# **Supplementary Information**

# Sequestration of Ruthenium Residues via Efficient Fluorous-

# enyne Termination

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General Procedures. All reactions were carried out under a nitrogen atmosphere with dry solvents using anhydrous conditions unless otherwise stated. Dry, degassed dichloromethane  $(CH_2Cl_2)$ , N,Ndimethylformamide (DMF), and tetrahydrofuran (THF) were obtained from a JC Meyer solvent purification system. Deionized (DI) water with a resistivity of 18.2 M $\Omega$ .cm was used for all experiments. CDCl<sub>3</sub> was stored under 4Å molecular sieves to remove water and acid. Unless otherwise stated, all other reagents were purchased at the highest commercial quality and used without further purification. Yields refer to chromatographically and spectroscopically (<sup>1</sup>H-NMR) homogeneous materials. Reactions were monitored by thin layer chromatography (TLC) carried out on 0.25 mm E. Merck silica gel plates (60F-254) using UV light as the visualizing agent and basic aqueous potassium permanganate (KMnO<sub>4</sub>), and heat as developing agents. SiliCyclic silica gel (60, particle size 0.043-0.063 mm) was used for flash column chromatography. NMR spectra were recorded on Bruker Avance 400 or 500 MHz instruments and calibrated using residual undeuterated solvent as an internal reference (CHCl<sub>3</sub> @ 7.26 ppm <sup>1</sup>H NMR, 77.16 ppm <sup>13</sup>C NMR). The following abbreviations (or combinations thereof) were used to explain the multiplicities: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet, br = broad. Mass spectra (MS) were recorded on LC/MS (Agilent Technologies 1260 Infinity II/6120 Quadrupole). Polymer samples were analyzed using a Tosoh EcoSEC HLC 8320GPC system with TSKgel SuperHZ-L columns eluting CHCl<sub>3</sub> containing 0.25% NEt<sub>3</sub> at a flow rate of 0.45 mL/min at 40 °C. All number-average molecular weights and dispersities were calculated from refractive index chromatograms using PStQuick Mp-M polystyrene standards. Melting points were measured on Bibby Scientific's MEL-TEMP Digital Melting Point Apparatus. Ruthenium content was determined by inductively coupled plasma mass spectrometry (IPC-MS, NexION 300Q, Perkin Elmer, USA). Anhydride endo-Tricyclo-[4.2.2.02,5]deca-3,9-diene,<sup>1</sup> N-tosylpropargylamine,<sup>2</sup> M2,<sup>3</sup> M3,<sup>3</sup> M4,<sup>4</sup> and M6<sup>5</sup> were prepared according to literature procedures.

#### **Experimental Procedures**



**Scheme S1:** Route for the synthesis of fluorous enyne (*Fenyne*)

<sup>C</sup><sub>8</sub>F<sub>17</sub> 2-Fluorobenzaldehyde (2.0 g, 16.1 mmol, 1.0 eq) was dissolved in DMF (10 ml) at room temperature, followed by addition of K<sub>2</sub>CO<sub>3</sub> powder (2.67 g, 19.3 mmol, 1.2 eq). The mixture was bubbled with nitrogen gas for 5 minutes, and *1H*,*1H*,*2H*,*2H*-perfluorodecanethiol (8.2 g, 16.9 mmol, 1.05 eq) in DMF (5 ml) was added via syringe. The reaction was placed in an oil bath that was preheated to 70 °C and stirred for 48 hours, then was quenched by H<sub>2</sub>O (20 ml). The aqueous layer was extracted by diethyl ether (3×30 ml). The combined organics were washed with brine (30 ml), dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by column chromatography (7% EtOAc in hexane) to give **F1** as a light-yellow solid (7.71 g, 82%).

