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Supporting Information

Highly asymmetric aldol reaction of isatins and ketones catalyzed by chiral bifunctional primary-amine organocatalyst on water

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1. General Experimental Details

The reactions were carried out in vials and stirred with a magnetic bar without inert atmosphere unless specified. All commercial reagents were purchased with the analysis purity grade. They were used without further purification unless specified.

The reactions were monitored by TLC (thin layer chromatography) method; column and preparative TLC purifications were carried out using silica gel. Melting points were uncorrected and recorded on XT-5 melting point apparatus.

NMR spectra were acquired on a Bruker 400/600 spectrometer, running at 400/600 MHz and 100/151 MHz for ¹H and ¹³C, respectively. NMR in CDCl₃, DMSO- d_6 with TMS as an internal standard, chemical shifts (δ) are reported in ppm relative to residual solvent signals (CDCl₃, 7.26 ppm for ¹H NMR and 77.00 ppm for ¹³C NMR; DMSO- d_6 , 2.50 ppm for ¹H NMR, 40.00 ppm for ¹³C NMR). The following abbreviations are used to describe peak patterns when appropriate: s (singlet), d (doublet), t (triplet), q (quartet), quint (quintet), sept (septuplet), m (multiplet), br (broad). High resolution mass spectra (HR-MS) were measured with ESI-Orbitrap mass spectrometer.

Enantiomeric excess (ee) were determined by chiral HPLC, Waters 1525 Binary HPLC.



2. HR-MS information of reaction intermediates Reaction mechanism

3. Preparation and characterization of catalysts

The catalyst C-H were reported in our previous work¹



3.1 General procedure for the synthesis of catalyst A-F.

1-hydroxyben-zotriazole (HOBt, 2.97 g, 22 mmol) and Boc-AA-OH **4** (20 mmol) were added to 50 mL dry CH₂Cl₂ in a 100 mL round bottom flask under argon. The reaction mixture was cooled to 0 °C, and DIC (*N*, *N*-diisopropylcarbodiimide) (3.4 mL, 22 mmol) was slowly added dropwise. After 30 min, 2-aminophenol **5a** (20 mmol, 2.18 g) was added and the stirring was continued at this temperature for another 30 min. Then the reaction was warmed to room temperature and the stirring was continued for 48 h. The insoluble material was filtered off and the filtrate was concentrated under reduced pressure. The residue was dissolved in ethyl acetate again and washed successively with 1.0 M NaOH solution, water, 1.0 M hydrochloric acid solution, saturated brine, dried with anhydrous Na₂SO₄, concentrated under reduced pressure and quickly prepared by column chromatography (petroleum ether: ethyl acetate = 8:1) to give the intermediate amides **6a-6f** (80~95% yield).

The amide **6a-6f** was dissolved in 20 mL CH_2Cl_2 . The reaction mixture was cooled to 0 °C, and trifluoroacetic acid (TFA, 20 mL) was slowly added dropwise. After stirring at room temperature for 6 h, the mixture was concentrated under reduced pressure to remove off TFA.

The residue was dissolved in dichloromethane, and added 1.0 M aqueous NaOH at 0 °C to the solution, adjust pH to 9.0, then the organic solution was extracted with dichloromethane (20 mL×3), and then washed with a small amount of saturated brine, dried with MgSO₄ and concentrated under reduced pressure to give target catalyst. The catalyst was directly obtained by recrystallization from petroleum ether and ethyl acetate.

(S)-2-Amino-N-(2-hydroxyphenyl)-3,3-dimethylbutanamide (A)¹



Yield 4.0 g, 90% (for 2 steps); White solid; M.p: 152-154 °C; $[\alpha]_D^{25}$ = -32.0 (*c* = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.44 (s, 1H), 7.13 – 7.09 (m, 1H), 7.02 (d, *J* = 7.8 Hz, 1H), 6.95 (d, *J* = 7.8 Hz, 1H), 6.87 – 6.82 (m, 1H), 3.35 (s, 1H), 1.08 (s, 9H).

(S)-2-amino-N-(2-hydroxyphenyl)-3-methylbutanamide (B)¹



Yield 3.66 g, 88% (for 2 steps); White solid; M.p:147.8-149.0 °C; $[\alpha]_D^{25}$ = - 39.0 (*c* = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.83 (s, 2H), 7.15 – 7.08 (m, 1H), 7.02 (dd, J = 8.1, 1.1 Hz, 1H), 6.94 (dd, J = 7.8, 1.2 Hz, 1H), 6.86 – 6.82 (m, 1H), 3.47 (d, J = 3.6 Hz, 1H), 2.44 – 2.50 (m, 1H), 1.57 (s, 2H), 1.06 (d, J = 7.2 Hz, 3H), 0.89 (d, J = 7.2 Hz, 3H).

(S)-2-Amino-N-(2-hydroxyphenyl)-4-methylpentanamide (C)¹



Yield 3.33 g, 75% (for 2 steps); White solid; M.p:143.4-144.5 °C; $[\alpha]_D^{25}$ = - 17.0 (*c* = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.85 (s, 1H), 7.11 (t, *J* = 7.6 Hz, 1H), 7.04 – 6.91, 6.84 (t, *J* = 7.2 Hz, 1H), 3.57 (d, *J* = 9.0 Hz, 1H), 1.87 – 1.74 (m, 2H), 1.49 – 1.41 (m, 1H), 0.99 (dd, *J* = 15.6, 6.0 Hz, 6H).

