

Transition Metal and Additives Free Intermolecular Hydroarylation of Alkenes with Indoles in Hexafluoroisopropanol

Changsheng Zhou^{a+}, Ming Huang^{a+}, Yufeng Yao^{a+}, Chunyu Chen^a, Xin Yi^a,
Kefang Yang^a, Guo-Qiao Lai^a, Wenjing Xuan^{*b}, Pinglu Zhang^{*a}

^aHangzhou Normal University, College of Material Chemistry and Chemical Engineering, Key Laboratory of Organosilicon Chemistry and Material Technology, Ministry of Education, Key Laboratory of Organosilicon Material Technology of Zhejiang Province, Hangzhou 311121, China

^bWestlake University, School of Engineering, Hangzhou 310030, China

+ Authors contribute equally

Table of Contents

1. General information	3
2. General procedures of substrates preparation.....	4
3. General procedure of the hydroarylation.....	6
4. Synthetic transformations	16
5. Gram scale reaction	20
6. In vitro cell cytotoxicity	21
7. Control experiments.....	22
8. X-Ray analysis.....	26
9. NMR spectra.....	34
10. Reference and notes.....	80

1. General information

Unless otherwise stated, all reactions were magnetically stirred and conducted in oven-dried (100 °C) or in anhydrous solvents under N₂, applying standard Schlenk techniques. Solvents and liquid reagents, as well as solutions of solid or liquid reagents were added through a weak N₂ counter-flow. Cooling baths were prepared in Dewar vessels, filled with ice/water (0 °C). Heated oil baths were used for reactions requiring elevated temperatures. Solvents were removed under reduced pressure at 40 °C using a rotary evaporator, and unless otherwise stated, the remaining compound was dried in vacuum at ambient temperature. All given yields are isolated yields of chromatographically and NMR spectroscopically pure materials, unless otherwise stated.

Chemicals

Chemicals were purchased from commercial suppliers (including Energy Chemical, Bidepharm, Aladdin, Meryer, Sinopharm) and used without further purification unless otherwise stated.

Solvents

Solvents (CH₂Cl₂, Et₂O, THF) were dried by distillation from an appropriate drying agent. In addition, more solvents were purchased from commercial suppliers.

Gas

Dry N₂ were purchased from Hangzhou Jingong Materials with > 99.9% purity.

Thin Layer chromatography

Thin-layer chromatography (TLC) was performed using silica gel pre-coated glass plates (SIL G-25, with fluorescent indicator UV254; Macherey-Nagel) with fluorescent indicator UV254; Macherey-Nagel), which were visualized by irradiation with UV light ($\lambda = 254$ or 366 nm), basic KMnO₄, and/or phosphomolybdic acid (PMA).

Column Chromatography

Column chromatography (CC) was carried out using silica gel (60 Å, 230–400 mesh, particle size 0.040–0.063 mm) using technical grade solvents. Elution was accelerated using compressed air. All reported yields, unless otherwise specified, refer to spectroscopically and chromatographically pure compounds.

Nuclear Magnetic Resonance Spectroscopy

¹H, ¹³C, nuclear magnetic resonance (NMR) spectra were recorded on a Bruker AV-500, AV-400 spectrometer in a suitable deuterated solvent. The solvent employed and respective measuring frequency are indicated for each experiment. Chemical shifts are reported with tetramethylsilane (TMS) serving as a universal reference of all nuclides

and with two or one digits after the comma. The resonance multiplicity is described as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), and bs (broad singlet). All spectra were recorded at 298 K unless otherwise noted, processed with program MestReNova 14.0, and coupling constants are reported as observed. The residual deuterated solvent signal relative to tetramethylsilane (TMS) was used as the internal reference in ^1H NMR spectra (CDCl_3 δ 7.26, THF δ 1.72, 3.58, C_6D_6 δ 7.16, CD_3OD δ 3.31), and are reported as follows: chemical shift δ in ppm (multiplicity, coupling constant J in Hz, number of protons). ^{13}C NMR spectra reported in ppm from tetramethylsilane (TMS) with the solvent resonance as the internal standard (CDCl_3 δ 77.2, THF δ 67.21, 25.31, C_6D_6 δ 128.1, CD_3OD δ 49.0). All spectra are broadband decoupled unless otherwise noted.

Representative biologically active compounds incorporating indole motif

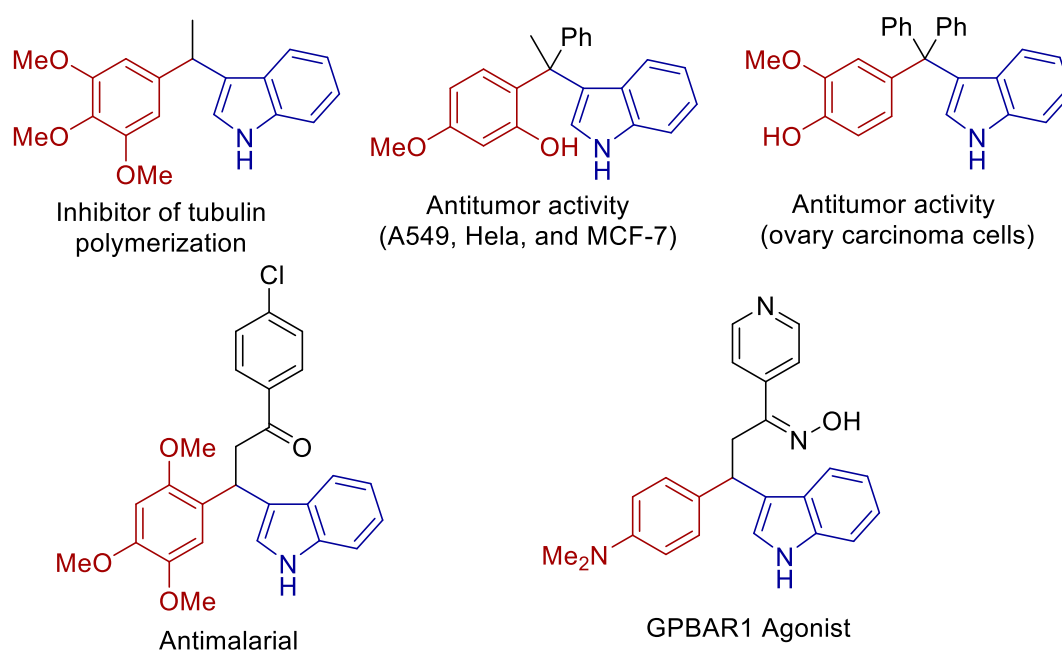
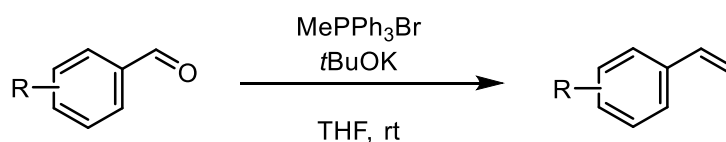


Figure S1. Representative multi-arylalkanes with indole motif

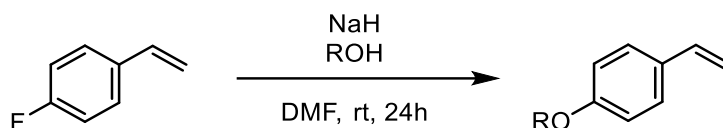
2. General procedures of substrates preparation

Method A:



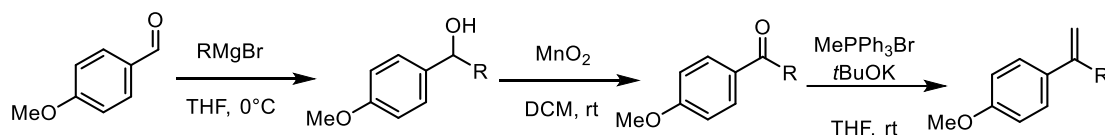
The alkenes **1a–1aa** were prepared according to method A.

Method B:



The alkenes **1ab–1ad** were prepared according to method B.

Method C:

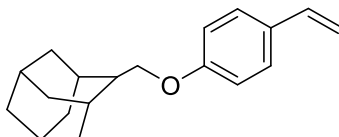


Scheme S1 General procedure of substrates preparation

The alkenes **1ae–1ai** were prepared according to method C.

Alkenes **1ab** and **1ad** are not reported in literature: The procedure was adapted from the literature report, **1ab** and **1ad** were afforded with 65% and 55% yields, respectively.

(1R,3S,5r)-2-((4-vinylphenoxy)methyl)adamantane (**1ab**)

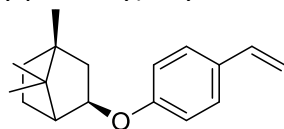


¹H NMR (400 MHz, CDCl₃) δ 7.22 (d, *J* = 8.7 Hz, 2H), 6.75 (d, *J* = 8.7 Hz, 2H), 6.56 (dd, *J* = 17.6, 10.9 Hz, 1H), 5.50 (d, *J* = 10.9 Hz, 1H), 5.01 (d, *J* = 10.9 Hz, 1H), 3.39 (s, 2H), 1.92 (s, 3H), 1.73–1.43 (m, 12H).

¹³C NMR (101 MHz, CDCl₃) δ 159.5, 136.4, 129.9, 127.5, 114.6, 110.9, 78.2, 39.4, 37.4, 33.6, 28.2.

GCMS (*m/z*): calculated for C₁₉H₂₄O [M]⁺: 268.18; found 268.1

(1R,3R)-1,7,7-trimethyl-3-(4-vinylphenoxy)bicyclo[2.2.1]heptane (**1ad**)

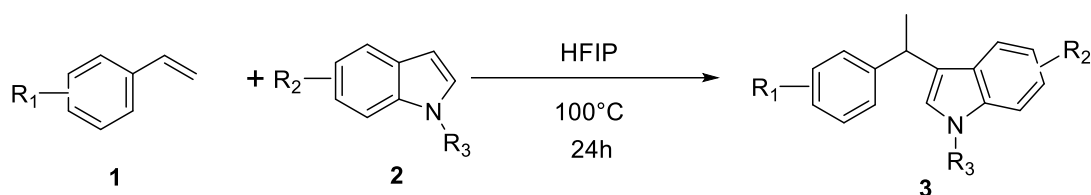


¹H NMR (400 MHz, CDCl₃) δ 7.34 (d, *J* = 8.6 Hz, 2H), 6.81 (d, *J* = 8.6 Hz, 2H), 6.67 (dd, *J* = 17.6, 10.9 Hz, 1H), 5.61 (d, *J* = 17.6 Hz, 1H), 5.12 (d, *J* = 10.9 Hz, 1H), 4.47–4.15 (m, 1H), 2.46–2.31 (m, 1H), 2.29–2.22 (m, 1H), 1.86–1.67 (m, 2H), 1.44–1.20 (m, 2H), 1.12 (dd, *J* = 10.3, 5.1 Hz, 1H), 0.97 (s, 3H), 0.94 (d, *J* = 3.4 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 159.2, 136.3, 129.9, 127.6, 115.4, 111.4, 82.9, 49.7, 47.7, 45.3, 36.9, 28.1, 26.9, 19.8, 19.0, 13.8.

GCMS (*m/z*): calculated for C₁₈H₂₄O [M]⁺: 256.18; found 256.1

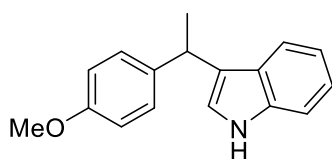
3. General procedure of the hydroarylation



Scheme S2 General procedure of the hydroarylation

A mixture of indoles (0.2 mmol, 1 equiv.) and alkenes (0.24 mmol, 1.2 equiv.) in HFIP (0.3 mL) were sealed in a Schlenk tube without cautious manipulations to exclusion of air or moisture. Then, the reaction mixture was stirred and heated at 100°C for 24h. After full consumption of the starting material, the HFIP was evaporated and the residue was purified by silica gel column chromatography (petroleum ether: ethyl acetate = 10 : 1) to afford the hydroarylation product **3**. The yields were indicated in Table 2 and Table 3.

3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3a**)

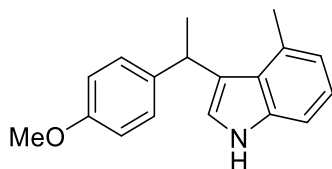


¹H NMR (400 MHz, CDCl₃) δ 7.93 (s, 1H), 7.39 (d, *J* = 8.2 Hz, 1H), 7.34 (d, *J* = 8.1 Hz, 1H), 7.22 (d, *J* = 8.5 Hz, 2H), 7.15 (t, *J* = 8.8 Hz, 1H), 7.06–6.96 (m, 2H), 6.86 (d, *J* = 8.5 Hz, 2H), 4.35 (q, *J* = 7.1 Hz, 1H), 3.85–3.70 (s, 3H), 1.68 (d, *J* = 6.7 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.9, 139.3, 136.7, 128.5, 127.0, 122.1, 121.9, 121.1, 119.9, 119.3, 113.8, 111.1, 55.4, 36.3, 22.7.

HRMS (ESI) (*m/z*): calculated for C₁₇H₁₈NO [M+H]⁺: 252.1383; found 252.1489

3-(1-(4-methoxyphenyl)ethyl)-4-methyl-1H-indole (**3b**)

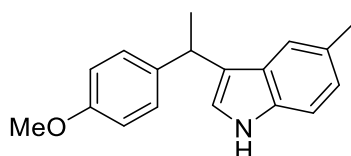


¹H NMR (400 MHz, CDCl₃) δ 7.98 (s, 1H), 7.19 (d, *J* = 10.3 Hz, 1H), 7.16–6.98 (m, 4H), 6.88–6.69 (m, 3H), 4.67 (q, *J* = 7.1 Hz, 1H), 3.79 (s, 3H), 2.48 (s, 3H), 1.66 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.8, 140.6, 137.1, 131.3, 128.5, 125.8, 122.2, 122.0, 121.7, 121.2, 113.8, 109.0, 55.3, 36.9, 24.6, 20.6.

HRMS (ESI) (*m/z*): calculated for C₁₈H₁₉NONa [M+Na]⁺: 288.1359; found 288.1408

3-(1-(4-methoxyphenyl)ethyl)-5-methyl-1H-indole (**3c**)

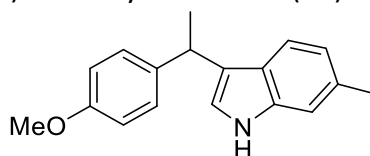


¹H NMR (400 MHz, CDCl₃) δ 7.82 (s, 1H), 7.25–7.13 (m, 4H), 6.97 (d, *J* = 8.2 Hz, 1H), 6.92 (d, *J* = 1.7 Hz, 1H), 6.80 (t, *J* = 9.9 Hz, 2H), 4.29 (q, *J* = 7.1 Hz, 1H), 3.77 (s, 3H), 2.41 (s, 3H), 1.64 (d, *J* = 7.1 Hz, 3H)

¹³C NMR (101 MHz, CDCl₃) δ 157.8, 139.2, 135.1, 128.7, 127.2, 123.7, 121.4, 119.3, 113.8, 110.6, 55.4, 36.4, 22.9, 21.5.

HRMS (ESI) (*m/z*): calculated for C₁₈H₂₀NO [M+H]⁺: 266.1539; found 266.1584

3-(1-(4-methoxyphenyl)ethyl)-6-methyl-1H-indole (**3d**)

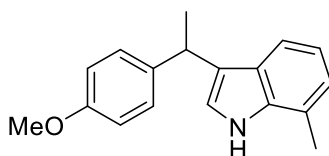


¹H NMR (400 MHz, CDCl₃) δ 7.79 (s, 1H), 7.25–7.15 (m, 4H), 6.89 (t, *J* = 8.5 Hz, 1H), 6.87–6.71 (m, 3H), 4.29 (q, *J* = 7.0 Hz, 1H), 3.76 (s, 3H), 2.40 (s, 3H), 1.64 (d, *J* = 7.3 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.9, 139.2, 137.2, 131.9, 128.3, 124.9, 121.8, 121.1, 120.5, 119.5, 113.8, 111.1, 55.3, 36.4, 22.7, 21.8.

HRMS (ESI) (*m/z*): calculated for C₁₈H₂₀NO [M+H]⁺: 266.1467; found 266.1631

3-(1-(4-methoxyphenyl)ethyl)-7-methyl-1H-indole (**3e**)

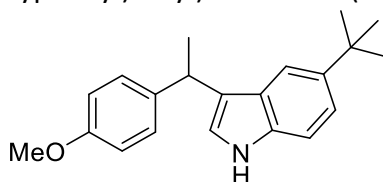


¹H NMR (400 MHz, CDCl₃) δ 7.85 (s, 1H), 7.25–7.15 (m, 4H), 6.98–6.93 (m, 2H), 6.79 (d, *J* = 8.4 Hz, 2H), 4.31 (q, *J* = 7.1 Hz, 1H), 3.76 (s, 3H), 2.46 (s, 3H), 1.65 (d, *J* = 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.9, 139.2, 136.3, 128.4, 126.5, 122.6, 122.5, 120.8, 120.3, 119.5, 117.6, 113.8, 55.4, 36.3, 22.6, 16.7.

HRMS (ESI) (*m/z*): calculated for C₁₈H₂₀NO [M+H]⁺: 266.1467; found 266.1558

5-(tert-butyl)-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3f**)

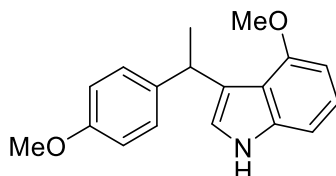


¹H NMR (400 MHz, CDCl₃) δ 7.82 (s, 1H), 7.33 (d, *J* = 1.7 Hz, 1H), 7.26–7.17 (m, 4H), 6.88 (d, *J* = 1.7 Hz, 1H), 6.80 (d, *J* = 8.7 Hz, 2H), 4.31 (q, *J* = 7.1 Hz, 1H), 3.76 (s, 3H), 1.65 (d, *J* = 7.0 Hz, 3H), 1.26 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 157.8, 142.1, 139.1, 134.9, 128.4, 126.7, 122.2, 121.3, 120.3, 115.6, 113.9, 110.5, 55.4, 36.1, 34.6, 32.0, 22.6.

HRMS (ESI) (m/z): calculated for C₂₁H₂₅NONa [M+Na]⁺: 330.1828; found 330.1849

4-methoxy-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3g**)

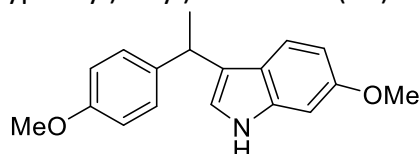


¹H NMR (400 MHz, CDCl₃) δ 8.18 (s, 1H), 7.18 (d, *J* = 9.0 Hz, 2H), 6.99–6.84 (m, 3H), 6.82–6.74 (m, 2H), 6.59 (d, *J* = 8.0 Hz, 1H), 4.29 (q, *J* = 7.1 Hz, 1H), 3.92 (s, 3H), 3.75 (s, 3H), 1.66 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.8, 145.9, 139.2, 128.2, 127.2, 122.3, 120.7, 119.6, 113.7, 112.7, 101.8, 55.3, 36.3, 22.7.

HRMS (ESI) (m/z): calculated for C₁₈H₂₀NO [M+H]⁺: 282.1489; found 282.1436

6-methoxy-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3h**)

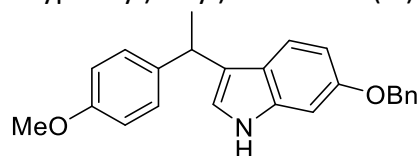


¹H NMR (400 MHz, CDCl₃) δ 7.83 (s, 1H), 7.18 (dd, *J* = 12.2, 9.0 Hz, 3H), 6.92–6.81 (m, 4H), 6.67 (dd, *J* = 8.7, 2.2 Hz, 1H), 4.28 (q, *J* = 7.1 Hz, 1H), 3.81 (s, 3H), 3.78 (s, 3H), 1.65 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.9, 156.7, 139.2, 137.4, 128.5, 122.1, 121.4, 120.5, 119.7, 113.8, 109.2, 94.6, 55.8, 55.3, 36.3, 22.6

HRMS (ESI) (m/z): calculated for C₁₈H₂₀NO₂ [M+H]⁺: 282.1489; found 282.1435

6-(benzyloxy)-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3i**)

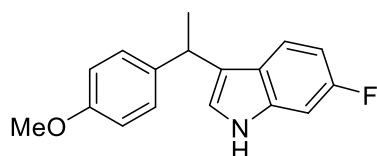


¹H NMR (400 MHz, CDCl₃) δ 7.79 (s, 1H), 7.43 (d, *J* = 6.7 Hz, 2H), 7.37 (dd, *J* = 9.9, 4.7 Hz, 2H), 7.30 (d, *J* = 4.9 Hz, 1H), 7.22–7.16 (m, 3H), 6.87–6.86 (m, 2H), 6.80 (d, *J* = 7.1 Hz, 2H), 6.74 (dd, *J* = 8.8, 2.1 Hz, 1H), 5.05 (d, *J* = 9.2 Hz, 2H), 4.26 (q, *J* = 7.0 Hz, 1H), 3.76 (s, 3H), 1.63 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.7, 155.6, 139.1, 137.5, 137.3, 128.6, 128.3, 127.9, 127.5, 121.8, 121.6, 120.4, 112.0, 113.7, 109.8, 95.9, 70.3, 55.0, 36.3, 23.0.

HRMS (ESI) (m/z): calculated for C₂₄H₂₄NO₂ [M+H]⁺: 358.1802; found 358.1807

6-fluoro-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3j**)



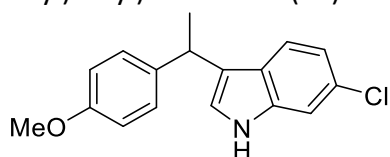
¹H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.31–7.11 (m, 3H), 7.08–6.92 (m, 2H), 6.81 (d, *J* = 8.0 Hz, 2H), 6.7 (td, *J* = 9.2, 2.1 Hz, 1H), 4.28 (q, *J* = 7.0 Hz, 1H), 3.78 (s, 3H), 1.65 (d, *J* = 9.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 161.4, 158.9, 157.9, 138.9, 136.7, 136.6, 128.4, 123.6, 122.0, 121.3, 121.3, 120.7, 120.6, 113.8, 108.2, 107.9, 97.6, 97.2, 55.4, 36.2, 22.7.

¹⁹F NMR (471 MHz, CDCl₃) δ -121.5

HRMS (ESI) (*m/z*): calculated for C₁₇H₁₆FNONa [M+Na]⁺: 292.1108; found 292.1327

6-chloro-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3k**)

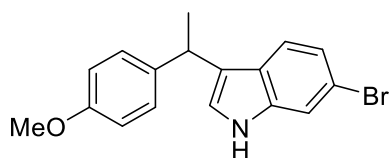


¹H NMR (400 MHz, CDCl₃) δ 7.96 (s, 1H), 7.34 (d, *J* = 1.7 Hz, 1H), 7.25 (d, *J* = 8.5 Hz, 1H), 7.23 – 7.16 (m, 2H), 7.02 (dd, *J* = 2.4, 0.7 Hz, 1H), 7.00 (dd, *J* = 5.4, 1.4 Hz, 1H), 6.88 – 6.82 (m, 2H), 4.31 (q, *J* = 7.1 Hz, 1H), 3.81 (s, 3H), 1.68 (d, *J* = 8.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 158.0, 138.8, 137.2, 128.5, 128.1, 125.7, 122.2, 121.7, 120.7, 119.9, 113.8, 110.9, 55.4, 36.1, 22.6.

HRMS (ESI) (*m/z*): calculated for C₁₇H₁₆ClNOK [M+K]⁺: 324.0552; found 324.0595

6-bromo-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3l**)

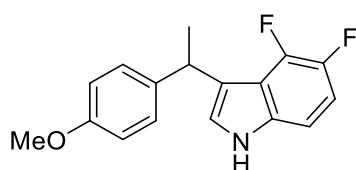


¹H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.49 (d, *J* = 11.4 Hz, 1H), 7.23–7.13 (m, 3H), 7.07 (dd, *J* = 16.2, 6.9 Hz, 1H), 6.98 (d, *J* = 1.2 Hz, 1H), 6.84 (t, *J* = 10.9 Hz, 2H), 4.29 (q, *J* = 7.1 Hz, 1H), 3.78 (s, 3H), 1.66 (d, *J* = 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 156.5, 137.6, 136.4, 127.3, 124.8, 121.6, 121.0, 120.6, 120.0, 114.5, 112.9, 112.7, 54.2, 34.9, 21.5.

HRMS (ESI) (*m/z*): calculated for C₁₇H₁₇BrNO [M+H]⁺: 330.0488; found 330.0496

4,5-difluoro-3-(1-(4-methoxyphenyl)ethyl)-1H-indole (**3m**)



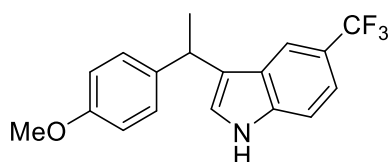
¹H NMR (400 MHz, CDCl₃) δ 7.98 (s, 1H), 7.22 (d, *J* = 8.6 Hz, 2H), 6.97-6.93 (m, 3H), 6.84 (d, *J* = 8.6 Hz, 2H), 4.51 (q, *J* = 7.1 Hz, 1H), 3.79 (s, 3H), 1.66 (d, *J* = 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.8, 145.3, 143.1, 138.9, 135.1, 134.8, 128.2, 122.5, 121.8, 116.7, 113.8, 112.1, 111.6, 106.1, 55.2, 36.3, 22.9.

¹⁹F NMR (471 MHz, CDCl₃) δ -148.2, -158.8.

GCMS (m/z): calculated for C₁₇H₁₅F₂NO [M]⁺: 287.11; found 287.1

3-(1-(4-methoxyphenyl)ethyl)-5-(trifluoromethyl)-1H-indole (**3n**)



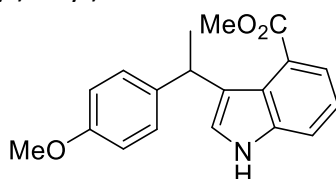
¹H NMR (400 MHz, CDCl₃) δ 8.17 (s, 1H), 7.62 (s, 1H), 7.40 (d, *J* = 11.1 Hz, 1H), 7.25–7.13 (m, 4H), 6.83 (dd, *J* = 6.8, 4.8 Hz, 2H), 4.33 (q, *J* = 7.1 Hz, 1H), 3.78 (s, 3H), 1.67 (d, *J* = 10.5 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 158.0, 138.6, 135.6, 128.4, 123.7, 122.3, 120.3, 113.9, 55.4, 36.1, 22.6.

¹⁹F NMR (471 MHz, CDCl₃) δ -60.6

HRMS (ESI) (m/z): calculated for C₁₈H₁₆F₃NOK [M+K]⁺: 358.0816; found 358.0844

methyl 3-(1-(4-methoxyphenyl)ethyl)-1H-indole-4-carboxylate (**3o**)

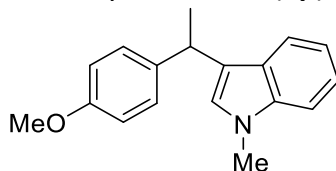


¹H NMR (400 MHz, CDCl₃) δ 7.86 (d, *J* = 7.5 Hz, 2H), 7.39 (d, *J* = 8.5 Hz, 1H), 7.17–7.12 (m, 3H), 7.08 (s, 1H), 6.85 (t, *d* = 9.6 Hz, 2H), 4.24 (q, *J* = 7.2 Hz, 1H), 3.99 (s, 3H), 3.78 (s, 3H), 1.73 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 168.3, 158.6, 146.2, 137.2, 136.2, 128.7, 128.4, 123.5, 120.9, 120.7, 115.4, 114.3, 100.7, 55.6, 51.6, 38.4, 21.2.

HRMS (ESI) (m/z): calculated for C₁₉H₂₀NO₃ [M+H]⁺: 310.1438; found 310.1440

3-(1-(4-methoxyphenyl)ethyl)-1-methyl-1H-indole (**3p**)

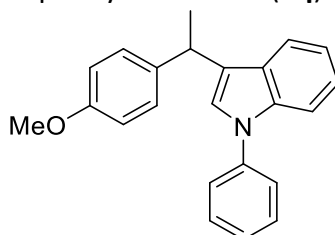


¹H NMR (400 MHz, CDCl₃) δ 7.36 (d, *J* = 7.9 Hz, 1H), 7.26 (d, *J* = 8.2 Hz, 1H), 7.24–7.11 (m, 3H), 6.99 (t, *J* = 7.5 Hz, 1H), 6.87–6.75 (m, 3H), 4.32 (q, *J* = 7.0 Hz, 1H), 3.74 (d, *J* = 11.9 Hz, 6H), 1.66 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.8, 139.4, 137.5, 128.4, 127.3, 125.9, 121.6, 120.4, 119.9, 118.7, 113.8, 109.2, 55.3, 36.3, 32.7, 22.8.

HRMS (ESI) (*m/z*): calculated for C₁₈H₂₀NO [M+H]⁺: 266.1539; found 266.1527

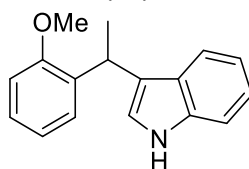
3-(1-(4-methoxyphenyl)ethyl)-1-phenyl-1H-indole (**3q**)



¹H NMR (400 MHz, CDCl₃) δ 7.54 (d, *J* = 6.7 Hz, 1H), 7.52–7.47 (m, 4H), 7.41 (d, *J* = 7.9 Hz, 1H), 7.37–7.29 (m, 1H), 7.28–7.22 (m, 2H), 7.21–7.11 (m, 2H), 7.09–7.01 (t, *J* = 6.9 Hz, 1H), 6.86–6.79 (m, 2H), 4.37 (q, *J* = 7.1 Hz, 1H), 3.76 (s, 3H), 1.71 (d, *J* = 7.1 Hz, 3H).
¹³C NMR (101 MHz, CDCl₃) δ 157.8, 139.98, 138.78, 136.4, 129.5, 128.4, 126.0, 125.0, 124.1, 123.0, 122.5, 120.2, 119.7, 113.7, 110.4, 55.2, 36.2, 22.5.

HRMS (ESI) (*m/z*): calculated for C₂₃H₂₂NO [M+H]⁺: 328.1696; found 328.1681.

3-(1-(2-methoxyphenyl)ethyl)-1H-indole (**3r**)

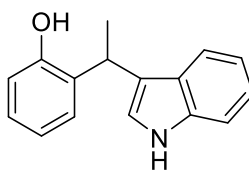


¹H NMR (400 MHz, CDCl₃) δ 7.93 (s, 1H), 7.35 (dd, *J* = 13.4, 8.1 Hz, 2H), 7.19–7.08 (m, 2H), 7.05–7.02 (m, 2H), 6.99 (t, *J* = 7.4 Hz, 1H), 6.88 (d, *J* = 11.1 Hz, 1H), 6.80 (t, *J* = 7.4 Hz, 1H), 4.84 (q, *J* = 6.9 Hz, 1H), 3.90 (s, 3H), 1.63 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 156.8, 136.7, 135.2, 128.0, 127.2, 126.9, 122.0, 121.6, 121.4, 120.7, 119.9, 119.2, 110.9, 110.5, 55.6, 29.2, 21.1.

HRMS (ESI) (*m/z*): calculated for C₁₇H₁₈NO [M+H]⁺: 252.1382; found 252.1241

2-(1-(1H-indol-3-yl)ethyl)phenol (**3s**)

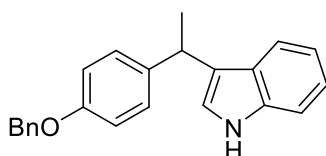


¹H NMR (400 MHz, CDCl₃) δ 8.05 (s, 1H), 7.39 (d, *J* = 7.0 Hz, 1H), 7.35 (d, *J* = 8.2 Hz, 1H), 7.31 (d, *J* = 7.6 Hz, 1H), 7.21 (d, *J* = 7.3 Hz, 1H), 7.18–7.12 (m, 1H), 7.05–7.02 (m, 2H), 6.95 (t, *J* = 7.5 Hz, 1H), 6.79 (d, *J* = 8.0 Hz, 1H), 5.27 (s, 1H), 4.54 (q, *J* = 7.1 Hz, 1H), 1.75 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 154.1, 136.9, 131.4, 128.4, 127.5, 126.6, 122.5, 121.4, 120.9, 119.7, 119.5, 119.4, 116.3, 111.3, 32.3, 20.2.

HRMS (ESI) (*m/z*): calculated for C₁₆H₁₆NO [M+H]⁺: 238.1232; found 238.1239

3-(1-(4-(benzyloxy)phenyl)ethyl)-1H-indole (**3t**)

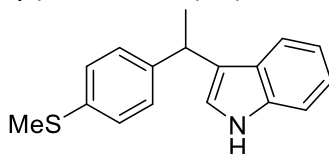


¹H NMR (400 MHz, CDCl₃) δ 7.96 (s, 1H), 7.49–7.31 (m, 7H), 7.22 (d, *J* = 8.6 Hz, 2H), 7.16 (dt, *J* = 11.6, 2.2 Hz, 1H), 7.06–6.98 (m, 2H), 6.94–6.87 (m, 2H), 5.04 (s, 2H), 4.34 (q, *J* = 7.1 Hz, 1H), 1.67 (dd, *J* = 18.0, 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.2, 139.6, 137.2, 136.7, 128.6, 128.4, 127.9, 127.6, 126.9, 121.9, 121.8, 121.0, 119.8, 119.2, 114.6, 111.0, 70.2, 36.3, 22.5.

GCMS (*m/z*): calculated for C₂₃H₂₁NO [M]⁺: 327.16; found 327.1

3-(1-(4-(methylthio)phenyl)ethyl)-1H-indole (**3u**)

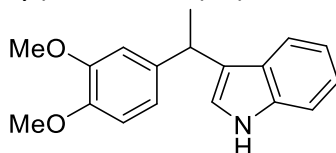


¹H NMR (400 MHz, CDCl₃) δ 7.95 (s, 1H), 7.34 (dd, *J* = 7.8, 4.5 Hz, 2H), 7.28–7.07 (m, 5H), 7.06–6.91 (m, 2H), 4.33 (q, *J* = 7.1 Hz, 1H), 2.44 (s, 3H), 1.67 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 143.9, 136.6, 135.2, 128.0, 127.0, 126.7, 122.0, 121.3, 121.0, 119.7, 119.3, 111.1, 36.5, 22.3, 16.2.

GCMS (*m/z*): calculated for C₁₇H₁₇NS [M]⁺: 267.11; found 267.1

3-(1-(3,4-dimethoxyphenyl)ethyl)-1H-indole (**3v**)

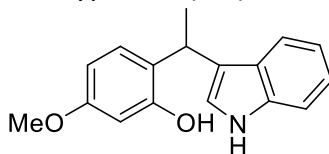


¹H NMR (400 MHz, CDCl₃) δ 8.00 (s, 1H), 7.39 (d, *J* = 9.4 Hz, 1H), 7.34 (d, *J* = 8.1 Hz, 1H), 7.16 (t, *J* = 7.4 Hz, 1H), 7.02 (t, *J* = 11.8 Hz, 1H), 6.98 (d, *J* = 1.7 Hz, 1H), 6.86–6.85 (m, 2H), 6.80 (d, *J* = 8.8 Hz, 1H), 4.34 (q, *J* = 7.1 Hz, 1H), 3.84 (s, 3H), 3.82 (s, 3H), 1.71 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 148.7, 147.2, 139.7, 136.6, 127.0, 122.0, 121.7, 121.1, 119.8, 119.2, 111.1, 111.1, 111.0, 55.9, 36.6, 22.6.

HRMS (ESI) (*m/z*): calculated for C₁₈H₂₀NO₂ [M+H]⁺: 282.1489; found 282.1469

2-(1-(1H-indol-3-yl)ethyl)-5-methoxyphenol (**3w**)

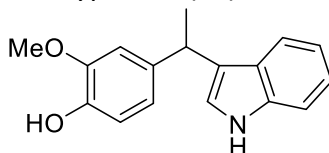


¹H NMR (400 MHz, CDCl₃) δ 8.07 (s, 1H), 7.37 (t, *J* = 7.8 Hz, 2H), 7.19 (t, *J* = 8.2 Hz, 2H), 7.07 (d, *J* = 1.3 Hz, 1H), 7.02 (t, *J* = 7.6 Hz, 1H), 6.49 (dt, *J* = 12.7, 6.3 Hz, 1H), 6.36 (d, *J* = 2.5 Hz, 1H), 5.23 (s, 1H), 4.43 (q, *J* = 7.1 Hz, 1H), 3.76 (s, 3H), 1.68 (d, *J* = 6.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 159.1, 154.9, 136.9, 128.7, 126.4, 123.6, 122.4, 121.0, 119.4, 110.9, 106.1, 102.4, 54.9, 31.4, 20.4.