 $\label{eq:LC-MS} \mbox{ (m/z): } [M+2H_2O-F]^+ \mbox{ calcd. for } C_{17}H_{13}F_{16}O_3S, \mbox{ 601.0, found, 601.1}$ 

#### Melting point: 52 °C.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 10.33 (s, 1H), 7.88 (dd, J = 7.6, 1.6 Hz, 1H), 7.59 (ddd, J = 8.0, 7.3, 1.6 Hz, 1H), 7.48 – 7.32 (m, 2H), 3.26 – 3.12 (m, 2H), 2.54 – 2.40 (m, 2H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>): δ 191.32, 139.52, 134.25, 134.11, 133.22, 127.44, 126.04, 121.60 - 120.18 (m), 119.52 (t, J = 32.1 Hz), 117.87 (dt, J = 93.9, 32.6 Hz), 116.95 - 114.51 (m), 114.26 - 111.97 (m), 111.61 – 109.46 (m), 109.13 – 107.61 (m), 106.20 (d, J = 39.8 Hz), 30.85 (t, J = 22.1 Hz), 23.58 (d, J = 4.4 Hz).

<sup>19</sup>**F NMR** (376 MHz, CDCl<sub>3</sub>) δ -80.72 (d, J = 9.5 Hz), -114.13 (d, J = 4.1 Hz), -121.63, -121.87 (d, J = 19.5 Hz), -122.67, -123.22, -126.06 (ddd, J = 18.1, 9.2, 3.6 Hz).



To a solution of **F1** (5.84 g, 10 mmol) in dry  $CH_2Cl_2$  (50 ml) was added  $Ph_3P=CHCO_2Me$  (3.68 g, 11 mmol, 1.1 equiv) at -10 °C. After stirring for 1 hour at 0 °C, the solution was placed at rt overnight. Then the reaction

mixture was concentrated under reduced pressure. The residue was purified by column chromatography (10% EtOAc in hexane) to give **F2** (a mixture of *trans* isomers and *cis* isomers, *trans/cis* = 99:1), which was used without further purification. Then, to a solution of **F2** in dry DCM (50 ml) at –30 °C was added dropwise DIBAL-H (20 ml of a 1.1 M solution in tol, 22 mmol, 2.2 equiv). The reaction was stirred further at 0 °C for 5 hours. Then the reaction was quenched by careful addition of a saturated solution of Rochelle salt (20 ml), and the mixture was allowed to warm to rt. The aqueous layer was extracted by DCM (3×30 ml). The combined organics were washed with brine (30 ml), dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated in vacuo. The residue was purified by column chromatography (20% EtOAc in hexane) to give **F3** as a light yellow liquid (4.1 g, 67% over 2 steps).

**LC-MS** (m/z):  $[M+2H_2O-F]^+$  calcd. for  $C_{19}H_{17}F_{16}O_3S$ , 629.1, found, 629.1

#### Melting point: 40 °C.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>): δ 7.58 – 7.49 (m, 1H), 7.46 – 7.35 (m, 1H), 7.31 – 7.23 (m, 1H), 7.14 (dt, J = 15.8, 1.7 Hz, 1H), 6.32 (dt, J = 15.8, 5.6 Hz, 1H), 4.37 (dd, J = 5.6, 1.7 Hz, 2H), 3.20 – 2.77 (m, 2H), 2.35 (tt, J = 17.6, 8.3 Hz, 2H).

<sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>): δ 138.54, 132.42, 131.40, 131.24, 128.19, 128.17, 127.72, 126.82, 121.40 (t, J = 33.0 Hz), 120.60 - 119.45 (m), 118.93 - 117.05 (m), 116.51 - 114.43 (m), 114.37 - 112.18 (m), 112.10 - 109.30 (m), 108.02 (dd, J = 69.1, 38.9 Hz), 105.67 (d, J = 38.6 Hz), 63.54, 31.46 (t, J = 22.0 Hz), 25.03 (t, J = 4.2 Hz).

<sup>19</sup>**F NMR** (376 MHz, CDCl<sub>3</sub>) δ -80.73, -112.45 - -117.08 (m), -121.70, -121.89, -122.69, -123.27, -126.07 (td, J = 13.2, 12.1, 5.5 Hz).



