m, 2H), 1.21 (m, 3H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 161.3, 132.5, 131.0, 128.6, 128.0, 79.4, 49.4, 32.9, 32.8, 25.3, 24.7; The data are consistent with those reported in previous literature.^[2]



1g was synthesized with the same procedure as compound **1a**. The obtained residue was purified over Al₂O₃ column chromatography (PE/ EA = 6:1) to yield the desired compound **1g** as a white solid (35 mg, 4% yield for two steps). ¹H NMR (400 MHz, CDCl₃) δ 7.40 (m, 10H), 6.36 (br, 1H), 5.04 (s, 1H), 4.47 (m, 2H). ¹³C{¹H} NMR (101 MHz, CDCl₃) δ 162.2, 137.0, 132.3, 131.1, 128.9, 128.7, 128.0, 127.8, 79.5, 44.4; The data are consistent with those reported in previous literature.^[2]

2.2 Synthesis of compound NapOx



The compound **NapOx** was synthesized with the same procedure as compound **1a** on page S3. The obtained crude was purified over Al₂O₃ chromatography with DCM as eluent to yield crude as a light yellow solid, which was further recrystallized from PE/DCM to provide the desired product **NapOx** as a white solid (140 mg, 24% yield for two steps). ¹H NMR (400 MHz, CDCl₃)

δ 7.93 (s, 1H), 7.76 (d, J = 8.8 Hz, 2H), 7.43 (dd, J = 8.8, 1.6 Hz, 1H), 7.16 (m, 2H), 6.09 (br, 1H), 5.13 (s, 1H), 3.93 (s, 3H), 3.35 (m, 2H), 1.23 (t, J = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 162.2, 158.8, 136.1, 129.8, 129.0, 128.1, 127.5, 127.4, 124.1, 119.5, 105.8, 79.9, 55.4, 35.4, 14.8; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₅H₁₇N₂O₃ 273.1234; found 273.1230.

2.3 Synthesis of compounds 2a-2g



General synthetic procedure for compounds 2a-2g: A solution of oxaziridine **1a-1g** (0.26 mmol, 2.0 equiv), SAH (0.13 mmol, 1.0 equiv) in ca. 20% MeCN aqueous solution was stirred at room temperature for approximately 1 h. The reaction mixture was washed with CH₂Cl₂ for three times. Then the aqueous layer was concentrated under vacuum, and the obtained crude was purified by column chromatography over reverse silica gel to afford the target compound.



Compound 2a was synthesized using the procedure described above. The obtained residue was purified over reverse phase silica gel column chromatography (10% MeOH in H₂O) to yield the desired compound **2a** (50 mg, 83% yield) as a white solid. ¹H NMR (400 MHz, D₂O) δ [8.12] 7.98 (s, 1H), [8.06] 7.96 (s, 1H), [5.89] 5.86 (d, *J* = 5.2 Hz, 1H), 4.73 (q, *J* = 5.2 Hz, 1H), 4.38 (m, 2H), 3.70 (q, *J* = 6.8 Hz, 1H), 3.47 (m, 2H), 3.12 (m, 2H), [2.96] 2.82 (q, *J* = 7.2 Hz, 2H), 2.09 (m, 2H), [0.91] 0.78 (t, *J* = 6.8 Hz, 3H). ¹³C NMR (101 MHz, D₂O) δ [173.1] 173.0, [167.7] 161.2, [155.2] 155.2, [152.6] 152.6, [148.6] 148.4, [140.2] 140.1, [118.6] 118.6, [88.3] 88.1, 79.0, 77.8, [72.9] 72.7, [72.8] 72.5, [53.3] 52.9, 49.5, [43.4] 43.0, [24.6] 24.5, [14.4] 14.3; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₇H₂₇N₈O₆S 471.1769; found 471.1780.