GCMS (m/z): calculated for C₁₇H₁₇NO₂ [M]⁺: 267.13; found 267.1

4-(1-(1H-indol-3-yl)ethyl)-2-methoxyphenol (**3x**)

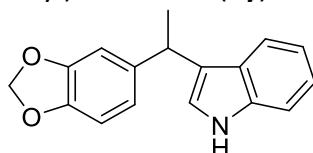


¹H NMR (400 MHz, CDCl₃) δ 7.96 (s, 1H), 7.40 (d, *J* = 10.4 Hz, 1H), 7.34 (d, *J* = 8.1 Hz, 1H), 7.17 (t, *J* = 7.6 Hz, 1H), 7.03 (t, *J* = 7.2 Hz, 1H), 6.98 (d, *J* = 2.5 Hz, 1H), 6.88–6.76 (m, 3H), 5.40 (s, 1H), 4.32 (q, *J* = 7.0 Hz, 1H), 3.81 (s, 3H), 1.70 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 146.3, 143.7, 138.9, 136.7, 126.8, 121.9, 121.8, 121.0, 120.0, 119.7, 119.2, 114.1, 111.0, 110.0, 55.9, 36.7, 22.5.

The structure was confirmed by comparison of the NMR spectra with the literature.¹

3-(1-(benzo[d][1,3]dioxol-5-yl)ethyl)-1H-indole (**3y**)

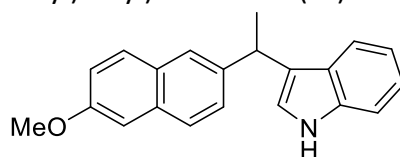


¹H NMR (400 MHz, CDCl₃) δ 7.96 (s, 1H), 7.37 (d, *J* = 7.6 Hz, 1H), 7.32 (d, *J* = 7.6 Hz, 1H), 7.16 (t, *J* = 7.6 Hz, 1H), 7.08–6.94 (m, 2H), 6.80 (d, *J* = 8.0 Hz, 1H), 6.74–6.71 (m, 2H), 5.88 (d, *J* = 7.8 Hz, 2H), 4.31 (q, *J* = 7.1 Hz, 1H), 1.66 (t, *J* = 7.6 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 147.6, 145.7, 141.2, 136.7, 127.0, 122.1, 121.6, 121.0, 120.3, 119.7, 119.3, 111.1, 108.1, 100.7, 36.8, 22.8.

GCMS (ESI) (m/z): calculated for C₁₇H₁₅NO₂ [M]⁺: 265.11; found 265.1

3-(1-(6-methoxynaphthalen-2-yl)ethyl)-1H-indole (**3z**)

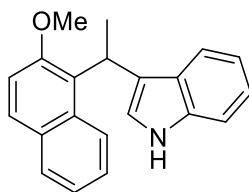


¹H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.71–7.58 (m, 3H), 7.35 (dd, *J* = 18.7, 8.1 Hz, 3H), 7.37–7.33 (m, 3H), 7.01 (d, *J* = 1.6 Hz, 1H), 6.96 (t, *J* = 7.5 Hz, 1H), 4.50 (q, *J* = 7.1 Hz, 1H), 3.89 (s, 3H), 1.72 (d, *J* = 6.9 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.3, 142.2, 136.7, 133.1, 129.3, 129.3, 127.2, 127.0, 126.9, 125.3, 122.1, 121.6, 121.3, 119.8, 119.3, 118.6, 111.1, 105.7, 55.4, 36.8, 22.3.

The structure was confirmed by comparison of the NMR spectra with the literature.¹

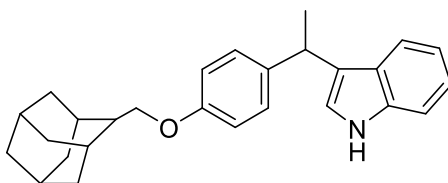
3-(1-(2-methoxynaphthalen-1-yl)ethyl)-1H-indole (**3aa**)



¹H NMR (400 MHz, CDCl₃) δ 8.23 (d, *J* = 9.0 Hz, 1H), 7.93 (s, 1H), 7.75 (d, *J* = 9.1 Hz, 2H), 7.35–7.21 (m, 4H), 7.17 (d, *J* = 8.0 Hz, 1H), 7.14–7.10 (m, 1H), 7.05 (t, *J* = 7.2 Hz, 1H), 6.84 (t, *J* = 7.2 Hz, 1H), 5.48 (q, *J* = 6.7 Hz, 1H), 3.92 (s, 3H), 1.93 (d, *J* = 13.4, 7.2 Hz, 3H).
¹³C NMR (101 MHz, CDCl₃) δ 157.7, 146.1, 139.1, 128.5, 128.0, 127.2, 122.3, 120.6, 119.6, 113.8, 112.5, 101.8, 55.4, 36.3, 22.6.

HRMS (ESI) (*m/z*): calculated for C₂₁H₂₀NO [M+H]⁺: 302.1539; found 302.1529

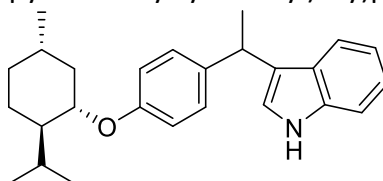
3-(1-(4-(((1R,3S,5r)-adamantan-2-yl)methoxy)phenyl)ethyl)-1H-indole (**3ab**)



¹H NMR (400 MHz, CDCl₃) δ 7.93 (s, 1H), 7.37 (d, *J* = 7.2 Hz, 1H), 7.32 (d, *J* = 9.3 Hz, 1H), 7.22–7.11 (m, 3H), 6.99–6.96 (m, 2H), 6.81 (d, *J* = 9.5 Hz, 2H), 4.33 (q, *J* = 7.1 Hz, 1H), 3.44 (s, 2H), 2.07–1.93 (m, 4H), 1.80–1.61 (m, 14H).
¹³C NMR (101 MHz, CDCl₃) δ 157.9, 138.5, 136.7, 128.4, 126.9, 121.9, 121.0, 119.8, 119.2, 114.2, 110.9, 78.3, 39.5, 37.2, 28.3.

HRMS (ESI) (*m/z*): calculated for C₂₇H₃₁NNaO [M+Na]⁺: 408.2298; found 408.2279

3-(1-(4-(((1S,2R,5S)-2-isopropyl-5-methylcyclohexyl)oxy)phenyl)ethyl)-1H-indole (**3ac**)

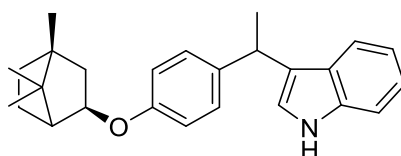


¹H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.40 (d, *J* = 8.0 Hz, 1H), 7.34 (d, *J* = 8.1 Hz, 1H), 7.23–7.11 (m, 3H), 7.05–6.95 (m, 2H), 6.80 (d, *J* = 8.5 Hz, 2H), 4.31 (q, *J* = 7.0 Hz, 1H), 3.96 (td, *J* = 10.5, 4.0 Hz, 1H), 2.23–2.13 (m, 2H), 1.71–1.66 (m, 4H), 1.53–0.82 (m, 12H), 0.75 (dd, *J* = 7.8, 2.0 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 156.5, 138.8, 136.8, 128.3, 122.0, 121.1, 119.9, 119.0, 115.8, 111.1, 48.2, 40.6, 36.1, 34.7, 31.6, 26.0, 23.9, 22.7, 22.1, 20.9, 16.6.

HRMS (ESI) (*m/z*): calculated for C₂₆H₃₃NONa [M+Na]⁺: 398.2454; found 398.2471

3-(1-(4-(((2R,4R)-4,7,7-trimethylbicyclo[2.2.1]heptan-2-yl)oxy)phenyl)ethyl)-1H-indole (**3ad**)

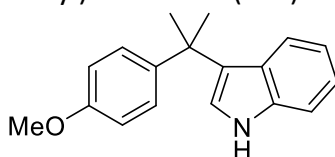


¹H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.40 (d, *J* = 7.8 Hz, 1H), 7.34 (d, *J* = 8.1 Hz, 1H), 7.17–7.14 (m, 3H), 7.04–6.97 (m, 2H), 6.74 (d, *J* = 8.6 Hz, 2H), 4.42–4.13 (m, 2H), 2.35–2.19 (m, 2H), 1.79–1.70 (m, 2H), 1.67 (d, *J* = 7.1 Hz, 3H), 1.35–1.32 (m, 2H), 1.15–1.05 (m, 1H), 0.90 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 157.4, 138.5, 136.7, 128.2, 127.0, 122.0, 121.0, 119.9, 119.1, 115.4, 111.1, 82.9, 49.3, 47.4, 45.0, 36.8, 36.0, 27.9, 26.7, 22.9, 19.8, 19.0, 13.6.

HRMS (ESI) (*m/z*): calculated for C₂₀H₃₁NNaO₂ [*M*+Na]⁺: 396.2370; found 396.2367

3-(2-(4-methoxyphenyl)propan-2-yl)-1H-indole (**3ae**)



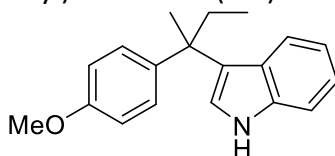
¹H NMR (400 MHz, CDCl₃) δ 7.89 (s, 1H), 7.31 (d, *J* = 8.1 Hz, 1H), 7.28–7.20 (m, 2H), 7.11–7.06 (m, 3H), 6.88 (td, *J* = 8.2, 3.5 Hz, 1H), 6.82–6.73 (m, 2H), 3.76 (s, 3H), 1.74 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 157.4, 142.3, 137.2, 127.5, 126.5, 126.0, 121.7, 121.5, 120.6, 118.9, 113.4, 111.2, 55.2, 38.3, 30.8.

HRMS (ESI) (*m/z*): calculated for C₁₈H₁₉NONa [*M*+Na]⁺: 288.1583; found 255.1581

The structure was confirmed by comparison of the NMR spectra with the literature.¹

3-(2-(4-methoxyphenyl)butan-2-yl)-1H-indole (**3af**)

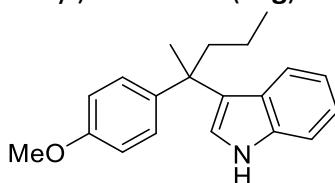


¹H NMR (400 MHz, CDCl₃) δ 7.89 (s, 1H), 7.27 (d, *J* = 7.8 Hz, 1H), 7.18 (d, *J* = 8.4 Hz, 2H), 7.10–7.05 (m, 2H), 7.01 (d, *J* = 8.0 Hz, 1H), 6.85 (dd, *J* = 7.9, 7.2 Hz, 1H), 6.77 (d, *J* = 7.8 Hz, 2H), 3.75 (s, 3H), 2.33–2.18 (m, 1H), 2.18–2.05 (m, 1H), 1.64 (s, 3H), 0.72 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.9, 139.3, 136.7, 128.5, 127.0, 122.1, 121.9, 121.1, 119.9, 119.3, 113.8, 111.1, 55.4, 36.3, 22.7.

HRMS (ESI) (*m/z*): calculated for C₁₉H₂₁NO [*M*+H]⁺: 280.1701; found 280.1671

3-(2-(4-methoxyphenyl)pentan-2-yl)-1H-indole (**3ag**)

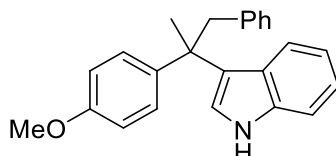


¹H NMR (400 MHz, CDCl₃) δ 7.90 (s, 1H), 7.31 (d, *J* = 8.2 Hz, 1H), 7.24–7.16 (m, 2H), 7.09–7.07 (m, 2H), 7.02 (d, *J* = 8.0 Hz, 1H), 6.85 (t, *J* = 7.5 Hz, 1H), 6.78 (d, *J* = 7.6 Hz, 2H), 3.76 (s, 3H), 2.24–2.11 (m, 1H), 2.11–1.97 (m, 1H), 1.67 (s, 3H), 1.20–1.00 (m, 2H), 0.85 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.3, 141.4, 137.0, 127.9, 126.1, 125.3, 121.6, 121.5, 121.2, 118.8, 113.1, 111.0, 55.2, 44.2, 41.7, 27.8, 18.0, 14.9.

HRMS (ESI) (*m/z*): calculated for C₂₀H₂₄NO [M+H]⁺: 294.1952; found 294.1951

3-(2-(4-methoxyphenyl)-1-phenylpropan-2-yl)-1H-indole (**3ah**)

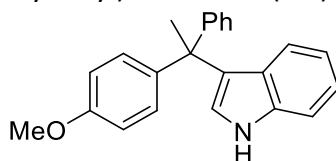


¹H NMR (400 MHz, CDCl₃) δ 7.92 (s, 1H), 7.37 (d, *J* = 8.1 Hz, 1H), 7.23–7.00 (m, 7H), 6.96 (d, *J* = 2.2 Hz, 1H), 6.91 (t, *J* = 7.5 Hz, 1H), 6.79 (d, *J* = 8.7 Hz, 2H), 6.60 (d, *J* = 7.2 Hz, 2H), 3.80 (s, 3H), 3.47 (d, *J* = 12.4 Hz, 1H), 3.41 (d, *J* = 12.4 Hz, 1H), 1.58 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.4, 140.1, 138.3, 137.0, 130.8, 128.2, 127.1, 126.2, 125.8, 124.3, 121.6, 121.5, 121.3, 119.0, 113.1, 111.1, 55.1, 46.9, 42.6, 27.2.

HRMS (ESI) (*m/z*): calculated for C₂₄H₂₄NO [M+H]⁺: 342.1852; found 342.1848

3-(1-(4-methoxyphenyl)-1-phenylethyl)-1H-indole (**3ai**)

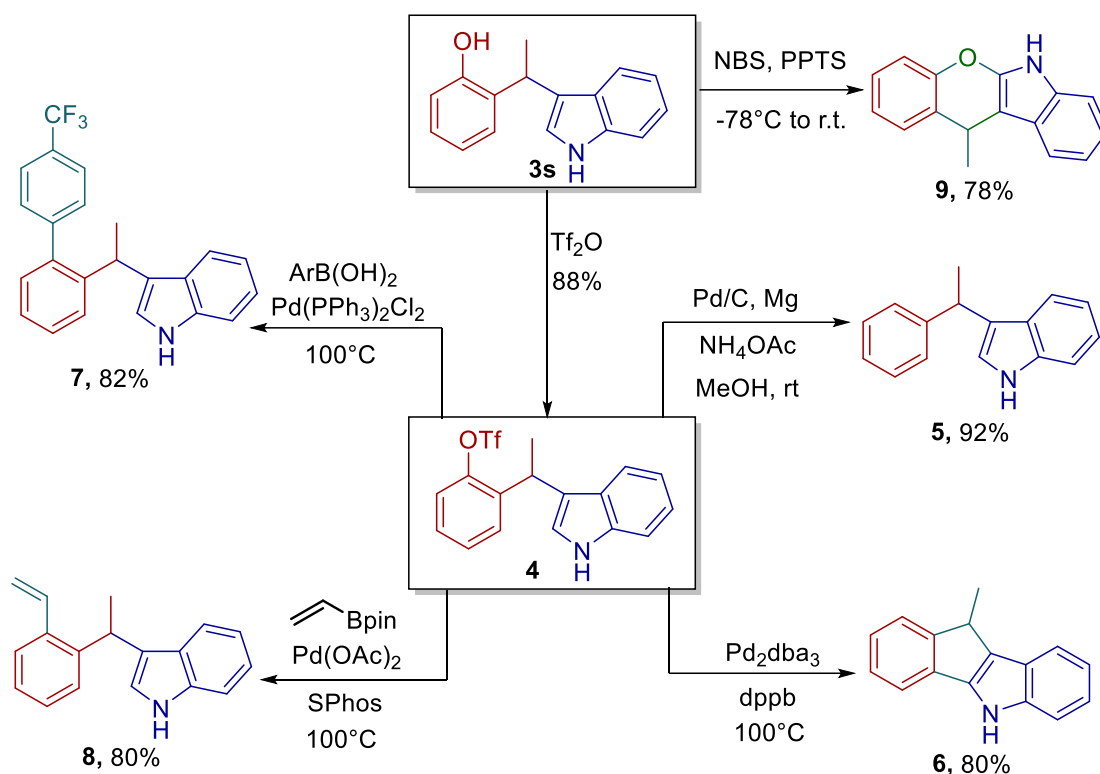


¹H NMR (400 MHz, CDCl₃) δ 7.77 (s, 1H), 7.22 (d, *J* = 10.8 Hz, 1H), 7.18–7.14 (m, 3H), 7.14–7.02 (m, 4H), 6.85 (t, *J* = 7.5 Hz, 1H), 6.71 (d, *J* = 8.0 Hz, 2H), 6.33 (d, *J* = 2.4 Hz, 2H), 3.69 (s, 3H), 2.15 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 157.6, 148.8, 140.8, 137.2, 129.4, 128.4, 127.9, 126.4, 125.8, 123.7, 122.2, 121.8, 119.1, 113.2, 111.2, 55.1, 47.4, 29.4.

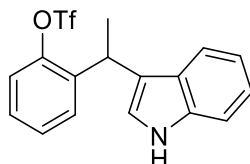
GC-MS (*m/z*): calculated for C₂₃H₂₁NO [M]⁺: 327.16; found 327.1

4. Synthetic transformations



Scheme S3 Synthetic transformations

2-(1-(1H-indol-3-yl)ethyl)phenyl trifluoromethanesulfonate (**4**)



In a dry two neck flask 239 mg (1 mmol, 1.0 equiv) of **3s** was dissolved in 15 mL DCM. 560 μ L of Et_3N (4 mmol, 4.0 equiv) was added to the reaction mixture at -20°C . At this temperature 200 μ L (1.2 mmol, 1.2 equiv) of Tf_2O was added drop wise. The reaction mixture was allowed to stir for 30 min at -20°C and slowly warmed to room temperature. The reaction was followed by TLC. After full consumption of starting material, it was quenched with saturated NH_4Cl aq. and extracted with DCM for 3 times. The combined organic layers were collected and dried over Na_2SO_4 , and concentrated under reduced pressure. The combined organic layers were collected and dried over Na_2SO_4 , and concentrated under reduced pressure. The residue was purified by silica column chromatography (PE:EtOAc = 10:1) to afford the product **4** (324 mg) in 88% yield.

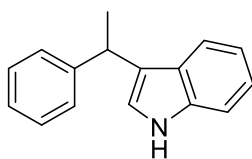
$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.97 (s, 1H), 7.24–7.20 (m, 3H), 7.18–7.03 (m, 4H), 6.99 (d, $J = 1.8$ Hz, 1H), 6.96–6.86 (m, 1H), 4.66 (q, $J = 7.0$ Hz, 1H), 1.61 (d, $J = 7.1$ Hz, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 147.5, 139.6, 136.7, 129.9, 128.6, 127.8, 126.6, 122.4, 121.8, 121.2, 119.6, 119.5, 119.3, 111.2, 30.6, 20.7.

$^{19}\text{F NMR}$ (471 MHz, CDCl_3) δ -73.8

HRMS (ESI) (m/z): calculated for $\text{C}_{17}\text{H}_{14}\text{F}_3\text{NNaO}_3\text{S}$ [$\text{M}+\text{Na}$] $^+$: 392.0539; found 392.0545

3-(1-phenylethyl)-1H-indole (**5**)



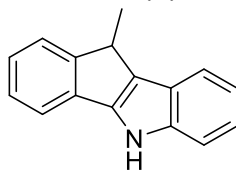
The reaction procedure was adapted from the previous literature²: Methanol (2 mL) was added to the mixture of compound **4** (37 mg, 0.1 mmol, 1 equiv.), magnesium powder (24 mg, 1 mmol, 10 equiv.), 10 % Pd/C (3.7 mg) and ammonium acetate (246 mg, 3.2 mmol, 32 equiv.). The reaction mixture was stirred at room temperature overnight. After evaporation of the methanol, the residue was purified by silica column chromatography (PE:EtOAc = 10:1) to afford the product **5** (20.3 mg) in 92% yield .

¹H NMR (400 MHz, CDCl₃) δ 7.99 (s, 1H), 7.27 (d, *J* = 8.2 Hz, 1H), 7.24–7.19 (m, 2H), 7.19–7.05 (m, 5H), 7.04 (d, *J* = 1.4 Hz, 1H), 6.93 (t, *J* = 7.4 Hz, 1H), 4.67 (q, *J* = 7.1 Hz, 1H), 1.63 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 147.4, 139.6, 136.7, 129.8, 128.7, 127.8, 126.5, 122.3, 121.7, 121.1, 119.6, 119.5, 119.3, 111.1, 30.4, 21.0.

HRMS (ESI) (*m/z*): calculated for C₁₆H₁₅N [M]⁺: 221.1205; found 222.1214

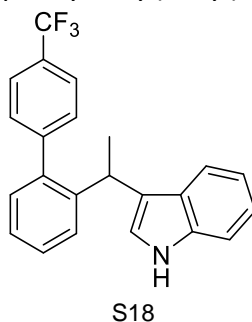
10-methyl-5,10-dihydroindeno[1,2-*b*]indole (**6**)



The reaction procedure was adapted from the previous literature: In a flame-dried Schlenk flask under N₂ and equipped with a magnetic stir bar, compound **4** (65 mg, 0.18 mmol, 1 equiv.), potassium carbonate (100 mg, 1.12 mmol, 4 equiv.), Pd₂(dba)₃ (16 mg, 0.018 mmol, 0.1 equiv.) and dppb (15mg, 0.036 mmol, 0.2 equiv.) were dissolved in DMA (1.0 mL). Then the reaction mixture was heated at 100 °C. 15 h later, the mixture was cooled to r.t., it was quenched with saturated NH₄Cl and extracted with DCM for 3 times. The combined organic layers were collected and dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica column chromatography (hexane:EtOAc = 10:1) to afford desired product **6** (34 mg) in 86 % yield.

The structure was confirmed by comparison of the NMR spectra with the literature.³

3-(1-(4'-(trifluoromethyl)-[1,1'-biphenyl]-2-yl)ethyl)-1H-indole (**7**)



In a flame-dried Schlenk flask under N₂ and equipped with a magnetic stir bar, compound **4** (104 mg, 0.28 mmol, 1 equiv.), 4-trifluoromethylphenylboronic acid (65 mg, 0.34 mmol, 1.2 equiv.), potassium carbonate (156 mg, 1.12 mmol, 4 equiv.), Pd(PPh₃)₂Cl₂ (10 mg, 0.014 mmol, 0.05 equiv.) were dissolved in a mixture solvent of 1,4-dioxane (0.8 mL) and degassed water (0.2 mL). Then the reaction mixture was heated at 100 °C. 15 h later, the mixture was cooled to r.t., it was quenched with saturated NH₄Cl and extracted with DCM for 3 times. The combined organic layers were collected and dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica column chromatography (hexane:EtOAc = 10:1) to afford desired product **7** (85mg) in 82 % yield.

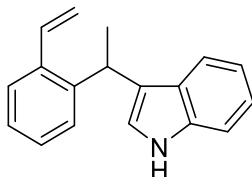
¹H NMR (400 MHz, CDCl₃) δ 7.94 (s, 1H), 7.65 (d, *J* = 8.0 Hz, 2H), 7.50 (d, *J* = 8.0 Hz, 2H), 7.38–7.18 (m, 5H), 7.13 (dd, *J* = 14.0, 7.3 Hz, 2H), 6.96 (t, *J* = 7.4 Hz, 2H), 4.39 (q, *J* = 6.9 Hz, 1H), 1.59 (t, *J* = 8.8 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 145.7, 144.0, 139.7, 136.7, 129.8, 128.4, 127.8, 126.5, 125.9, 125.0, 121.9, 121.6, 121.5, 119.5, 119.2, 111.2, 32.9, 22.7.

¹⁹F NMR (471 MHz, CDCl₃) δ -62.3

HRMS (ESI) (*m/z*): calculated for C₂₃H₁₉F₃N [M+H]⁺: 366.1464; found 366.1458

3-(1-(2-vinylphenyl)ethyl)-1H-indole (**8**)



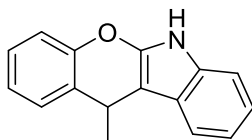
In a flame-dried Schlenk flask under N₂ and equipped with a magnetic stir bar, compound **4** (86 mg, 0.233 mmol, 1 equiv.), pinacol vinylboronate (79 μL, 0.466 mmol, 2 equiv.), potassium carbonate (128 mg, 0.932 mmol, 4 equiv.), Pd(OAc)₂ (3 mg, 0.011 mmol, 0.05 equiv.) and Sphos (9.6 mg, 0.0233 mmol, 0.1 equiv.) were dissolved in a mixture solvent of THF (0.8 mL) and degassed water (0.2 mL). Then the reaction mixture was heated at 100 °C. 15 h later, the mixture was cooled to r.t., it was quenched with saturated NH₄Cl and extracted with DCM for 3 times. The combined organic layers were collected and dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica column chromatography (hexane:EtOAc = 10:1) to afford desired product **8** (46 mg) in 80 % yield.

¹H NMR (400 MHz, CDCl₃) δ 7.96 (s, 1H), 7.54 (d, *J* = 8.1 Hz, 1H), 7.35 (dd, *J* = 15.9, 8.1 Hz, 2H), 7.28–7.13 (m, 5H), 7.03 (t, *J* = 8.1 Hz, 1H), 6.97 (d, *J* = 1.4 Hz, 1H), 5.70 (dd, *J* = 17.3, 1.5 Hz, 1H), 5.34 (dd, *J* = 10.9, 1.5 Hz, 1H), 4.72 (q, *J* = 7.0 Hz, 1H), 1.70 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 143.6, 136.8, 136.3, 135.1, 128.0, 127.2, 126.9, 126.2, 122.2, 121.5, 121.5, 119.7, 119.4, 116.2, 111.1, 32.6, 21.9.

HRMS (ESI) (*m/z*): calculated for C₁₈H₁₈N [M+H]⁺: 248.1434; found 248.1432

11-methyl-6,11-dihydrochromeno[2,3-*b*]indole (**9**)



The reaction procedure was adapted from the previous literature: In a dry two neck flask 37 mg (0.1 mmol, 1.0 equiv) of **3s** was dissolved in 3 mL DCM. PPTS (28mg, 0.11 mmol, 1.1 equiv) was added to the reaction mixture was cooled to -78 °C. At this temperature NBS (20 mg, 0.11 mmol, 1.1 equiv) in THF (2 mL) was added drop wise. The reaction mixture was allowed to stir for 1h at -78 °C. The reaction was followed by TLC. After full consumption of starting material, it was quenched with saturated NH₄Cl aq. and extracted with DCM for 3 times. The combined organic layers were collected and dried over Na₂SO₄, and concentrated under reduced pressure. The combined organic layers were collected and dried over Na₂SO₄, and concentrated under reduced pressure. The residue was purified by silica column chromatography (PE:EtOAc = 10:1) to afford the product **9** (18 mg) in 80% yield .

The structure was confirmed by comparison of the NMR spectra with the literature.³

5. Gram scale reaction

A mixture of indoles (10 mmol, 1 equiv.) and alkenes **1a** and **1s** (12 mmol, 1.2 equiv.) in HFIP (15 mL) were sealed in a Schlenk tube without cautious manipulations to exclusion of air or moisture. Then, the reaction mixture was stirred and heated at 100°C for 24h. After full consumption of the starting material, the HFIP was evaporated and the residue was purified by silica gel column chromatography (petroleum ether: ethyl acetate = 10 : 1) to afford the hydroarylation products **3a** 1.79g and **3s** 1.89g in 71% and 80% yields, respectively.

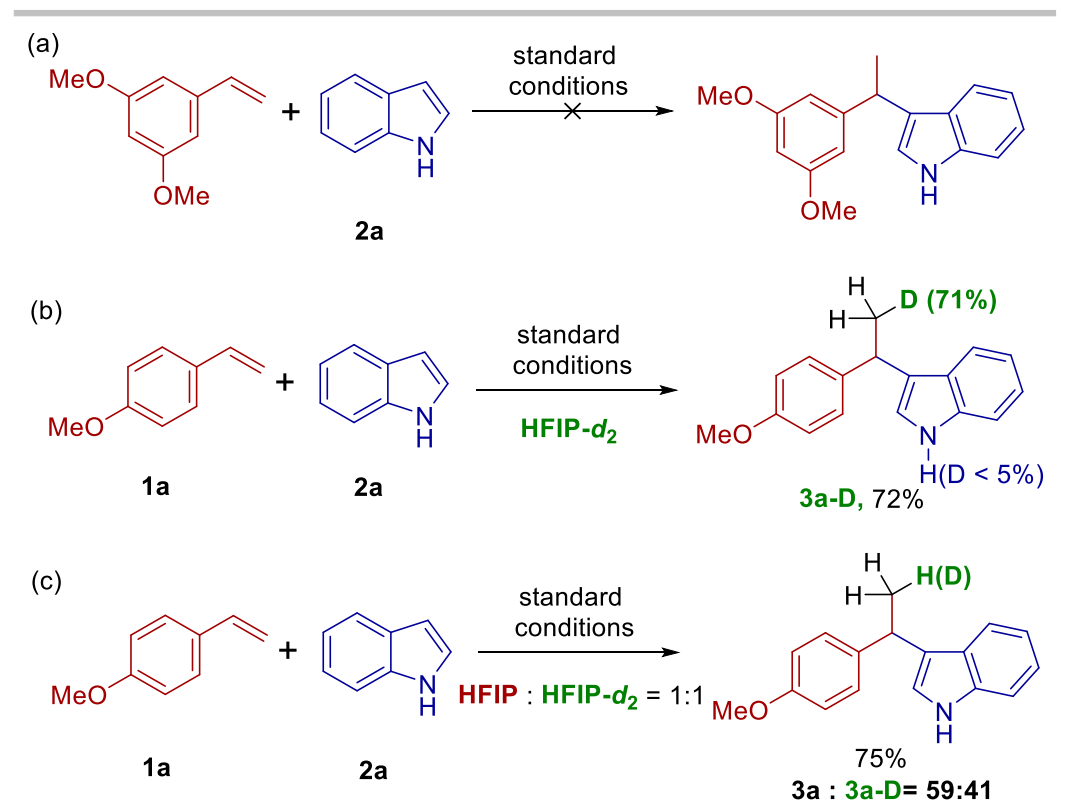
6. In vitro cell cytotoxicity

Method and materials:

The following reagents were obtained from commercial sources as indicated: DMEM or DMEM/F12 containing 2.5 mM L-glutamine and 15 mM HEPES buffer (HyClone); fetal bovine serum (FBS) was heat inactivated (HyClone); 100x penicillin-streptomycin-glutamine (PS) (Gibco); Molecular biology grade DMSO (Sigma); 0.05% Trypsin-EDTA (Gibco); Initial Evaluation of Compounds 3ai, 3v, 3w and 3x-: MCF-7, MDA-MB-231 or GL-261 cells were seeded at 4,000 cells per well in a 96 well flat bottom plate with DMEM or DMEM/F12 media supplemented with 10% FBS, and incubated with 5% CO₂ at 37 °C overnight. The compounds, were dissolved in molecular biology grade DMSO to achieve a 40 mM stock and then subsequently diluted by a factor of four using DMEM or DMEM/F12 media supplemented with 10 % FBS. The media was aspirated from the 96 well plate containing the MCF-7, MDA-MB-231, and GL-261 cells and the media containing the compounds or DMSO vehicle controls were added. After culturing for 24-72 h, cell proliferation was measured by metabolic reduction of MTT/Thiazolyl Blue Tetrazolium Bromide or CCK8 assay. All cells were confirmed to be *Mycoplasma*-free and were maintained at 37 °C and 5% CO₂.

	IC ₅₀ (μM)		
	MCF-7	MDA-MB-231	GL-261
3ai	31.3	5	3.9
3v	36.3	8.7	nd
3w	2.5	11.3	2.8
3x	10.8	16.8	8.1

7. Control experiments



Scheme S4. Control experiments

With the aim of better understanding the role of HFIP, several control experiments were performed to shed light onto this reaction. First, the hydroarylation of 1,3-dimethoxy-5-vinylbenzene with indole in HFIP did not deliver any of the desired products (Scheme S4a), implying the quinone methides structure might be involved as the key intermediate in this transformation. Second, when deuterated HFIP- d_2 was used as the promoter in the model reaction, it afforded the corresponding product 3a-D with 71% incorporation of the deuterium on the terminal position of olefinic bond, clearly indicating the protonation of double bond by HFIP (Scheme S4b and Figure S2). Notably, N-H of the indole did not show significant incorporation of the deuterium, while the proton at C-2 position of indole exhibit deuterium incorporation by dissolving indole in HFIP- d_2 under standard conditions (Figure S5-S6). Intermolecular competition experiment between HFIP and HFIP- d_2 was also carried out (Scheme S4c), a ratio of 59:41 between 3a and 3a-D was afforded with an estimated KIE of 1.4 (Supporting information Figure S3-S4), indicating the protonation step might not be the rate-determining step, which was not fully consistent with the previous report involving additional Lewis acids catalysts.