(2S)-2-amino-N-(2-hydroxyphenyl)-3-methylpentanamide (D)¹



Yield 3.1 g, 70% (for 2 steps); White solid; M.p:115 -117 °C; $[\alpha]_D^{25}$ = - 22.0 (*c* = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.86 (s, 2H), 7.13 – 7.07 (m, 1H), 7.01 (d, *J* = 8.4 Hz, 1H), 6.98 – 6.93 (m, 1H), 6.87 – 6.80 (m, 1H), 3.49 (d, *J* = 3.6 Hz, 1H), 2.20 – 2.10 (m, 1H), 1.59 (s, 2H), 1.45 – 1.38 (m, 1H), 1.19 – 1.12 (m, 1H), 1.04 (d, *J* = 7.2 Hz, 3H), 0.92 (t, *J* = 7.2 Hz, 3H).

(S)-2-amino-N-(2-hydroxyphenyl)-3-phenylpropanamide (E)¹



Yield 4.25 g, 83% (for 2 steps); White solid; M.p:133.8-135.2 °C; $[\alpha]_D^{25} = -83$ (c = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.81 – 9.49 (m, 2H), 7.35 (t, J = 7.2 Hz, 2H), 7.31 – 7.23 (m, 3H), 7.15 – 7.10 (m, 2H), 7.06 – 7.00 (m, 1H), 6.92 (dd, J = 7.8, 1.2 Hz, 1H), 6.86 – 6.82 (m, 1H), 3.82 (dd, J = 9.0, 4.2 Hz, 1H), 3.36 (dd, J = 13.8, 4.2 Hz, 1H), 2.85 (dd, J = 13.8, 9.0 Hz, 1H), 1.59 (s, 2H).

(S)-N-(2-hydroxyphenyl) pyrrolidine-2-carboxamide (F)¹



Yield 3.13 g, 76% (for 2 steps); White solid; M.p:169.5-170.8 °C; $[\alpha]_D^{25}$ = - 24.0 (*c* = 1.0, CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 9.98 (s, 2H), 7.13 – 7.08 (m, 1H), 7.00 (dd, *J* = 8.2, 1.2 Hz, 1H), 6.91 (dd, *J* = 7.9, 1.6 Hz, 1H), 6.86 – 6.81 (m, 1H), 3.93 (dd, *J* = 9.6, 4.8 Hz, 1H), 3.14 – 3.08 (m, 1H), 3.04 – 3.99 (m, 1H), 2.36 – 2.16 (m, 2H), 2.08 – 2.02 (m, 1H), 1.86 – 1.72 (m, 2H), 1.59 – 1.63 (m, 1H)

3.2 The procedure for the synthesis of catalyst G-K



1-hydroxyben-zotriazole (HOBt, 2.97 g, 22 mmol) and Boc-Tle-OH **4a** (20 mmol, 4.620 g) were added to 50 mL dry CH₂Cl₂ in a 100 mL round bottom flask under argon. The reaction mixture was cooled to 0 °C, and DIC (*N*, *N*-diisopropylcarbodiimide) (3.4 mL, 22 mmol) was slowly added dropwise. After 0.5 h, the amine **5** (20 mmol) was added and the stirring was continued at this temperature for another 0.5 h. Then the reaction was warmed to room temperature and the stirring was continued for 48 h. The insoluble material was filtered off and the filtrate was concentrated under reduced pressure. The residue was dissolved in ethyl acetate again and washed successively with 1.0 M NaOH solution, water, 1.0 M hydrochloric acid solution, saturated brine, dried with anhydrous Na₂SO₄, concentrated under reduced pressure and quickly prepared by column chromatography (petroleum ether: ethyl acetate = 8:1) to give the intermediate amides **6g-6k**.

The amide **6g-6k** were dissolved in 20 mL CH₂Cl₂. The reaction mixture was cooled to 0 °C, and trifluoroacetic acid (TFA, 20 mL) was slowly added dropwise. After stirring at room temperature for 6 h, the mixture was concentrated under reduced pressure to remove off TFA. The residue was dissolved in dichloromethane, and added 1.0 M aqueous NaOH at 0 °C to the solution, adjust pH to 9.0, then the organic solution was extracted with dichloromethane (20 mL×3), and then washed with a small amount of saturated brine, dried with MgSO₄ and concentrated under reduced pressure to give target catalyst. The catalyst was directly obtained by recrystallization from petroleum ether and ethyl acetate.



Yield 4.01 g, 85% (for 2 steps); White solid; M.p:134.6-135.4 °C; $[\alpha]_D^{25}$ = + 62.08 (*c* = 0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.14 (s, 1H), 8.38 (s, 3H), 7.46 (d, *J* = 7.8 Hz, 2H), 7.30 - 7.15 (m, 2H), 4.54 (s, 2H), 4.02 (d, *J* = 5.2 Hz, 1H), 3.60 (t, *J* = 6 Hz, 1H), 1.10 (s, 9H); ¹³C NMR (100 MHz, DMSO) δ 167.0, 136.4, 134.4, 128.2, 127.4, 126.0, 125.1, 67.5, 60.9, 60.0, 40.6, 40.4, 40.2, 40.0, 39.8, 39.6, 33.6, 26.9, 25.6; HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₃H₂₀N₂O₂Na 259.1422, found: 259.1415.

