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Supporting Information

То

Conversion of Amino-Terephthalonitriles to Multi-Substituted Single Benzene Fluorophores with Utility in Bioimaging

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Materials and methods

Solvents, reagents, and other materials were purchased from commercial suppliers and used without further purification. TLC: pre-coated silica gel thin layer sheets ALUGRAM Xtra SIL G/UV₂₅₄ (Macherey-Nagel) and detection by UV-lamp. Flash chromatography: silica gel 60 Å, 60-120 mesh from Loba Chemie Pvt. Ltd. NMR spectra were recorded on Bruker Avance Neo 500 MHz NMR spectrometer at Central Sophisticated Instrumentation Facility, Birla Institute of Technology and Science, Pilani, K.K. Birla, Goa Campus, India and Bruker 400 MHz NMR spectrometer at Birla Institute of Technology and Science, Pilani, Hyderabad Campus, India with DMSO-d₆ and CDCl₃ as internal standards. Chemicals shifts (δ) for d₆-DMSO are expressed in ppm relative to the residual DMSO signal at 2.50 ppm for ¹H and at 39.65 ppm for ¹³C nuclei. Chemicals shifts (δ) for CDCl₃ are expressed in ppm relative to the residual CHCl₃ signal at 7.25 ppm for ¹H and at 77 ppm for ¹³C nuclei. UV-visible spectra were recorded on a Jasco V-570 spectrophotometer; fluorescence spectra were recorded on a Jasco FP-8500 spectrofluorimeter in the Department of Chemistry, Birla Institute of Technology and Science, K.K. Birla, Goa campus, India. Liquid Chromatography-Mass spectrometry (LC-MS) data were obtained from Agilent triplequad 6460 instrument to determine the molecular mass of compounds in the electrospray ionization (ESI+) and (ESI-) modes at Central Sophisticated Instrumentation Facility, BITS Pilani, K.K. Birla, Goa, India. High-Resolution Mass Spectrometry (HRMS) data were obtained using Agilent Technologies 6545 Q-TOF LC/MS instrument at HRMS Facility at BITS Pilani, Pilani Campus.

Experimental Procedures

8-cyano-6,7-difluoro-1,2,3,4-tetrahydroquinoxaline-5-carboxamide, **9**: Potassium hydroxide (89.26 mg, 1.59 mmol) was taken in a 100 mL round bottom flask and dissolved in 1 mL distilled water. To this ethanol (10 mL) was added followed by addition of 6,7-difluoro-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile **6** (70 mg, 0.318 mmol) to the above reaction mixture, which was kept at reflux for 5 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (40 mL) and washed with brine (5 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated *in vacuo*. The crude compound was purified by column chromatography using ethyl acetate as eluent. The yield of **9** was 20% (weight: 15 mg). ¹H NMR (500 MHz, *d*₆-DMSO): δ 7.85-7.84 (m, 2H), 6.68 (s, 1H), 6.47 (s, 1H), 3.28 (m, 2H), 3.24 (m, 2H); ¹³C NMR (125 MHz, *d*₆-DMSO): δ 165.08, 136.07, 130.88, 113.27, 110.63, 82.95, 38.88; ¹⁹F NMR (470 MHz, *d*₆-DMSO): δ -153.31 (d, ³*J* = 24.44 Hz, 1F), -154.88 (d, ³*J* = 25.38 Hz, 1F). LCMS-ESI (+) (M+1): 238.90 and LCMS-ESI (-) (M-1): 236.80.