To a stirred and cooled (0 °C) solution of **F3** (613 mg, 1.0 mmol, 1 equiv), N-tosyl-propargylamine (209 mg, 1.0 mmol, 1 equiv) and PPh3 (420 mg, 1.6 mmol, 1.6 equiv) in anhydrous THF (8 ml) was added

dropwise DIAD (0.3 ml, 1.5 mmol, 1.5 equiv). The reaction was placed in a water bath and stirred for 24 hours at room temperature. Then the reaction was concentrated in vacuo. Chromatography gave the target enyne  $\mathbf{F}_{enyne}$  (0.7 g, 87%) as a light yellow oil.

LC-MS (m/z): [M+2H<sub>2</sub>O-F]<sup>+</sup> calcd. for C<sub>29</sub>H<sub>26</sub>F<sub>16</sub>NO<sub>4</sub>S<sub>2</sub>, 820.1, found, 820.1

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.79 (d, J = 8.4 Hz, 2H), 7.53 – 7.45 (m, 1H), 7.45 – 7.38 (m, 1H), 7.34 (d, J = 7.9 Hz, 2H), 7.32 – 7.28 (m, 1H), 7.15 (d, J = 15.7 Hz, 1H), 6.08 (dt, J = 15.7, 6.9 Hz, 1H), 4.17 (d, J = 2.5 Hz, 2H), 4.05 (dd, J = 6.9, 1.3 Hz, 2H), 3.10 – 2.94 (m, 2H), 2.46 (s, 3H), 2.34 (dp, J = 17.5, 8.1 Hz, 2H), 2.07 (t, J = 2.4 Hz, 1H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>): δ 143.51, 137.90, 135.72, 132.28, 132.14, 131.50, 129.35, 128.42, 127.74, 127.62, 126.77, 125.61, 120.22 (d, J = 32.6 Hz), 119.38 (t, J = 31.5 Hz), 118.06 (t, J = 33.1 Hz), 117.34 (t, J = 31.7 Hz), 116.44 - 114.63 (m), 113.44 - 111.89 (m), 111.50 - 109.43 (m), 108.34 (t, J = 32.4 Hz), 76.18, 73.78, 48.50, 35.82, 31.27 (t, J = 22.0 Hz), 25.04 (d, J = 4.3 Hz), 21.26.

<sup>19</sup>**F NMR** (376 MHz, CDCl<sub>3</sub>) δ -80.73, -109.85 - -116.63 (m), -121.70, -121.89 (d, J = 19.6 Hz), -122.68, -123.23, -126.07 (td, J = 13.1, 11.9, 4.1 Hz).

Scheme S2: Route for the synthesis of M5



Anhydride endo-Tricyclo-[4.2.2.02.5]deca-3,9-diene (1.00 g, 4.94 mmol) and benzylamine (0.28 ml, 4.94 mmol) was purged with argon gas. Then 20 mL of anhydrous toluene was added, and the flask was fitted with a Dean-Stark trap and a condenser. The reaction mixture was refluxed at 120 °C for 16 h. After completion of reflux, the solvent was removed on a rotary evaporator. The residue was purified by column chromatography (25% EtOAc in hexane) to give **M5** as a white solid (1.11 g, 77%).

**LC-MS** (m/z): [M+H]<sup>+</sup> calcd. for C<sub>19</sub>H<sub>18</sub>NO<sub>2</sub>, 292.1, found, 292.2.

Melting point: 150-151 °C.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>): δ 7.35 – 7.20 (m, 5H), 5.87 (s, 2H), 5.79 (dd, J = 4.7, 3.2 Hz, 2H), 4.57 (s, 2H), 3.16 (ddt, J = 3.5, 2.4, 1.3 Hz, 2H), 2.83 – 2.79 (m, 2H), 2.79 (t, J = 1.6 Hz, 2H).

<sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>): δ 178.24, 137.76, 135.66, 128.31, 128.24, 128.08, 127.50, 43.95, 43.18, 41.99, 36.49.