(S)-2-amino-N-((S)-2-hydroxy-1-phenylethyl)-3,3-dimethylbutanamide(H)



Yield 3.90 g, 78% (for 2 steps); White solid; M.p: 114.6-115.9 °C; $[\alpha]_D^{25}$ = 124.48 (*c* = 0.25, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 7.50 (d, *J* = 6.8 Hz, 1H), 7.40 – 7.22 (m, 5H), 5.10 – 5.00 (m, 1H), 3.91 – 3.78 (m, 2H), 3.13 (s, 1H), 2.25 (s, 2H), 0.97 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ 174.3, 139.2, 128.8, 127.8, 126.8, 66.7, 64.3, 55.8, 34.26, 26.8; HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₄H₂₂N₂O₂Na 273.1579, found:273.1577.

(S)-2-amino-3,3-dimethyl-N-phenylbutanamide (I)¹



Yield 3.3 g, 80% (for 2 steps); White solid; M.p: 222 - 224 °C; $[\alpha]_D^{25}$ = +38 (*c* = 1.0, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.97 (s, 1H), 8.38 (s, 2H), 7.68 – 7.62 (m, 2H), 7.33 – 7.28 (m, 2H), 7.07 (dd, *J* = 8.8, 4.9 Hz, 1H), 3.92 (s, 1H), 3.34 (s, 1H), 1.04 (s, 9H).

(S)-2-amino-N-(2-methoxyphenyl)-3,3-dimethylbutanamide (J)



Yield 4.48 g, 95% (for 2 steps); White solid; M.p: 168-170 °C; $[\alpha]_D^{25}$ = 16.40 (*c* = 0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 9.69 (s, 1H), 8.16 (d, *J* = 8 Hz, 1H), 7.04 (d, *J* = 4 Hz, 2H), 6.94 - 6.87 (m, 1H), 3.84 (s, 3H), 3.19 (s, 1H), 2.64 (s, 2H), 0.97 (s, 9H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 172.5, 149.2, 127.7, 124.1, 120.9, 120.3, 111.4, 64.2, 56.3, 39.38, 34.5, 27.2; HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₃H₂₀N₂O₂Na 259.1422, found:259.1420.

(S)-2-amino-N-(2-hydroxyphenyl)-N,3,3-trimethylbutanamide (K)



Yield 3.61 g, 82% (for 2 steps); White solid; M.p: 188.2-190.1 °C; $[\alpha]_D^{25} = -273.36$ (c = 0.25, MeOH); ¹H NMR (400 MHz, CDCl₃) δ 8.73 (s, 3H), 7.70 – 7.30 (m, 5H), 3.94 (d, J = 5.6 Hz, 1H), 3.34 (s, 3H), 1.06 (s, 9H); ¹³C NMR (100 MHz, CDCl₃) δ 167.9, 142.7, 130.2, 128.3, 128.0, 57.6, 38.5, 34.6, 26.8; HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₄H₂₂N₂ONa 243.1473, found:243.1469.

4. General procedure for the Aldol reaction

4.1 The effect of the amount of water on the stereoselectivity and yield of the reaction



S-Table 1 Optimization of the amount of water.

[a] **1a** (0.5 mmol), **2a** (1.0 mmol) were used; [b] isolated yields are reported; [c] dr (*syn: anti*) was determined by chiral HPLC; [d] the ee of *syn-***3a** was determined by chiral HPLC (absolute configuration is inferred from the single-crystal structure of **3d**).

4.2. General procedure for synthesis of product 3



In a 5 mL reaction flask equipped with a magnetic stirrer, catalyst A (11 mg, 0.05 mmol, 10%) and isatins 1 (0.5 mmol) were added, then 1.0 mL water and ketone (1.0 mmol) was added dropwise then. The reaction mixture was stirred at room temperature and detected by TLC. After the reaction was completed, the reaction mixture was purified by column chromatography on silica gel (CH₂Cl₂: MeOH = 100:1~50:1) to give 3a - 3s.

4.3. Scale-up reaction and reuse of catalyst



In a 50 mL round bottomed flask equipped with a magnetic stirrer, catalyst A (222 mg, 1.0 mmol, 10%) and isatins 1a (1.47g, 10 mmol) were added, then 20 mL water and cyclohexanone (2 mL, 20 mmol) was added dropwise then. The reaction mixture was stirred at room temperature until completed. After the reaction was completed, the mixture was filtered directly to give product 3a, and the solution is directly used in the next cycle.



(a) Reaction in progress

(b) The reaction was completed

Run	time (h)	yield %	dr	ee %
1	48	98	99:1	96
2	48	96	99:1	93
3	72	85	99:1	93

S-Table 1 Recovery and Reuse of Catalyst A

5. Characterization of Aldol reaction products

(R)-3-hydroxy-3-((S)-2-oxocyclohexyl) indolin-2-one (3a)²

3a: white solid, 120 mg, 98% yield, 98:2dr, 99% ee, $[\alpha]_D^{25} = 44.64$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.16 (s, 1H), 7.19 (d, *J* = 7.2 Hz, 1H), 7.16 – 7.12 (m, 1H), 6.85 – 6.81 (m, 1H), 6.76 (d, *J* = 7.2 Hz, 1H), 5.78 (s, 1H), 3.08 – 3.03 (m, 1H), 2.61 – 2.55 (m, 1H), 2.33 – 2.24 (m, 1H), 2.04 – 1.87 (m, 3H), 1.85 – 1.76 (m, 1H), 1.70 – 1.59 (m, 1H), 1.49 – 1.38 (m, 1H); The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 254 nm; retention time: 17.2 min (minor) and 12.1 min (major).