6,7-difluoro-1,2,3,4-tetrahydroquinoxaline-5,8-dicarboxamide, **10**: Potassium hydroxide (89.26 mg, 1.59 mmol) was taken in a 100 mL round bottom flask and dissolved in 1 mL distilled water. To this ethanol (10 mL) was added followed by addition of 6,7-difluoro-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile **6** (70 mg, 0.318 mmol) to the above reaction mixture, which was kept at reflux for 5 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (40 mL) and washed with brine (5 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated *in vacuo*. The crude compound was purified by column chromatography using ethyl acetate as eluent. The yield of **10** was 6 % (weight: 5 mg). ¹H NMR (500 MHz, *d*₆-DMSO): δ 7.70 (m, 2H), 6.49 (s, 1H), 3.25 (s, 2H); ¹³C NMR (125 MHz, *d*₆-DMSO): δ 166.02, 137.97, 131.21, 107.65; ¹⁹F NMR (470 MHz, *d*₆-DMSO): δ -155.63 (s, 1F). LCMS-ESI (+) (M+1): 257.00.

6,9-dimethyl-2,3,6,7,8,9-hexahydro-[1,4]dioxino[2,3-g]quinoxaline-5,10-dicarbonitrile,

11: Potassium hydroxide (226 mg, 4.03 mmol) was taken in a 100 mL round bottom flask and dissolved in 0.5 mL distilled water. To this ethylene glycol (8 mL) was added followed by addition of 6,7-difluoro-1,4-dimethyl-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile 5 (100 mg, 0.40 mmol) to the above reaction mixture, which was kept at reflux for 1 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCI was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (50 mL) and washed with brine (6 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated in vacuo. The crude compound was purified by column chromatography using chloroform and petroleum ether (1:1) as eluent. The yield of **11** was 39% (weight: 42 mg). ¹H NMR (400 MHz, CDCl₃): δ 4.28 (s, 4H), 3.04 (s, 4H), 3.03 (s, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 138.83, 137.81, 114.69, 96.32, 64.86, 46.78, 44.30. LCMS-ESI (+) (M+1): 271.1000. ESI-HRMS (+) (M+1) for C₁₄H₁₅N₄O₂ calculated: 271.1190, found: 271.1192.

6,7-diethoxy-1,4-dimethyl-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile, 12:

Potassium hydroxide (452.08 mg, 8.05 mmol) was taken in a 100 mL round bottom flask and dissolved in 1 mL distilled water. To this ethanol (20 mL) was added followed by addition of 6,7-difluoro-1,4-dimethyl-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile **5** (200 mg,

0.805 mmol) to the above reaction mixture, which was kept at reflux for 6 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (70 mL) and washed with brine (8 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated *in vacuo*. The crude compound was purified by column chromatography using chloroform and petroleum ether (1:1) as eluent. The yield of **12** was 25 % (weight: 60 mg). ¹H NMR (400 MHz, d_6 -DMSO): δ 4.09 (q, ³*J* = 8.00 Hz, 4H), 3.12 (s, 4H), 3.05 (s, 6H), 1.32 (t, ³*J* = 8.00 Hz, 6H); ¹³C NMR (100 MHz, d_6 -DMSO): δ 145.70, 140.75, 115.76, 101.57, 70.63, 46.60, 44.28, 15.85. LCMS-ESI (+) (M+1): 301.1995. ESI-HRMS (+) (M+1) for C₁₆H₂₁N₄O₂ calculated: 301.1660, found: 301.1661.

8-cyano-6,7-diethoxy-1,4-dimethyl-1,2,3,4-tetrahydroquinoxaline-5-carboxamide, 13:

Potassium hydroxide (339.06 mg, 6.04 mmol) was taken in a 100 mL round bottom flask and dissolved in 1 mL distilled water. To this ethanol (20 mL) was added followed by addition of 6,7-difluoro-1,4-dimethyl-1,2,3,4-tetrahydroguinoxaline-5,8-dicarbonitrile **5** (150 mg,