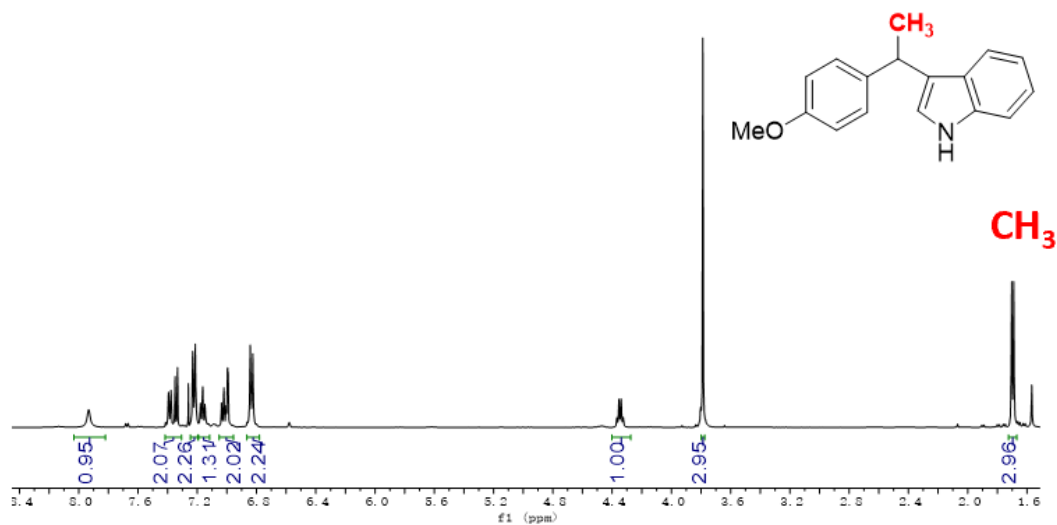


Figure S1 ^1H NMR of **3a**, CDCl_3 , 298K

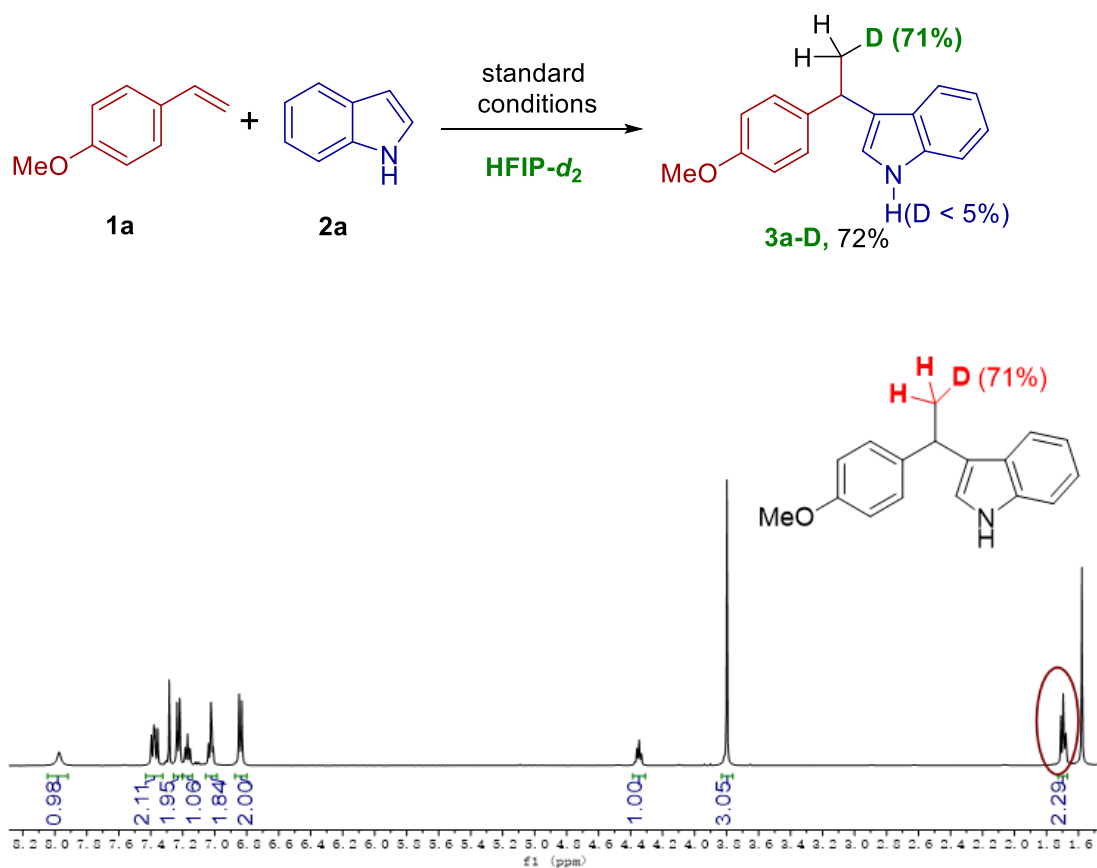


Figure S2 ^1H NMR of **3a-D**, CDCl_3 , 298K

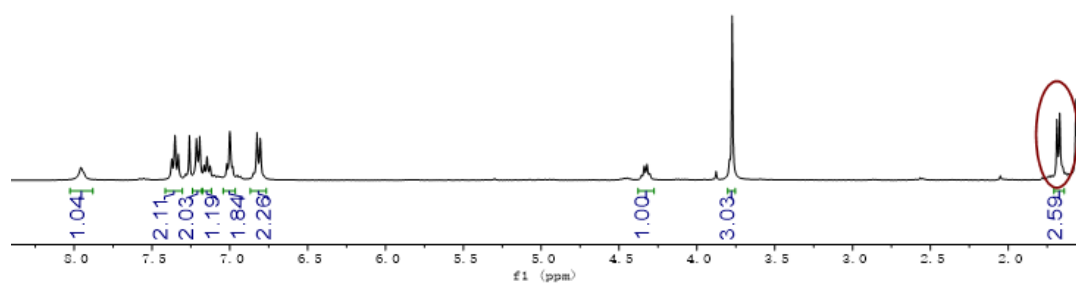
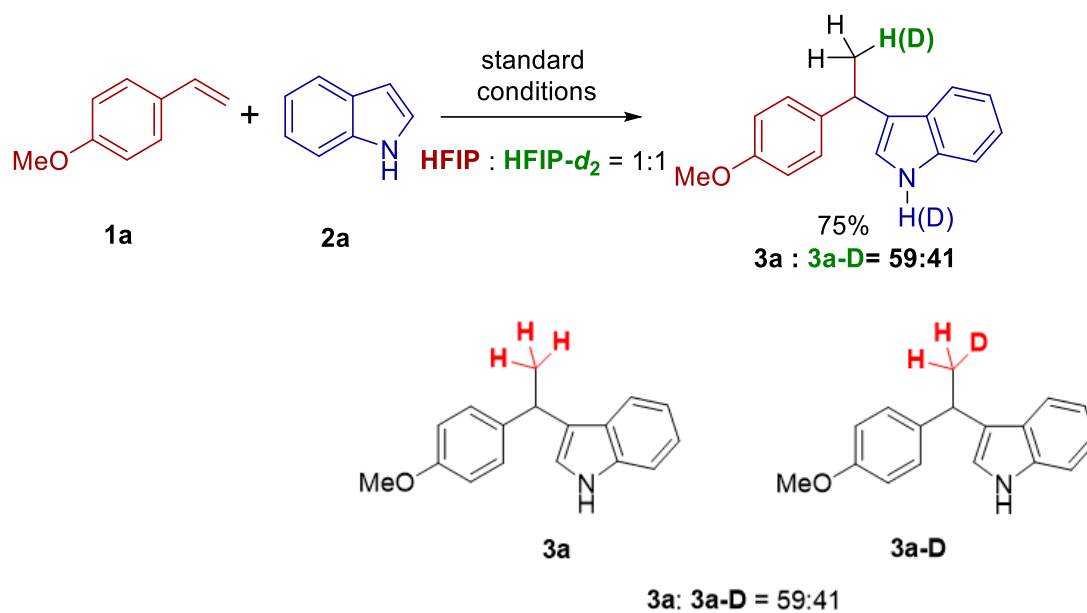


Figure S3 ^1H NMR of 3a and 3a-D, CDCl_3 , 298K

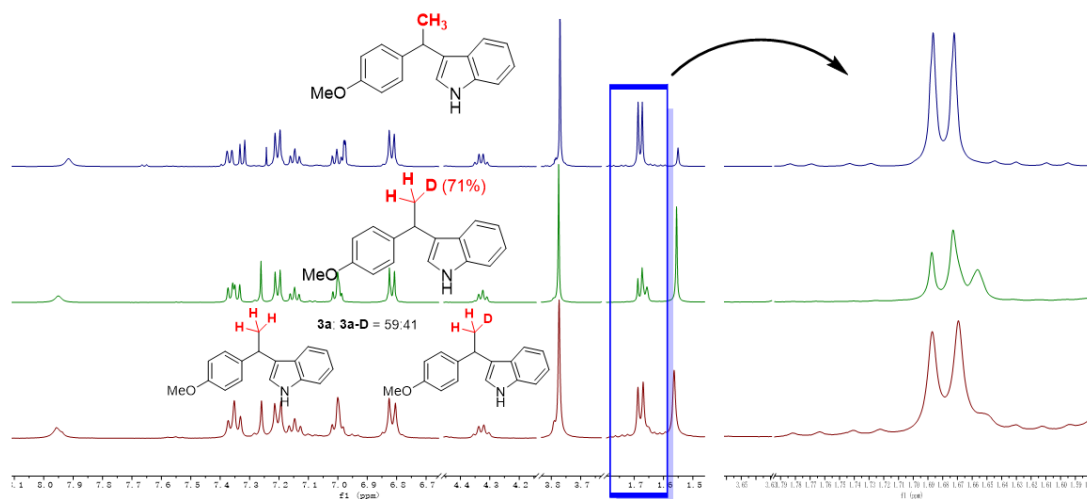


Figure S4 Stacked ^1H NMR of 3a and 3a-D, CDCl_3 , 298K

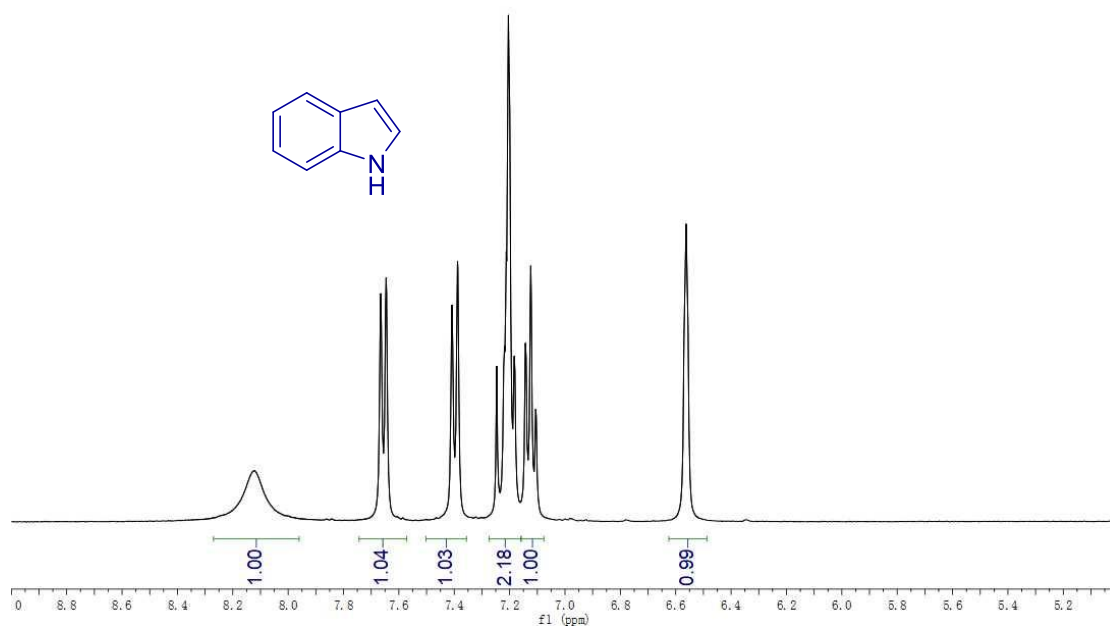


Figure S5 ^1H NMR of 3a and 3a-D, CDCl_3 , 298K

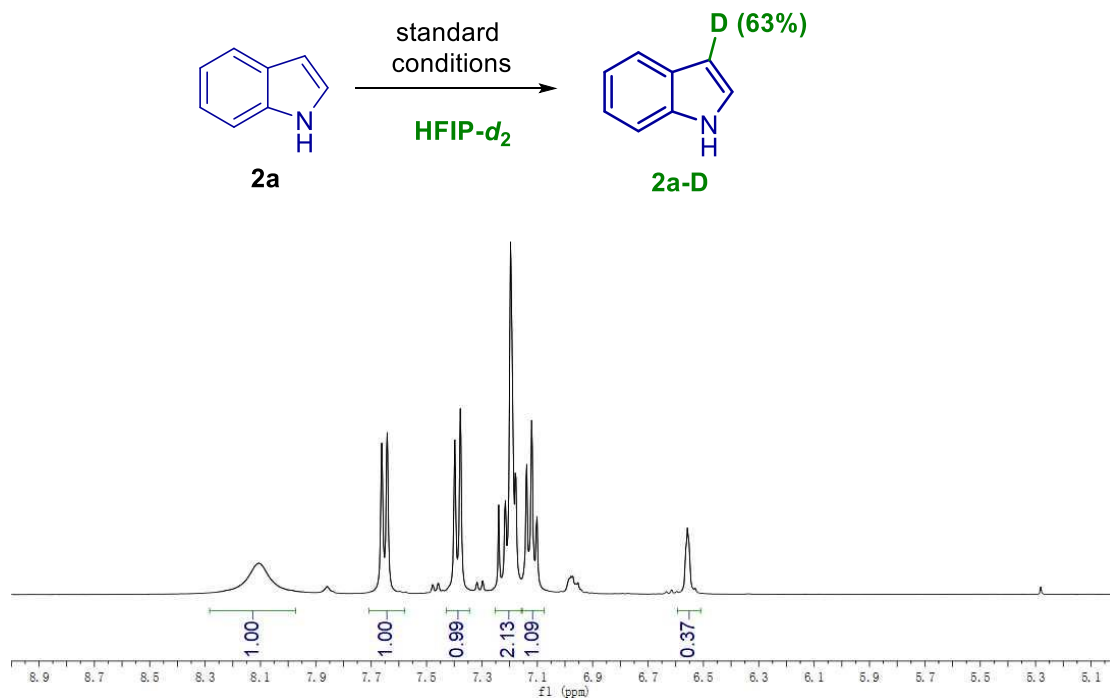


Figure S6 ^1H NMR of 3a and 3a-D, CDCl_3 , 298K

8. X-Ray analysis

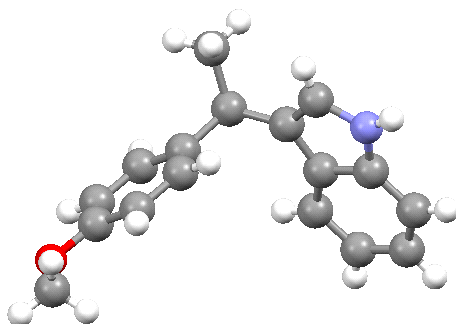


Figure S Single-crystal X-Ray diffraction of **3a**

Table S1 Crystal data and structure refinement

Identification code	mo230428a
Empirical formula	C ₁₇ H ₁₇ NO
Formula weight	251.31
Temperature/K	296.15
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	6.9302(9)
b/Å	10.5748(14)
c/Å	19.067(3)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1397.4(3)
Z	4
ρ _{calc} /cm ³	1.195
μ/mm ⁻¹	0.074
F(000)	536.0
Crystal size/mm ³	0.15 × 0.15 × 0.12
Radiation	MoKα (λ = 0.71073)
2θ range for data collection/°	4.272 to 61
Index ranges	-9 ≤ h ≤ 9, -14 ≤ k ≤ 15, -26 ≤ l ≤ 26
Reflections collected	20151
Independent reflections	3983 [R _{int} = 0.0436, R _{sigma} = 0.0356]
Data/restraints/parameters	3983/0/174
Goodness-of-fit on F ²	1.024
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0434, wR ₂ = 0.0987
Final R indexes [all data]	R ₁ = 0.0649, wR ₂ = 0.1102
Largest diff. peak/hole / e Å ⁻³	0.15/-0.16
Flack parameter	0.4(6)

Table S2 Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230428a. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U(eq)
O(1)	5046(2)	9214.0(13)	3371.0(8)	54.4(4)
N(1)	6534(3)	1701.5(16)	3861.0(11)	58.3(5)
C(2)	6138(3)	2594.8(18)	4367.5(13)	52.8(5)
C(3)	7438(3)	3552.2(16)	4330.0(9)	39.6(4)
C(4)	8730(3)	3242.2(17)	3769.9(10)	40.6(4)
C(5)	10358(3)	3822(2)	3482.9(13)	56.4(6)
C(6)	11296(4)	3237(3)	2932.1(15)	75.6(8)
C(7)	10650(5)	2090(3)	2661.2(14)	78.1(9)
C(8)	9083(4)	1489(2)	2936.0(12)	64.4(7)
C(9)	8123(3)	2074.9(19)	3490.4(11)	47.4(5)
C(10)	7526(3)	4733.7(17)	4770.2(10)	42.8(4)
C(11)	6865(3)	5907.8(17)	4370.1(9)	39.6(4)
C(12)	7780(3)	7057(2)	4460.3(11)	50.3(5)
C(13)	7139(3)	8135.3(19)	4125.0(12)	52.4(5)
C(14)	5559(3)	8086.6(17)	3687.3(10)	41.5(4)
C(15)	4604(3)	6957.6(19)	3590.9(12)	50.3(5)
C(16)	5272(3)	5884.4(19)	3933.2(12)	50.3(5)
C(17)	6377(4)	4605(2)	5450.1(11)	63.6(6)
C(18)	3597(4)	9171(2)	2843.3(14)	67.5(7)

Table S3 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230428a. The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^2U_{11}+2hka^*b^*U_{12}+\dots]$.

Atom	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
O(1)	65.8(9)	30.5(7)	67.0(9)	2.3(6)	-17.4(8)	0.1(7)
N(1)	62.5(12)	32.8(8)	79.7(13)	-2.5(8)	-4.5(10)	-8.8(8)
C(2)	52.6(12)	39.0(10)	66.7(14)	5.8(10)	7.7(11)	-2.6(9)
C(3)	43.6(10)	31.2(8)	44.0(9)	6.0(7)	-0.9(8)	2.1(8)
C(4)	46.5(10)	32.1(9)	43.3(9)	7.6(8)	-2.5(8)	4.6(8)
C(5)	53.4(12)	47.2(12)	68.6(14)	9.7(10)	9.3(11)	3.0(10)
C(6)	74.3(18)	72.6(17)	79.8(17)	20.9(14)	32.0(15)	18.9(15)
C(7)	109(2)	67.2(17)	58.3(14)	8.7(13)	21.2(16)	42.7(17)
C(8)	98(2)	44.1(12)	51.3(12)	-3.6(10)	-7.3(13)	25.3(13)
C(9)	60.1(13)	32.7(9)	49.4(10)	4.1(8)	-8.1(10)	9.3(9)
C(10)	45.1(10)	38.9(10)	44.2(9)	1.4(8)	-3.4(9)	2.6(9)
C(11)	44.1(10)	33.9(8)	40.7(9)	-3.8(8)	-2.1(8)	1.1(8)
C(12)	51.6(12)	42.2(10)	57.2(12)	-2.0(9)	-19.1(10)	-4.0(9)
C(13)	58.0(13)	32.7(9)	66.6(12)	-1.6(9)	-15.5(10)	-10.4(9)
C(14)	48.8(11)	30.1(9)	45.7(9)	-2.4(8)	-4.1(8)	0.3(8)
C(15)	52.3(12)	36.9(10)	61.7(12)	-1.3(9)	-19.3(10)	-4.1(9)
C(16)	55.2(12)	30.1(9)	65.7(13)	-1.3(9)	-15.3(11)	-9.5(9)

Table S3 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230428a.
The Anisotropic displacement factor exponent takes the form: $-2 \pi^2 [h^2 a^2 U_{11} + 2 h k a^2 b^* U_{12} + \dots]$.

Atom	U_{11}	U_{22}	U_{33}	U_{23}	U_{13}	U_{12}
C(17)	83.7(17)	59.4(13)	47.8(11)	4.1(10)	9.9(12)	15.8(13)
C(18)	77.3(17)	48.9(12)	76.2(16)	8.9(12)	-27.6(14)	5.6(13)

Table S4 Bond Lengths for mo230428a.

Atom	Atom	Length/ \AA	Atom	Atom	Length/ \AA
O(1)	C(14)	1.383(2)	C(7)	C(8)	1.363(4)
O(1)	C(18)	1.422(3)	C(8)	C(9)	1.394(3)
N(1)	C(2)	1.379(3)	C(10)	C(11)	1.528(3)
N(1)	C(9)	1.367(3)	C(10)	C(17)	1.528(3)
C(2)	C(3)	1.357(3)	C(11)	C(12)	1.382(3)
C(3)	C(4)	1.432(3)	C(11)	C(16)	1.383(3)
C(3)	C(10)	1.506(3)	C(12)	C(13)	1.381(3)
C(4)	C(5)	1.396(3)	C(13)	C(14)	1.378(3)
C(4)	C(9)	1.409(3)	C(14)	C(15)	1.377(3)
C(5)	C(6)	1.381(4)	C(15)	C(16)	1.389(3)
C(6)	C(7)	1.393(4)			

Table S5 Bond Angles for mo230428a.

Atom	Atom	Atom	Angle/ $^\circ$	Atom	Atom	Atom	Angle/ $^\circ$
C(14)	O(1)	C(18)	117.55(16)	C(8)	C(9)	C(4)	122.2(2)
C(9)	N(1)	C(2)	108.95(18)	C(3)	C(10)	C(11)	112.56(15)
C(3)	C(2)	N(1)	110.0(2)	C(3)	C(10)	C(17)	112.20(17)
C(2)	C(3)	C(4)	106.48(17)	C(17)	C(10)	C(11)	109.84(17)
C(2)	C(3)	C(10)	128.06(19)	C(12)	C(11)	C(10)	120.99(17)
C(4)	C(3)	C(10)	125.45(17)	C(12)	C(11)	C(16)	117.20(17)
C(5)	C(4)	C(3)	134.22(19)	C(16)	C(11)	C(10)	121.72(17)
C(5)	C(4)	C(9)	118.6(2)	C(13)	C(12)	C(11)	121.39(18)
C(9)	C(4)	C(3)	107.21(18)	C(14)	C(13)	C(12)	120.37(18)
C(6)	C(5)	C(4)	118.8(2)	C(13)	C(14)	O(1)	115.87(17)
C(5)	C(6)	C(7)	121.4(3)	C(15)	C(14)	O(1)	124.45(17)
C(8)	C(7)	C(6)	121.3(2)	C(15)	C(14)	C(13)	119.68(18)
C(7)	C(8)	C(9)	117.7(2)	C(14)	C(15)	C(16)	119.04(18)
N(1)	C(9)	C(4)	107.36(18)	C(11)	C(16)	C(15)	122.31(18)
N(1)	C(9)	C(8)	130.4(2)				

Table S6 Torsion Angles for mo230428a.

A	B	C	D	Angle/ $^\circ$	A	B	C	D	Angle/ $^\circ$
O(1)	C(14)	C(15)	C(16)	179.4(2)	C(6)	C(7)	C(8)	C(9)	1.2(4)
N(1)	C(2)	C(3)	C(4)	-0.2(2)	C(7)	C(8)	C(9)	N(1)	179.8(2)
N(1)	C(2)	C(3)	C(10)	178.60(19)	C(7)	C(8)	C(9)	C(4)	-0.4(3)

Table S6 Torsion Angles for mo230428a.

A	B	C	D	Angle/°	A	B	C	D	Angle/°
C(2)	N(1)	C(9)	C(4)	-0.5(2)	C(9)	N(1)	C(2)	C(3)	0.4(2)
C(2)	N(1)	C(9)	C(8)	179.3(2)	C(9)	C(4)	C(5)	C(6)	0.7(3)
C(2)	C(3)	C(4)	C(5)	-178.8(2)	C(10)	C(3)	C(4)	C(5)	2.3(3)
C(2)	C(3)	C(4)	C(9)	-0.1(2)	C(10)	C(3)	C(4)	C(9)	-178.93(17)
C(2)	C(3)	C(10)	C(11)	-105.7(2)	C(10)	C(11)	C(12)	C(13)	-177.1(2)
C(2)	C(3)	C(10)	C(17)	18.8(3)	C(10)	C(11)	C(16)	C(15)	177.1(2)
C(3)	C(4)	C(5)	C(6)	179.4(2)	C(11)	C(12)	C(13)	C(14)	-0.2(3)
C(3)	C(4)	C(9)	N(1)	0.3(2)	C(12)	C(11)	C(16)	C(15)	0.4(3)
C(3)	C(4)	C(9)	C(8)	-179.49(19)	C(12)	C(13)	C(14)	O(1)	-179.3(2)
C(3)	C(10)	C(11)	C(12)	-141.6(2)	C(12)	C(13)	C(14)	C(15)	0.7(3)
C(3)	C(10)	C(11)	C(16)	41.9(3)	C(13)	C(14)	C(15)	C(16)	-0.7(3)
C(4)	C(3)	C(10)	C(11)	72.9(2)	C(14)	C(15)	C(16)	C(11)	0.1(4)
C(4)	C(3)	C(10)	C(17)	-162.59(19)	C(16)	C(11)	C(12)	C(13)	-0.4(3)
C(4)	C(5)	C(6)	C(7)	0.0(4)	C(17)	C(10)	C(11)	C(12)	92.6(2)
C(5)	C(4)	C(9)	N(1)	179.32(18)	C(17)	C(10)	C(11)	C(16)	-83.9(2)
C(5)	C(4)	C(9)	C(8)	-0.5(3)	C(18)	O(1)	C(14)	C(13)	171.5(2)
C(5)	C(6)	C(7)	C(8)	-1.0(4)	C(18)	O(1)	C(14)	C(15)	-8.5(3)

Table S7 Hydrogen Atom Coordinates ($\text{\AA} \times 10^4$) and Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230428a.

Atom	x	y	z	U(eq)
H(1)	5886.56	1019.73	3789.78	70
H(2)	5128.13	2548.18	4687.72	63
H(5)	10803.51	4588.39	3659.37	68
H(6)	12382.21	3617.86	2738.01	91
H(7)	11301.19	1726.34	2285.42	94
H(8)	8668	715.75	2759.71	77
H(10)	8879.94	4865.66	4899.93	51
H(12)	8851.02	7105.64	4753.11	60
H(13)	7776.97	8898.65	4195.03	63
H(15)	3527.11	6914.7	3300.73	60
H(16)	4625.58	5123.16	3866.37	60
H(17A)	5050.09	4433.01	5341.53	95
H(17B)	6462.77	5378.75	5711.98	95
H(17C)	6895.4	3923.82	5724.14	95
H(18A)	3988.53	8600.81	2478.32	101
H(18B)	3417.02	10001.2	2650.46	101
H(18C)	2406.66	8881.75	3045.01	101

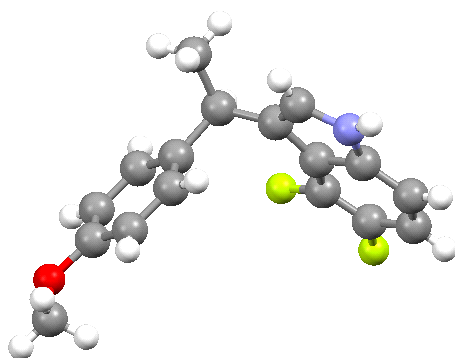


Figure S Single-crystal X-Ray diffraction of **3m**

Table S8 Crystal data and structure refinement.

Identification code	mo230505b
Empirical formula	C ₁₇ H ₁₅ F ₂ NO
Formula weight	287.311
Temperature/K	296.15
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	7.4260(14)
b/Å	10.475(2)
c/Å	18.684(4)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1453.4(5)
Z	4
ρ _{calc} /cm ³	1.313
μ/mm ⁻¹	0.099
F(000)	600.4
Crystal size/mm ³	0.13 × 0.13 × 0.1
Radiation	Mo Kα (λ = 0.71073)
2θ range for data collection/°	4.36 to 60.96
Index ranges	-8 ≤ h ≤ 10, -14 ≤ k ≤ 14, -19 ≤ l ≤ 25
Reflections collected	10744
Independent reflections	4011 [R _{int} = 0.0710, R _{sigma} = 0.1065]
Data/restraints/parameters	4011/0/192
Goodness-of-fit on F ²	1.046
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0543, wR ₂ = 0.1127
Final R indexes [all data]	R ₁ = 0.1470, wR ₂ = 0.1556
Largest diff. peak/hole / e Å ⁻³	0.30/-0.34
Flack parameter	0.3(11)

Table S9 Fractional Atomic Coordinates ($\times 10^4$) and Equivalent Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230505b. U_{eq} is defined as 1/3 of the trace of the orthogonalised U_{ij} tensor.

Atom	x	y	z	U_{eq}
F(1)	4263(2)	5083.3(18)	3835.5(11)	80.8(6)
O(1)	10150(3)	667.4(16)	3432.1(11)	57.7(6)
F(2)	2417(3)	6220(2)	2792.7(14)	115.0(9)
N(1)	8583(4)	8234(2)	3835.2(18)	76.5(8)
C(13)	9602(4)	1813(2)	3730.1(14)	46.0(7)
C(3)	6494(4)	6713(2)	3809.3(15)	47.8(7)
C(2)	7756(4)	6394(2)	4357.9(15)	49.7(7)
C(10)	8350(4)	4026(2)	4401.6(14)	45.1(7)
C(11)	9787(4)	4050(2)	3936.4(16)	54.5(8)
C(12)	10421(4)	2959(3)	3598.2(16)	55.5(8)
C(9)	7740(5)	5209(3)	4812.0(14)	52.9(7)
C(15)	7552(4)	2856(3)	4520.2(17)	59.3(8)
C(8)	7038(5)	7875(3)	3494.1(18)	60.7(9)
C(4)	4892(5)	6182(3)	3551.6(16)	57.1(8)
C(14)	8170(4)	1762(3)	4188.6(17)	61.0(9)
C(1)	8992(5)	7352(3)	4351(2)	67.6(9)
C(5)	3985(5)	6764(4)	3021(2)	73.1(10)
C(7)	6105(6)	8457(3)	2943(2)	80.3(11)
C(17)	11410(5)	705(3)	2859.9(18)	72.6(10)
C(16)	8877(5)	5361(3)	5482.1(17)	79.0(11)
C(6)	4574(7)	7882(4)	2700(2)	89.2(14)

Table S10 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230505b. The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^{*2}U_{11}+2hka^*b^*U_{12}+\dots]$.

Atom	U_{11}	U_{22}	U_{33}	U_{12}	U_{13}	U_{23}
F(1)	73.3(14)	67.7(12)	101.4(14)	-21.1(10)	-12.3(11)	3.2(11)
O(1)	67.9(14)	35.9(10)	69.2(12)	-1.2(10)	17.6(12)	0.8(8)
F(2)	97.1(19)	126(2)	121.9(19)	25.6(15)	-46.3(16)	-23.8(15)
N(1)	82(2)	36.8(13)	111(2)	-9.6(14)	21.6(19)	-1.3(15)
C(13)	53.7(19)	34.4(13)	49.7(15)	-2.3(13)	1.9(15)	0.7(11)
C(3)	56.6(19)	34.7(13)	52.0(16)	4.5(13)	12.6(15)	-6.2(12)
C(2)	54.1(19)	36.9(14)	58.1(17)	-3.4(13)	0.4(16)	-6.5(12)
C(10)	50.2(18)	41.3(14)	43.8(15)	0.7(13)	0.5(15)	2.6(11)
C(11)	64(2)	32.8(13)	66.5(18)	-11.1(14)	14.2(17)	3.3(12)
C(12)	57(2)	41.8(16)	67.3(19)	-6.1(14)	18.6(16)	2.8(13)
C(9)	59(2)	47.7(16)	52.3(16)	-0.3(15)	1.0(15)	-4.4(13)
C(15)	62(2)	46.5(16)	70(2)	-10.8(16)	22.7(17)	-5.2(14)
C(8)	74(2)	36.1(14)	72(2)	9.6(16)	22.0(19)	-1.2(14)
C(4)	61(2)	46.7(16)	63.9(19)	0.6(15)	-2.9(18)	-4.7(14)
C(14)	62(2)	41.6(15)	79(2)	-14.1(15)	21.4(17)	0.1(14)

Table S10 Anisotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230505b.
The Anisotropic displacement factor exponent takes the form: $-2\pi^2[h^2a^*U_{11}+2hka^*b^*U_{12}+\dots]$.

Atom	U_{11}	U_{22}	U_{33}	U_{12}	U_{13}	U_{23}
C(1)	67(2)	45.9(16)	90(2)	-6.5(16)	0(2)	-13.2(17)
C(5)	68(3)	76(2)	76(2)	25(2)	-15(2)	-17(2)
C(7)	113(3)	53(2)	75(2)	28(2)	27(2)	12.3(18)
C(17)	81(3)	55.7(19)	81(2)	9.6(18)	30(2)	-9.2(17)
C(16)	96(3)	81(2)	60(2)	15(2)	-15(2)	-12.1(17)
C(6)	123(4)	81(3)	64(2)	52(3)	6(3)	2(2)

Table S11 Bond Lengths for mo230505b.

Atom	Atom	Length/ \AA	Atom	Atom	Length/ \AA
F(1)	C(4)	1.351(3)	C(2)	C(1)	1.360(4)
O(1)	C(13)	1.384(3)	C(10)	C(11)	1.377(4)
O(1)	C(17)	1.421(3)	C(10)	C(9)	1.526(4)
F(2)	C(5)	1.365(4)	C(10)	C(15)	1.379(4)
N(1)	C(8)	1.365(4)	C(11)	C(12)	1.388(4)
N(1)	C(1)	1.369(4)	C(9)	C(16)	1.518(4)
C(13)	C(12)	1.368(4)	C(15)	C(14)	1.381(4)
C(13)	C(14)	1.367(4)	C(8)	C(7)	1.383(5)
C(3)	C(2)	1.429(4)	C(4)	C(5)	1.345(4)
C(3)	C(8)	1.411(4)	C(5)	C(6)	1.386(5)
C(3)	C(4)	1.398(4)	C(7)	C(6)	1.365(5)
C(2)	C(9)	1.504(4)			

Table S12 Bond Angles for mo230505b.

Atom	Atom	Atom	Angle/ $^\circ$	Atom	Atom	Atom	Angle/ $^\circ$
C(17)	O(1)	C(13)	118.2(2)	C(16)	C(9)	C(2)	112.0(3)
C(1)	N(1)	C(8)	109.2(3)	C(16)	C(9)	C(10)	109.5(2)
C(12)	C(13)	O(1)	123.9(2)	C(14)	C(15)	C(10)	121.5(3)
C(14)	C(13)	O(1)	116.6(2)	C(3)	C(8)	N(1)	106.5(3)
C(14)	C(13)	C(12)	119.5(2)	C(7)	C(8)	N(1)	130.3(3)
C(8)	C(3)	C(2)	108.3(3)	C(7)	C(8)	C(3)	123.2(4)
C(4)	C(3)	C(2)	135.4(3)	C(3)	C(4)	F(1)	119.9(3)
C(4)	C(3)	C(8)	116.3(3)	C(5)	C(4)	F(1)	120.1(3)
C(9)	C(2)	C(3)	126.4(2)	C(5)	C(4)	C(3)	120.0(3)
C(1)	C(2)	C(3)	105.2(3)	C(15)	C(14)	C(13)	120.5(3)
C(1)	C(2)	C(9)	128.3(3)	C(2)	C(1)	N(1)	110.8(3)
C(9)	C(10)	C(11)	122.2(2)	C(4)	C(5)	F(2)	118.0(4)
C(15)	C(10)	C(11)	116.8(2)	C(6)	C(5)	F(2)	119.1(4)
C(15)	C(10)	C(9)	120.9(2)	C(6)	C(5)	C(4)	122.9(4)
C(12)	C(11)	C(10)	122.4(2)	C(6)	C(7)	C(8)	118.1(4)
C(11)	C(12)	C(13)	119.3(3)	C(7)	C(6)	C(5)	119.5(4)
C(10)	C(9)	C(2)	112.6(2)				

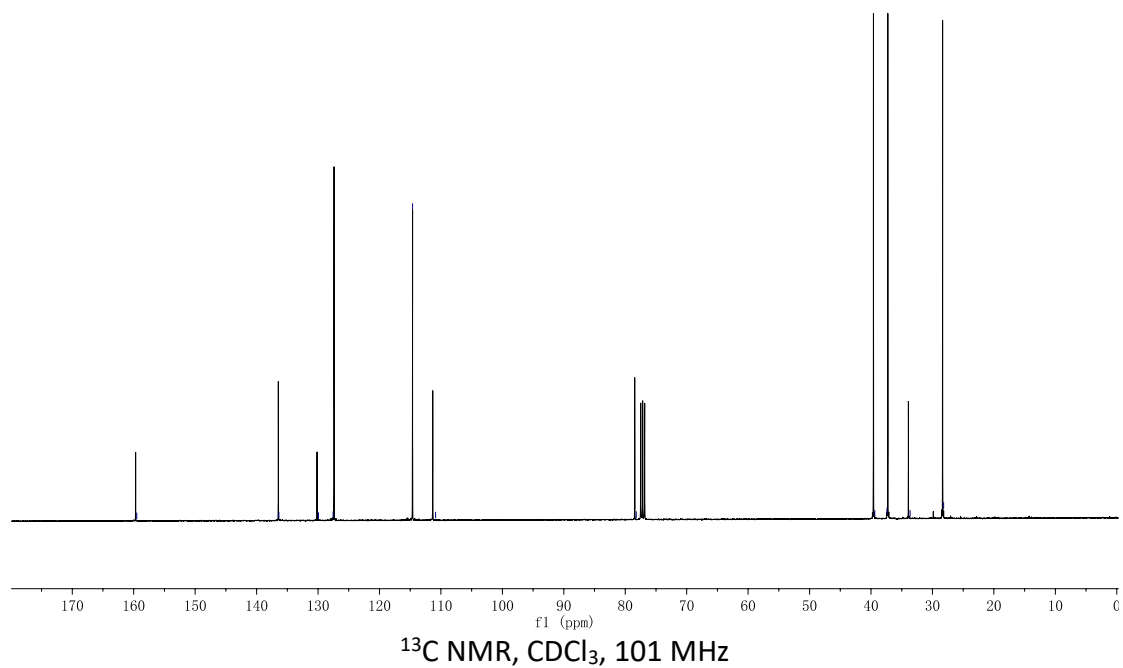
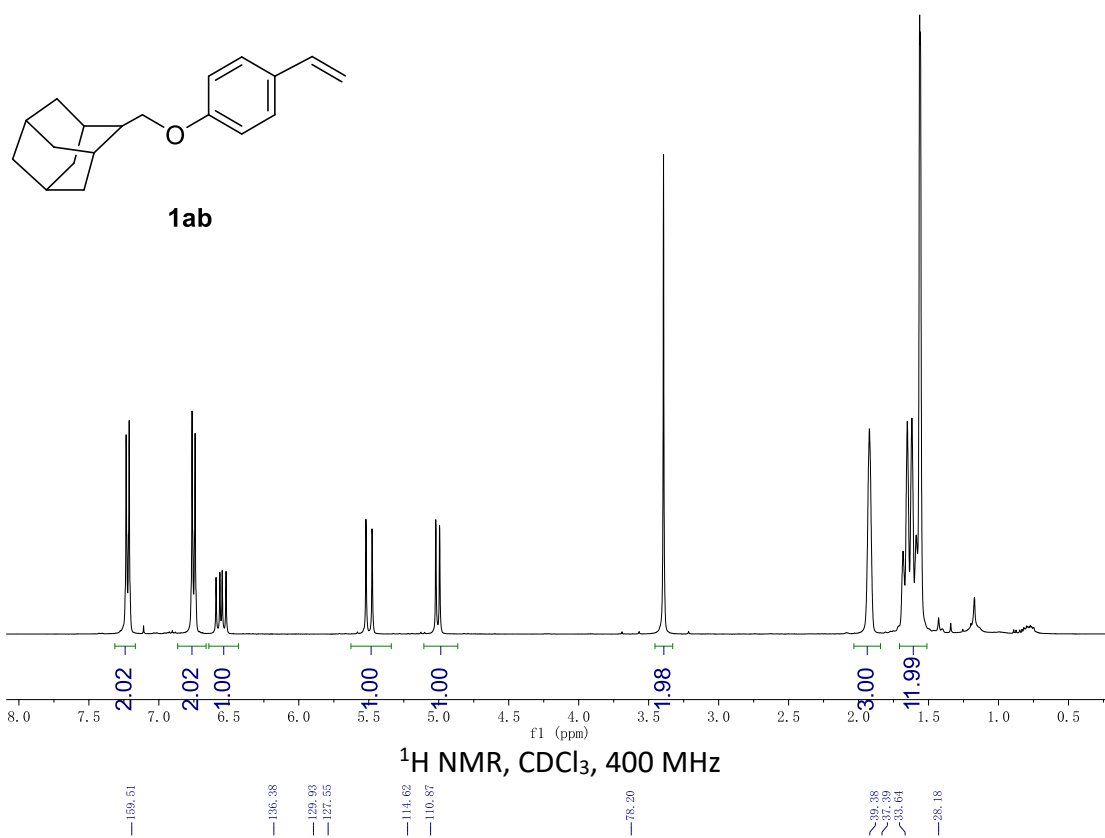
Table S13 Torsion Angles for mo230505b.