(S)-3-hydroxy-3-((R)-2-oxocyclohexyl) indolin-2-one (ent-3a)²

*ent-3***a**: white solid, 120 mg, 98% yield, 99:1 dr, 99% ee, $[\alpha]_D^{25} = -44.72$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.16 (s, 1H), 7.19 (d, *J* = 7.2 Hz, 1H), 7.16 – 7.12 (m, 1H), 6.85 – 6.81 (m, 1H), 6.76 (d, *J* = 7.2 Hz, 1H), 5.78 (s, 1H), 3.08 – 3.03 (m, 1H), 2.61 – 2.55 (m, 1H), 2.33 – 2.24 (m, 1H), 2.04 – 1.87 (m, 3H), 1.85 – 1.76 (m, 1H), 1.70 – 1.59 (m, 1H), 1.49 – 1.38 (m, 1H); The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 254 nm; retention time: 12.3 min (minor) and 17.2 min (major).



(R)-3-hydroxy-1-methyl-3-((S)-2-oxocyclohexyl) indolin-2-one (3b)²

3b: white solid, 117 mg, 90% yield, 96:4 dr, 64% ee, $[\alpha]_D^{25} = 59.84$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.37 – 7.20 (m, 2H), 7.02 – 6.87 (m, 2H), 5.89 (s, 1H), 3.20 – 3.02 (m, 3H), 2.68 – 2.56 (m, 1H), 2.35 – 2.25 (m, 1H), 2.08 – 1.75 (m, 4H), 1.75 – 1.60 (m, 1H), 1.51 – 1.36 (m, 1H); The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 254 nm; retention time: 6.6 min (major) and 7.4 min (minor).



(*R*)-3-hydroxy-3-((*R*)-2-oxocyclohexyl)-1-phenylindolin-2-one (3c)²

3c: white solid, 138 mg, 86% yield, 99:1 dr, 28% ee, $[\alpha]_D^{25} = -53.28$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.58 (t, *J* = 7.6 Hz, 2H), 7.50 – 7.35 (m, 3H), 7.22 (t, *J* = 7.6 Hz, 1H), 7.00 (t, *J* = 7.5 Hz, 1H), 6.68 (d, *J* = 7.9 Hz, 1H), 6.16 (s, 1H), 3.28 – 3.18 (m, 1H), 2.77 – 2.21 (m, 3H), 2.11 – 1.84 (m, 3H), 1.80 – 1.65 (m, 1H), 1.55– 1.40 (m, 1H). The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=80:20; flow rate 1.0 mL/min; 254 nm; retention time: 10.1 min (major) and 13.4 min (minor).



(*R*)-4-chloro-3-hydroxy-3-((*S*)-2-oxocyclohexyl) indolin-2-one (3d)³

3d: white solid, 133 mg, 95% yield, 99:1 dr, 94% ee, $[\alpha]_D^{25} = -36.80$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.35 (s, 1H), 7.15 (t, *J* = 8.0 Hz, 1H), 6.82 (d, *J* = 8.0 Hz, 1H), 6.71 (d, *J* = 7.6 Hz, 1H), 6.04 (s, 1H), 3.79 (dd, *J* = 12.8, 5.2 Hz, 1H), 2.40 – 2.11 (m, 2H), 2.10 – 1.79 (m, 3H), 1.71 – 1.39 (m, 2H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 7.2 min (minor) and 7.5 min (major).



(*R*)-4-bromo-3-hydroxy-3-((*S*)-2-oxocyclohexyl) indolin-2-one (3e)³

3e: white solid, 153 mg, 95% yield, 99:1 dr, 98% ee, $[\alpha]_D^{25} = -54.08$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.59 (s, 1H), 7.21 – 7.09 (m, 2H), 6.82 (d, *J* = 7.6 Hz, 1H), 6.18 (s, 1H), 2.44 – 2.26 (m, 2H), 1.96 – 1.69 (m, 3H), 1.64 – 1.41 (m, 3H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 7.4 min (minor) and 8.3 min (major).



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(R)-5--fluoro-3-hydroxy-3-((S)-2-oxocyclohexyl) indolin-2-one (3f)³

3f: white solid, 125 mg, 95% yield, 99:1 dr, 75% ee, $[\alpha]_D^{25} = 57.92$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-d₆) δ 7.56 (t, *J* = 7.8 Hz, 2H), 7.39 (m, 4H), 7.20 (t, *J* = 7.8 Hz, 1H), 6.98 (t, *J* = 7.8 Hz, 1H), 6.66 (d, *J* = 7.8 Hz, 1H), 6.14 (d, *J* = 4.2 Hz, 1H), 3.21 (dd, *J* = 4.8, 13.2Hz, 1H), 2.70 – 2.61 (m, 1H), 2.33 (m, 1H), 2.10 – 2.00 (m, 1H), 1.91 (m, 3H), 1.70 (m, 1H), 1.52 – 1.41 (m, 1H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol= 90:10; flow rate 1.0 mL/min; 254 nm; retention time: 31.2 min (minor) and 36.8 min (major).