0.60 mmol) to the above reaction mixture, which was kept at reflux for 24 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (60 mL) and washed with brine (7 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated *in vacuo*. The crude compound was purified by column chromatography using ethyl acetate as eluent. The yield of **13** was 27 % (weight: 52 mg). ¹H NMR (500 MHz, *d*₆-DMSO): δ 7.66 (s, 1H), 7.50 (s, 1H), 4.09 (q, ³*J* = 7.00 Hz, 2H), 3.93 (q, ³*J* = 7.00 Hz, 2H), 3.12 (m, 2H), 3.06 (s, 3H), 2.94 (m, 2H), 2.75 (s, 3H), 1.32 (t, ³*J* = 5.00 Hz, 3H), 1.24 (t, ³*J* = 5.00 Hz, 3H); ¹³C NMR (125 MHz, *d*₆-DMSO): δ 167.85, 149.15, 141.55, 140.83, 133.55, 116.77, 96.48, 70.09, 47.05, 45.78, 44.36, 43.88, 15.99. LCMS-ESI (+) (M+1): 319.1800. ESI-HRMS (+) (M+1) for C₁₆H₂₃N₄O₃ calculated: 319.1770, found: 319.1766.

6-fluoro-7-isopropoxy-1,4-dimethyl-1,2,3,4-tetrahydroquinoxaline-

5,8-dicarbonitrile, **14**: Potassium hydroxide (113.02 mg, 2.01 mmol) was taken in a 100 mL round bottom flask and dissolved in 0.5 mL distilled water. To this isopropanol (10 mL) was added followed by addition of 6,7-difluoro-1,4-dimethyl-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile **5** (50 mg, 0.201 mmol) to the above reaction, which was kept at reflux for 6 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (30 mL) and washed with brine (5 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated *in vacuo*. The crude compound was purified by column chromatography using chloroform and petroleum ether (1:1) as eluent. The yield of **14** was 36 % (weight: 21 mg). ¹H NMR (400 MHz, *d*₆-DMSO): δ 4.39 (m, 1H), 3.23 (m, 2H), 3.17 (s, 3H), 3.15 (m, 2H), 3.06

(s, 3H), 1.29 (d, ³*J* = 8.00 Hz, 6H); ¹³C NMR (100 MHz, *d*₆-DMSO): δ 141.04, 140.73, 115.54, 114.12, 101.29, 78.80, 47.17, 46.94, 44.35, 43.73, 22.61. LCMS-ESI (+) (M+Acetonitrile): 329.20.

6-fluoro-1,4-dimethyl-7-(octyloxy)-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile, **17**: Potassium hydroxide (226.04 mg, 4.03 mmol) was taken in a 100 mL round bottom flask and dissolved in 1 mL distilled water. To this octanol (637 µL, 4.03 mmol) was added followed by addition of 6,7-difluoro-1,4-dimethyl-1,2,3,4-tetrahydroquinoxaline-5,8-dicarbonitrile **5** (100 mg, 0.40 mmol) to the above reaction mixture in 7 mL DMF, which was kept at reflux for 12 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (60 mL) and washed with brine (7 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated *in vacuo*. The crude compound was purified by column chromatography using ethyl acetate as eluent. The yield of **17** was 45 % (weight: 65 mg). ¹H NMR (500 MHz, *d*₆-DMSO): δ 4.06 (t, ³*J* = 7.50 Hz, 2H), 3.21 (m, 2H), 3.15 (s, 5H), 3.06 (s, 3H), 1.67 (m, 2H), 1.42 (m, 2H), 1.25 (m, 8H), 0.86 (t, ³*J* = 7.50 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 147.05, 140.03, 139.41, 114.53, 113.26, 100.81, 75.44, 47.15, 46.95, 44.02, 43.56, 31.55, 29.68, 29.00, 25.42, 22.41, 13.95. LCMS-ESI (+) (M+1): 359.2004.