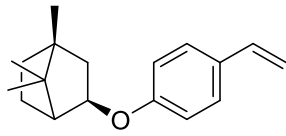
A	B	C	D	Angle/°	A	B	C	D	Angle/°
F(1)	C(4)	C(3)	C(2)	-1.9(4)	N(1)	C(1)	C(2)	C(9)	176.8(2)
F(1)	C(4)	C(3)	C(8)	-178.7(3)	C(13)	C(12)	C(11)	C(10)	-0.3(3)
F(1)	C(4)	C(5)	F(2)	1.0(3)	C(13)	C(14)	C(15)	C(10)	-0.0(4)
F(1)	C(4)	C(5)	C(6)	-179.0(3)	C(3)	C(2)	C(9)	C(10)	74.1(3)
O(1)	C(13)	C(12)	C(11)	-178.7(3)	C(3)	C(2)	C(9)	C(16)	-161.9(3)
O(1)	C(13)	C(14)	C(15)	178.9(3)	C(3)	C(8)	C(7)	C(6)	0.8(3)
F(2)	C(5)	C(4)	C(3)	-179.2(3)	C(3)	C(4)	C(5)	C(6)	0.8(4)
F(2)	C(5)	C(6)	C(7)	177.6(3)	C(2)	C(9)	C(10)	C(11)	41.8(3)
N(1)	C(8)	C(3)	C(2)	0.7(3)	C(2)	C(9)	C(10)	C(15)	-142.7(3)
N(1)	C(8)	C(3)	C(4)	178.3(2)	C(8)	C(7)	C(6)	C(5)	1.5(4)
N(1)	C(8)	C(7)	C(6)	-180.0(4)	C(4)	C(5)	C(6)	C(7)	-2.4(4)
N(1)	C(1)	C(2)	C(3)	-0.3(3)					

Table S14 Hydrogen Atom Coordinates ($\text{\AA} \times 10^4$) and Isotropic Displacement Parameters ($\text{\AA}^2 \times 10^3$) for mo230505b.

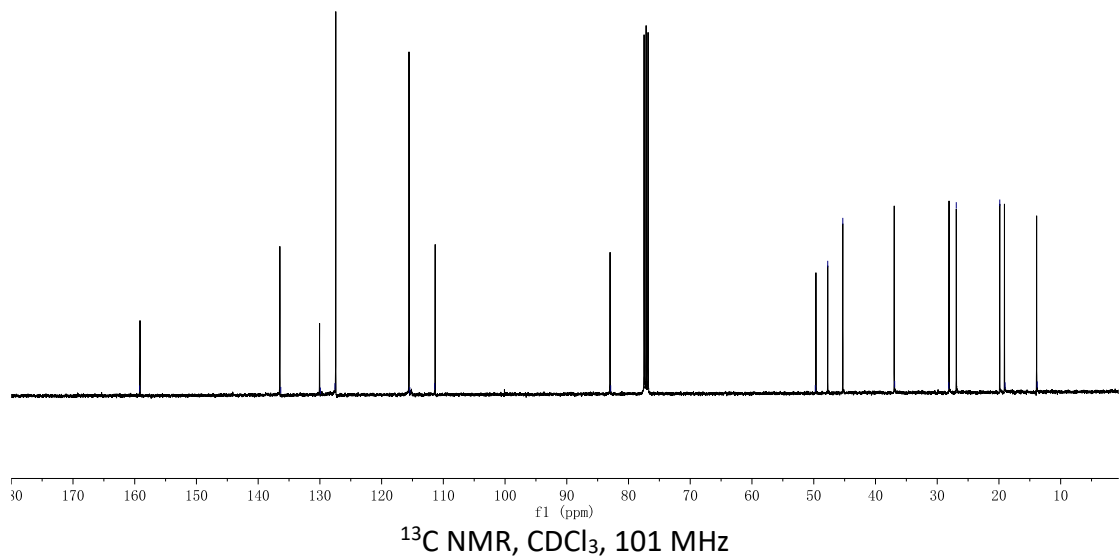
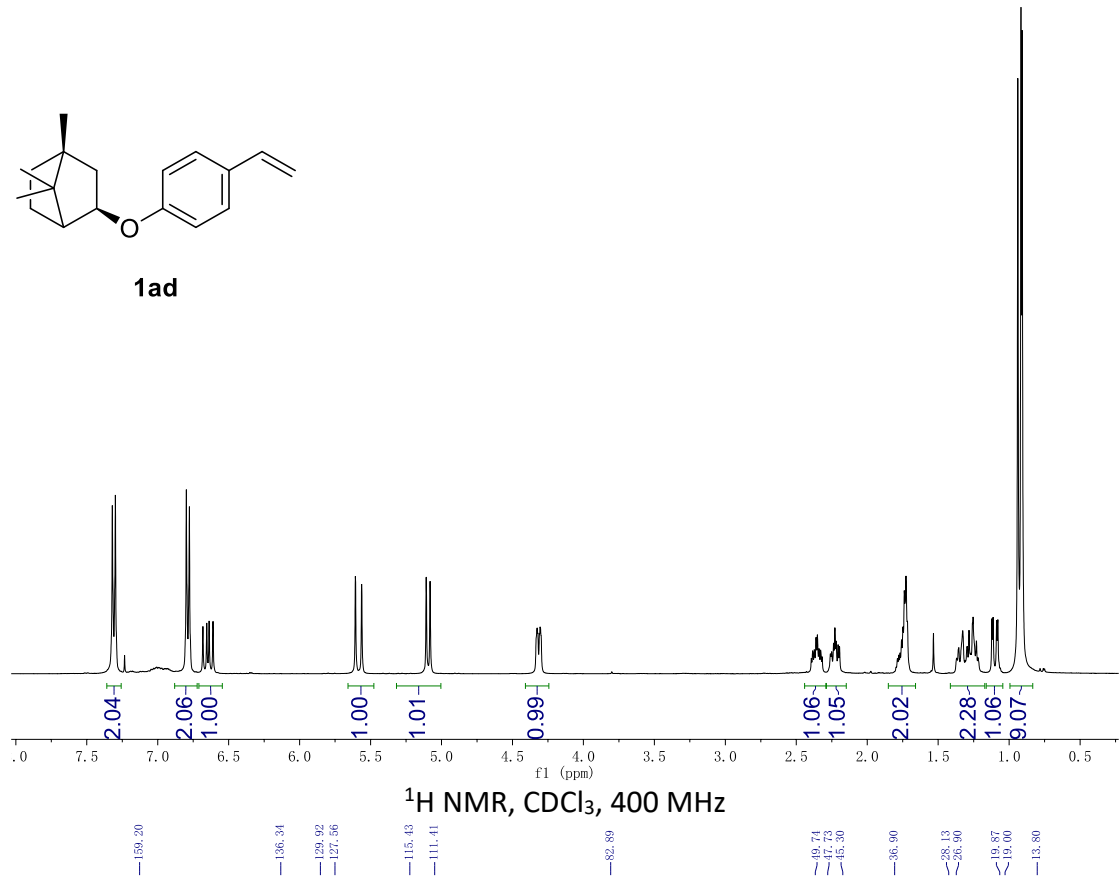
Atom	x	y	z	U(eq)
H(1)	9201(4)	8907(2)	3740.9(18)	91.8(10)
H(11)	10354(4)	4825(2)	3845.5(16)	65.4(9)
H(12)	11392(4)	3007(3)	3285.0(16)	66.6(10)
H(9)	6495(5)	5063(3)	4965.0(14)	63.5(9)
H(15)	6576(4)	2802(3)	4830.4(17)	71.1(10)
H(14)	7608(4)	985(3)	4278.1(17)	73.2(10)
H(1a)	9980(5)	7402(3)	4655(2)	81.2(11)
H(7)	6510(6)	9218(3)	2743(2)	96.4(14)
H(17a)	10988(16)	1282(18)	2497(6)	108.9(15)
H(17b)	12554(9)	990(20)	3036(3)	108.9(15)
H(17c)	11540(20)	-134(6)	2659(9)	108.9(15)
H(16a)	10110(8)	5500(20)	5349.3(17)	118.5(16)
H(16b)	8450(20)	6077(15)	5754(7)	118.5(16)
H(16c)	8790(30)	4600(10)	5767(7)	118.5(16)
H(6)	3930(7)	8237(4)	2322(2)	107.0(16)

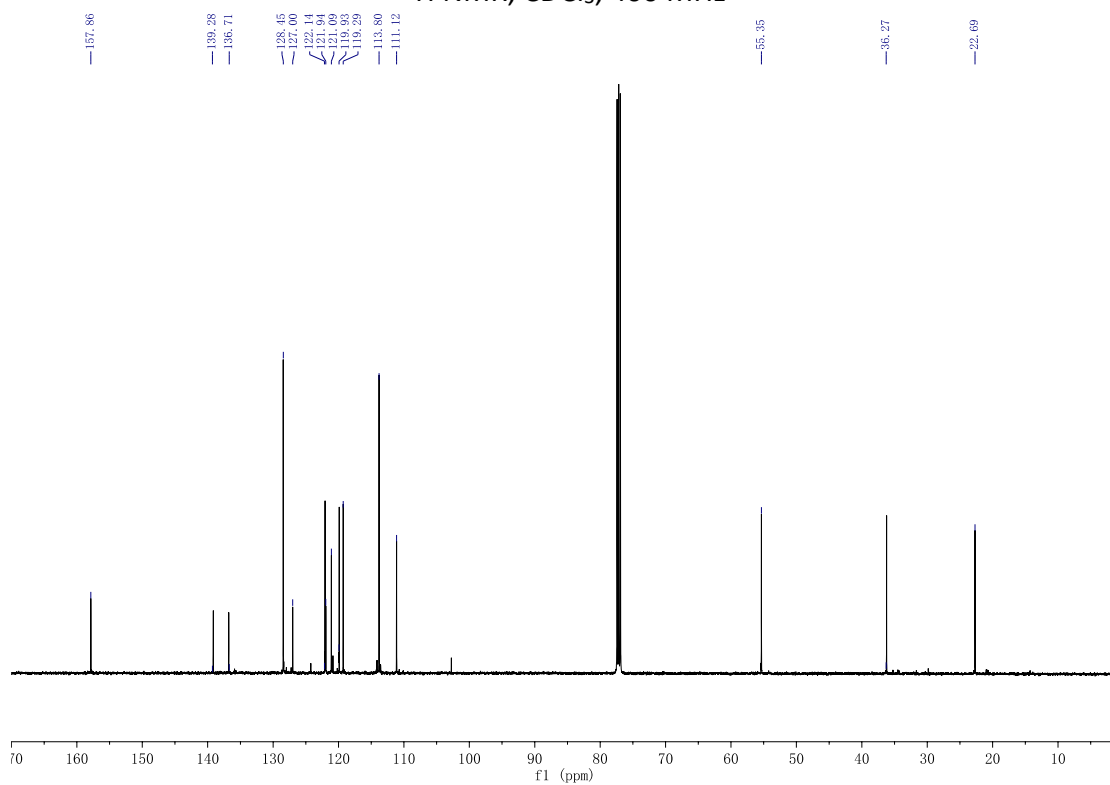
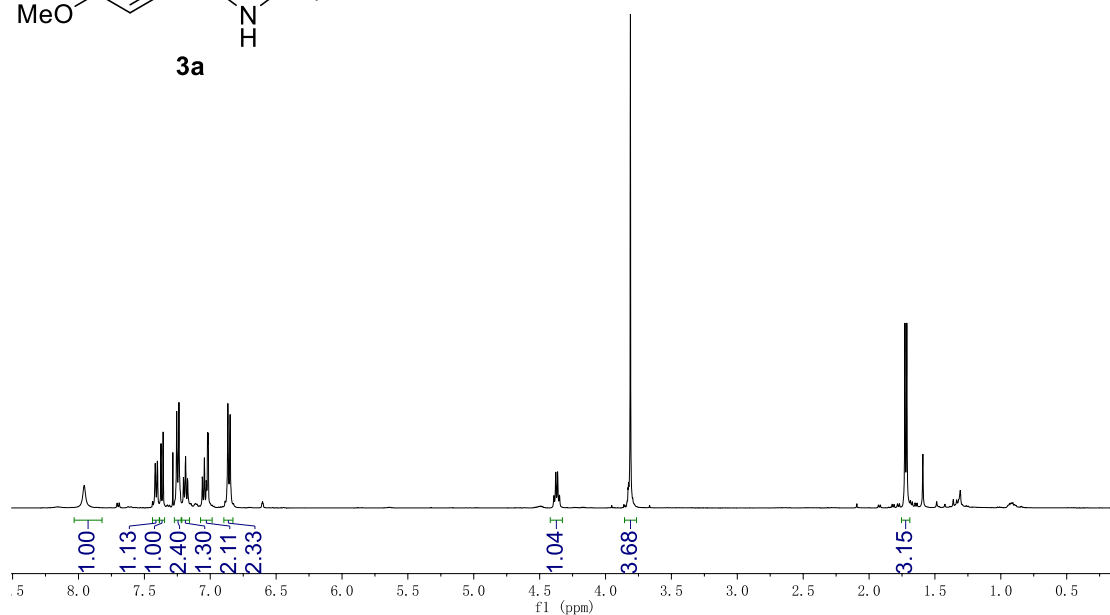
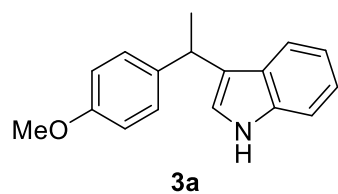
9.NMR spectra

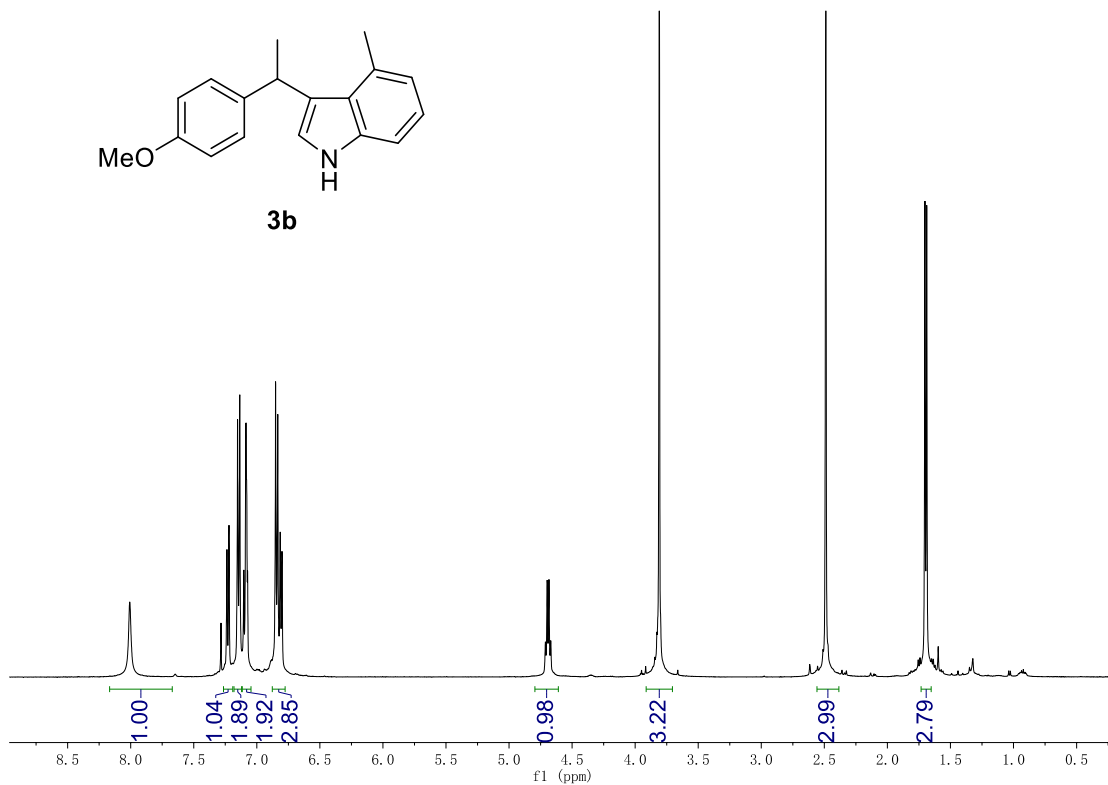
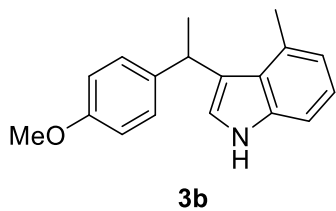




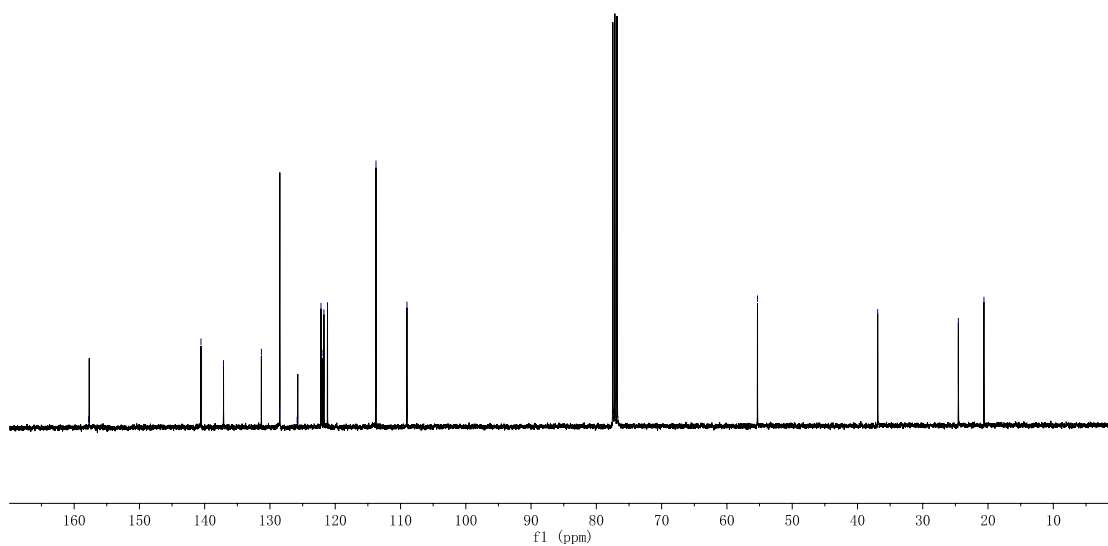
1ad

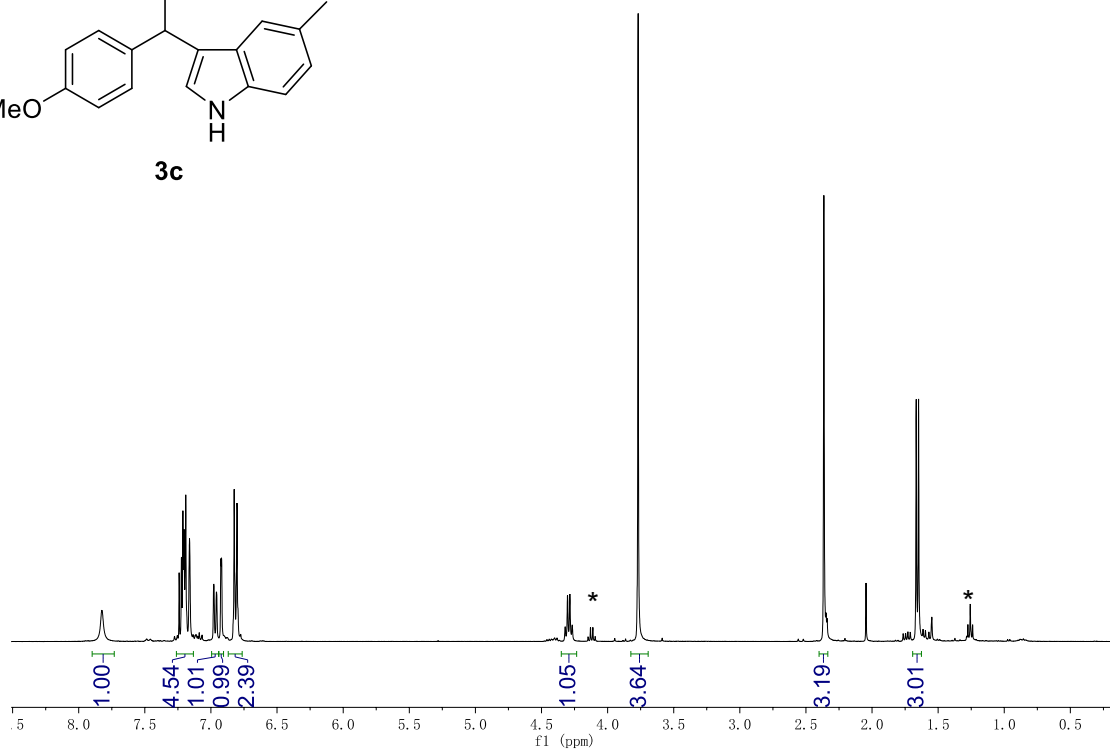
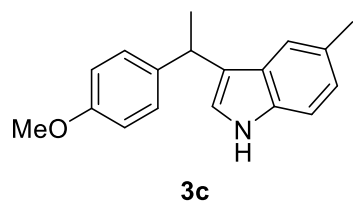




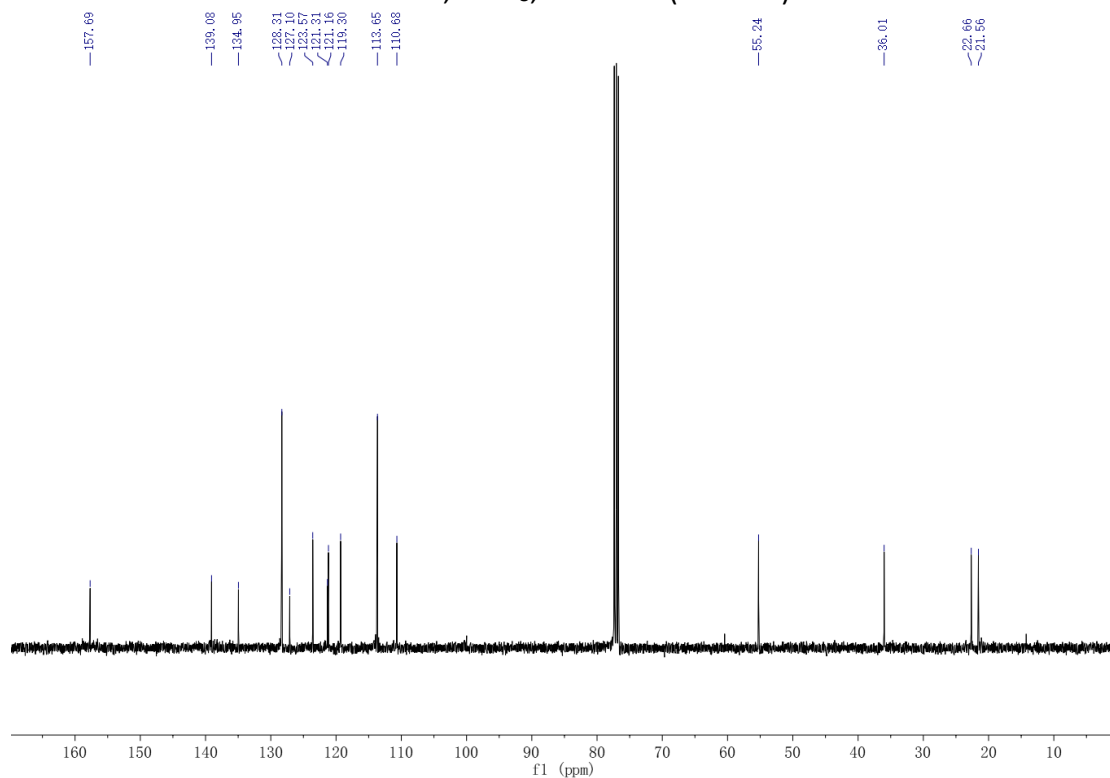


-157.78
 -140.57
 -137.11
 -131.32
 -128.54
 -125.84
 -122.02
 -121.73
 -121.19
 -113.76
 -109.01
 -55.32
 -38.90
 -21.55
 -20.63

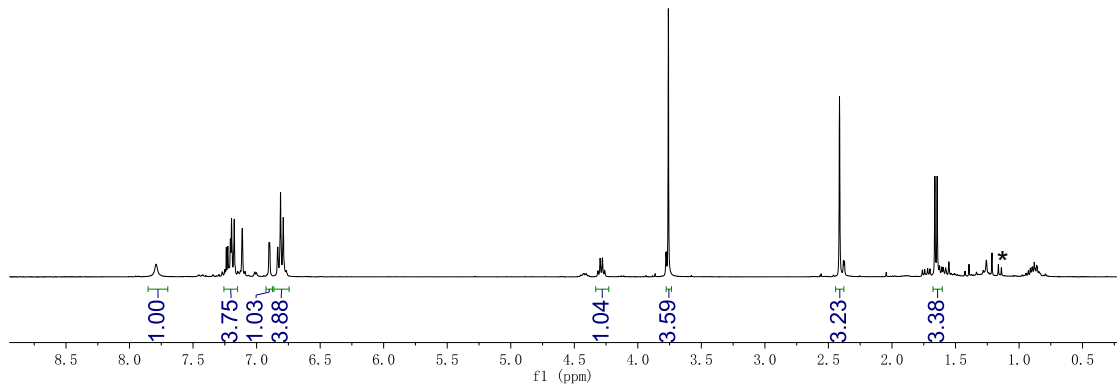
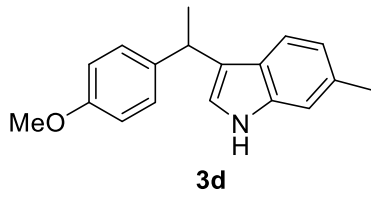




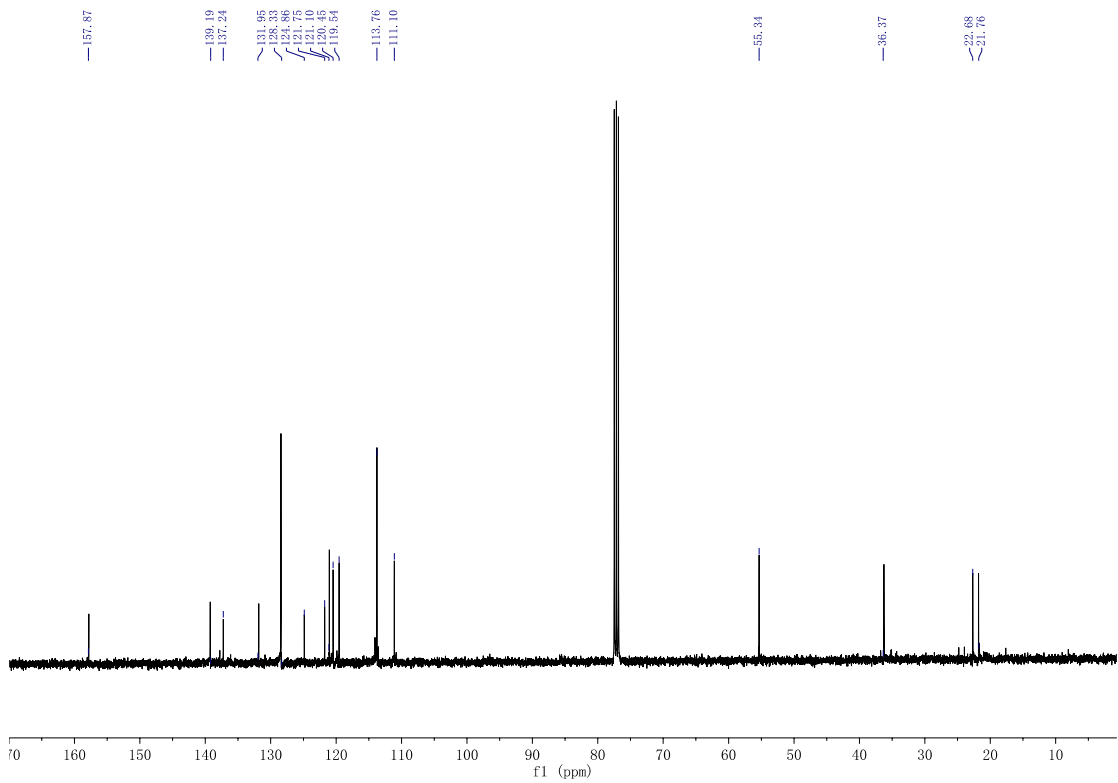
¹H NMR, CDCl₃, 400 MHz (* EtOAc)



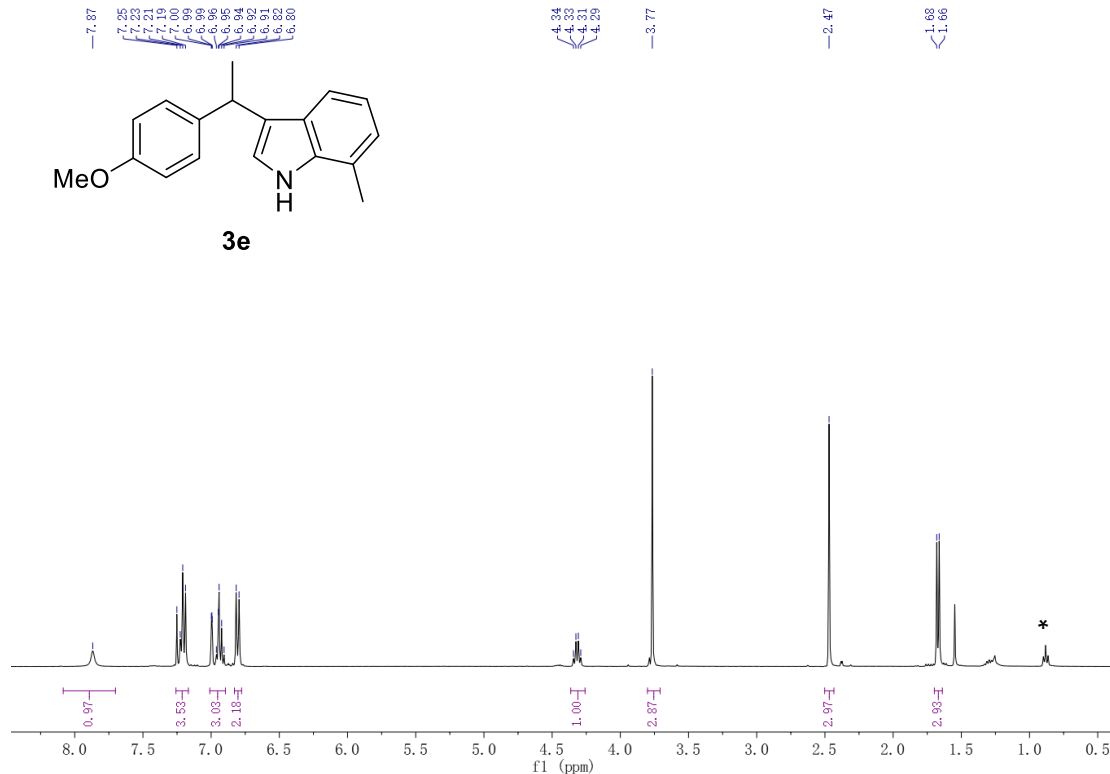
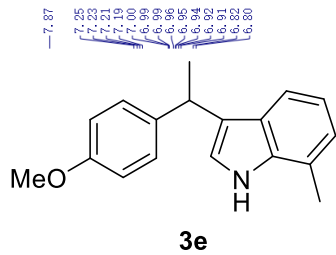
¹³C NMR, CDCl₃, 101 MHz



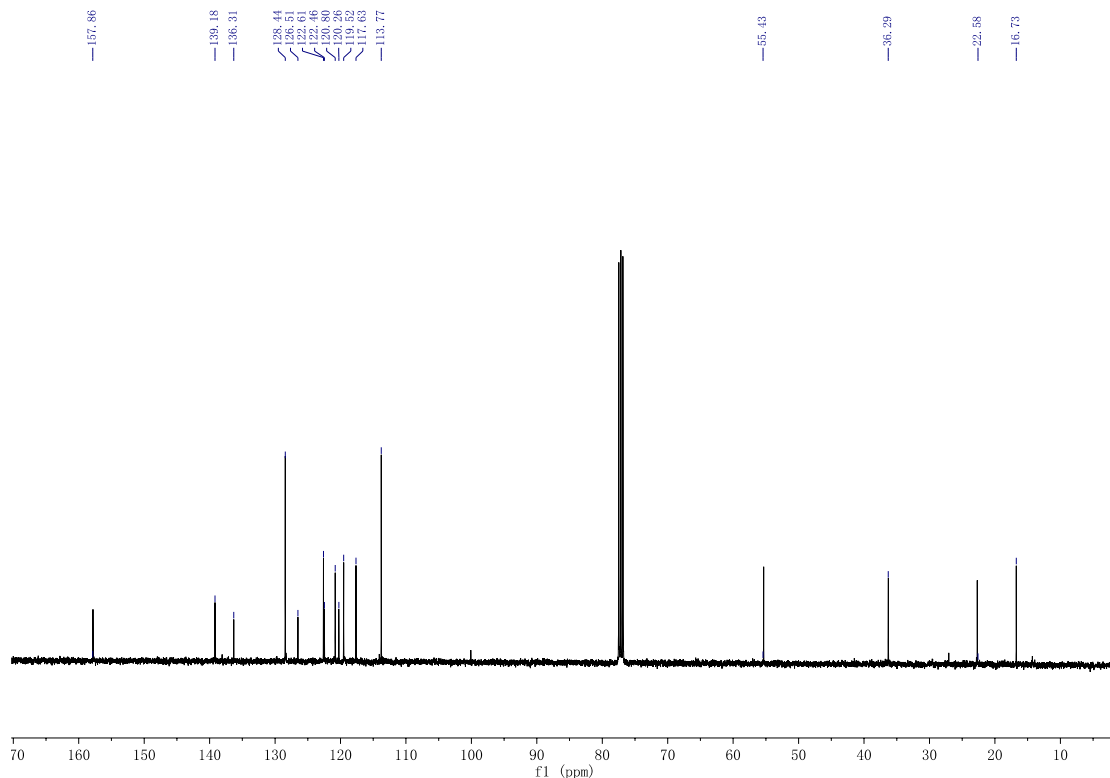
¹H NMR, CDCl₃, 400 MHz (* grease)