(*R*)-5-chloro-3-hydroxy-3-((*S*)-2-oxocyclohexyl) indolin-2-one (3g)³

3g: white solid, 134 mg, 96% yield, 99:1 dr, 87% ee, $[\alpha]_D^{25} = 36.16$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.35 (s, 1H), 7.25 – 7.17 (m, 2H), 6.81 (d, *J* = 7.8 Hz, 1H), 6.00 (s, 1H), 3.13 – 3.07 (m, 1H), 2.62 – 2.55 (m, 1H), 2.37 – 2.28 (m, 1H), 2.07 – 2.01 (m, 1H), 1.98 – 1.88 (m, 2H), 1.83 – 1.74 (m, 1H), 1.73 – 1.62 (m, 1H), 1.56 – 1.45 (m, 1H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=90:10; flow rate 1.0 mL/min; 254 nm; retention time: 29.8 min (minor) and 39.0 min (major).



(R)-5-bromo-3-hydroxy-3-((S)-2-oxocyclohexyl) indolin-2-one (3h)³

3h: white solid, 149 mg, 92% yield, 99:1 dr, 74% ee, $[\alpha]_D^{25} = 22.24$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.36 (s, 1H), 7.36 (dd, *J* = 8.4, 2.4 Hz, 1H), 7.30 (d, *J* = 1.8 Hz, 1H), 6.77 (d, *J* = 8.4 Hz, 1H), 6.01 (s, 1H), 3.10 (dd, *J* = 13.2, 5.4 Hz, 1H), 2.63 – 2.54 (m, 1H), 2.33 (td, *J* = 13.8, 6.0 Hz, 1H), 2.07 – 1.45 (m, 6H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 8.3 min (minor) and 10.7 min (major).



(R)-3-hydroxy-5-nitro-3-((S)-2-oxocyclohexyl) indolin-2-one (3i)²

3i: white solid, 131 mg, 91% yield, 96:4 dr, 96% ee, $[\alpha]_D^{25} = 71.92$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.36 (s, 1H), 7.41 – 7.23 (m, 2H), 6.76 (d, *J* = 8.0 Hz, 1H), 6.01 (s, 1H), 3.09 (dd, *J* = 13.2, 5.1 Hz, 1H), 2.57 (d, *J* = 12.0 Hz, 1H), 2.41 – 2.23 (m, 1H), 2.11 – 1.39 (m, 6H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 8.5 min (minor). and 10.3 min (major).



(R)-3-hydroxy-5-methyl-3-((S)-2-oxocyclohexyl) indolin-2-one (3j)³

3j: white solid, 123 mg, 95% yield, 98:2 dr, 60% ee, $[\alpha]_D^{25} = 29.44$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.07 (s, 1H), 7.08 – 6.92 (m, 2H), 6.67 (d, *J* = 7.8 Hz, 1H), 5.75 (s, 1H), 3.06 (dd, *J* = 13.2, 5.4 Hz, 1H), 2.58 (s, 1H), 2.31 (td, *J* = 13.8, 6.0 Hz, 1H), 2.22 (s, 3H), 2.08 – 1.78 (m, 4H), 1.73 – 1.61 (m, 1H), 1.57 – 1.41 (m, 1H); The ee was determined by HPLC analysis. CHIRALPAK IC; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 15.5 min (major) and 28.4 min (minor).



(*R*)-3-hydroxy-5-methyl-3-((*S*)-2-oxocyclohexyl) indolin-2-one (3k)²

3k: white solid, 129 mg, 94% yield, 97:3 dr, 70% ee, $[\alpha]_D^{25} = 38.0$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.02 (s, 1H), 6.87 – 6.61 (m, 3H), 5.82 (s, 1H), 3.68 (s, 3H), 3.06 (dd, J = 13.2, 5.2 Hz, 1H), 2.60 – 2.53 (m, 1H), 2.31 (td, J = 13.6, 6.0 Hz, 1H), 2.07 – 1.45 (m, 6H).; The ee was determined by HPLC analysis. CHIRALPAK IC; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 15.5 min (major) and 28.4 min (minor).



(R)-6-bromo-3-hydroxy-3-((S)-2-oxocyclohexyl) indolin-2-one (31)⁴

31: white solid, 147 mg, 91% yield, 99:1 dr, 92% ee, $[\alpha]_D^{25} = 57.04$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.35 (s, 1H), 7.15 (d, *J* = 7.8 Hz, 1H), 7.04 (dd, *J* = 7.8, 1.8 Hz, 1H), 6.94 - 6.92 (m, 1H), 5.94 (s, 1H), 3.13 - 3.04 (m, 1H), 2.62 - 2.55 (m, 1H), 2.36 - 2.28 (m, 1H), 2.06 - 1.41 (m, 6H); The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 9.6 min (minor) and 13.1 min (major).