10-cyano-6,9-dimethyl-2,3,6,7,8,9-hexahydro-[1,4]dioxino[2,3-g]quinoxaline-5-

carboxamide, **18**: Potassium hydroxide (62.34 mg, 1.11 mmol) was taken in a 100 mL round bottom flask and dissolved in 0.5 mL distilled water. To this isopropanol (10 mL) was added followed by addition of 6,9-dimethyl-2,3,6,7,8,9-hexahydro-[1,4]dioxino[2,3-g]quinoxaline-5,10-dicarbonitrile **11** (30 mg, 0.11 mmol) to the above reaction mixture and continued refluxing for 24 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (30 mL) and washed with brine (4 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated *in vacuo*. The crude compound was purified by column chromatography using ethyl acetate as eluent. The yield of **18** was 27 % (weight: 8.5 mg). ¹H NMR (400 MHz, *d*₆-DMSO): δ 7.65 (s, 1H), 7.46 (s, 1H), 4.30 (m, 2H), 4.18 (m, 2H), 3.09 (m, 2H), 3.01 (s, 3H), 2.90 (m, 2H), 2.70 (s, 3H); ¹³C NMR (100 MHz, *d*₆-DMSO): δ 167.06, 141.58, 139.57, 133.17, 130.88, 127.39, 124.90, 116.28, 90.25, 65.38, 64.24, 47.10, 45.63, 44.49. LCMS-ESI (+) (M+1): 289.100. ESI-HRMS (+) (M+1) for C₁₄H₁₇N₄O₃ calculated: 289.1296, found: 289.1298.

12-cyano-1,4-dimethyl-1,2,3,4-tetrahydrobenzo[5,6][1,4]dioxino[2,3-g]quinoxaline-5-

carboxamide, 20: Potassium hydroxide (176.45 mg, 3.14 mmol) was taken in a 100 mL round bottom flask and dissolved in 0.5 mL distilled water. To this ethanol (10 mL) was added 1,4-dimethyl-1,2,3,4-tetrahydrobenzo[5,6][1,4]dioxino[2,3followed by addition of g]quinoxaline-5,12-dicarbonitrile **19** (100 mg, 0.314 mmol) to the above reaction mixture and continued refluxing for 12 h monitored by TLC. After complete consumption of starting material, the reaction mixture was cooled to room temperature and then 6M HCl was added to reaction mixture till the pH became approximately 2. The reaction mixture was dissolved in ethyl acetate (30 mL) and washed with brine (7 × 25 mL). The combined organic extracts were dried over sodium sulphate and concentrated in vacuo. The crude compound was purified by column chromatography using ethyl acetate as eluent. The yield of 20 was 31 % (weight: 32 mg). ¹H NMR (500 MHz, *d*₆-DMSO): δ 7.90 (s, 1H), 7.67 (s, 1H), 7.01 (m, 4H), 3.14 (m, 2H), 3.08 (s, 3H), 2.95 (m, 2H), 2.76 (s, 3H); ¹³C NMR (125 MHz, d_θ-DMSO): δ 165.90, 141.40, 140.95, 138.43, 132.81, 130.48, 125.61, 125.00, 116.94, 115.17, 89.15, 47.09, 45.97, 44.17, 43.72. LCMS-ESI (+) (M+1) for C₁₈H₁₇N₄O₃ calculated: 337.1295, found:337.50.

Cell culture and confocal imaging protocol for compound 17

HeLa cells were procured from National Centre for Cell Sciences (Pune, India) and maintained in Dulbecco's Modified Eagle Medium (HiMedia), supplemented with 10% (v/v) Fetal Bovine Serum (HiMedia), antibiotics (Penicillin10000U/mL+ Streptomycin 10000 μ g/mL) (GIBCO) and maintained at 37°C incubator with humidified atmosphere of CO₂ (5%) and air (95%). Cells were cultured in vented T-25 cell culture flasks.

For confocal imaging, HeLa cells ($8x10^3$) were seeded on a clean cover slip and grown overnight. Next day, cells were fixed with ice-cold methanol for 10 minutes and then washed with cold 1x PBS buffer (pH 7.4). The cells were then stained with 100 μ M of compound **17** for 30 minutes. Then the cells were washed thrice with cold 1x PBS buffer (pH 7.4). To stain the nuclei, DAPI was added to the cells in fresh 1x PBS buffer for 10 minutes. The cover slip was mounted on clean glass slide with glycerol and viewed under confocal microscope at 80X magnification and 1% laser power. The excitation laser user for **17** was 488 nm with an emission window between 500-550 nm.