Compound 2b was synthesized using the procedure described above. The obtained residue was purified over reverse phase silica gel column chromatography (10% MeOH in H₂O) to yield the desired compound **2b** (30 mg, 51% yield) as a white solid. ¹H NMR (400 MHz, D₂O) δ [8.14] 8.03 (s, 1H), [8.09] 8.02 (s, 1H), [5.92] 5.90 (d, *J* = 5.2 Hz, 1H), 4.77 (m, 1H), 4.37 (m, 2H), 3.69 (q, *J* = 6.4 Hz, 1H), 3.47 (m, 2H), 3.08 (m, 2H), [2.54] 2.39 (s, 3H), 2.09 (m, 2H). ¹³C NMR (101 MHz, D₂O) δ 173.3, 168.4, [155.3] 155.3, [152.7] 152.7, [148.7] 148.5, [140.3] 140.1, [118.8] 118.7, [88.4] 88.1, 78.9, 77.8, [72.8] 72.6, [72.8], 72.5, [53.3] 52.9, 49.6, [43.5] 43.1, [24.7] 24.6; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₆H₂₅N₈O₆S 457.1612; found 457.1622.



Compound 2c was synthesized using the procedure described above. The obtained residue was purified over reverse phase silica gel column chromatography (30% MeOH in H₂O) to yield the desired compound **2c** (28 mg, 45% yield) as a white solid. ¹H NMR (400 MHz, D₂O) δ [8.19] 8.09 (s, 1H), [8.13] 8.08 (s, 1H), [5.94] 5.92 (d, *J* = 5.2 Hz, 1H), 4.78 (m, 1H), 4.38 (m, 2H), 3.65 (t, *J* = 6.4 Hz, 1H), 3.48 (m, 2H), 3.06 (m, 2H), [2.89] 2.74 (t, *J* = 7.2 Hz, 2H), 2.08 (m, 2H), [1.29] 1.11 (q, *J* = 7.2 Hz, 2H), [0.71] 0.58 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, D₂O) δ 173.9, 167.8, 155.5, [152.8] 152.8, [148.8] 148.7, [140.4] 140.2, [118.9] 118.8, [88.3] 88.0, 79.3, 77.9, [72.9] 72.7, [72.8] 72.5, [53.4] 53.1, 49.5, [43.4] 43.1, [25.0] 24.9, [22.7] 22.6, [10.5] 10.3; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₈H₂₉N₈O₆S 485.1925; found 485.1936.



2d

Compound 2d was synthesized using the procedure described above. The obtained residue was purified over reverse phase silica gel column chromatography (10% MeOH in H₂O) to yield the desired compound **2d** (33 mg, 53% yield) as a white solid. ¹H NMR (400 MHz, D₂O) δ [8.17] 8.07 (s, 1H), [8.12] 8.06 (s, 1H), [5.94] 5.91 (d, *J* = 5.2 Hz, 1H), 4.77 (q, *J* = 5.2 Hz, 1H), 4.40 (m, 2H), 3.72 (m, 2H), 3.61 (s, 1H), 3.50 (m, 2H), 3.13 (m, 2H), [2.45] 2.32 (t, *J* = 2.4 Hz, 1H), 2.12 (m, 2H). ¹³C NMR (101 MHz, D₂O) δ 173.2, 167.1, [155.4] 155.4, [152.8] 152.7, [148.7] 148.6, [140.4] 140.2, [118.9] 118.8, [88.4] 88.1, [81.4] 81.3, 79.0, 77.8, [72.8] 72.8, [72.8] 72.5, [71.1] 70.9, [53.2] 52.9, 49.4, [43.3] 43.0, [24.7] 24.6; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₈H₂₅N₈O₆S 481.1612; found 481.1643.



Compound 2e was synthesized using the procedure described above. The obtained residue was purified over reverse phase silica gel column chromatography (5% MeOH in H₂O) to yield the desired compound **2e** (36 mg, 57% yield) as a white solid. ¹H NMR (400 MHz, D₂O) δ [8.14] 8.02 (s, 1H), [8.09] 8.01 (s, 1H), [5.92] 5.90 (d, *J* = 4.8 Hz, 1H), 4.76 (t, *J* = 5.2 Hz, 1H), 4.38 (m, 2H), 3.71 (q, *J* = 6.8 Hz, 1H), 3.48 (m, 4H), 3.13 (m, 2H), 3.02 (m, 2H), 2.11 (m, 2H). ¹³C NMR (101 MHz, D₂O) δ 173.1, 167.5, [155.3] 155.2, [152.7] 152.6, [148.6] 148.4, [140.3] 140.1, [118.7] 118.6, [88.4] 88.1, 78.8, 77.8, [72.8] 72.6, 72.7, [60.8] 60.8, [53.2] 52.9, [49.6] 49.3, [43.4] 43.0, [24.6] 24.5; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₁₇H₂₇N₈O₇S 487.1718; found 487.1739.