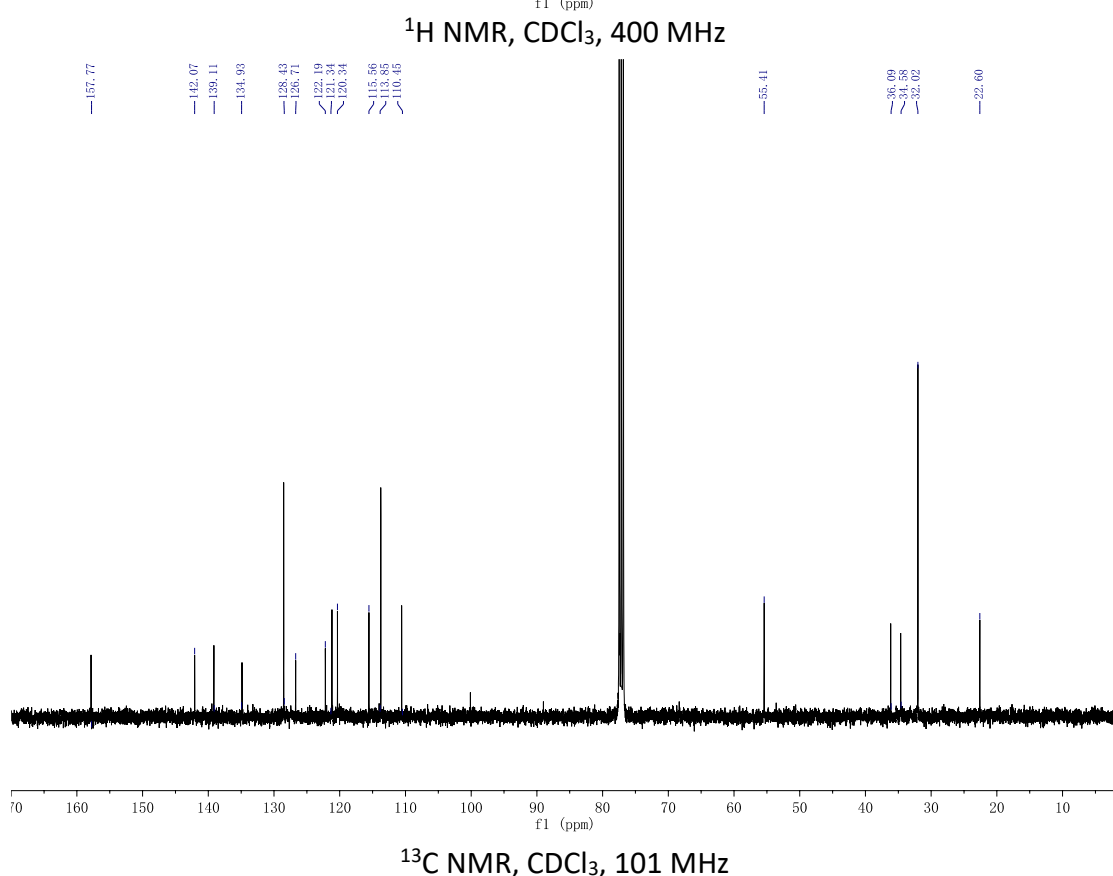
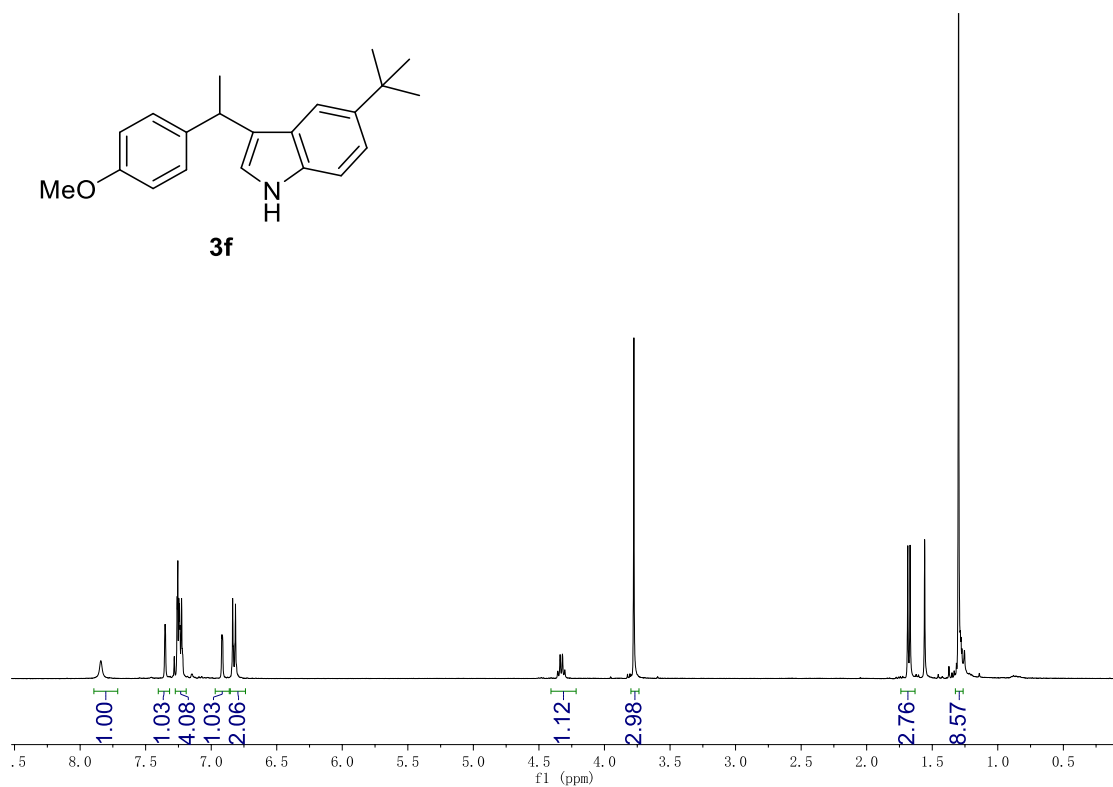
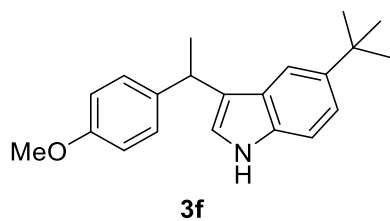
¹³C NMR, CDCl₃, 101 MHz

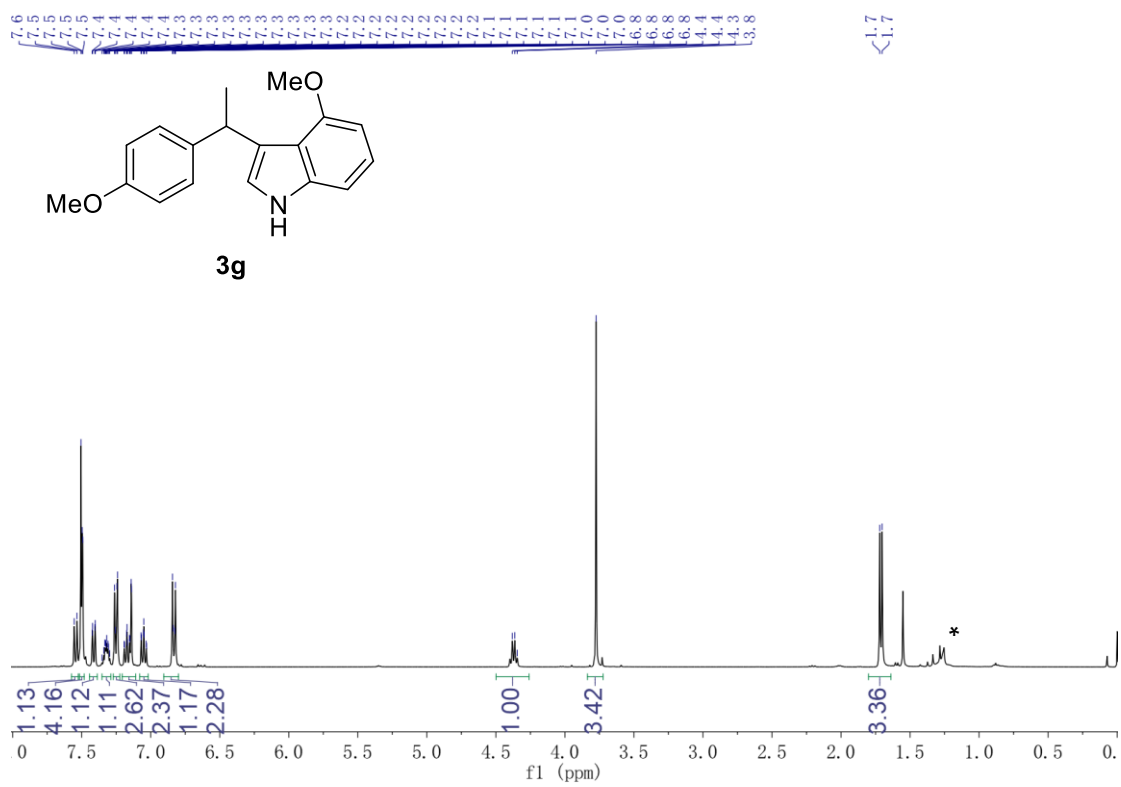


^1H NMR, CDCl_3 , 400 MHz (* grease)



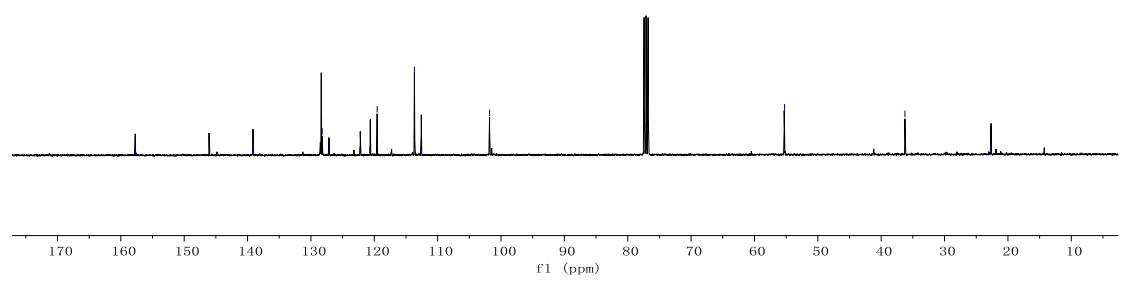
^{13}C NMR, CDCl_3 , 101 MHz



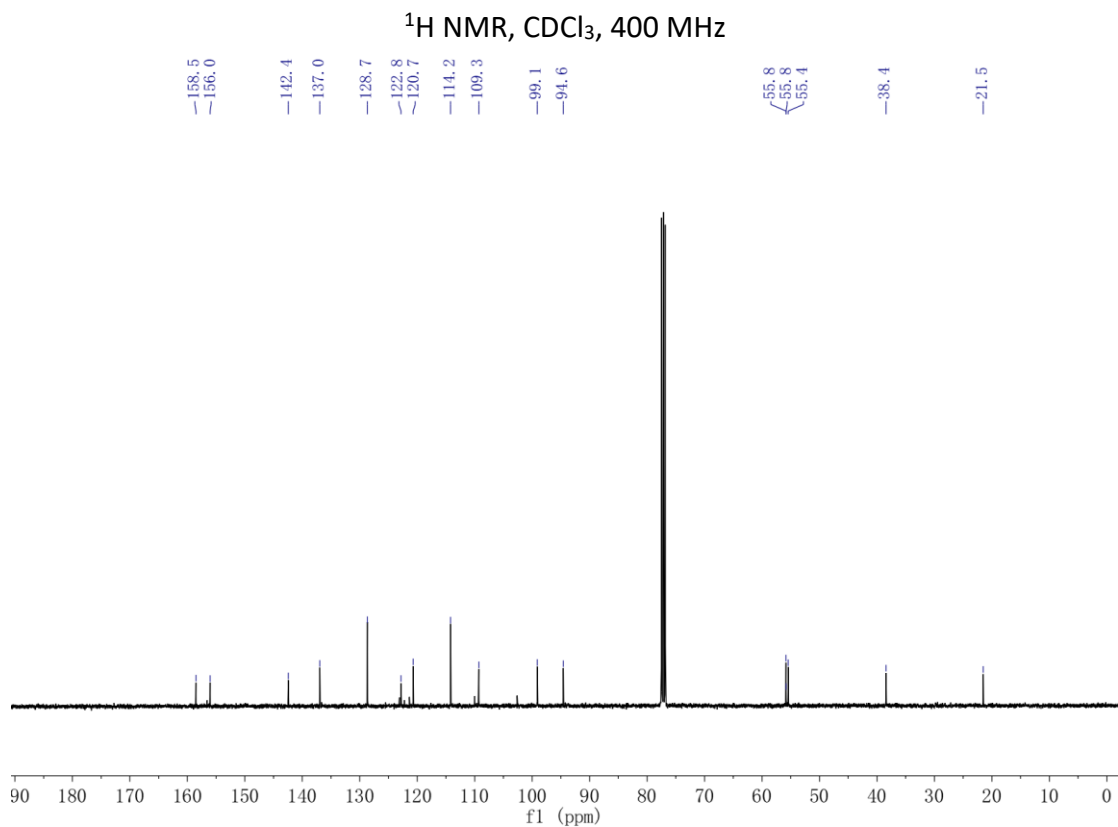
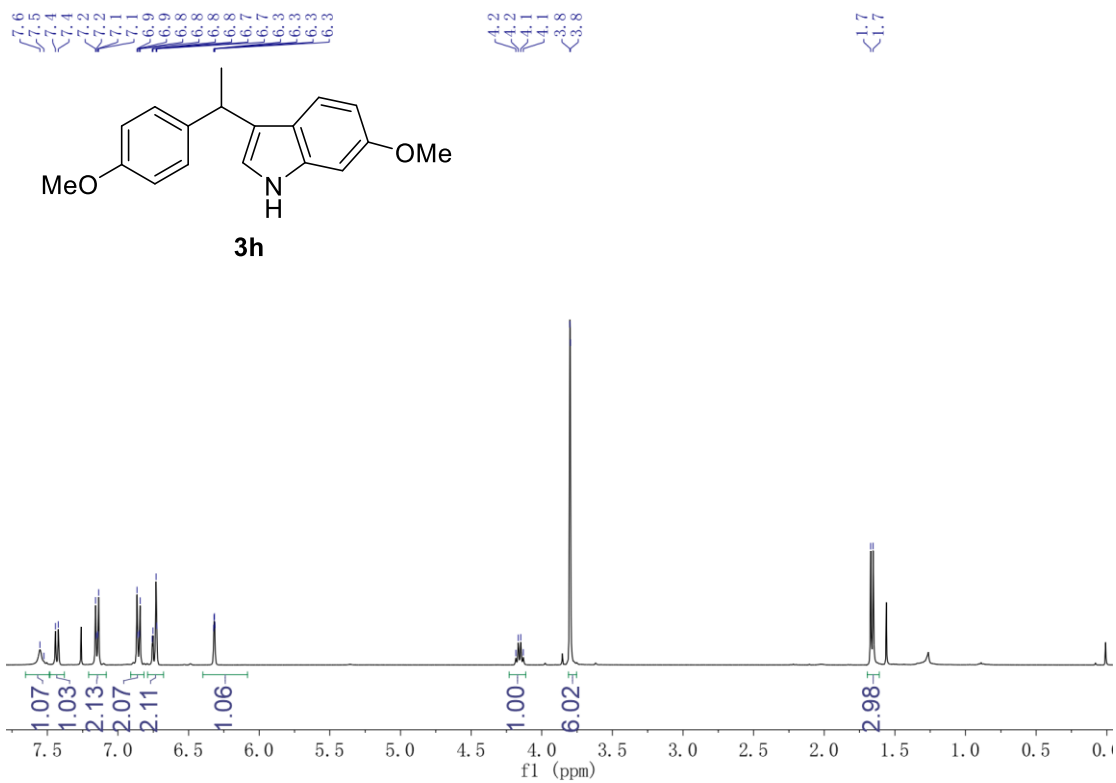


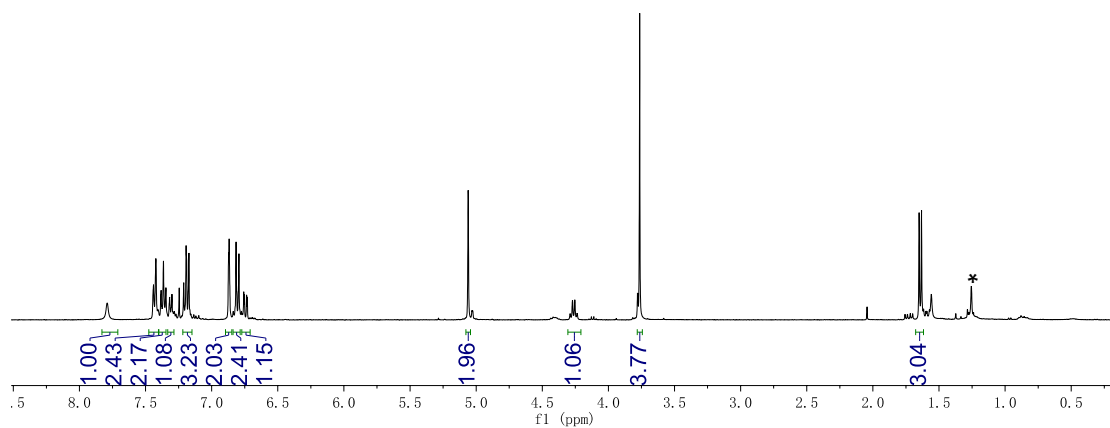
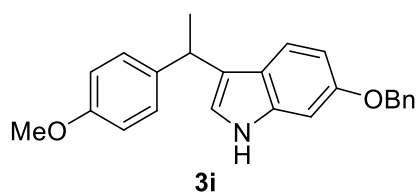
¹H NMR, CDCl₃, 400 MHz (* grease)

- 157.81
- 145.97
- 139.21
- ∨ 128.22
- ∨ 127.17
- ∨ 122.25
- ∨ 120.69
- ∨ 119.55
- ∨ 115.67
- ∨ 112.66
- 101.80
- 55.25
- 36.26
- 22.72

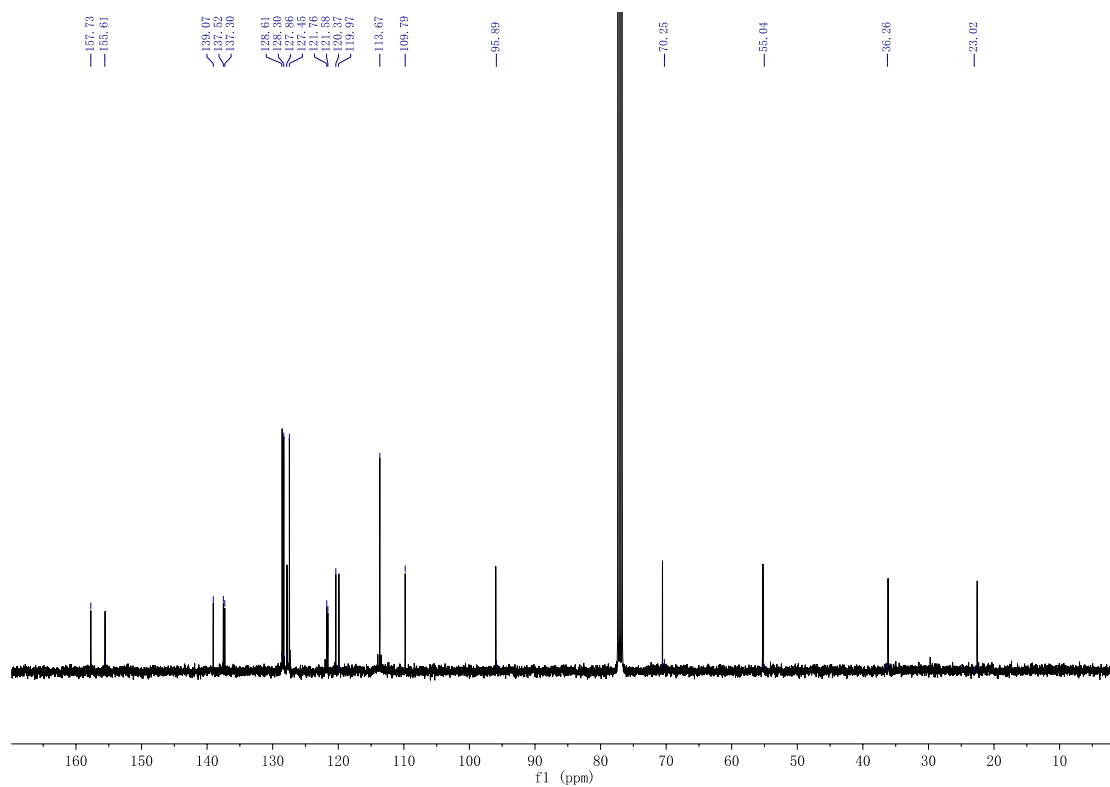


¹³C NMR, CDCl₃, 101 MHz

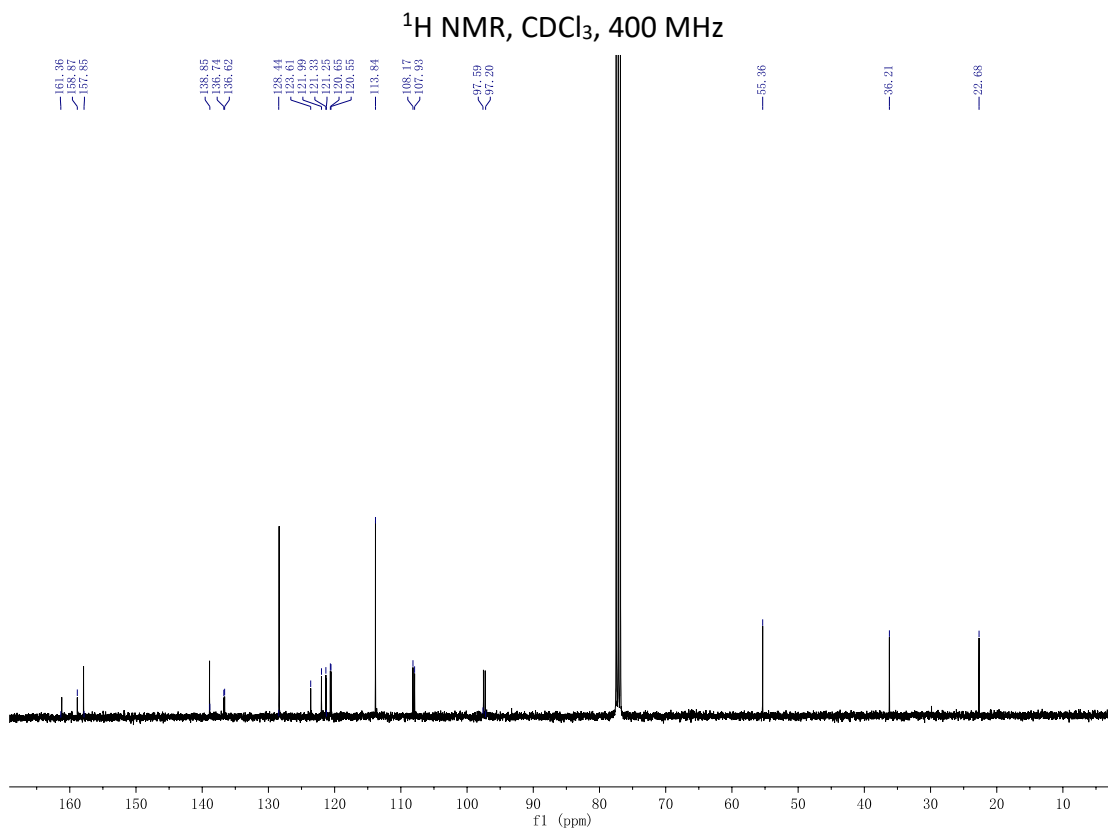
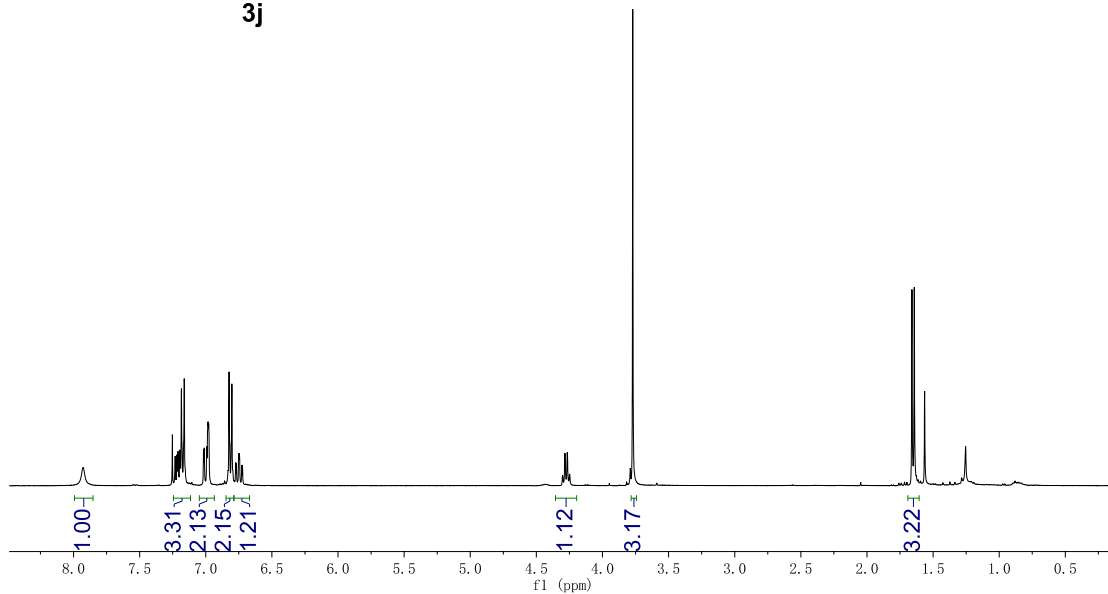
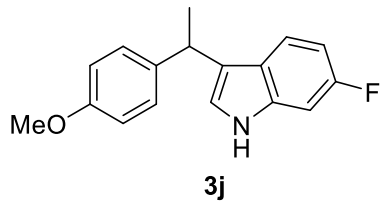


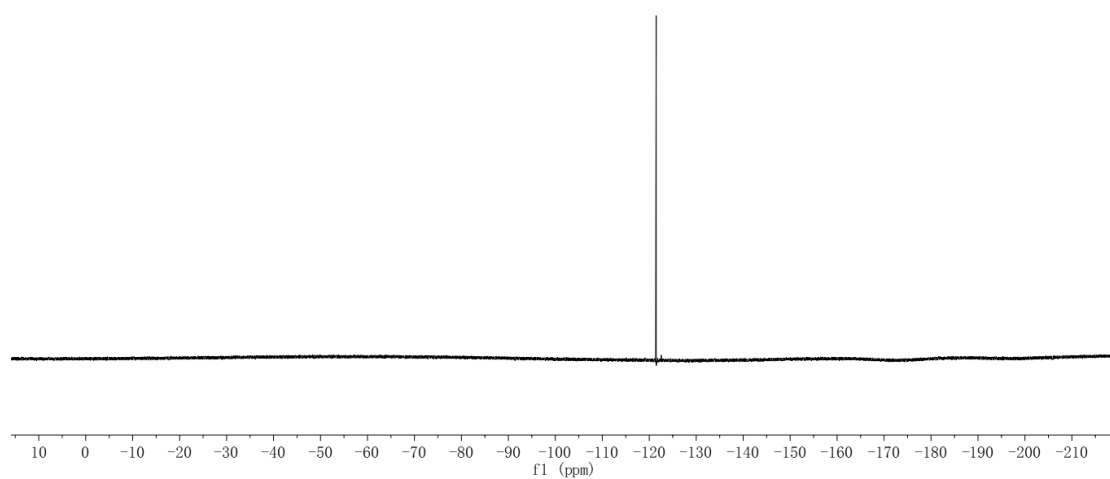


^1H NMR, CDCl_3 , 400 MHz (* grease)

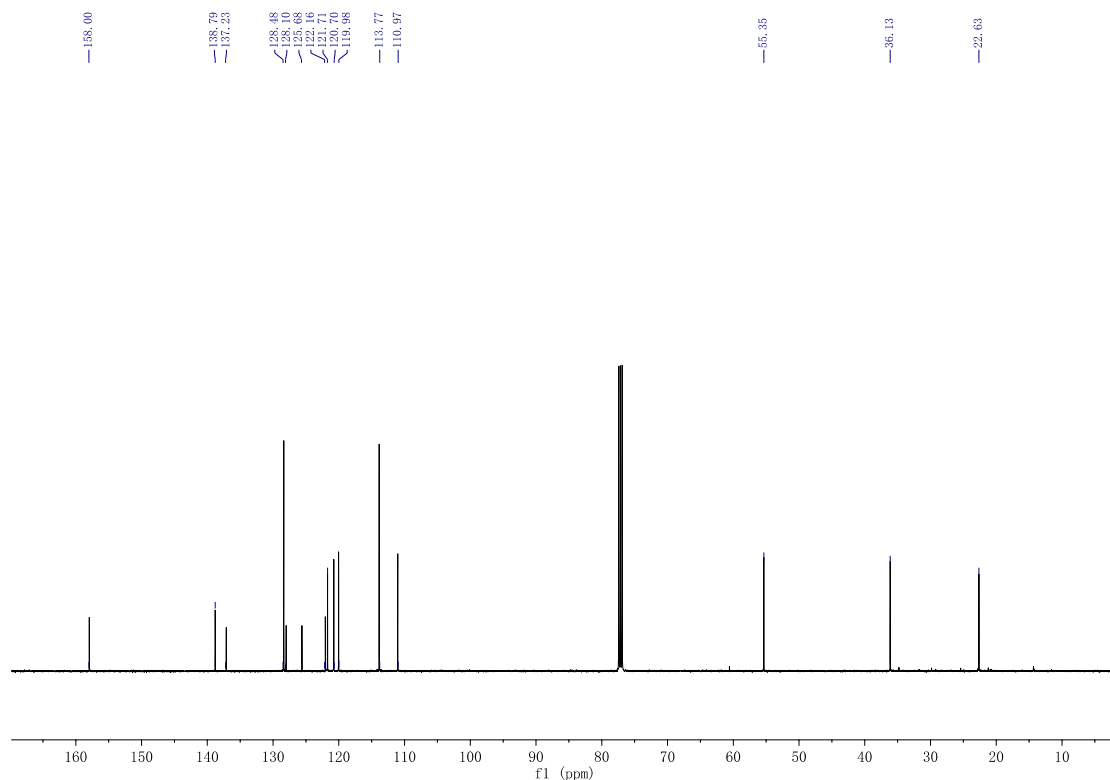
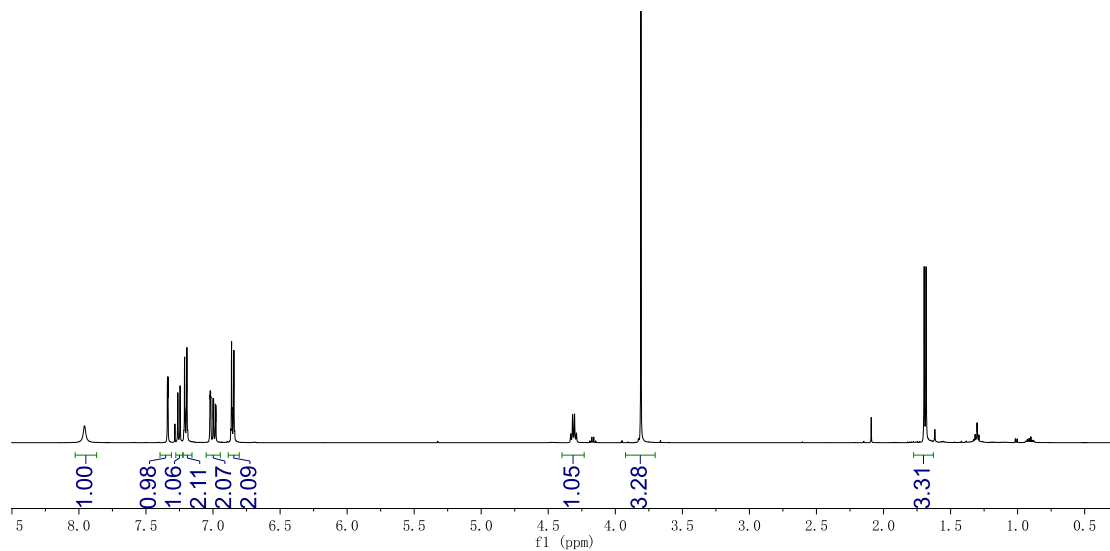
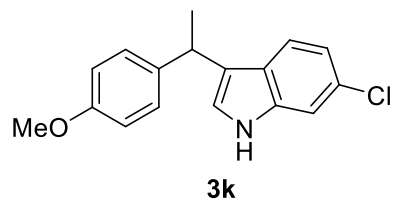


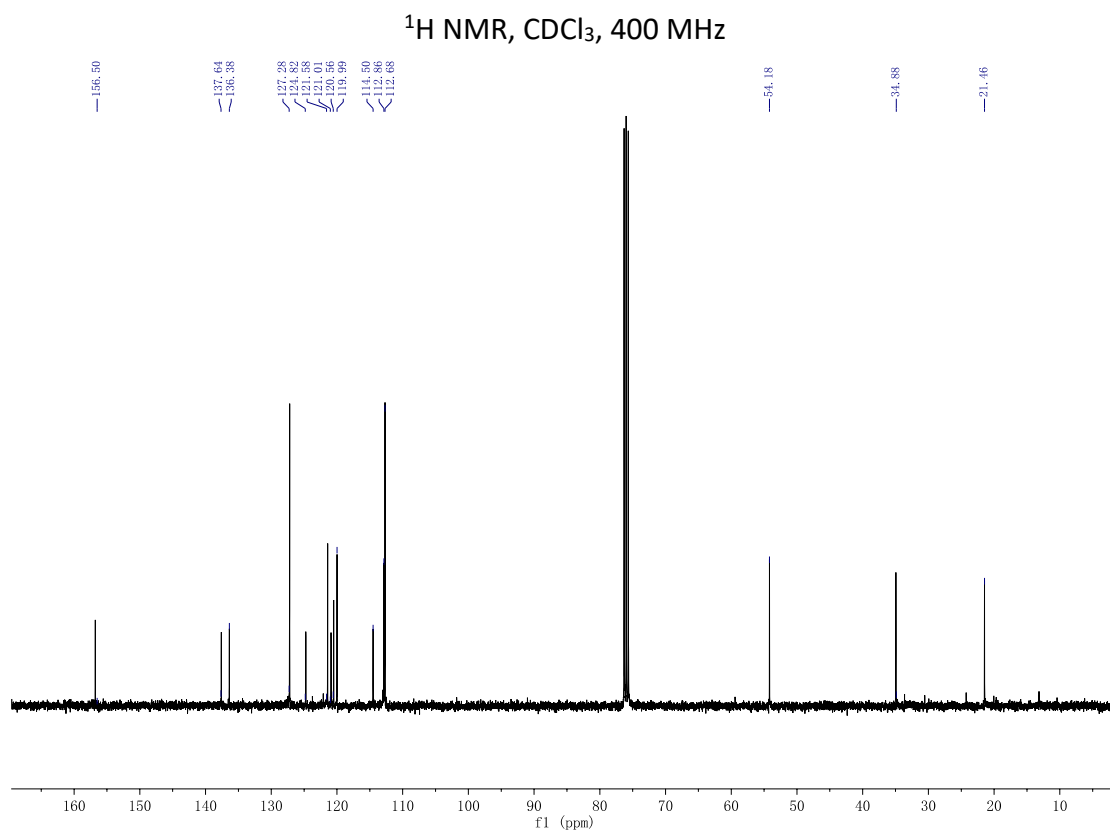
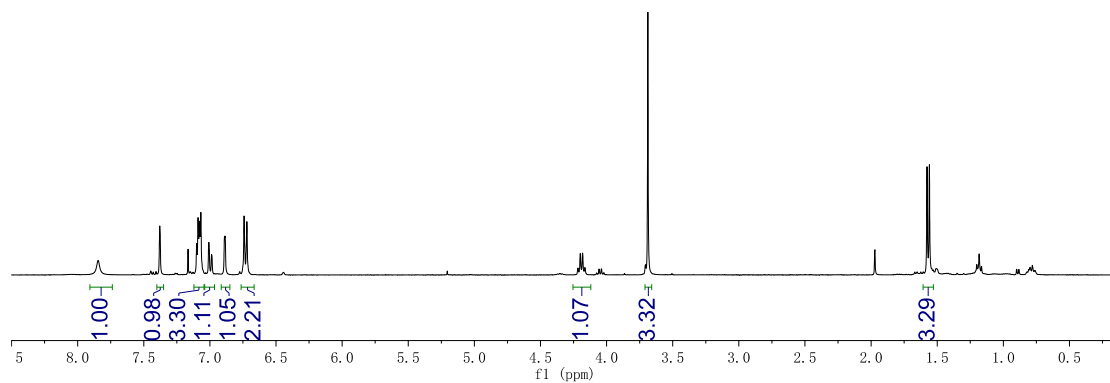
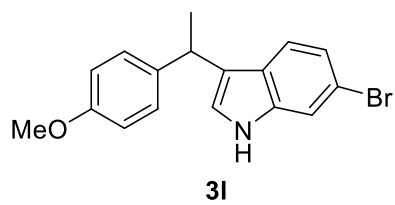
^{13}C NMR, CDCl_3 , 101 MHz