Figure S1: ¹H NMR spectrum (500 MHz) of compound 9 in DMSO-d₆.



Figure S2: ¹³C NMR spectrum (125 MHz) of compound 9 in DMSO-d₆



Figure S3: ¹⁹F NMR spectrum (470 MHz) of compound 9 in DMSO-d₆.



Figure S4: (A) LCMS-ESI (-) (M-1) compound 9 shows peak at 236.80. (B) LCMS-ESI (+) (M+1) of compound 9 shows peak at 238.90.



Figure S5: ¹H NMR spectrum (500 MHz) of compound **10** in DMSO-d₆.



Figure S6: ¹³C NMR spectrum (125 MHz) of compound **10** in DMSO-d₆.



Figure S7: ¹⁹F NMR spectrum (470 MHz) of compound **10** in DMSO-d₆.



Figure S8: LCMS-ESI (+) of compound 10 shows peak at 257.00.



Figure S9: ¹H NMR spectrum (400 MHz) of compound 11 in CDCI₃.



Figure S10: ¹³C NMR spectrum (100 MHz) of compound 11 in CDCl₃.



Figure S11: HRMS-ESI (+) of compound 11.



Figure S12: LCMS-ESI (+) (M+1) of compound 11 shows peak at 271.10.



Figure S13: ¹H NMR spectrum (400 MHz) of compound 12 in DMSO-d₆.



Figure S14: ¹³C NMR spectrum (100 MHz) of compound 12 in DMSO-d₆.



Figure S15: HRMS-ESI (+) of compound 12.



Figure S16: LCMS-ESI (+) (M+1) of compound 12 shows peak at 301.1995.



Figure S17: ¹H NMR spectrum (500 MHz) of compound 13 in DMSO-d₆.



Figure S18: ¹³C NMR spectrum (125 MHz) of compound 13 in DMSO-d₆.



Figure S19: HRMS-ESI (+) of compound 13.



Figure S20: LCMS-ESI (+) (M+1) of compound 13 shows peak at 319.18.



Figure S21: ¹H NMR spectrum (400 MHz) of compound 14 in DMSO-d₆.



Figure S22: ¹³C NMR spectrum (100 MHz) of compound **14** in DMSO-d₆.



m/z	Z	Abund
230.1006	50 S 20 Z	23693.28
244.1195	100	33606.69
245.1	1.0	30215.23
286.1402		11159.64
287.1899	1	27920.58
288.1902	1	5665.32
328.1704		44659.61
329.2	1	583718.81
330.1999	1	121733
331.2091	1	13598.47

Figure S23: LCMS-ESI (+) of compound 14 (M+Acetonitrile at m/z 329.20).



Figure S24: ¹H NMR spectrum (400 MHz) of mixture of compounds 15 and 16 in DMSO-d₆.



Figure S25: ¹³C NMR spectrum (100 MHz) of mixture of compounds 15 and 16 in DMSO-d₆.



Figure S26: LCMS-ESI (+) of compound mixture of compounds **15** and **16** shows peak at 307.20 and 347.20.



Figure S27: ¹H NMR spectrum (500 MHz) of compound 17 in DMSO-d₆.



Figure S28: ¹³C NMR spectrum (100 MHz) of compound **17** in CDCl₃.



Figure S29: LCMS-ESI (+) (M+1) of compound 17 shows peak at 359.2004.



Figure S30: ¹H NMR spectrum (400 MHz) of compound 18 in DMSO-d₆.



Figure S31: ¹³C NMR spectrum (100 MHz) of compound 18 in DMSO-d₆.



Figure S32: HRMS-ESI (+) of compound 18.



Figure S33: LCMS-ESI (+) (M+1) of compound 18 shows peak at 289.100.





Figure S35: ¹³C NMR spectrum (100 MHz) of compound 20 in CDCI₃.



Figure S36: LCMS-ESI (+) (M+1) of compound 20 shows peak at 337.50.