6523136

218319

95.89

13.070

(R)-7-bromo-3-hydroxy-3-((S)-2-oxocyclohexyl) indolin-2-one (3m)

3m: white solid, 155 mg, 96% yield, 93:7 dr, 57% ee, $[\alpha]_D^{25} = 44.08$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.48 (s, 1H), 7.37 (d, *J* = 8.0 Hz, 1H), 7.22 (d, *J* = 7.2 Hz, 1H), 6.83 (t, *J* = 7.6 Hz, 1H), 6.02 (s, 1H), 3.11 (dd, *J* = 13.2, 5.2 Hz, 1H), 2.58 (d, *J* = 12.0 Hz, 1H), 2.41 – 2.26 (m, 1H), 2.11 – 1.36 (m, 6H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 209.7, 179.0, 143.2, 133.4, 131.99, 124.3, 123.1, 102.5, 75.1, 58.0, 41.9, 27.1, 24.9; The ee was determined by HPLC analysis. CHIRALPAK IC; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 13.0 min (minor) and 14.9 min (major). HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₄H₁₄NO₃NaBr, 346.0055, found: 346.0049.



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(R)-3-hydroxy-3-((S)-4-oxotetrahydro-2H-thiopyran-3-yl) indolin-2-one (3n)²

3n: white solid, 124 mg, 94% yield, 72:28 dr, 80% ee, $[\alpha]_D^{25} = 57.04$ (*c* =0.25, MeOH); ¹H NMR (600 MHz, DMSO-*d*₆) δ 10.26 (s, 1H), 7.26 (dd, *J* = 7.2, 1.2 Hz, 1H), 7.17 (td, *J* = 7.8, 1.2 Hz, 1H), 6.89 - 6.84 (m, 1H), 6.79 (d, *J* = 7.8 Hz, 1H), 6.04 (s, 1H), 3.50 - 3.45 (m, 1H), 3.23 (t, *J* = 12.6 Hz, 1H), 2.94 - 2.85 (m, 2H), 2.60 - 2.53 (m, 1H), 2.44 - 2.38 (m, 1H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 11.7 min (major) and 13.7min (minor).



(R)-3-hydroxy-3-((1S,5R)-5-methyl-2-oxocyclohexyl) indolin-2-one (3o)²

3o: white solid, 56 mg, 43% yield, 97:3 dr, 67% ee, $[\alpha]_D^{25} = 57.04$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-d₆) δ 10.14 (s, 1H), 7.26 (d, *J* = 7.4 Hz, 1H), 7.14 (t, *J* = 7.6 Hz, 1H), 6.93 -8.82 (m, 1H), 6.74 (d, *J* = 7.7 Hz, 1H), 5.71 (s, 1H), 3.30 (d, *J* = 4.9 Hz, 1H), 2.42 (dd, *J* = 13.6, 5.6 Hz, 1H), 2.26 - 2.15 (m, 1H), 2.05 - 1.86 (m, 3H), 1.53 (q, *J* = 12.8 Hz, 1H), 1.27 - 1.12 (m, 1H), 0.94 (d, *J* = 6.4 Hz, 3H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 7.8 min (minor) and 9.1 min (major).



(*R*)-3-hydroxy-3-((*1S*,4*R*)-4-methyl-2-oxocyclohexyl) indolin-2-one (3p)

Yellow solid, 123 mg, 95% yield, 99:1 dr, 29% ee, $[\alpha]_D^{25} = 15.84$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.18 (s, 1H), 7.23 – 7.11 (m, 2H), 6.88 – 6.76 (m, 2H), 5.81 (s, 1H), 3.03 (dd, *J* = 13.6, 5.2 Hz, 1H), 2.61 – 2.52 (m, 1H), 2.08 – 1.98 (m, 2H), 1.96 – 1.77 (m, 2H), 1.75 – 1.61 (m, 1H), 1.47 – 1.34 (m, 1H), 0.94 (d, *J* = 6.4 Hz, 3H); ¹³C NMR (100 MHz, DMSO-*d*₆) δ 209.1, 179.2, 143.9, 131.3, 129.0, 125.3, 121.3, 109.9, 74.3, 57.1, 50.0, 34.7, 33.5, 26.1, 22.4. The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=90:10; flow rate 1.0 mL/min; 254 nm; retention time: 16.6 min (minor) and 18.3 min (major). HRMS (ESI-Orbitrap) m/z: [M+Na]⁺ Calcd for C₁₅H₁₇NO₃Na, 282.1106, found: 282.1099.



	name	Retention	Area(µV*s)	Height	% Area
1		16.566	2775336	78229	35.57
2		18.251	5027280	133494	64.43

(R)-3-hydroxy-3-((S)-2-oxocycloheptyl) indolin-2-one (3q)²

3p: white solid, 128 mg, 99% yield, 99:1 dr, 32% ee, $[\alpha]_D^{25} = -0.48$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.16 (s, 1H), 7.29 – 7.11 (m, 2H), 6.94 – 6.71 (m, 2H), 5.95 (s, 1H), 3.09 (dd, *J* = 11.3, 2.8 Hz, 1H), 2.48 – 2.36 (m, 1H), 2.29 – 2.11 (m, 2H), 2.08 – 1.80 (m, 2H), 1.79 – 1.58 (m, 2H), 1.53 – 1.21 (m, 3H); The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 8.4 min (minor) and 9.9 min (major).