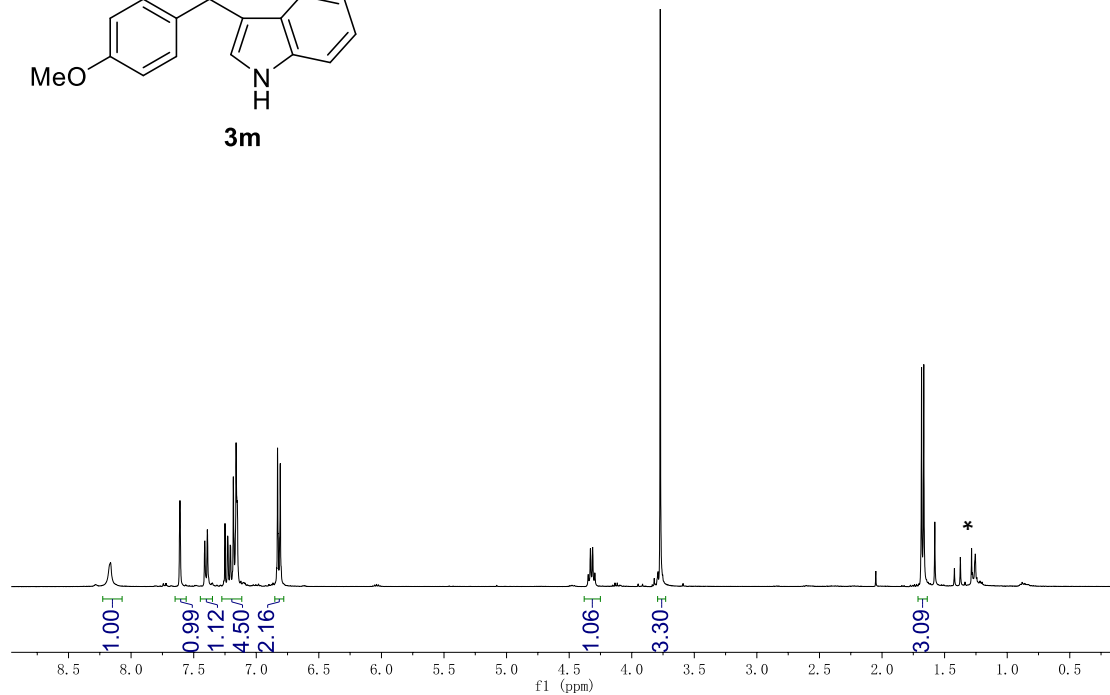
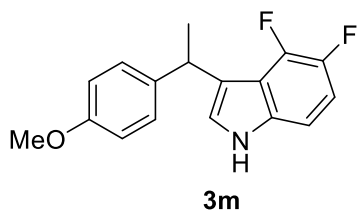




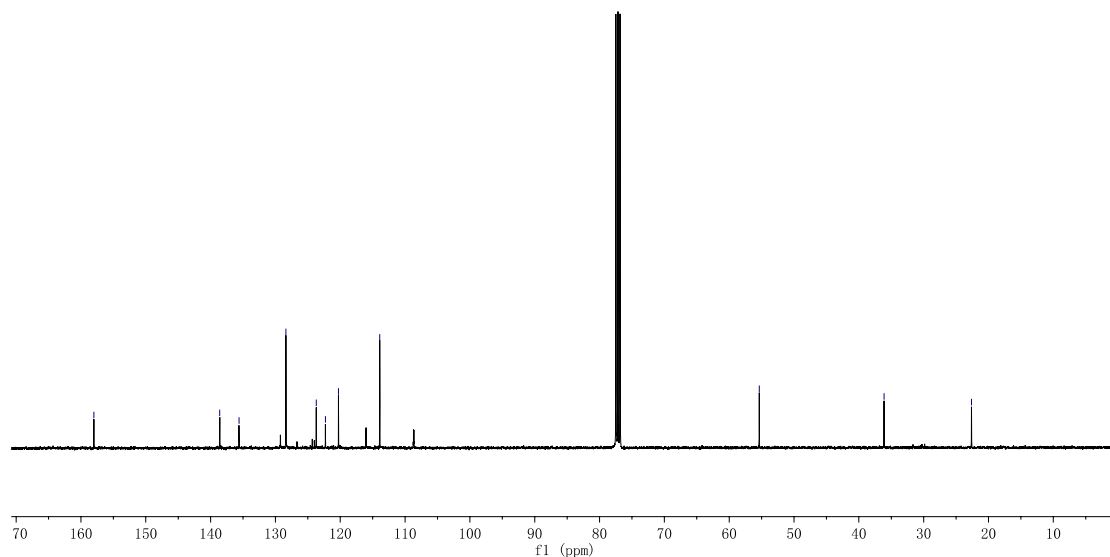
^{19}F NMR, CDCl_3 , 471 MHz

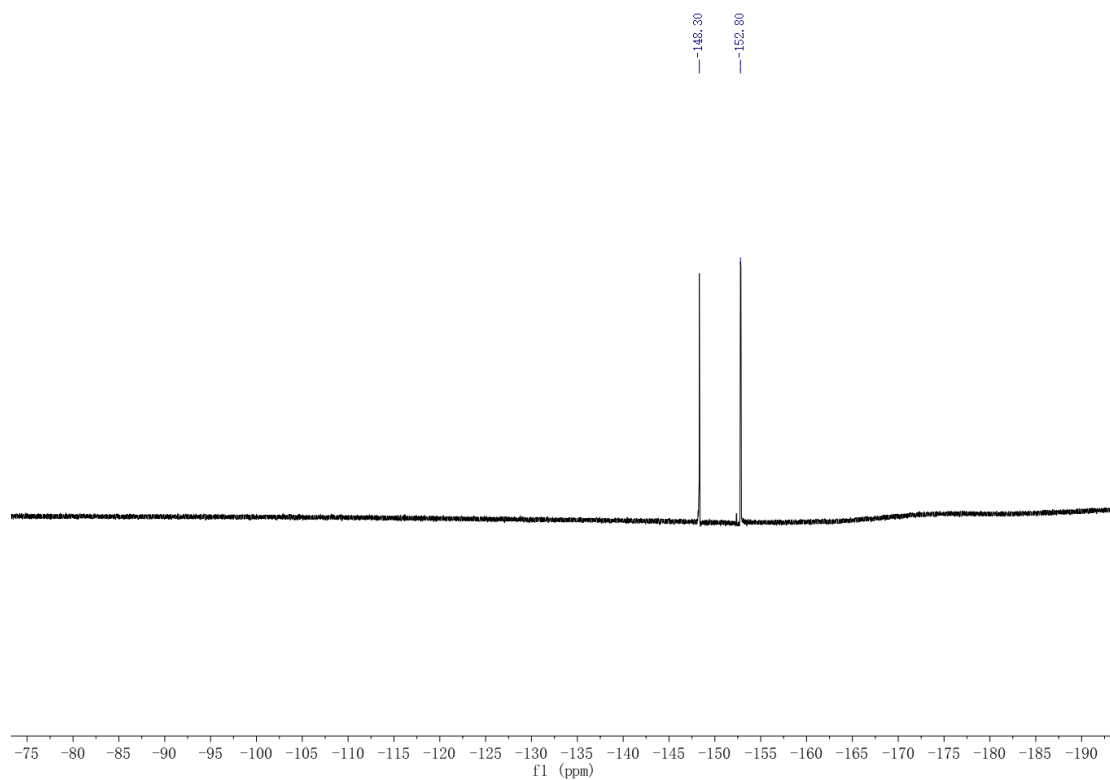




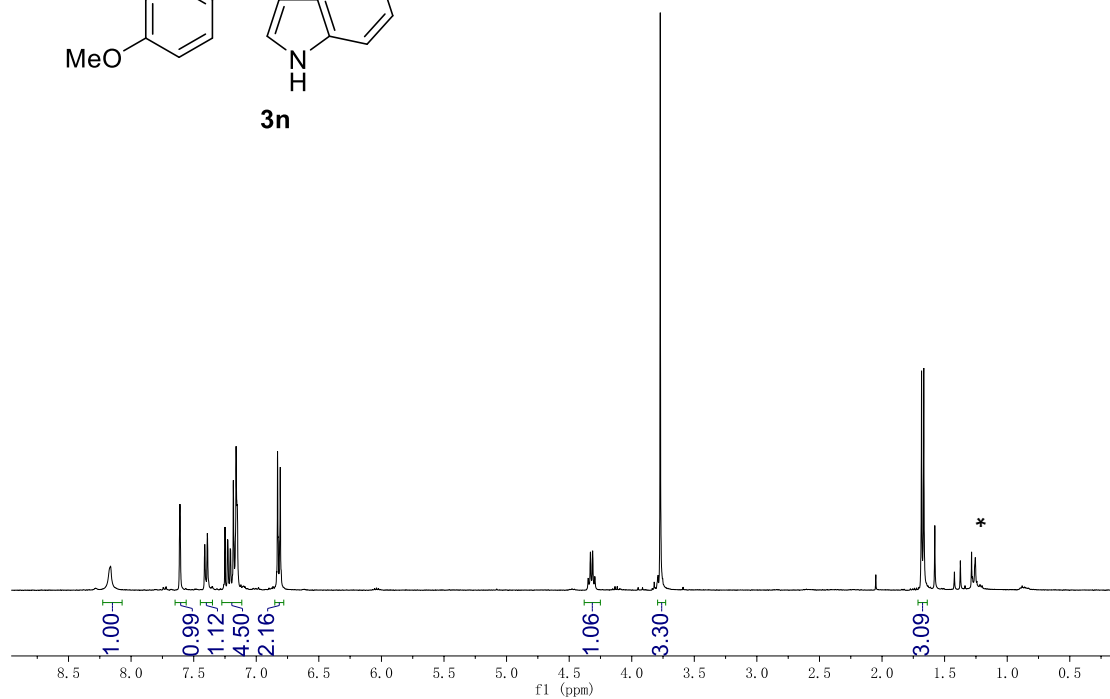
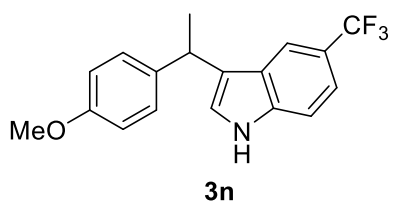


— 158.01 — 138.59 — 135.62 — 128.38 — 125.70 — 122.76 — 120.27 — 113.90 — 55.36 — 36.12 — 22.62

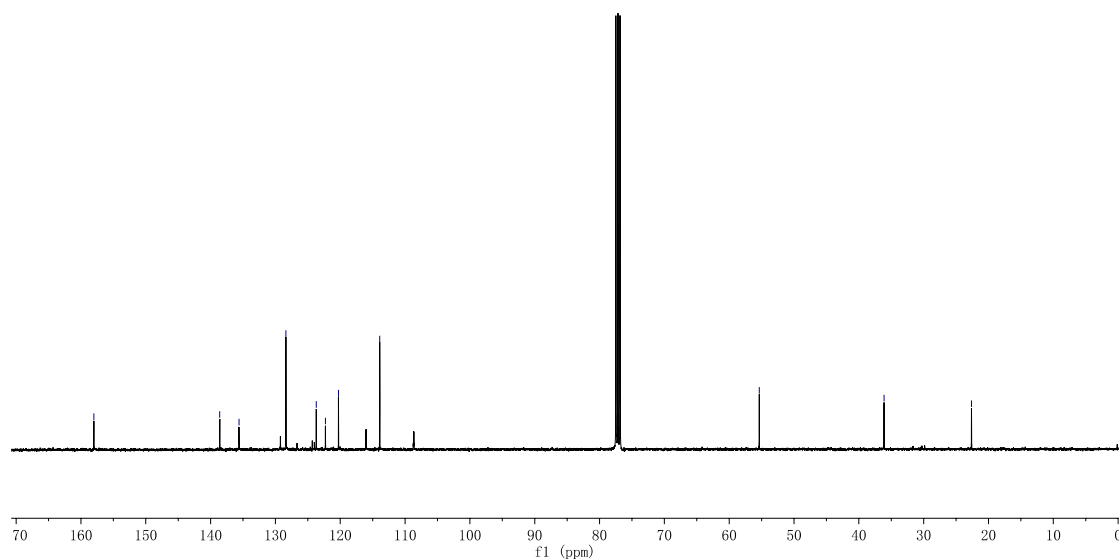




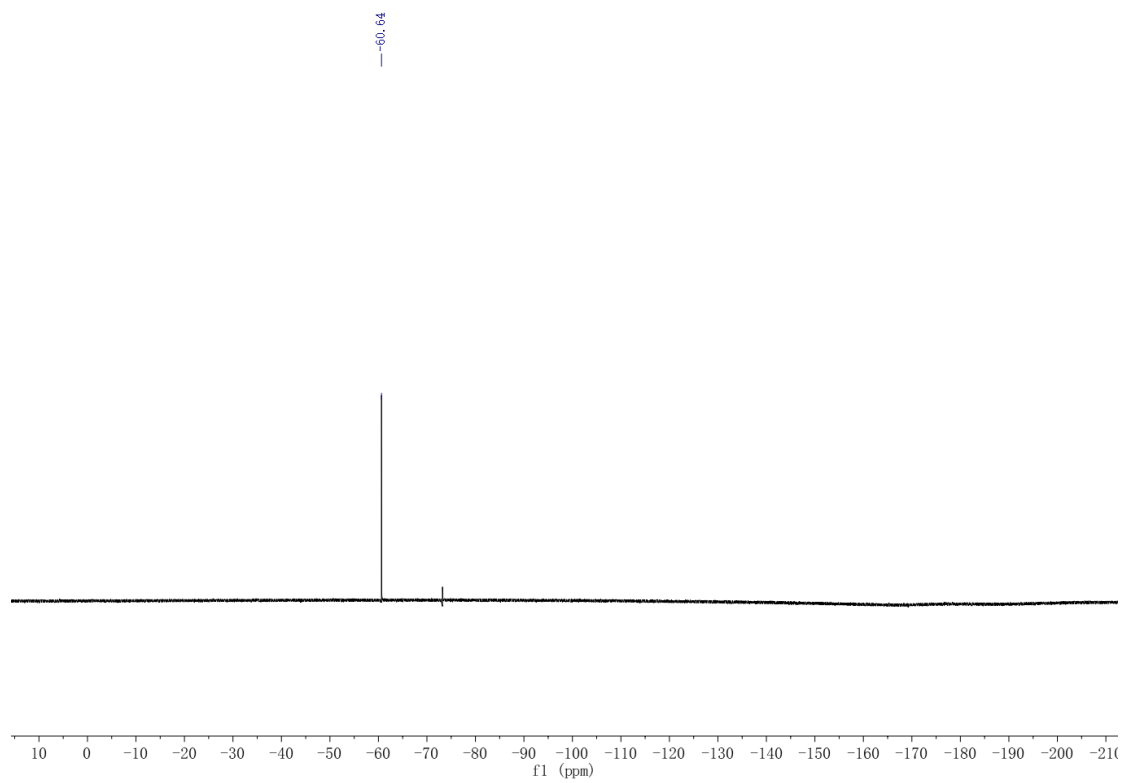
^{19}F NMR, CDCl_3 , 471 MHz



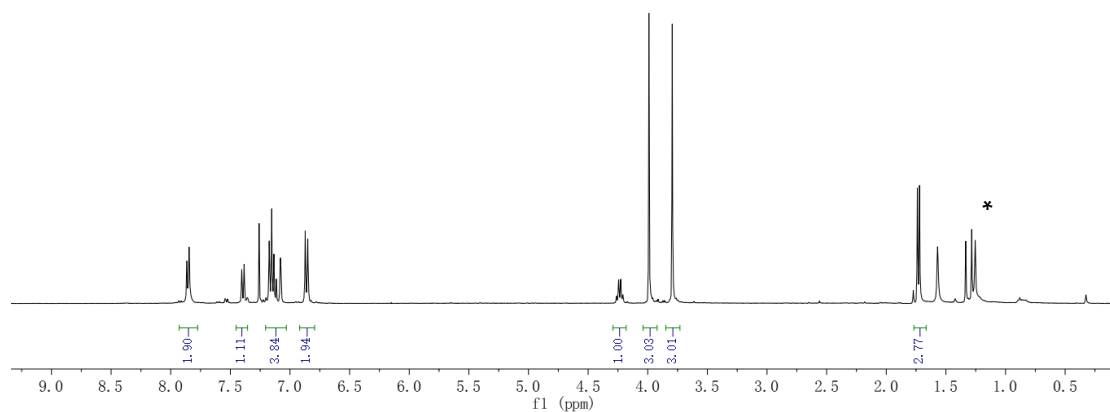
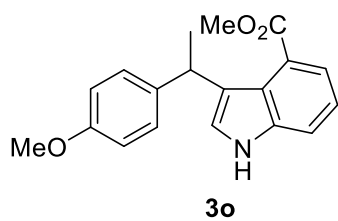
¹H NMR, CDCl₃, 400 MHz (* grease)



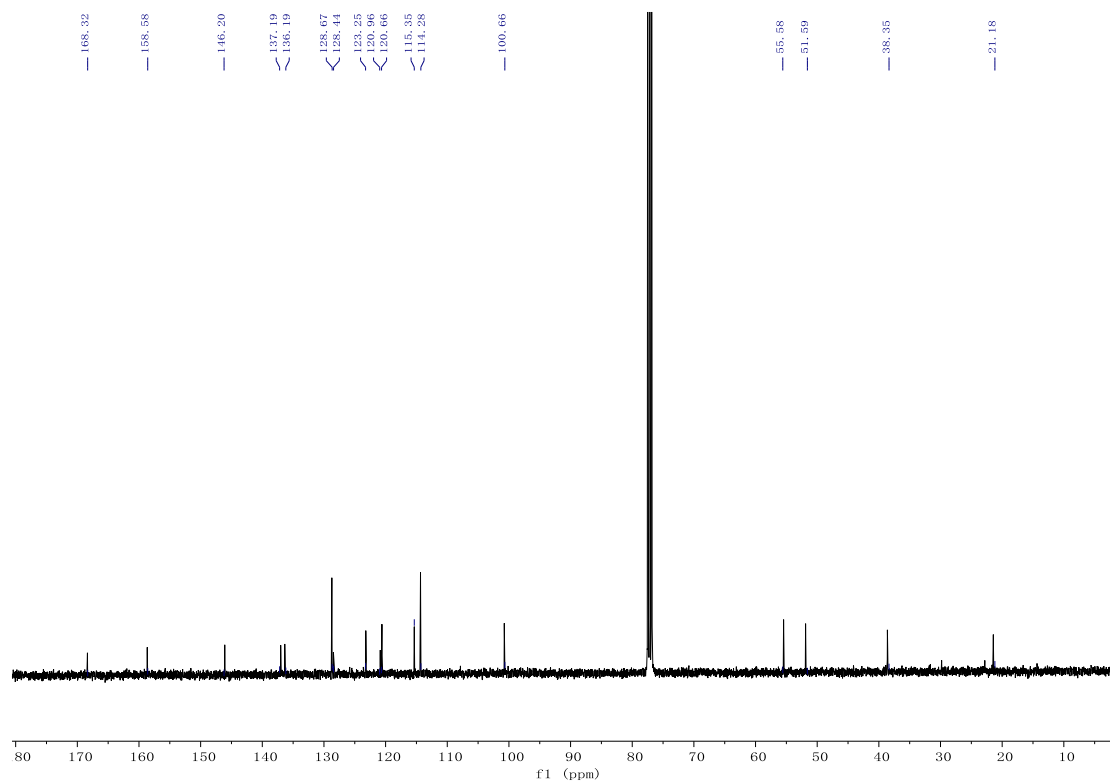
¹³C NMR, CDCl₃, 101 MHz



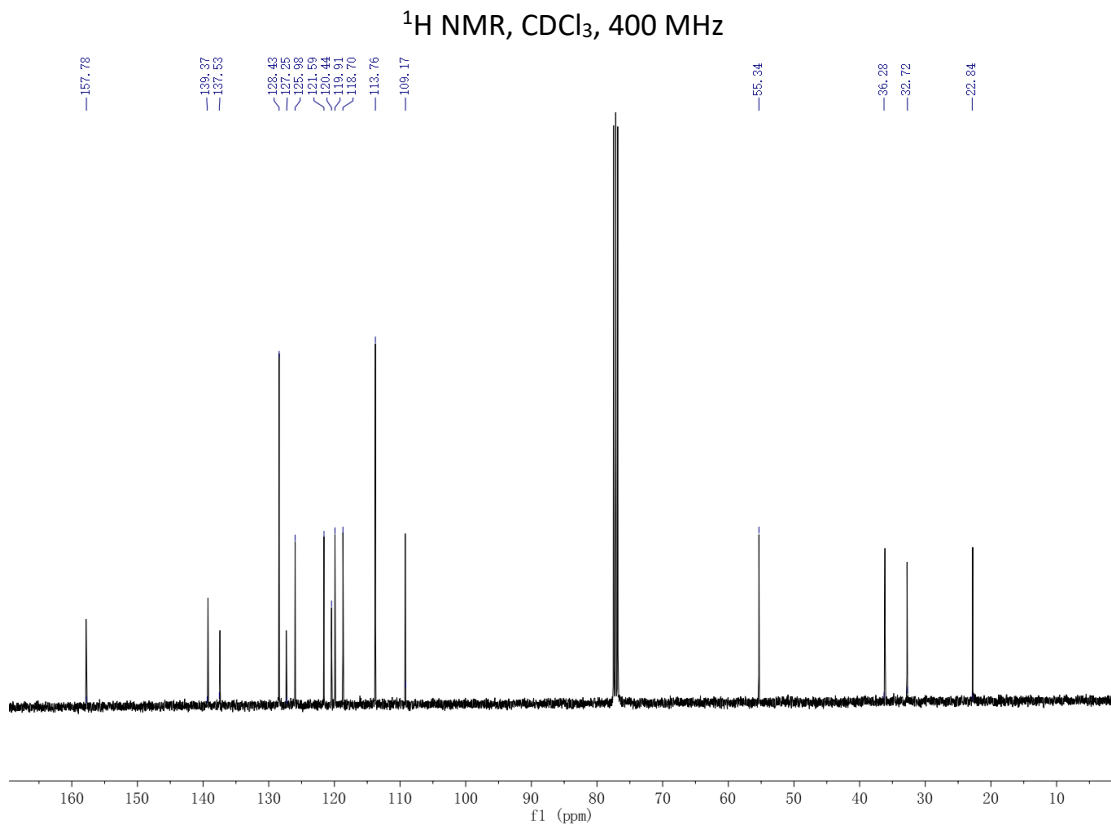
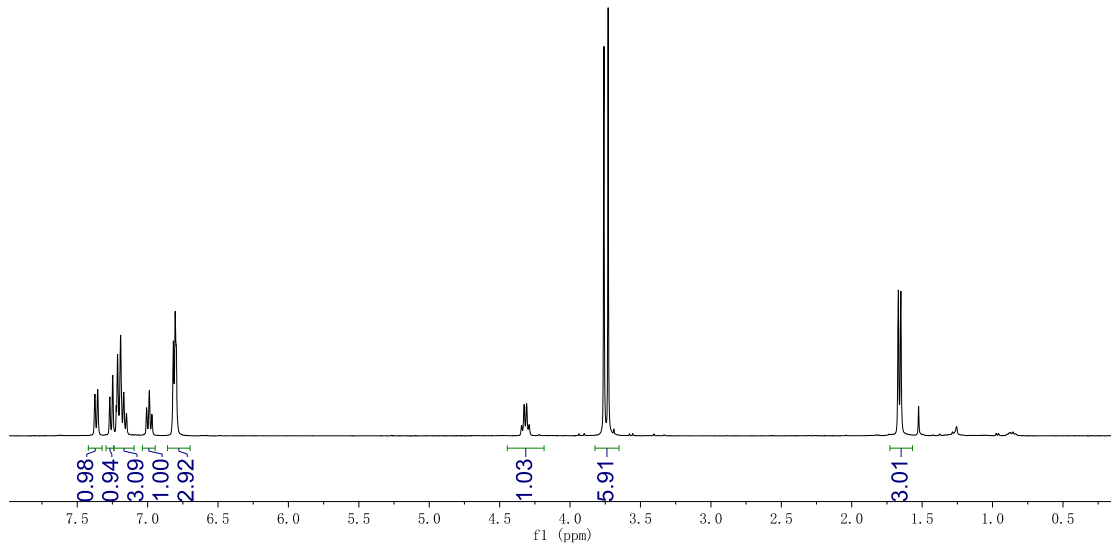
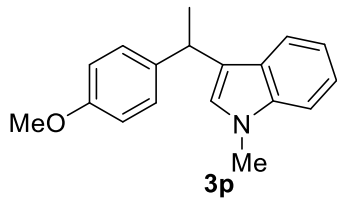
^{19}F NMR, CDCl_3 , 471 MHz

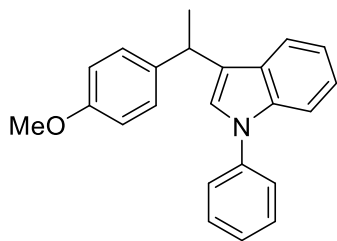


¹H NMR, CDCl₃, 400 MHz(* grease)

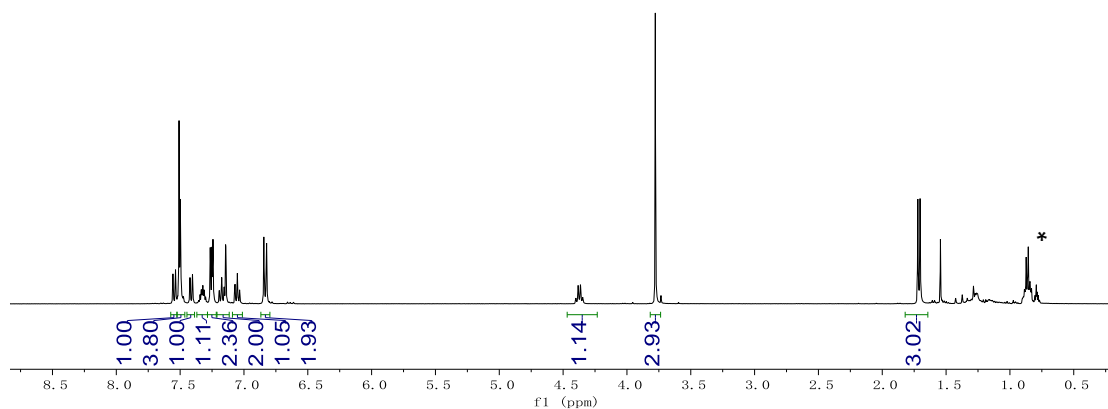


¹³C NMR, CDCl₃, 101 MHz

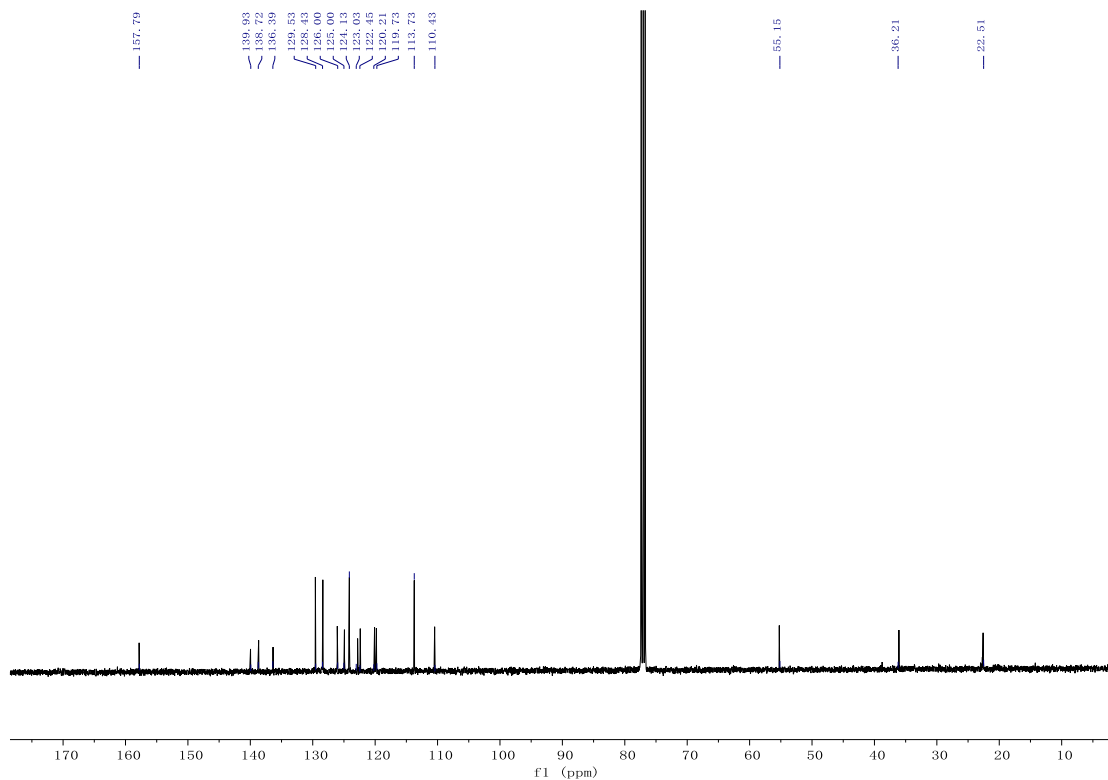




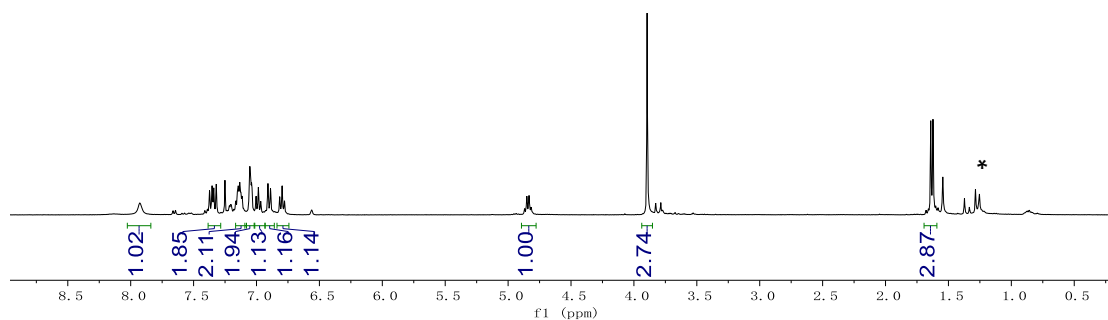
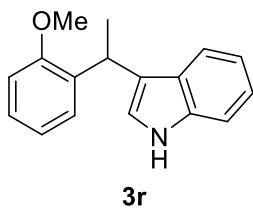
3q



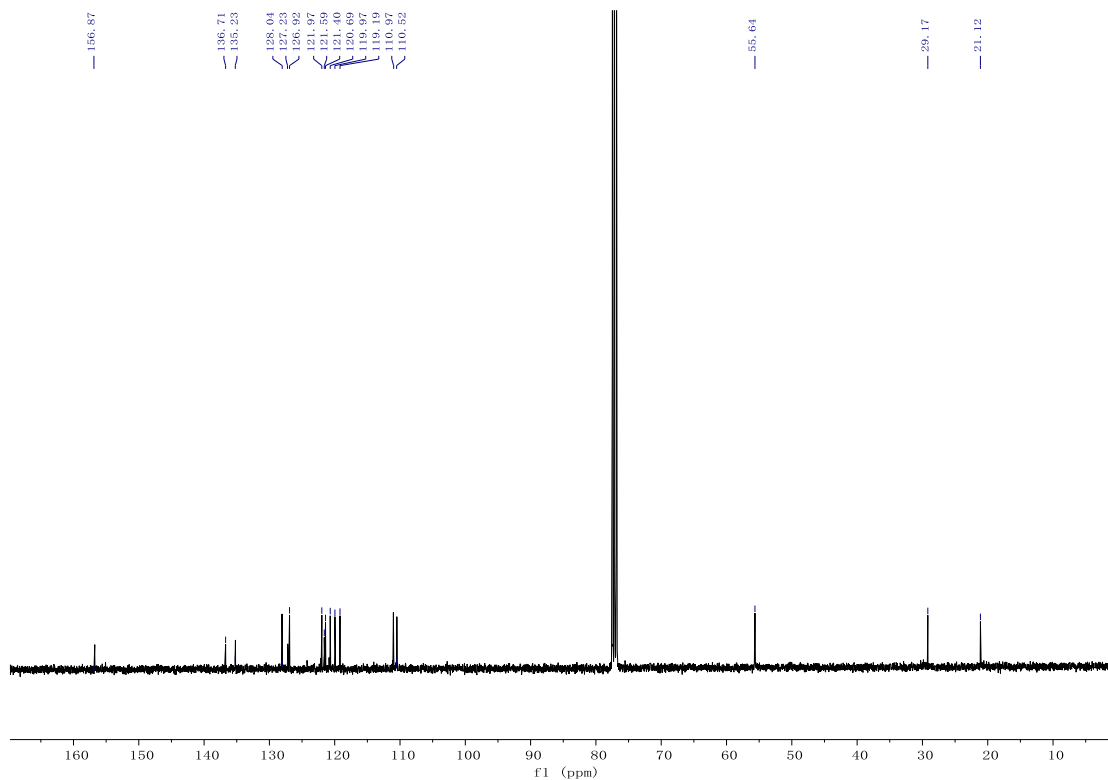
¹H NMR, CDCl₃, 400 MHz(* grease)



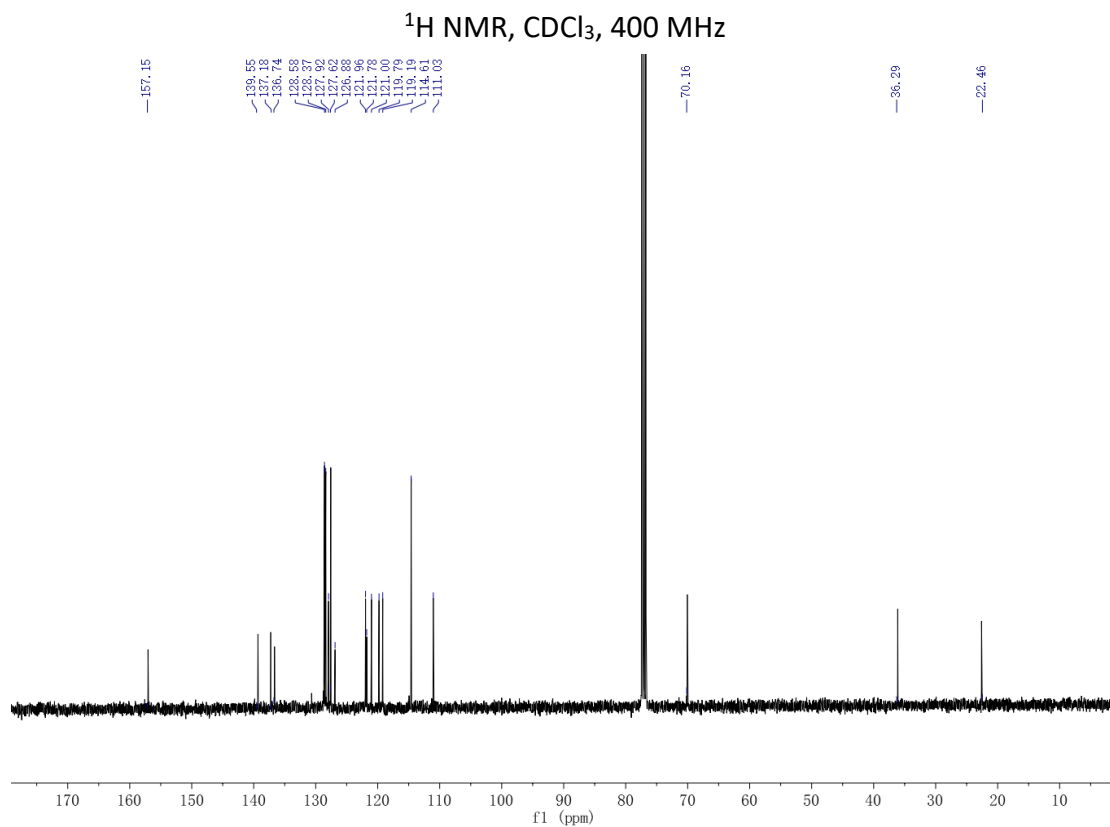
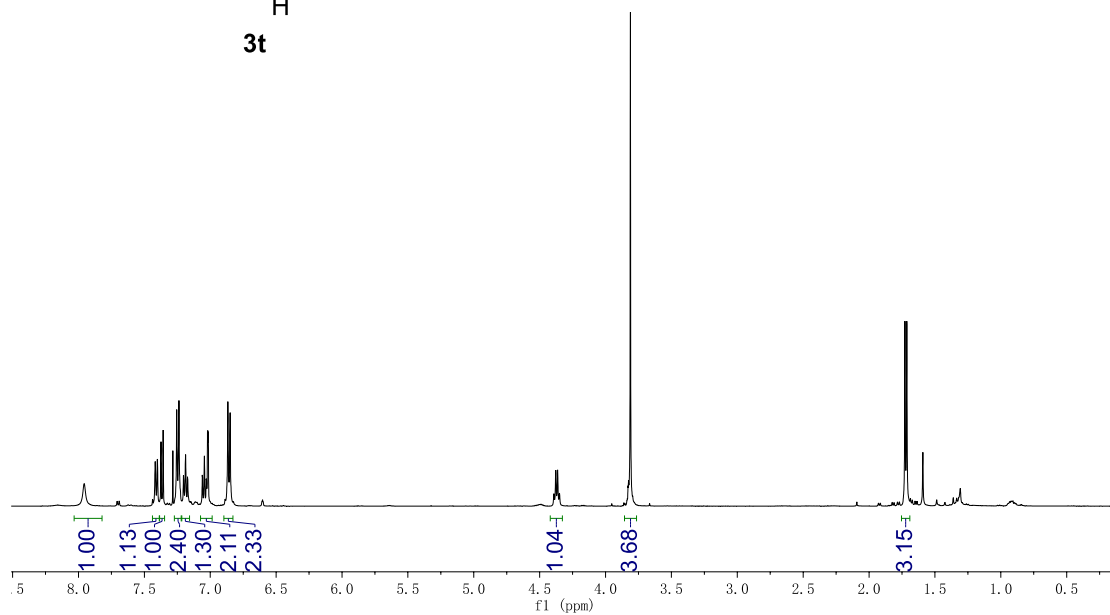
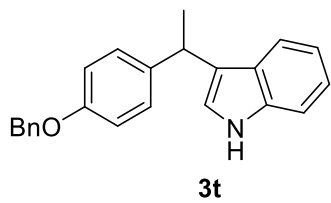
¹³C NMR, CDCl₃, 101 MHz