#### **General Polymerization Procedure**

Grubbs 3 catalyst with desired amount was added to an 8 mL vial (Chemglass, CG-4909-03), placed under a nitrogen atmosphere, at room temperature. Then the desired amount of DCM or THF was

added to make a stock solution (0.015 M). In another 8 mL vial equipped with a stir bar, a solution of **M1-M6** (30-500 eq) in degassed DCM or THF was prepared. Appropriate volume of Grubbs 3 solution is rapidly added to the vial containing monomers using a microliter syringe. After the desired time, the polymerization was stopped by addition of ethyl vinyl ether (**EVE**, 0.2 ml) or  $\mathbf{F}_{enyne}$  (1.5eq for **M1-M4** and **M6**, 5 eq for **M5**). The reaction mixture was further stirred for 10 minutes (**M1-M4**) or 4 hours (**M5**) before it was precipitated using 15-fold volume of methanol or purified via Fluorous Solid-phase Extraction (**F-SPE**). Polymer parts were dried under vacuum for analysis using GPC and NMR.

#### **Polymerization Data**

Entry	Monome	[M]	Solvent	חח	Torminator	Mothod <sup>b</sup>	Recovery	$M_{n,SEC}^{c}$	Ð
Entry	r	(mol/L)	Solvent	DP	Terminator	Method	(%)	(kDa)	$\mathcal{D}^{*}$
1	M1	1	DCM	30	EVE	MeOH	85	8.5	2.03
2	M1	1	DCM	30	F <sub>enyne</sub>	MeOH	84	9.2	1.98
3	M1	1	DCM	30	F <sub>enyne</sub>	CF <sub>3</sub> -OH	82	9.3	1.97
4	M1	1	DCM	30	F <sub>enyne</sub>	F-SPE	47	8.1	1.96
5	M2	0.2	DCM	30	EVE	MeOH	91	8.9	1.11
6	M2	0.2	DCM	30	F <sub>enyne</sub>	MeOH	91	9.2	1.11
7	M2	0.2	DCM	30	F <sub>enyne</sub>	F-SPE	91	9.3	1.10
8	M2	0.2	DCM	60	F <sub>enyne</sub>	MeOH	92	17.6	1.09
9	M2	0.2	DCM	60	F <sub>enyne</sub>	F-SPE	92	17.6	1.09
10	M2	0.2	DCM	100	Fenyne	F-SPE	92	29.9	1.09
11	M2	0.2	DCM	500	F <sub>enyne</sub>	F-SPE	92	116	1.11
12	M3	0.2	DCM	30	EVE	MeOH	90	8.3	1.12
13	M3	0.2	DCM	30	F <sub>enyne</sub>	MeOH	91	8.8	1.11
14	M3	0.2	DCM	30	F <sub>enyne</sub>	F-SPE	91	8.8	1.11
15	M4	0.2	DCM	30	EVE	MeOH	82	7.2	1.19
16	M4	0.2	DCM	30	F <sub>enyne</sub>	MeOH	81	7.9	1.18
17	M4	0.2	DCM	30	F <sub>enyne</sub>	F-SPE	85	7.9	1.18
18	M5	0.2	DCM	30	EVE	MeOH	92	7.4	1.12
19 <sup>d</sup>	M5	0.2	DCM	30	F <sub>enyne</sub>	MeOH	92	8.0	1.13
20 <sup>d</sup>	M5	0.2	DCM	30	F <sub>enyne</sub>	F-SPE	93	8.1	1.13
21	M6	0.2	THF	30	EVE	MeOH	84	9.4	1.39
22	M6	0.2	THF	30	F <sub>enyne</sub>	MeOH	84	10	1.32
23	M6	0.2	THF	30	F <sub>enyne</sub>	F-SPE	88	10	1.31
24	M2	0.2	THF	30	F <sub>enyne</sub>	MeOH	90	9.6	1.12
25	M2	0.2	THF	30	F <sub>enyne</sub>	F-SPE	92	9.6	1.12
26	M2	0.2	EtOAc	30	F <sub>enyne</sub>	MeOH	88	6.6	1.38