(R)-3-hydroxy-3-((S)-2-oxocycloheptyl) indolin-2-one (3r)²

3q: white solid, 114 mg, 99% yield, 80:20 dr, 4% ee, $[\alpha]_D^{25} = 2.64$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.28 (d, *J* = 6.8 Hz, 1H), 7.36 (d, *J* = 7.2 Hz, 1H), 7.22 - 7.10 (m, 1H), 6.95 - 6.85 (m, 1H), 6.81 - 6.72 (m, 1H), 5.98 (d, *J* = 20.4 Hz, 1H), 2.93 - 2.78 (m, 1H), 2.29 - 1.54 (m, 6H).; The ee was determined by HPLC analysis. CHIRALPAK AD-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 8.4 min (minor) and 8.7 min (major).



(R)-3-hydroxy-3-(2-oxopropyl) indolin-2-one (3s)⁵

3r: white solid, 97 mg, 95% yield, 3% ee, $[\alpha]_D^{25} = 0.80$ (*c* =0.25, MeOH); ¹H NMR (400 MHz, DMSO-*d*₆) δ 10.20 (s, 1H), 7.30 – 7.11 (m, 2H), 6.90 (t, *J* = 7.6 Hz, 1H), 6.77 (d, *J* = 7.6 Hz, 1H), 5.97 (s, 1H), 3.35 – 3.23 (m, 1H), 3.00 (d, *J* = 16.4 Hz, 1H), 2.00 (s, 3H); The ee was determined by HPLC analysis. CHIRALPAK OJ-H; Hexane/2-propanol=70:30; flow rate 1.0 mL/min; 254 nm; retention time: 16.4 min (minor) and 19.5 min (major).



6. Copies of NMR spectra



Figure S1. ¹H NMR (600 MHz, CDCl₃) of Catalyst A



Figure S2. ¹H NMR (600 MHz, CDCl₃) of Catalyst B



Figure S3. ¹H NMR (600 MHz, CDCl₃) of Catalyst C



Figure S4. ¹H NMR (600 MHz, CDCl₃) of Catalyst **D**



Figure S5. ¹H NMR (600 MHz, CDCl₃) of Catalyst E



Figure S6. ¹H NMR (600 MHz, CDCl₃) of Catalyst F



Figure S7. ¹H NMR (400 MHz, DMSO-d₆) and ¹³C NMR (100 MHz, DMSO-d₆) of Catalyst G



Figure S8. ¹H NMR (400 MHz, CDCl₃) and ¹³C NMR (100 MHz, CDCl₃) of Catalyst H



Figure S9. ¹H NMR (600 MHz, DMSO-*d*₆) of Catalyst K



Figure S10. ¹H NMR (400 MHz, CDCl₃) and ¹³C NMR (600 MHz, CDCl₃) of Catalyst K



Figure S11. ¹H NMR (400 MHz, CDCl₃) and ¹³C NMR (100 MHz, CDCl₃) of Catalyst K



Figure S12. ¹H NMR (600 MHz, DMSO- d_6) of **3a**.



Figure S13. ¹H NMR (400 MHz, DMSO-*d*₆) of **3b**.







Figure S15. ¹H NMR (400 MHz, DMSO-d₆) of **3d**



Figure S16. ¹H NMR (400 MHz, DMSO-*d*₆) of **3e**.



Figure S17. ¹H NMR (600 MHz, DMSO-*d*₆) of **3f**.



Figure S18. ¹H NMR (400 MHz, DMSO-*d*₆) of **3g**.



Figure S19. ¹H NMR (600 MHz, DMSO-*d*₆) of **3h**.



Figure S20. ¹H NMR (600 MHz, DMSO-*d*₆) of **3i**.



Figure S21. ¹H NMR (400 MHz, DMSO-*d*₆) of **3j**.



Figure S22. ¹H NMR (400 MHz, DMSO-*d*₆) of **3**k.



Figure S23. ¹H NMR (600 MHz, DMSO-*d*₆) of **31**.



Figure S24. ¹H NMR (400 MHz, DMSO-*d*₆) and ¹³C NMR (100 MHz, DMSO-*d*₆) of **3m**.



Figure S25. ¹H NMR (600 MHz, DMSO-*d*₆) of **3n**.



Figure S26. ¹H NMR (400 MHz, DMSO-*d*₆) of **30**.



Figure S27. ¹H NMR (400 MHz, DMSO-*d*₆) and ¹³C NMR (100 MHz, DMSO-*d*₆) of **3p**.



Figure S28. ¹H NMR (400 MHz, DMSO- d_6) of **3q**.



Figure S29. ¹H NMR (400 MHz, DMSO-*d*₆) of **3r**.



Figure S30. ¹H NMR (400 MHz, DMSO-*d*₆) of **3s**.

7. Single crystal X-ray crystallographic data

7.1. Single crystal data, structural refinement and measurement of catalyst A



C: Crystal Data for C₁₂H₁₈N₂O₂ (M =222.28 g/mol): monoclinic, space group P2₁ (no. 4), a = 6.0452(3) Å, b = 10.0440(5) Å, c = 9.6566(4) Å, β = 90.901(2)°, V = 586.26(5) Å³, Z = 2, T =

173.0 K, $\mu(CuK\alpha) = 0.698 \text{ mm}^{-1}$, $Dcalc = 1.259 \text{ g/cm}^3$, 12901 reflections measured (9.158° $\leq 2\Theta \leq 149.614^\circ$), 2372 unique ($R_{int} = 0.0540$, $R_{sigma} = 0.0338$) which were used in all calculations. The final R_1 was 0.0369 (I > $2\sigma(I)$) and wR_2 was 0.0936 (all data). A suitable crystal was selected on a Bruker D8 Venture diffractometer. The crystal was kept at 173.00 K during data collection. Using Olex2^[1], the structure was solved with the olex2.solve ^[2] structure solution program using Charge Flipping and refined with the olex2.refine ^[3] refinement package using Gauss-Newton minimization.

