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## Supplementary Information

## Synthesis of the full-length hepatitis B virus core protein and its capsid formation

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**Fig. S1** Screening concept for HBV capsid assembly modulators from a virtual mirror-image library of natural products. Screening of the natural product library using the mirror-image HBV core protein corresponds to the mirror-image library of natural products using the natural HBV core protein in a mirror. The HBV core protein structure was obtained from the Protein Data Bank (PDB ID: 6HTX).



Fig. S2 Previous synthetic studies on Cp149.<sup>S1</sup> Reagents and conditions: (a) MPAA, TCEP, 6 M guanidine HCl, and 200 mM phosphate buffer (pH 7.0); (b) NaNO<sub>2</sub>, 6 M guanidine HCl, 200 mM phosphate buffer, then MESNa and TCEP; (c) MPAA, TCEP, 6 M guanidine HCl, and 200 mM phosphate buffer (pH 7.0) containing 25% NMP; (d) 1,2,4-triazole, TCEP, 6 M guanidine HCl, and 100 mM phosphate buffer (pH 7.1); (e) Trt(OH)-K<sub>10</sub> and TFA; (f) methoxyamine, 6 M guanidine HCl, and 200 mM phosphate buffer (pH 4.0); (g) TFA/TIS (95:5). Abbreviations: Dbz, 3,4-diaminobenzoic *N*-acyl-*N*'-methylacylurea; acid; MeNbz, MES, 2-mercaptethanesulfonate; MPA. 3mercaptopropionic acid; MPAA, 4-mercaptophenylacetic TCEP. tris(2acid; carboxyethyl)phosphine; Thz, thiazolidine carboxylic acid.



**Fig. S3** CD analysis of the synthetic HBV core protein under various conditions. (A) CD spectra of synthetic Cp183(C183A) at various concentrations of guanidine. (B) Comparison of synthetic Cp183(C183A) CD spectra in a 1.5M guanidine or NaCl solution.

# **<u>References</u>**

S1 S. Tsuda, M. Mochizuki, H. Ishiba, K. Yoshizawa-Kumagaye, H. Nishio, S. Oishi and T. Yoshiya, *Angew. Chem. Int. Ed.*, 2018, **57**, 2105–2109.

#### Analytical HPLC Chromatograms and Mass Spectrometry Data of Synthetic Peptides



**HPLC conditions:** Cosmosil 5C18-AR300 column (Nacalai Tesque, 4.6 × 250 mm), linear gradient of 35–55% CH<sub>3</sub>CN containing 0.1% TFA at a flow rate of 1 mL/min over 20 min.

**MS analysis**: Expected mass based on the sequence: 5493.45; Major observed ions:  $[M+6H]^{6+} m/z = 916.85$ ,  $[M+5H]^{5+} m/z = 1099.99$ ,  $[M+4H]^{4+} m/z = 1374.55$ ,  $[M+3H]^{3+} m/z = 1831.68$ .

[Ala<sup>183</sup>]-Cp183<sup>154–183</sup> (4)



**HPLC conditions:** Cosmosil 5C18-AR300 column (Nacalai Tesque,  $4.6 \times 250$  mm), linear gradient of 0–20% CH<sub>3</sub>CN containing 0.05% TFA at a flow rate of 1 mL/min over 20 min.

**MS analysis**: Expected mass based on the sequence: 3732.18; Major observed ions:  $[M+6H]^{6+} m/z = 622.99$ ,  $[M+5H]^{5+} m/z = 747.13$ ,  $[M+4H]^{4+} m/z = 933.93$ ,  $[M+3H]^{3+} m/z = 1245.15$ ,  $[M+2H]^{2+} m/z = 1866.85$ .

[Ala<sup>183</sup>]-Cp183<sup>107-183</sup> (8)



**HPLC conditions:** Cosmosil 5C4-AR300 column (Nacalai Tesque,  $4.6 \times 250$  mm), linear gradient of 30–50% CH<sub>3</sub>CN containing 0.1% TFA at a flow rate of 1 mL/min over 20 min.

**MS analysis**: Expected mass based on the sequence: 9045.41; Major observed ions:  $[M+8H]^{8+} m/z = 1131.63$ ,  $[M+7H]^{7+} m/z = 1293.24$ ,  $[M+6H]^{6+} m/z = 1508.22$ ,  $[M+5H]^{5+} m/z = 1809.56$ .

[Ala<sup>183</sup>]-Cp183<sup>1-183</sup> (9)



**HPLC conditions:** Cosmosil 5C4-AR300 column (Nacalai Tesque,  $4.6 \times 250$  mm), linear gradient of 30–60% CH<sub>3</sub>CN containing 0.1% TFA at a flow rate of 1 mL/min over 30 min.

**MS analysis**: Expected mass based on the sequence: 21010.06; Major observed ions:  $[M+15H]^{15+}$ m/z = 1402.08,  $[M+14H]^{14+} m/z = 1502.13$ ,  $[M+13H]^{13+} m/z = 1617.73$ ,  $[M+12H]^{12+} m/z = 1752.00$ ,  $[M+11H]^{11+} m/z = 1912.00$ . [Ala<sup>183</sup>]-Cp183<sup>61-183</sup> (11)



**HPLC conditions:** Cosmosil 5C4-AR300 column (Nacalai Tesque,  $4.6 \times 250$  mm), linear gradient of 30–60% CH<sub>3</sub>CN containing 0.1% TFA at a flow rate of 1 mL/min over 30 min.

**MS analysis**: Expected mass based on the sequence: 14295.46; Major observed ions:  $[M+11H]^{11+}$ m/z = 1300.95,  $[M+10H]^{10+} m/z = 1431.64$ ,  $[M+9H]^{9+} m/z = 1589.93$ .