#### Table S1. Polymerization of M1-M5.

27	M2	0.2	Tol	30	F <sub>enyne</sub>	MeOH	89	3.8	1.82
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<sup>*a*</sup>Polymerization Condition: 3 µmol **G3** catalyst, termination using 0.2 ml **EVE** or 1.5 eq **F**<sub>enyne</sub>. <sup>*b*</sup>MeOH or CF3-OH: precipitation using 6 ml of MeOH or CF<sub>3</sub>CH<sub>2</sub>OH, F-SPE: Flourous Solid Phase Extraction. <sup>*c*</sup>Determined by CHCl<sub>3</sub> size-exclusion chromatography (SEC) calibrated using polystyrene standards. <sup>*d*</sup>Termination using 5 eq **F**<sub>enyne</sub>.

Characterization Data of Polymers (P1 - P6,  $P1_F - P6_F$ )



**P1**: Prepared according to Polymerization General Procedure using **M1** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>):  $\delta$  7.37 (d, J = 1.7 Hz, 1H), 7.35 (d, J = 1.3 Hz, 1H), 7.33 (d, J = 2.5 Hz, 1H), 7.30 (d, J = 1.8 Hz, 1H), 7.26 - 7.10 (m, 1H), 6.55 - 6.34 (m, 1H), 6.24 (dt, J = 15.8, 6.8 Hz, 1H), 5.83 (ddt, J = 16.9, 10.1, 6.6 Hz, 1H), 5.64 - 5.19 (m, 63H), 5.14 - 4.83 (m, 2H), 2.13 - 1.86 (m, 126H), 1.41 - 1.12 (m, 256H). **SEC** (CHCl<sub>3</sub>, RI):  $M_n$  = 8.5 kg/mol, D = 2.03.



**P1**<sub>F:</sub> Prepared according to Polymerization General Procedure using **M1** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.84 – 7.69 (m, 2H), 7.46 – 7.29 (m, 5H), 7.26 – 7.17 (m, 1H), 6.39 (d, J = 15.8 Hz, 1H), 6.25 (dt, J = 15.9, 6.9 Hz, 1H), 5.61 – 5.23 (m, 67H), 4.44 – 3.97 (m, 4H), 2.52 – 2.37 (m, 3H), 2.28 – 1.84 (m, 134H), 1.53 – 1.04 (m, 268H).

**SEC** (CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 9.2 kg/mol, *Đ* = 1.98.



Figure S1. SEC traces for P1 and P1<sub>F</sub>.



**P2**: Prepared according to Polymerization General Procedure using **M2** and dihydrofuran with targeted degrees of polymerization of 30.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>): δ 7.58 – 6.94 (m, 153H), 6.66 – 6.42 (m, 1H), 6.00 – 5.87 (m, 1H), 5.91 – 5.65 (m, 30H), 5.61 – 5.34 (m, 30H), 5.18 (dd, J = 34.7, 13.7 Hz, 2H), 4.86 – 4.25 (m, 60H), 3.38 – 2.59 (m, 121H), 2.33 – 1.93 (m, 30H), 1.75 – 1.45 (m, 30H).

**SEC** (CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 8.9 kg/mol, *Đ* = 1.11.



**P2**<sub>F:</sub> Prepared according to Polymerization General Procedure using **M2** and dihydrofuran with targeted degrees of polymerization of 30, 60, 100, and 500.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>): δ 7.75 (dd, J = 8.2, 3.0 Hz, 2H), 7.50 – 7.02 (m, 160H), 6.73 – 6.49 (m, 1H), 6.39 – 6.16 (m, 1H), 5.97 – 5.64 (m, 32H), 5.60 – 5.42 (m, 32H), 4.71 – 4.43 (m, 64H), 4.37 – 3.93 (m, 4H), 3.53 – 2.56 (m, 127H), 2.44 (d, J = 5.1 Hz, 3H), 2.24 – 2.01 (m, 32H), 1.75 – 1.43 (m, 32H).

**SEC** (DP30, CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 9.2 kg/mol, *Đ* = 1.11.



Figure S2. SEC trace for P2 and P2<sub>F</sub>.