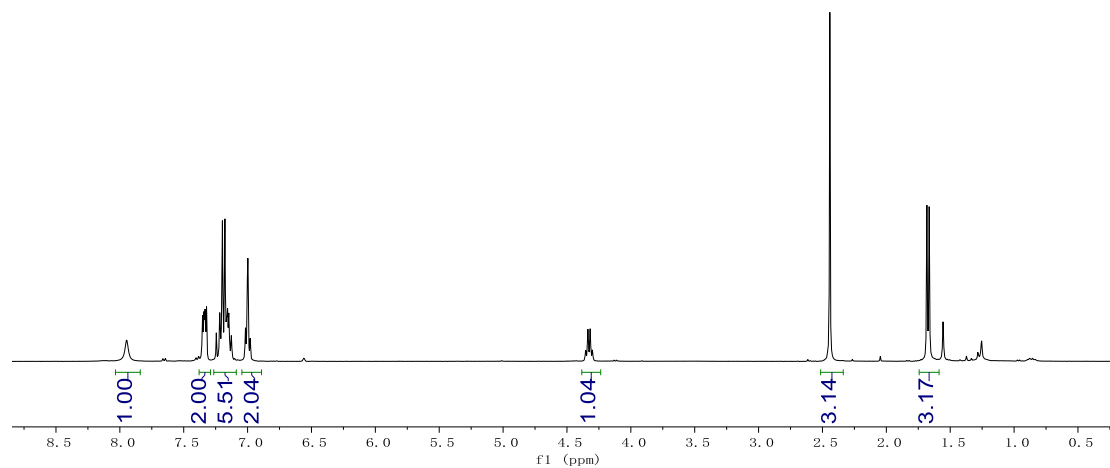
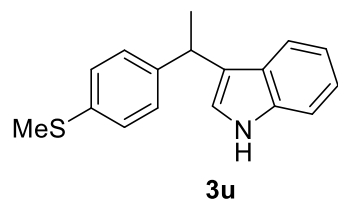


¹H NMR, CDCl₃, 400 MHz(* grease)



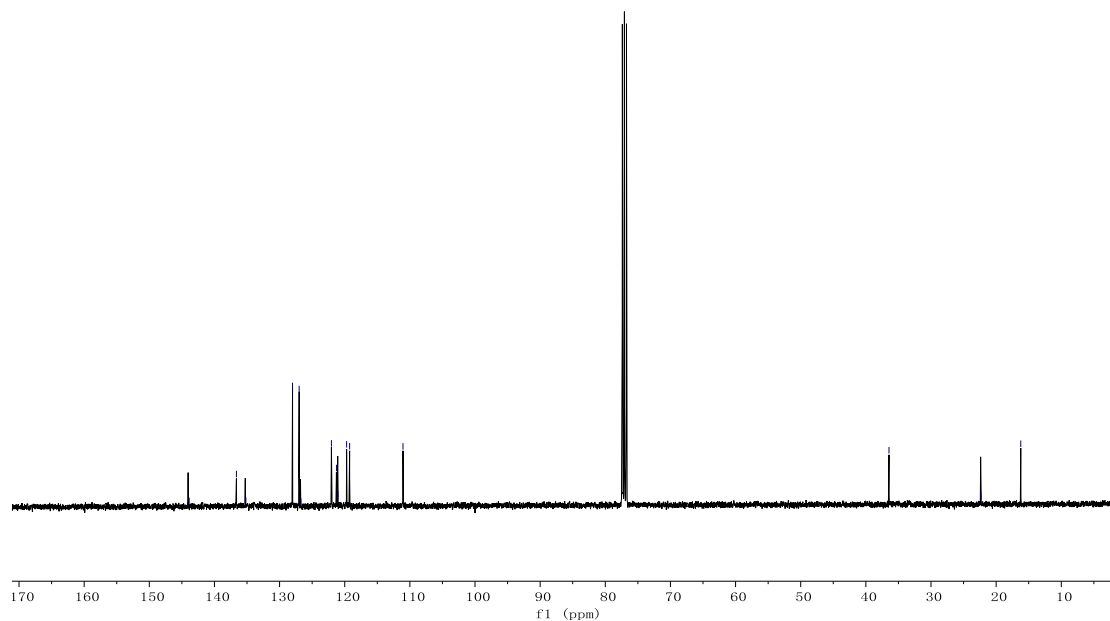
¹³C NMR, CDCl₃, 101 MHz

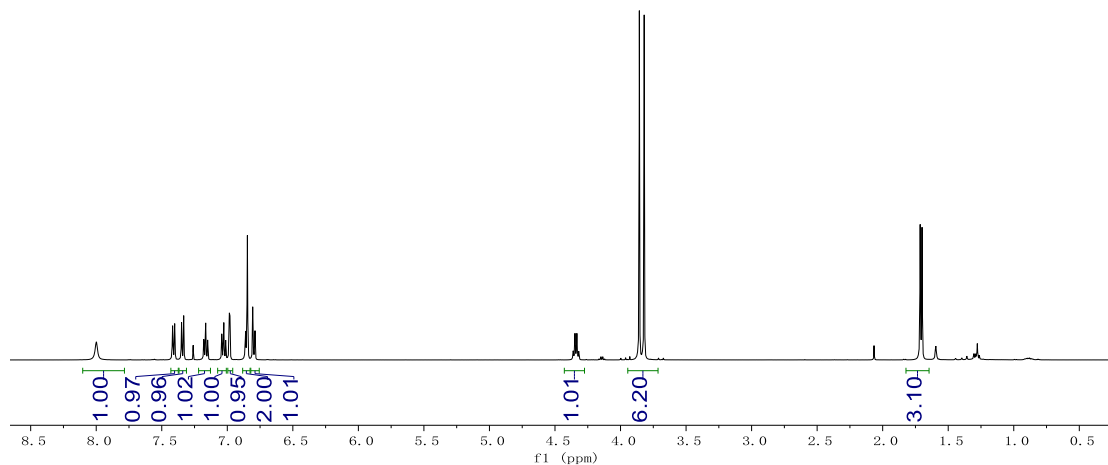
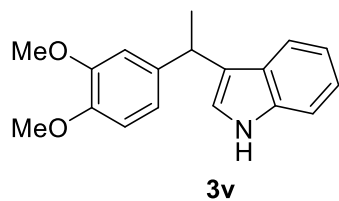




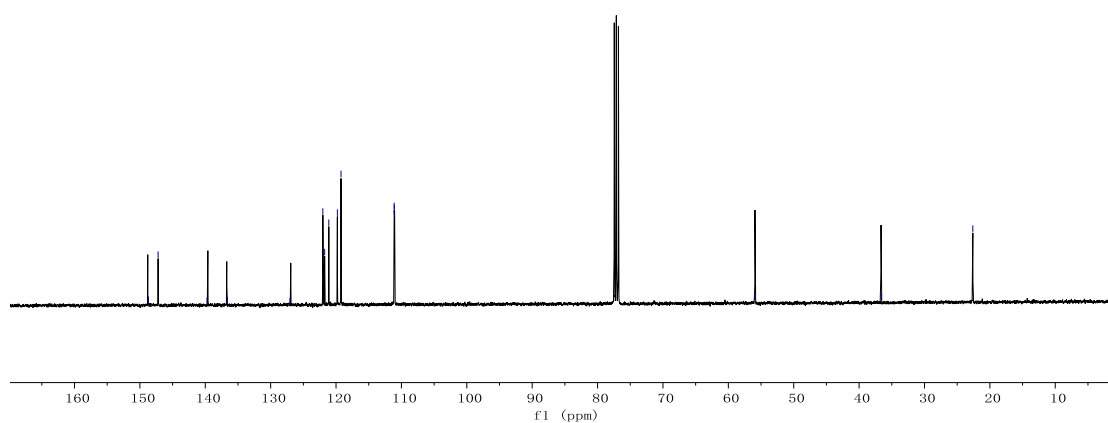
143.88
136.63
135.18
129.01
127.00
126.72
122.04
121.27
121.04
119.51
119.23
111.06

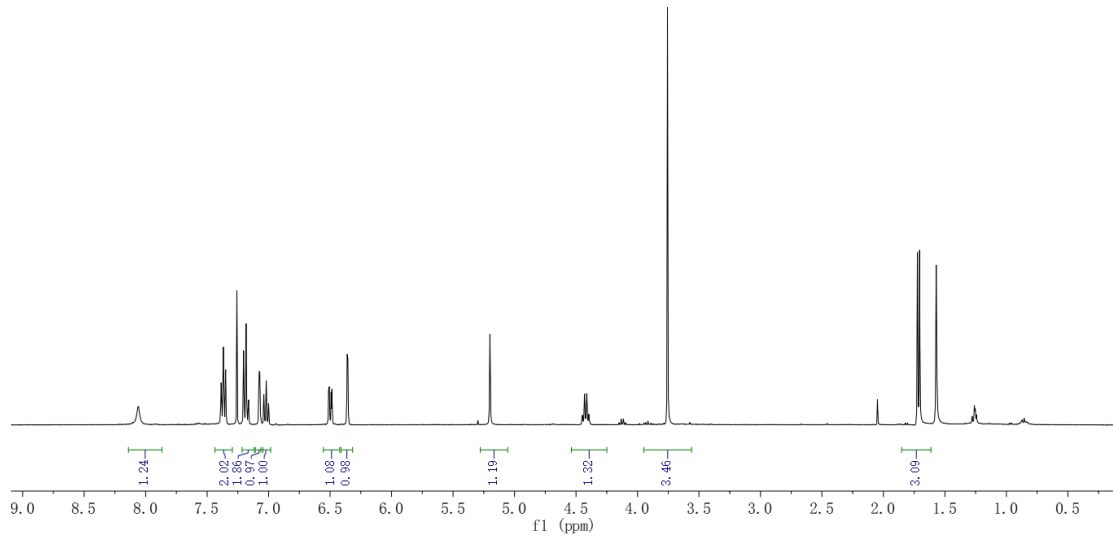
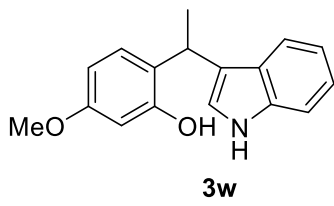
36.45
22.34
16.22



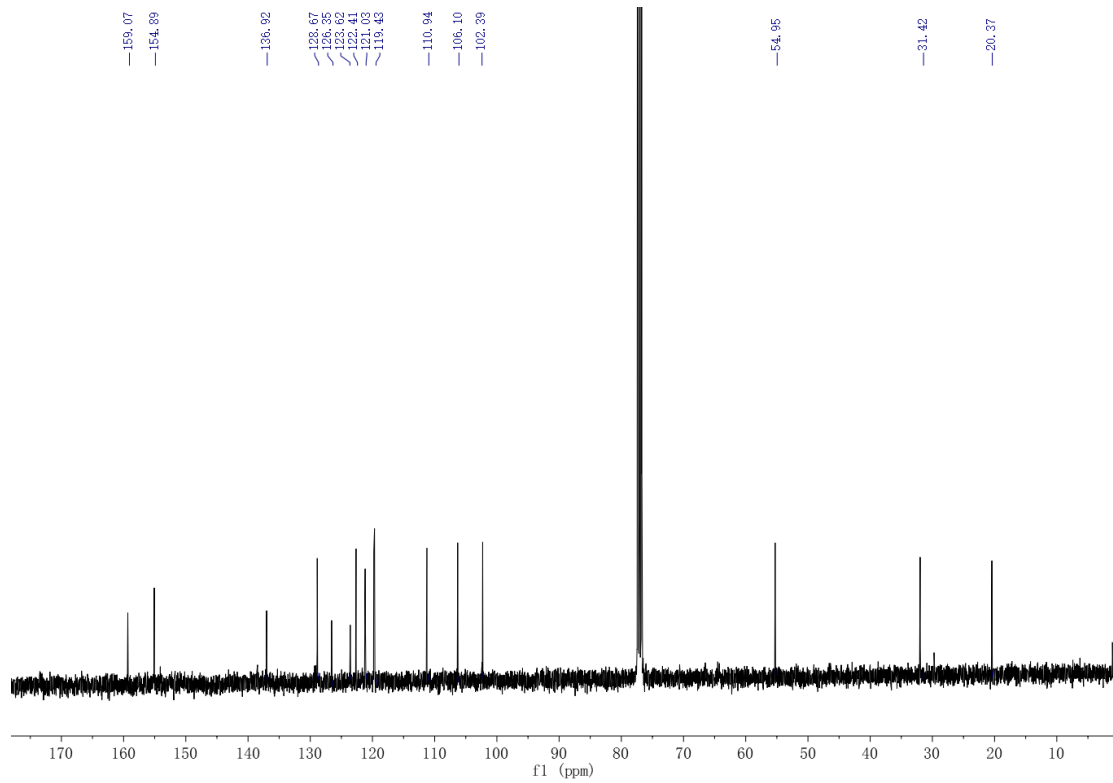


148.69
147.21
136.72
136.64
127.04
122.02
121.74
121.10
118.80
116.24
111.11
111.09
111.04
55.99
36.59
22.61

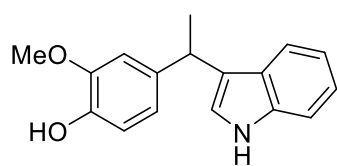




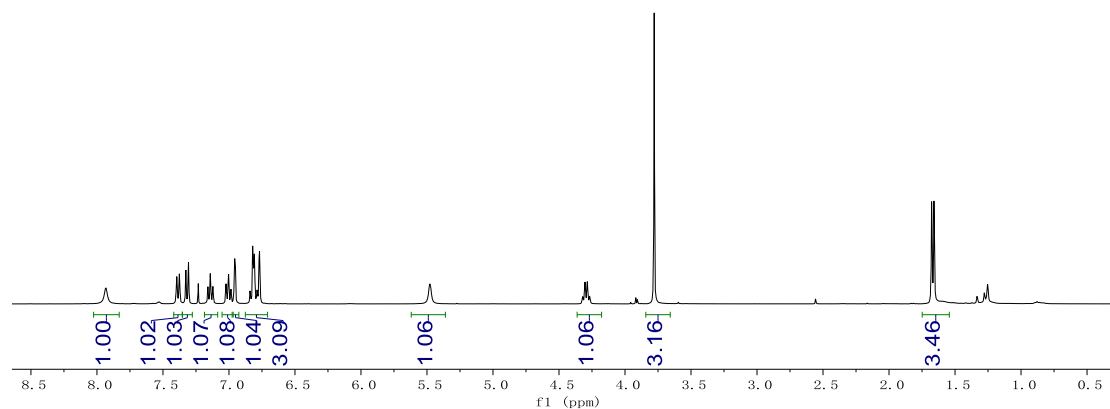
¹H NMR, CDCl₃, 400 MHz



¹³C NMR, CDCl₃, 101 MHz



3x



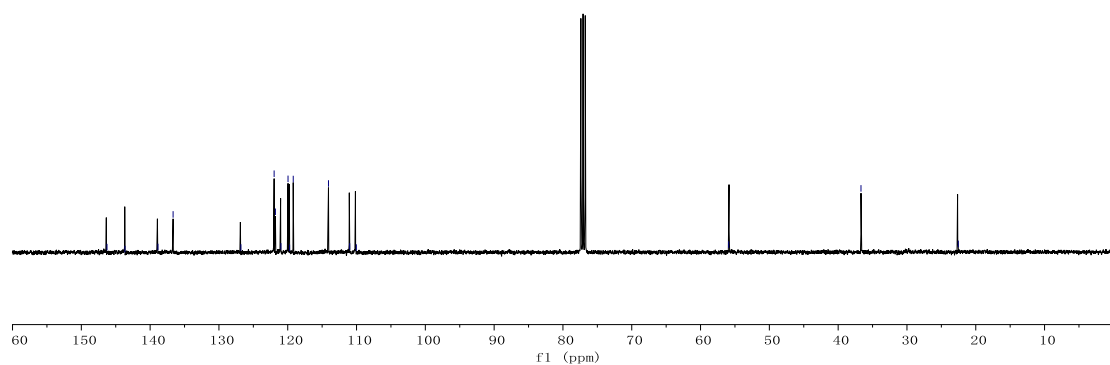
^1H NMR, CDCl_3 , 400 MHz

146.27
143.66
138.86
136.66
126.81
121.98
121.79
120.99
118.96
118.72
118.52
114.68
110.99
110.03

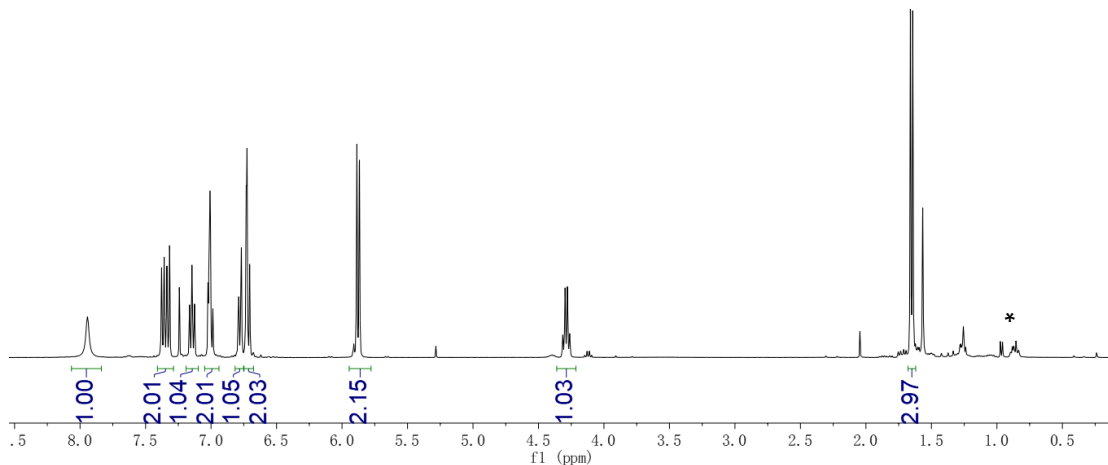
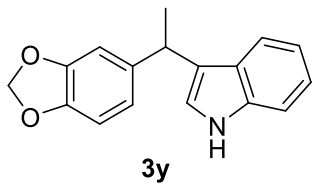
65.91

36.68

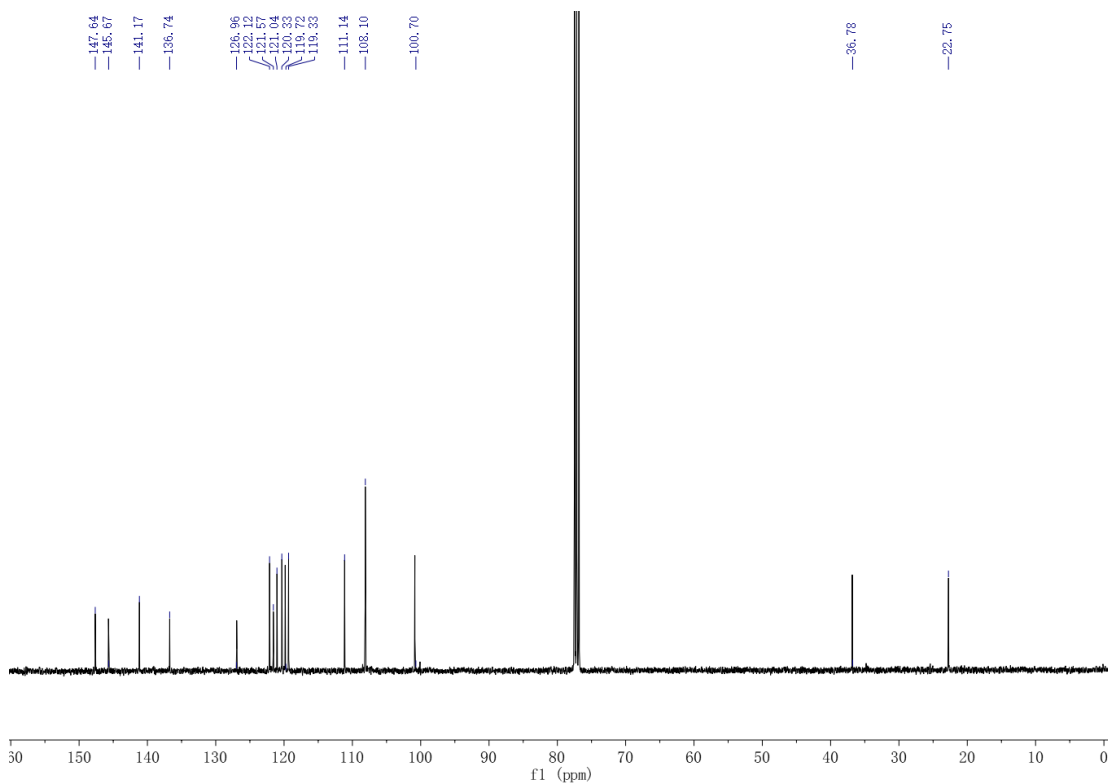
22.54



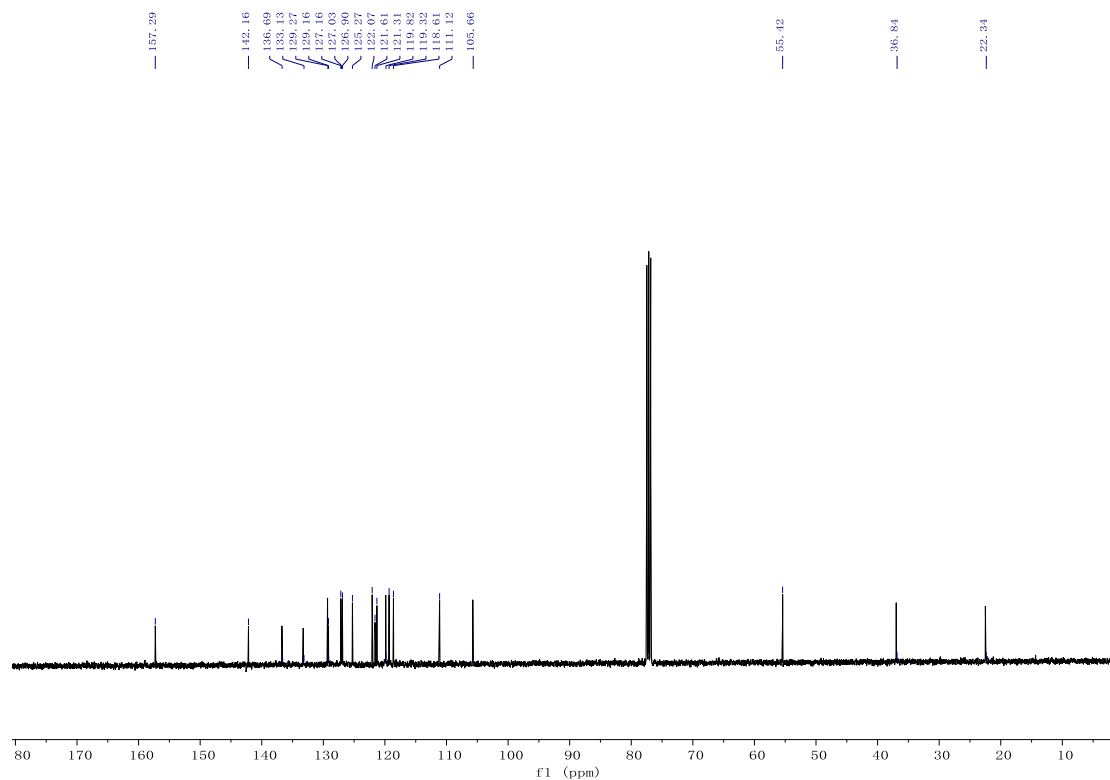
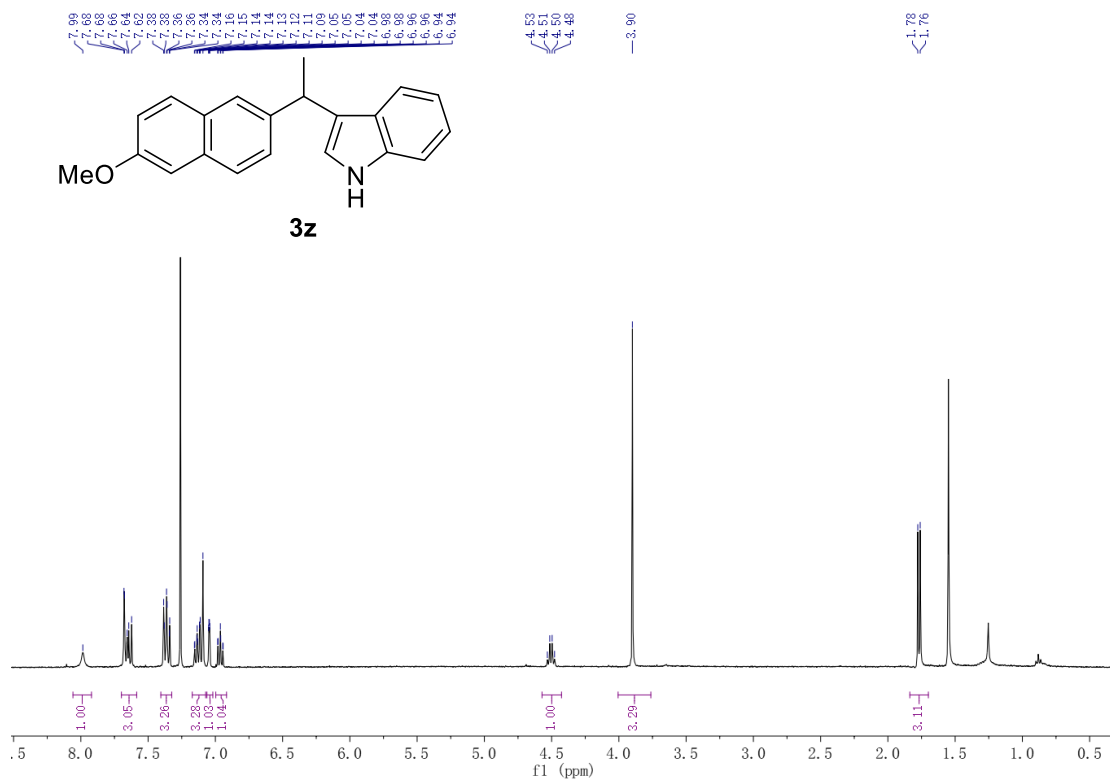
^{13}C NMR, CDCl_3 , 101 MHz

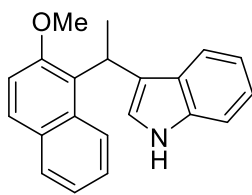


¹H NMR, CDCl₃, 400 MHz(* grease)

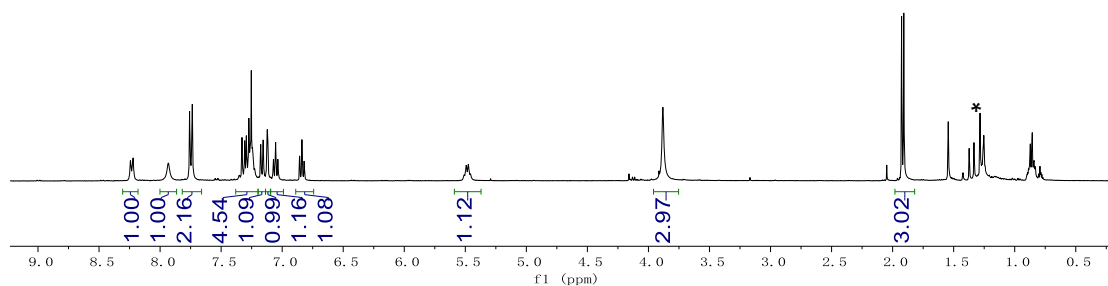


¹³C NMR, CDCl₃, 101 MHz





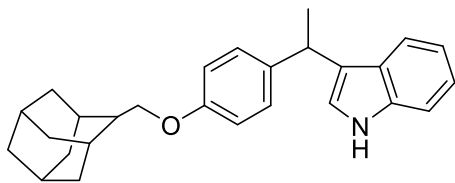
3aa



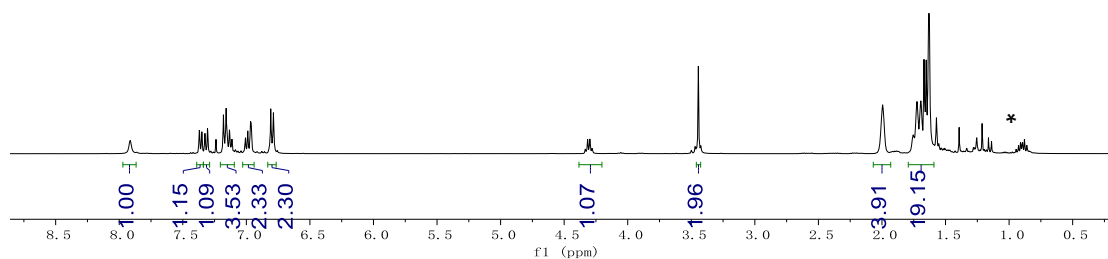
¹H NMR, CDCl₃, 400 MHz (*grease)



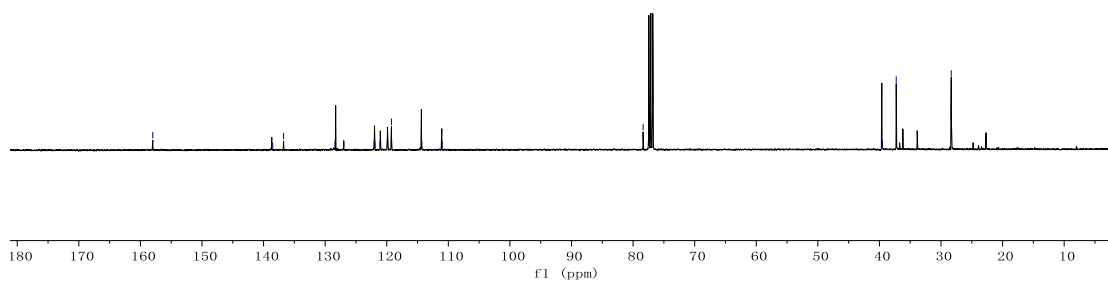
¹³C NMR, CDCl₃, 101 MHz



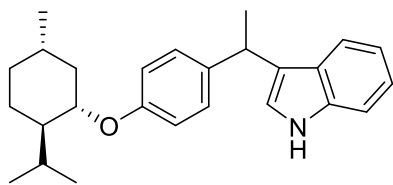
3ab



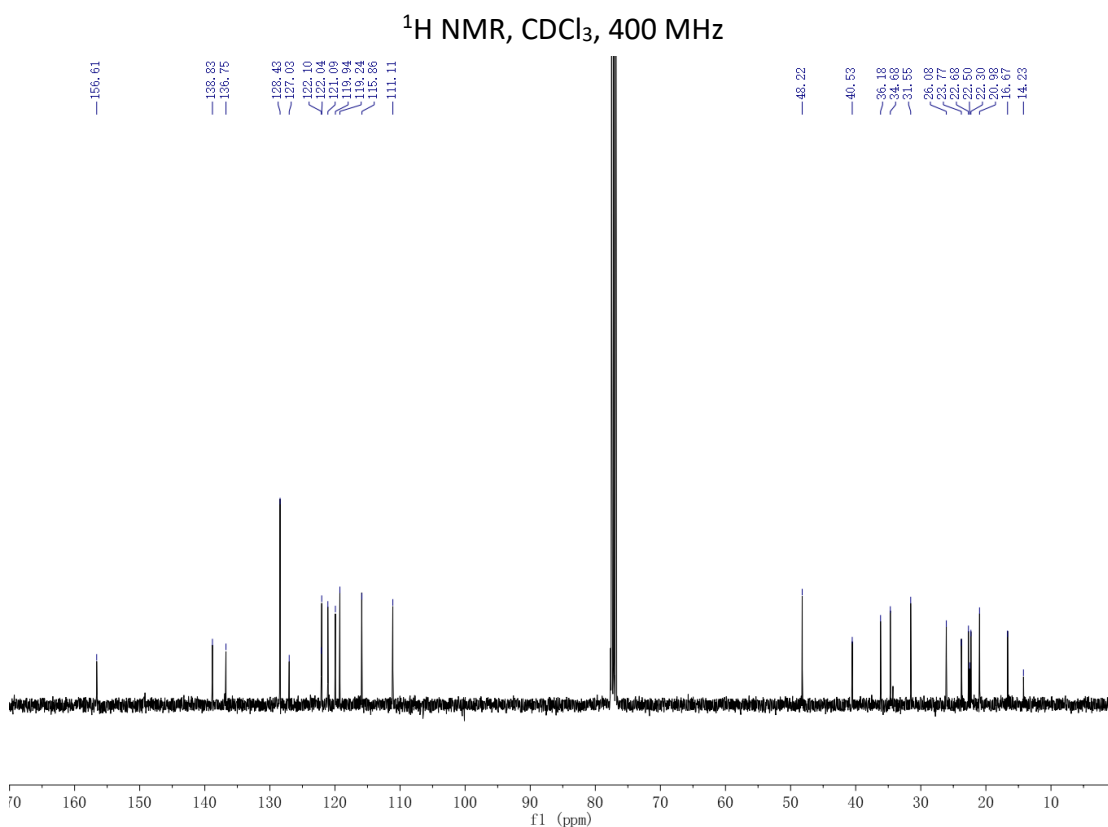
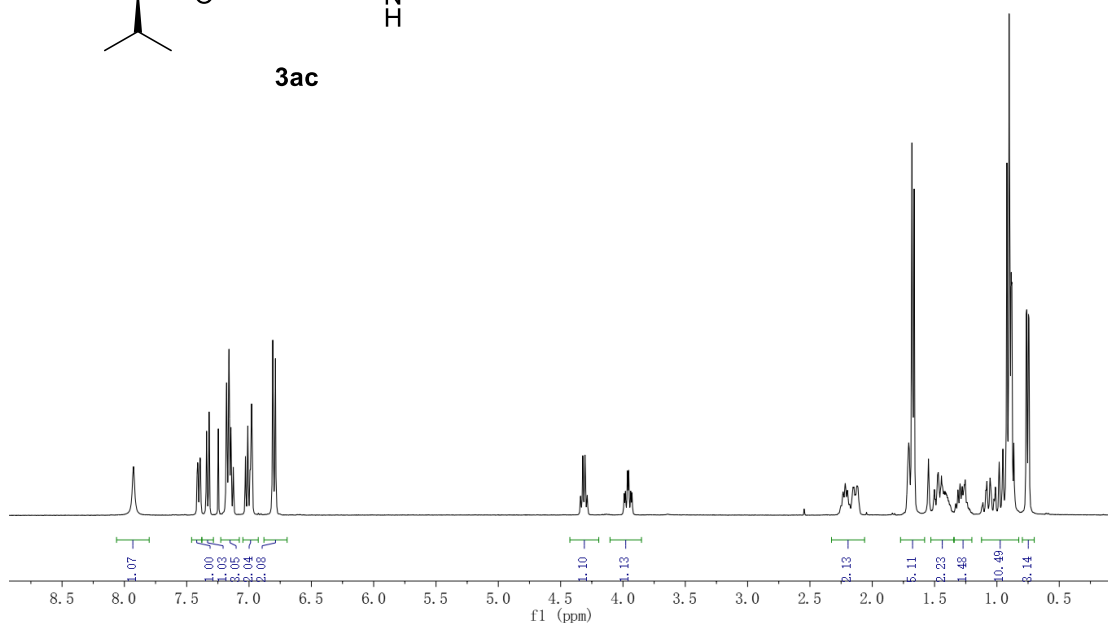
¹H NMR, CDCl₃, 400 MHz (*grease)