Figure S37: UV-Visible spectra (A) of compounds **5**, **11**, **12**, **14**, and **17** in DMSO and UV-Visible spectra (B) of compounds **5**, **13**, **18**, and **20** in DMSO.



Figure S38: Excited state geometries and frontier molecular orbitals of compound 5 and 6.



Figure S39: Ground state geometries of dimer of compound 9.



Figure S40: Ground state geometries of compounds 14 and 16.



Figure S41: Ground state (GS) and excited state (ES) geometries of compound 12.



15a, Energy = -1052.5676 Hartree



First one more stable by 5.4 kcal/mol

Figure S42. Ground state geometries of compounds 15a and 15b.



Figure S43: Solvatochromism plots for absorption spectra of compounds 9 (A), 10 (C), and fluorescence spectra of compounds 9 (B), and 10 (D).

Table S1 Absorption peak positions

Compounds	Predicted peak position	Experimental
		peak position
12	438 (0.29) H-L (0.70)	428
13	375(0.20) H-L (0.70)	374
14	431 (0.15), H-L(0.70)	427
9	424 (0.22), H-L (0.70)	406
10	429 (0.22), H-L (0.70)	392

Table S2 Fluorescence peak positions

.

Compounds	Predicted peak position	Experimental peak
		position
12	548	537
13	571	546
9	510	519
10	525	523

5		12		13			14				
MO	Energy (eV)										
NO.			NO.			NO.			NO.		
62	-7.954494912	HOMO-2	78	-0.27937	HOMO-2	83	-7.122364184	HOMO-2	74	-7.783061832	HOMO-2
63	-6.799634608	HOMO-1	79	-0.23797	HOMO-1	84	-6.304383488	HOMO-1	75	-6.67772664	HOMO-1
64	-5.772124592	НОМО	80	-0.19476	НОМО	85	-5.12802602	номо	76	-5.66817628	НОМО
65	-2.420199704	LUMO	81	-0.07228	LUMO	86	-1.74018182	LUMO	77	-2.291760952	LUMO
66	-0.530354084	LUMO+1	82	-0.01111	LUMO+1	87	-0.13197626	LUMO+1	78	-0.453345256	LUMO+1
67	-0.002993276	LUMO+2	83	0.01464	LUMO+2	88	0.50749634	LUMO+2	79	0.198100448	LUMO+2
6		9		10							
MO	Energy (eV)		MO	Energy (eV)		MO	Energy (eV)				
NO.			NO.			NO.					
54	-8.659275352	HOMO-2	83	-7.122364184	HOMO-2	64	-7.450263964	HOMO-2			
55	-6.957734004	HOMO-1	84	-6.304383488	HOMO-1	65	-6.562349456	HOMO-1			
56	-5.704367708	номо	85	-5.12802602	HOMO	66	-5.170204	НОМО			
57	-2.286046516	LUMO	86	-1.74018182	LUMO	67	-1.798142528	LUMO			
58	-0.266129448	LUMO+1	87	-0.13197626	LUMO+1	68	0.0748319	LUMO+1			
59	-0.045443372	LUMO+2	88	0.50749634	LUMO+2	69	0.394023968	LUMO+2			

 Table S3. Energies of Molecular Orbitals

Table S4. Absorption peak positions

	Theoretically predicted	Experimental		
Compounds	Predicted longest wave	Oscillator	Excitations	peak position
	length absorption peak	strengths of S ₀ -S ₁	dominating the	
	position	transitions	S_0 - S_1 transitions	
	(S_0-S_1) transition at		(contribution)	
	optimized S ₀)			
12	426	0.22	H→L (0.70)	428
13	375	0.20	H→L (0.70)	374
14	431	0.15	H→L (0.70)	427
9	411	0.16	H→L (0.70)	406
10	416	0.15	H→L (0.70)	392







Figure S44: Left graphs (A), (C), (E), (G), (I) shows the TDDFT calculated absorption spectra and the right graphs (B), (D), (F), (H), (J) shows the experimental absorption spectra of compounds 9, 10, 12,13, and 14 respectively.