Compound 2f was synthesized using the procedure described above. The obtained residue was purified over reverse phase silica gel column chromatography (40% MeOH in H₂O) to yield the desired compound **2f** (42 mg, 69% yield) as a light yellow solid. ¹H NMR (400 MHz, D₂O) δ [8.15] 8.00 (s, 1H), [8.07] 7.97 (s, 1H), [5.89] 5.86 (d, *J* = 4.8 Hz, 1H), 4.72 (br, 1H), 4.36 (m, 2H), 3.72 (t, *J* = 6.4 Hz, 1H), 3.48 (m, 2H), 3.15 (m, 2H), 3.01 (m,1H), 2.12 (m,2H), [1.45] 0.97 (m, 10H). ¹³C NMR (101 MHz, D₂O) δ [173.0] 173.0, [166.9] 166.6, [155.3] 155.2, [152.7] 152.6, [148.6] 148.4, [140.2] 140.1, [118.6] 118.5, [88.2] 87.9, 79.5, 77.8, [72.9] 72.5, 72.9, [53.3] 52.9, [50.0] 49.9, 49.3, [43.2] 43.0, [32.8] 32.8, [32.8] 32.6, [25.0] 24.9, [24.6] 24.5, 24.4; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₁H₃₃N₈O₆S 525.2238; found 525.2248.



Compound 2g was synthesized using the procedure described above. The obtained residue was purified over reverse phase silica gel column chromatography (30% MeOH in H₂O) to yield the desired compound **2g** (30 mg, 44% yield) as a light yellow solid. ¹H NMR (400 MHz, D₂O) δ [8.01] 7.96 (s, 1H), [8.99] 7.95 (s, 1H), [7.08] 6.89 (m,5H), 5.80 (t, *J* = 4.4 Hz, 1H), 4.64 (t, *J* = 5.2 Hz, 1H), [4.48] 4.22 (t, *J* = 5.2 Hz, 1H), [4.42] 4.29 (m, 1H), 4.04 (m, 2H), 3.71 (q, *J* = 6.4 Hz, 1H), 3.46 (m, 2H), 3.10 (m, 2H), 2.12 (m, 2H). ¹³C NMR (101 MHz, D₂O) δ [172.9] 172.9, 167.3, 155.2, [152.5] 152.5, [148.3] 148.2, [140.1] 139.5, [139.6] 139.5, [129.1] 127.1, [128.6] 126.7, [128.4] 128.1, [126.8] 126.6, 126.5, 118.7, [88.5] 88.2, 79.3, 77.8, [73.3] 72.4, 72.9, [53.2] 52.9, [49.3] 48.8, 43.3, [24.6] 24.5; HRMS (ESI) m/z: [M + H]⁺ Calcd for C₂₂H₂₉N₈O₆S 533.1925; found 533.1943.

3. Fluorescence measurement

3.1 SAH concentration measurement

A solution of **NapOx** in IPA (20 μ L, 100 μ M) and SAH in ddH₂O (20 μ L, 0 - 2 mM, 2-fold dilution) was added to a mixture of IPA/ddH₂O (7:1, 160 μ L) in 96-well plate, with a final concentration of 10 μ M for **NapOx** and 0-200 μ M for SAH. The mixture was incubated at 37 °C for 30 min. Then it was directly used for fluorescence measurement.

3.2 Fluorescence profiling of NapOx reaction with thiol or thioether compound

A solution of **NapOx** in IPA (20 μ L, 2 mM) and thiol or thioether compound in 1X PBS (20 μ L, 8 mM) was added to a mixture of IPA/1XPBS (7:1, 160 μ L) in 96-well plate, with a final concentration of 200 μ M for **NapOx** and 800 μ M for thiol or thioether compound. The mixture was allowed to incubate at room temperature for 3 h. Then it was directly analyzed by fluorescence measurement.

3.3 NapOx selectivity under neutral or acidic condition

For the reaction performed under neutral condition, a solution of **NapOx** in IPA (20 μ L, 100 μ M) and analyte in ddH₂O (20 μ L, 2 mM) was incubated in a mixture of IPA/ddH₂O (7:1, 160 μ L) in 96-well plate at 37 °C for 30 min, then it was directly used for fluorescence measurement. For the reaction under acidic condition, the procedure was the same except that 1 mM HCl aqueous solution was used to replace ddH₂O.