Identification code	А
Empirical formula	$C_{12}H_{18}N_2O_2$
Formula weight	222.28
Temperature/K	173.0
Crystal system	monoclinic
Space group	P21
a/Å	6.0452(3)
b/Å	10.0440(5)
c/Å	9.6566(4)
α/°	90
β/°	90.901(2)
γ/°	90
Volume/Å ³	586.26(5)
Z	2
$\rho_{calc}g/cm^3$	1.259
μ/mm^{-1}	0.698
F(000)	240.0
Crystal size/mm ³	$0.25 \times 0.22 \times 0.19$
Radiation	$CuK\alpha$ ($\lambda = 1.54178$)
2Θ range for data collection/°	9.158 to 149.614
Index ranges	$-7 \le h \le 7, -12 \le k \le 12, -12$
index ranges	$\leq 1 \leq 12$
Reflections collected	12901
Independent reflections	2372 [$R_{int} = 0.0540, R_{sigma} =$
independent reflections	0.0338]
Data/restraints/parameters	2372/1/156
Goodness-of-fit on F ²	1.057
Final R indexes [I>= 2σ (I)]	$R_1 = 0.0369, wR_2 = 0.0915$
Final R indexes [all data]	$R_1 = 0.0386, wR_2 = 0.0936$
Largest diff. peak/hole / e Å ⁻³	0.22/-0.18

Table 1	Crystal	data and	l structure	refinement	for	A.
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Flack parameter

0.06(9)



7.2 Single crystal data, structural refinement and measurement of 3d

3d: Single crystals of C₁₄H₁₄ClNO₃ (M=279.725 g/mol): orthorhombic, space group P2₁2₁2₁ (no. 19), a = 7.2587(2) Å, b = 11.3133(3) Å, c = 15.3314(4) Å, V = 1259.01(6) Å³, Z = 4, T = 173.00 K, μ (Cu K α) = 2.730 mm⁻¹, Dcalc = 1.476 g/cm³, 30428 reflections measured (9.72° $\leq 2\Theta \leq 144.08^{\circ}$), 2467 unique (R_{int} = 0.0533, R_{sigma} = 0.0207) which were used in all calculations. The final R_1 was 0.0298 (I>=2u(I)) and wR_2 was 0.0768 (all data). A suitable crystal was selected on a Bruker D8 Venture diffractometer. The crystal was kept at 173.00 K during data collection. Using Olex2^[1], the structure was solved with the olex2.solve ^[2] structure solution program using Charge Flipping and refined with the olex2.refine ^[3] refinement package using Gauss-Newton minimization.

Table 1 Crystal data and structure refinement for 3d.

Identification code	3d
Empirical formula	$C_{14}H_{14}ClNO_3$
Formula weight	279.725
Temperature/K	173.00
Crystal system	orthorhombic
Space group	$P2_{1}2_{1}2_{1}$
a/Å	7.2587(2)
b/Å	11.3133(3)
c/Å	15.3314(4)
α/°	90

β/°	90
$\gamma/^{\circ}$	90
Volume/Å ³	1259.01(6)
Z	4
$\rho_{calc}g/cm^3$	1.476
μ/mm^{-1}	2.730
F(000)	587.4
Crystal size/mm ³	$0.19 \times 0.17 \times 0.16$
Radiation	Cu Ka ($\lambda = 1.54178$)
2Θ range for data collection/°	9.72 to 144.08
Index ranges	$\begin{array}{l} -8 \leq h \leq 8, -13 \leq k \leq 13, -18 \\ \leq l \leq 17 \end{array}$
Reflections collected	30428
Independent reflections	2467 [$R_{int} = 0.0533$, $R_{sigma} = 0.0207$]
Data/restraints/parameters	2467/0/173
Goodness-of-fit on F ²	1.086
Final R indexes [I>=2 σ (I)]	$R_1 = 0.0298, wR_2 = 0.0761$
Final R indexes [all data]	$R_1 = 0.0307, wR_2 = 0.0768$
Largest diff. peak/hole / e Å ⁻³	0.29/-0.18
Flack parameter	0.027(6)

8. Copies of HRMS spectra



Chemical Formula: C₁₃H₂₀N₂O₂

C: 13-13 H: 20-20 N: 0-100 O: 0-100 Na: 0-3







н





DBE 4.5



Chemical Formula: C₁₃H₂₀N₂O₂



Figure S33. HRMS spectra of J.



Κ

Chemical Formula: C₁₃H₂₀N₂O



Figure S34. HRMS spectra of K



Chemical Formula: C₁₄H₁₄BrNO₃



Figure S35. HRMS spectra of 3m



Chemical Formula: C₁₅H₁₇NO₃



Figure S36. HRMS spectra of **3p**

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