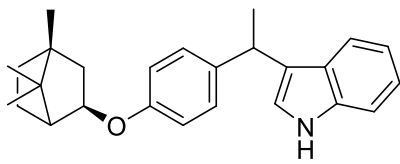


¹³C NMR, CDCl₃, 101 MHz

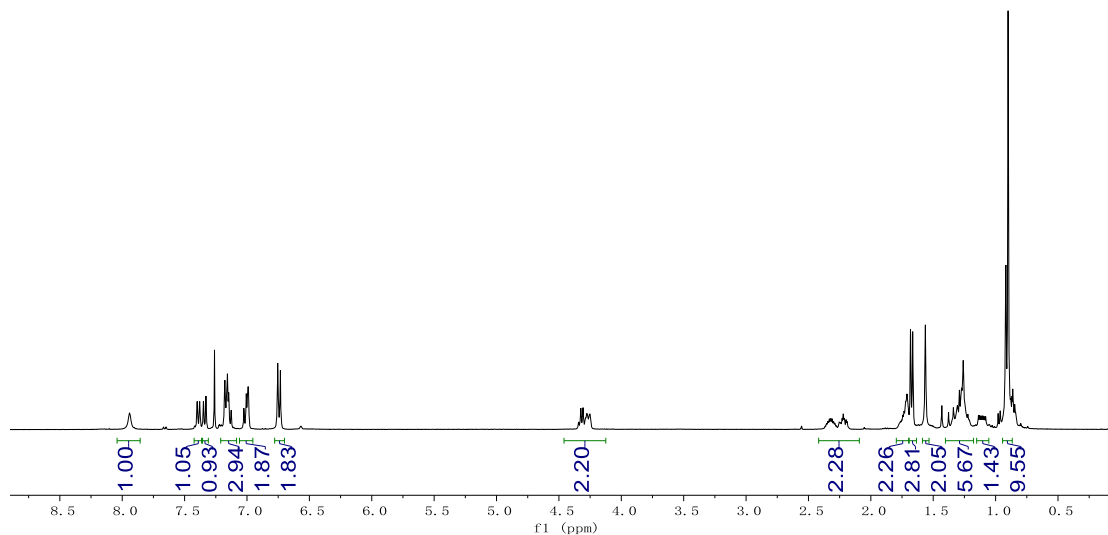


3ac

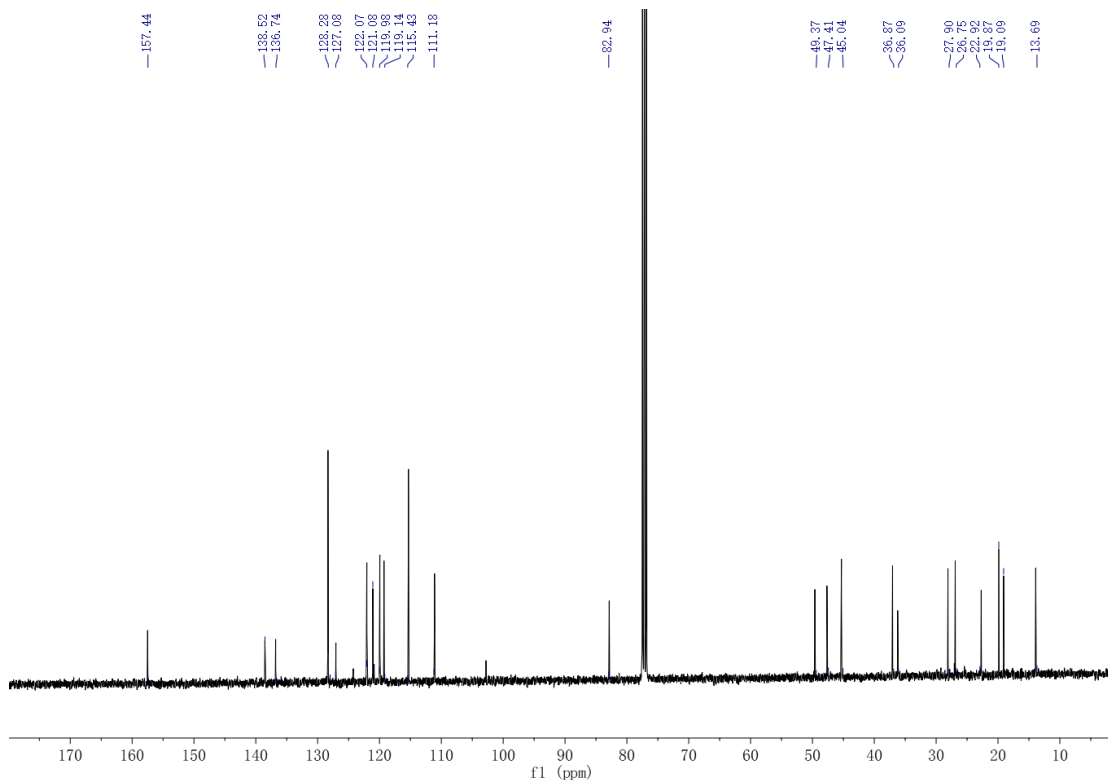




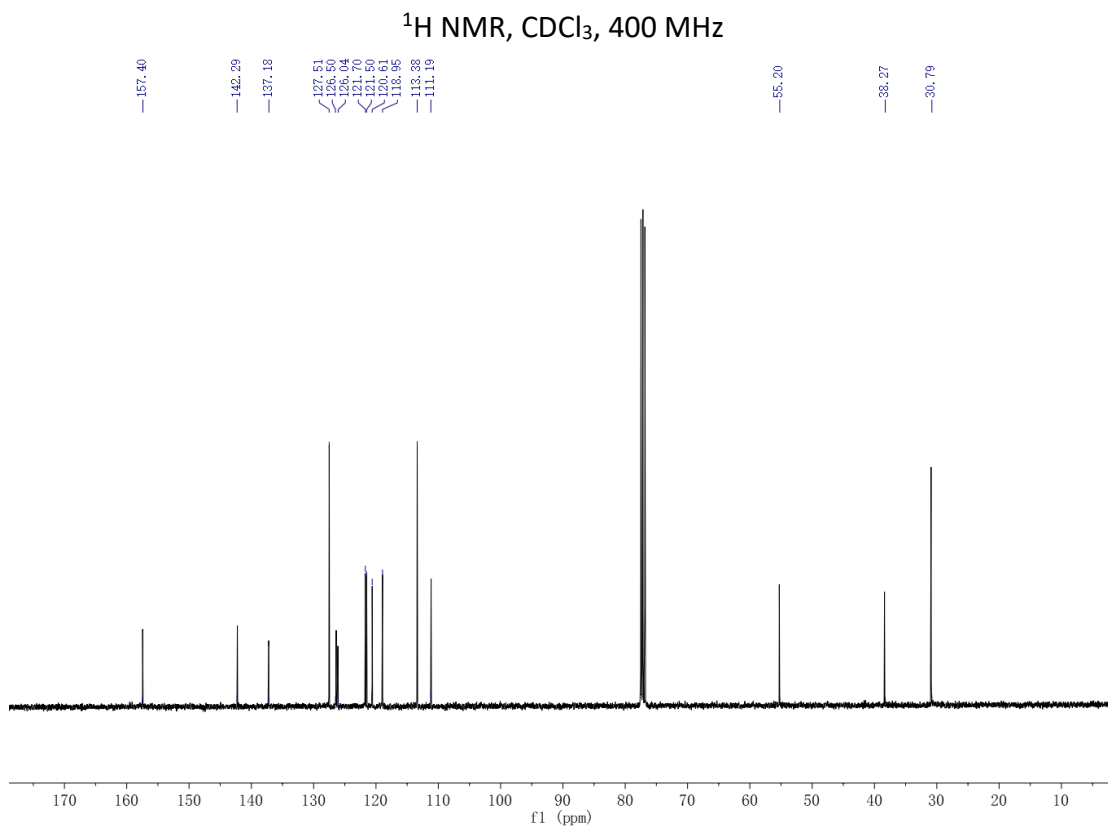
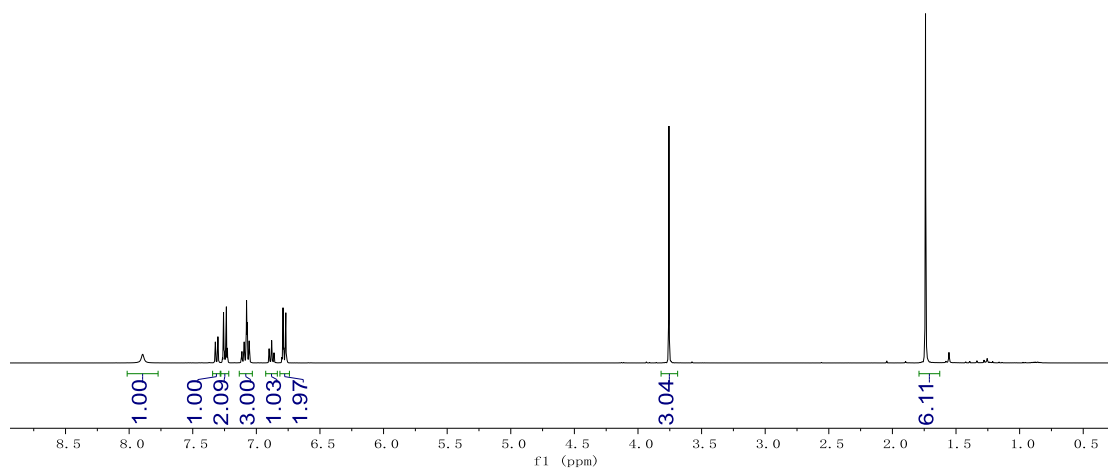
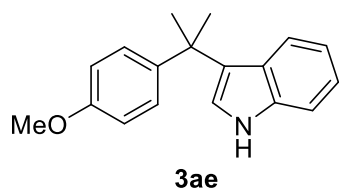
3ad

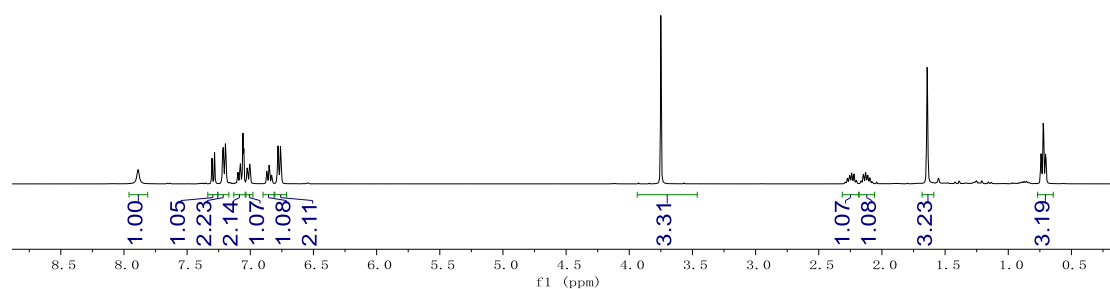
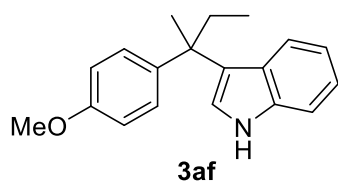


¹H NMR, CDCl₃, 400 MHz

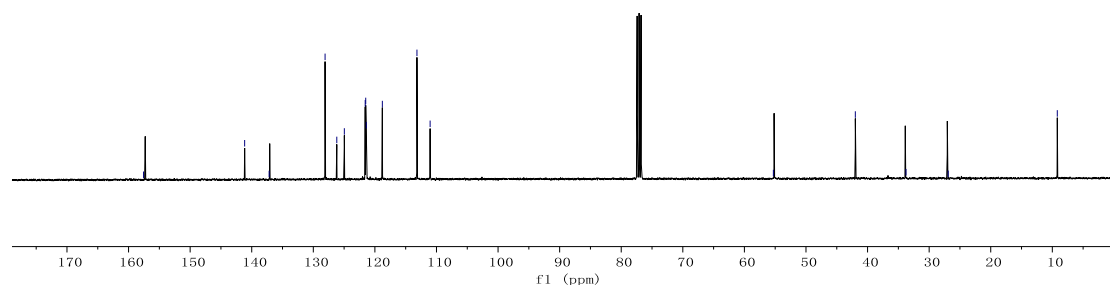


¹³C NMR, CDCl₃, 101 MHz

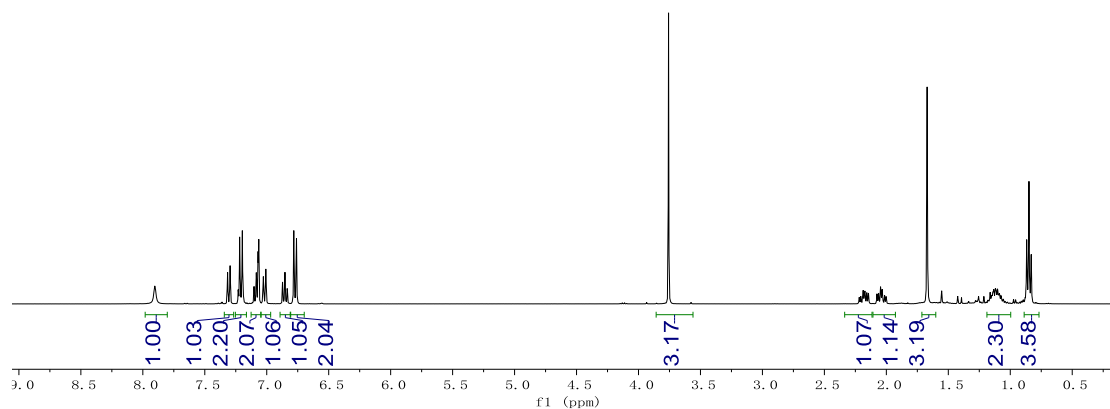
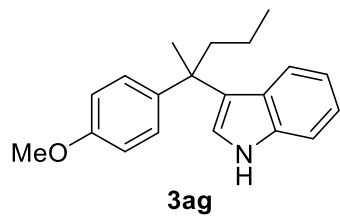




¹H NMR, CDCl₃, 400 MHz



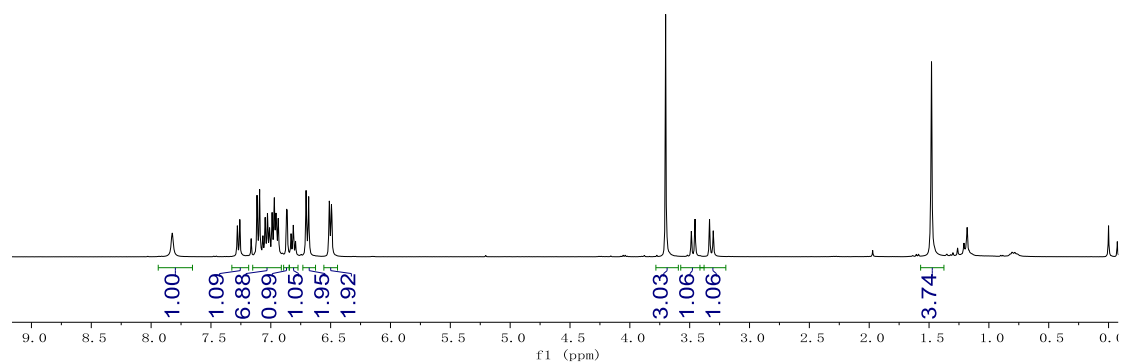
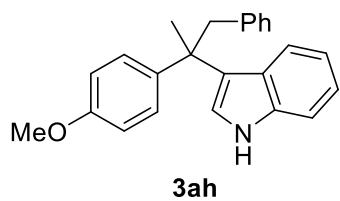
¹³C NMR, CDCl₃, 101 MHz



¹H NMR, CDCl₃, 400 MHz



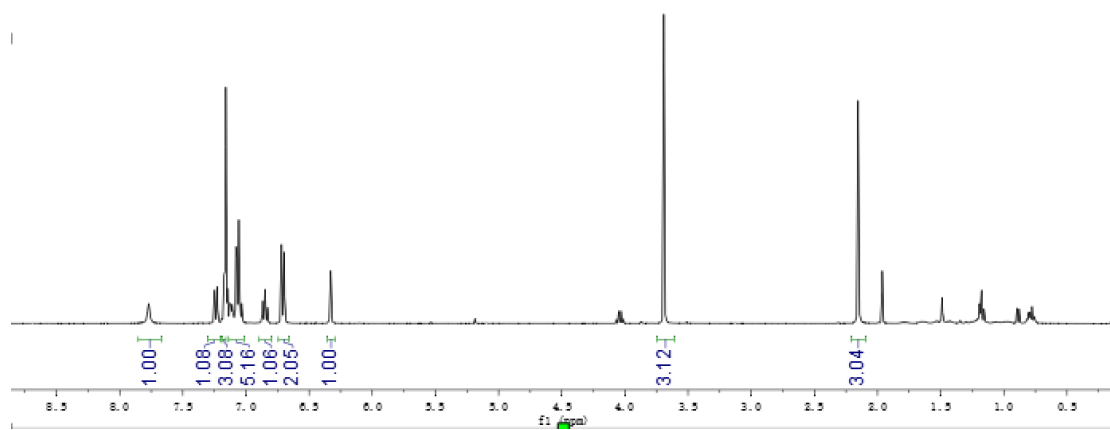
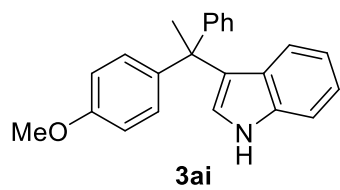
¹³C NMR, CDCl₃, 101 MHz



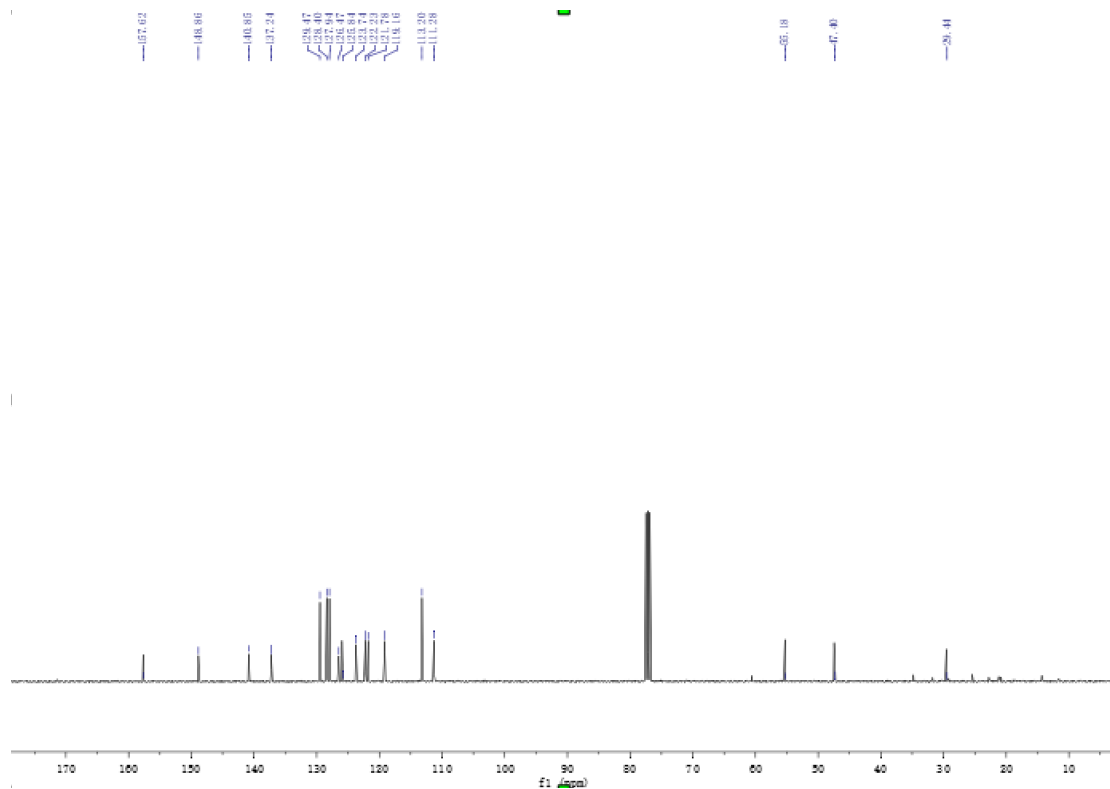
¹H NMR, CDCl₃, 400 MHz



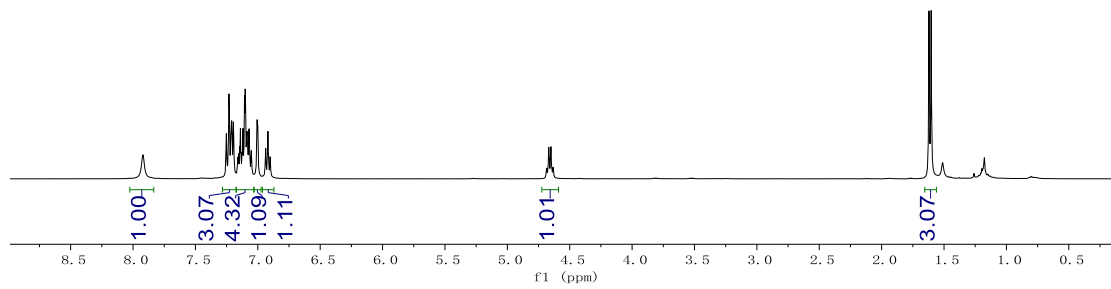
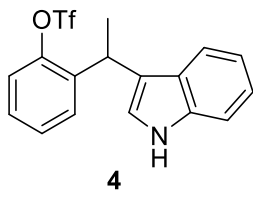
¹³C NMR, CDCl₃, 101 MHz



$^1\text{H NMR}$, CDCl_3 , 400 MHz



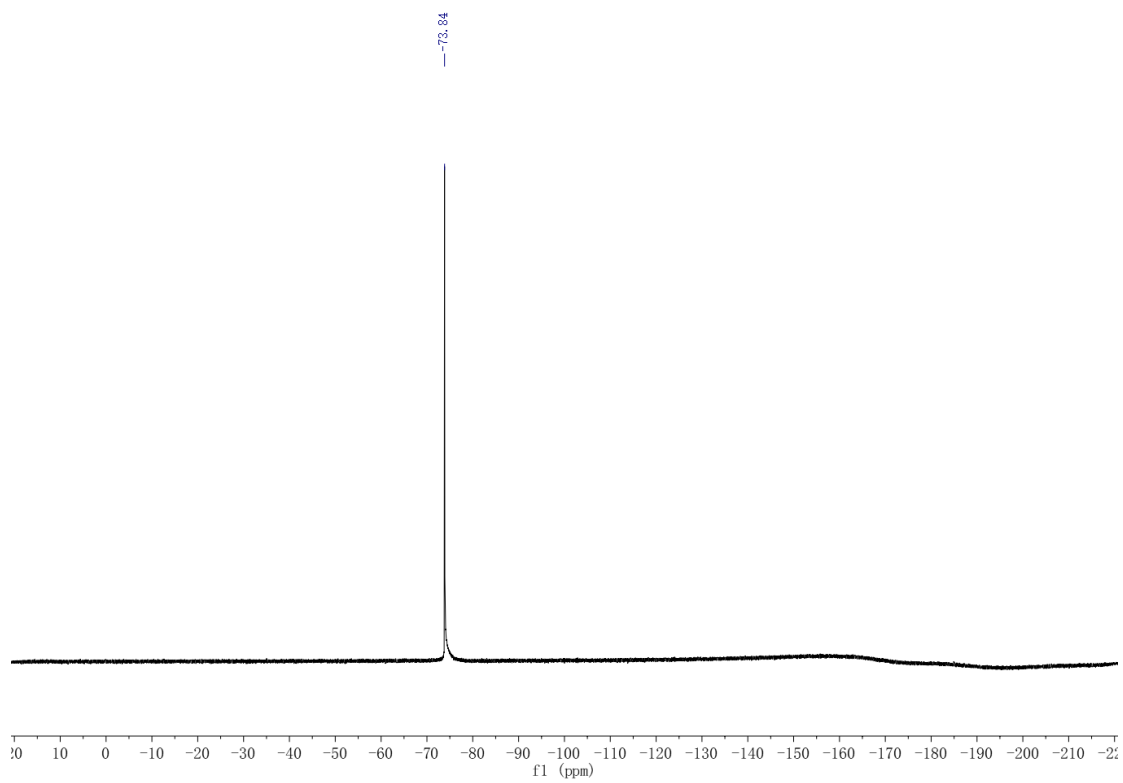
$^{13}\text{C NMR}$, CDCl_3 , 101 MHz



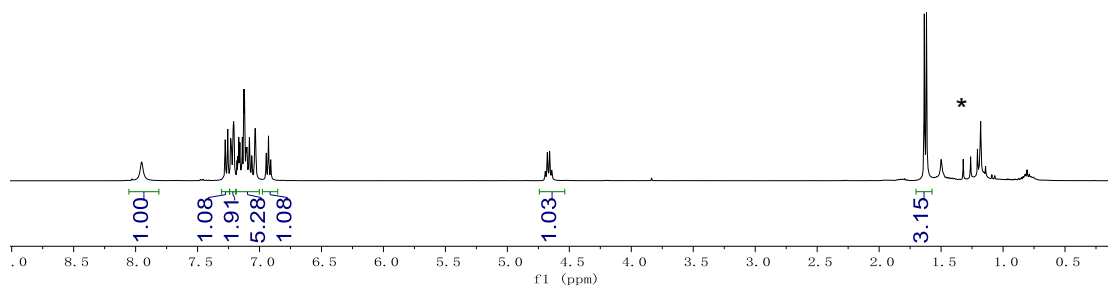
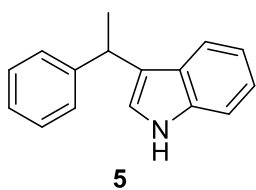
¹H NMR, CDCl₃, 400 MHz



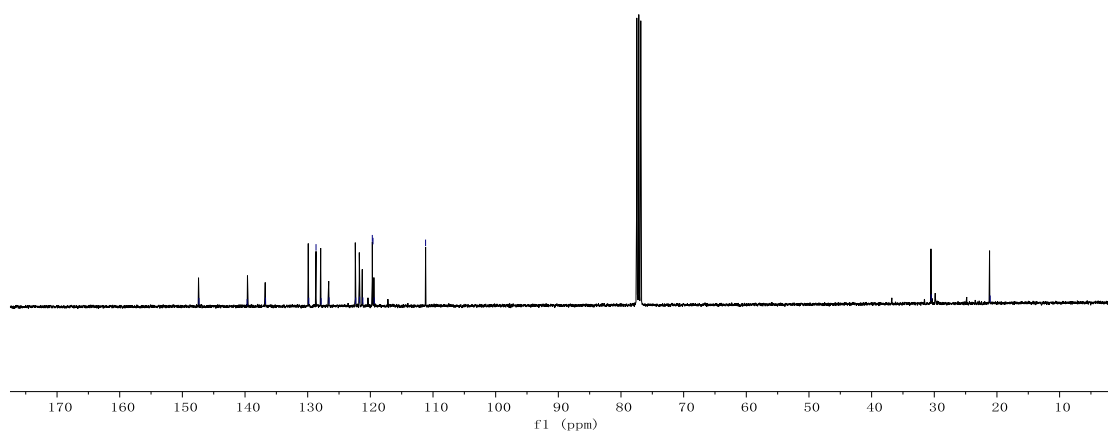
¹³C NMR, CDCl₃, 101 MHz



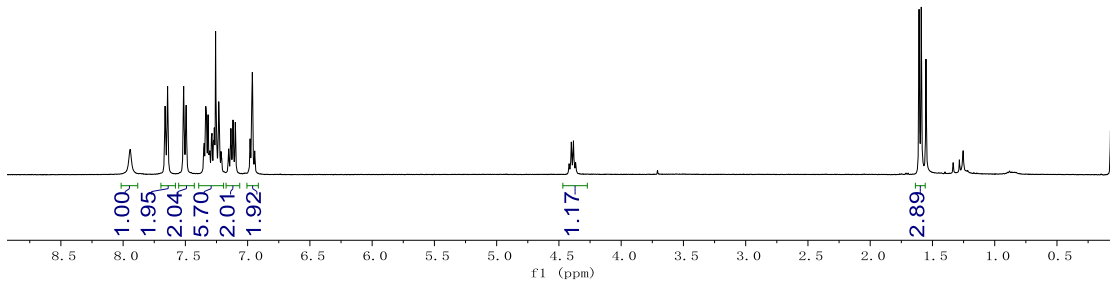
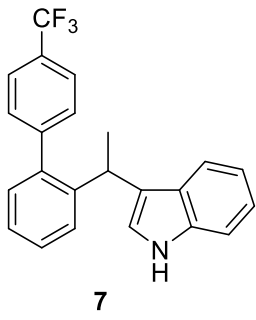
^{19}F NMR, CDCl_3 , 471 MHz



¹H NMR, CDCl₃, 400 MHz (* grease)

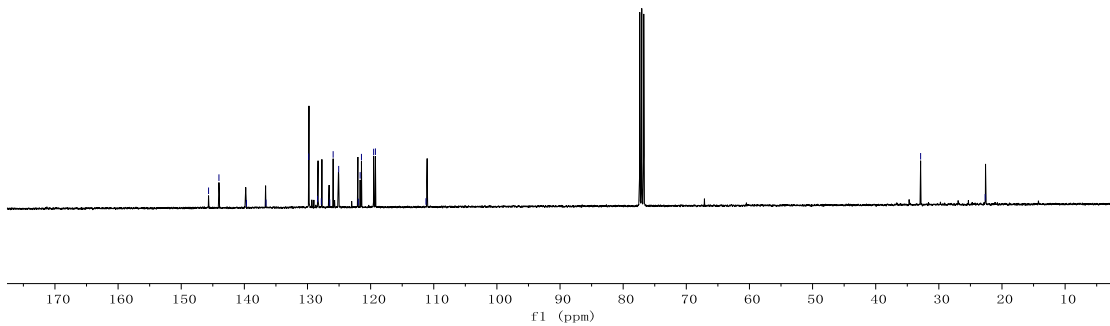


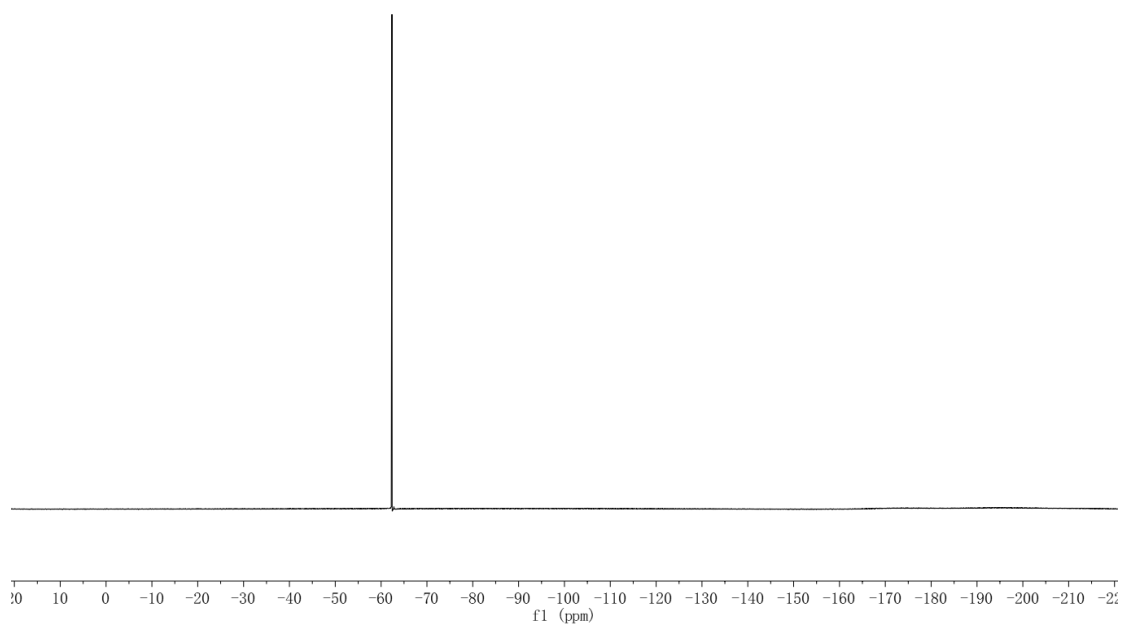
¹³C NMR, CDCl₃, 101 MHz



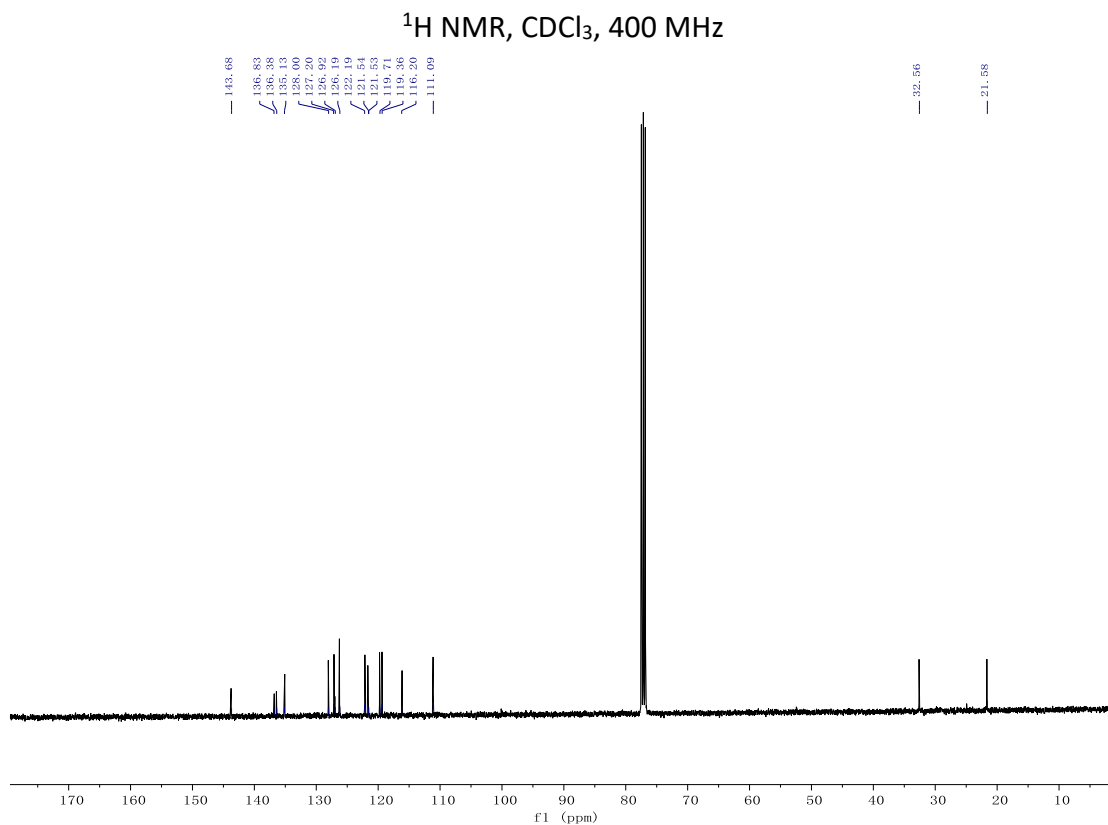
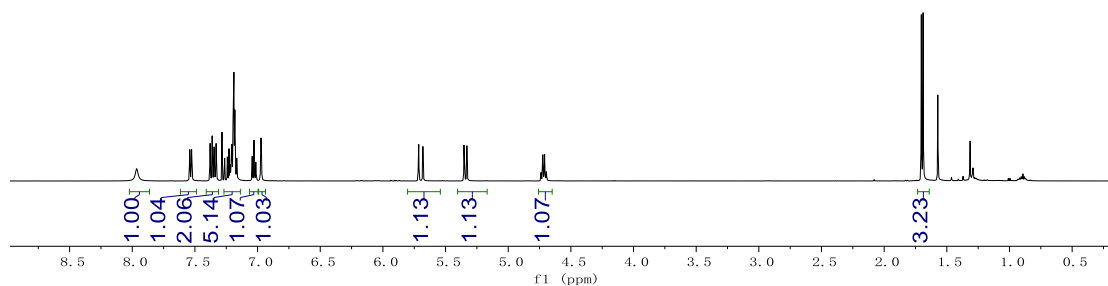
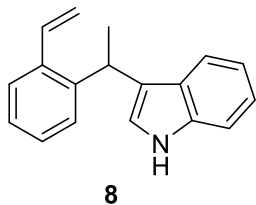
145.67
144.03
136.57
128.81
127.81
126.47
125.94
125.06
121.66
121.45
119.52
119.24
111.24

32.89
22.66





^{19}F NMR, CDCl_3 , 471 MHz



10. Reference and notes

(1) Esezobor, O. Z.; Zeng, W.; Niederegger, L.; Grübel, M.; Hess, C. R. *J. Am. Chem. Soc.* **2022**, *144*, 2994–3004.

(2) Jiang, F.; Zhang, Y-C.; Sun, S-B.; Zhou, L-J.; Shi, F. *Adv. Synth. Catal.* **2015**, *357*, 1283–1292.

(3) Pathak, T. P.; Sigman, M. S. *Org Lett.* **2011**, *13*, 2774-2777.