4. HPLC experiment

Compound **1a** (2.0 equiv) was mixed with SAH (1.0 equiv) in a mixture of H₂O/CH₃OH (1:1), and the mixture was allowed to incubate at room temperature for 10 min. Then, 100 μ L of reaction mixture was taken, and analyzed by HPLC.

5. NMR spectra of compounds

04 4 4 4 4 4 4 6 6 4 4 4 8 8 9 4 4 4 6 6 4 4 4 8 8 8 8 4 4 4 6 6 4 4 4 8 8 8 8	8	37 37 35 35 33 33 33 33 33 33 33 33 33 33 33	5 23 23
~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~~	LO I	ෆ්ෆ්ෆ්ෆ්ෆ්ෆ්ෆ්ෆ්ෆ්ෆ්	- + + +
	1		

¹H NMR: CDCI₃, 400 MHz





¹³C{¹H} NMR: CDCI₃, 101 MHz







¹H NMR: CDCI₃, 400 MHz





¹³C{¹H} NMR: CDCI₃, 101 MHz



200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 ( f1 (ppm)



![](_page_13_Figure_1.jpeg)

![](_page_14_Figure_0.jpeg)

NH₂

¹H NMR: *d*₆-DMSO, 400 MHz

![](_page_14_Figure_3.jpeg)

![](_page_15_Figure_0.jpeg)

Л

¹H NMR: CDCI₃, 400 MHz

![](_page_15_Figure_3.jpeg)

[—]и́

¹³C{¹H} NMR: CDCI₃, 101 MHz

![](_page_15_Figure_6.jpeg)

200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 f1 (ppm)

533 533 533 533 533 533 533 533 54 54 54 54 54 54 54 54 54 54 54 54 54	04 77 76 44 44 45 44 44 44 44 44 43	20 19
NNNNNNNNNNNNNN	ທີ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່ຕໍ່	N N N
		$\checkmark$

~ ∑^∬ Ҷ┐҈ѻӈ

¹H NMR: CDCI₃, 400 MHz

![](_page_16_Figure_3.jpeg)

00 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 C f1 (ppm)

![](_page_17_Figure_0.jpeg)

¹H NMR: CDCI₃, 400 MHz

![](_page_17_Figure_3.jpeg)

 $^{13}\text{C}\{^{1}\text{H}\}$  NMR: CDCI₃, 101 MHz

![](_page_17_Figure_6.jpeg)

![](_page_18_Figure_0.jpeg)

5.04 4.52 4.51 4.48 4.47 4.47 4.46 4.43 4.42

¹H NMR: CDCI₃, 400 MHz

![](_page_18_Figure_4.jpeg)

![](_page_19_Figure_0.jpeg)

![](_page_20_Figure_0.jpeg)

![](_page_20_Figure_1.jpeg)

![](_page_21_Figure_0.jpeg)

S22

![](_page_22_Figure_0.jpeg)

![](_page_23_Figure_0.jpeg)

![](_page_24_Figure_0.jpeg)

![](_page_25_Figure_0.jpeg)

S26

![](_page_26_Figure_0.jpeg)

#### 6. HRMS spectra of compounds

![](_page_27_Figure_1.jpeg)

![](_page_28_Figure_0.jpeg)

![](_page_28_Figure_1.jpeg)

![](_page_28_Figure_2.jpeg)

![](_page_29_Figure_0.jpeg)

![](_page_29_Figure_1.jpeg)

![](_page_29_Figure_2.jpeg)

![](_page_30_Figure_0.jpeg)

### 7. Reference

- S. Lin, X. Yang, S. Jia, A. M. Weeks, M. Hornsby, P. S. Lee, R. V. Nichiporuk, A. T. Iavarone, J. A. Wells, F. D. Toste, C. J. Chang, *Science* 2017, 355, 597-602.
- [2] A. H Christian, S. Jia, W. Cao, P. Zhang, A. T Meza, M. S Sigman, C. J Chang, F. D Toste. J. Am. Chem. Soc. 2019, 141, 12657-12662.