**P3**: Prepared according to Polymerization General Procedure using **M3** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>): δ 7.57 – 7.08 (m, 152H), 6.78 – 6.23 (m, 1H), 5.94 (dt, J = 17.5, 8.4 Hz, 1H), 5.80 – 5.12 (m, 61H), 5.29 – 4.92 (m, 2H), 4.91 – 4.17 (m, 62H), 3.57 – 2.54 (m, 124H), 2.25 – 1.45 (m, 31H), 1.40 – 0.75 (m, 32H).

**SEC** (CHCl<sub>3</sub>, RI):  $M_n = 8.3 \text{ kg/mol}$ , D = 1.12.



**P3**<sub>F</sub>: Prepared according to Polymerization General Procedure using **M3** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>): δ 7.75 (q, J = 6.8, 5.7 Hz, 2H), 7.65 – 6.98 (m, 178H), 6.72 – 6.21 (m, 1H), 6.14 – 5.84 (m, 1H), 5.89 – 5.14 (m, 70H), 5.07 – 4.07 (m, 70H), 4.45 – 4.06 (m, 4H), 3.39 – 2.67 (m, 141H), 2.59 – 2.30 (m, 3H), 2.08 – 1.43 (m, 32H), 1.54 – 0.83 (m, 33H).

**SEC** (CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 8.8 kg/mol, *Đ* = 1.11.



Figure S3. SEC trace for P3 and P3<sub>F</sub>.



**P4**: Prepared according to Polymerization General Procedure using **M4** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.73 – 7.08 (m, 140 H), 6.91 – 6.65 (m, 1H), 6.47 – 6.24 (m, 1H), 6.19 – 5.92 (m, 27H), 5.89 – 5.51 (m, 28H), 5.49 – 5.19 (m, 2H), 5.11 – 4.85 (m, 27H), 4.72 – 4.49 (m, 56H), 4.52 – 4.32 (m, 28H), 3.59 – 2.91 (m, 54H).

**SEC** (CHCl<sub>3</sub>, RI):  $M_n = 7.2 \text{ kg/mol}$ , D = 1.19.



**P4**<sub>F:</sub> Prepared according to Polymerization General Procedure using **M4** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.84 – 7.64 (m, 2H), 7.57 – 7.14 (m, 144H), 6.96 – 6.69 (m, 1H), 6.53 – 6.26 (m, 1H), 6.16 – 5.90 (m, 27H), 5.82 – 5.59 (m, 28H), 5.13 – 4.83 (m, 28H), 4.77 – 4.49 (m, 57H), 4.49 – 4.27 (m, 28H), 4.29 – 4.03 (m, 4H), 3.44 – 2.92 (m, 58H), 2.51 – 2.26 (m, 3H).

**SEC** (CHCl<sub>3</sub>, RI):  $M_n = 7.9 \text{ kg/mol}$ , D = 1.18.



Figure S4. SEC trace for P4 and P4<sub>F</sub>.



**P5**: Prepared according to Polymerization General Procedure using **M5** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.47 – 7.07 (m, 162H), 6.59 – 6.43 (m, 1H), 6.44 – 5.98 (m, 64H), 5.49 (dt, J = 18.0, 9.4 Hz, 1H), 5.38 – 5.21 (m, 1H), 5.25 – 4.89 (m, 63H), 4.69 – 4.11 (m, 63H), 3.42 – 2.40 (m, 190H). **SEC** (CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 7.4 kg/mol, *Đ* = 1.12.



**P5**<sub>F</sub>: Prepared according to Polymerization General Procedure using **M5** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.88 – 7.68 (m, 2H), 7.42 – 6.95 (m, 165H), 6.61 – 5.96 (m, 64H), 5.95 – 5.78 (m, 2H), 5.40 – 4.64 (m, 64H), 4.77 – 4.21 (m, 64H), 4.32 – 4.00 (m, 4H), 3.41 – 2.54 (m, 192H), 2.48 – 2.23 (m, 3H).

**SEC** (CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 8.0 kg/mol, *Đ* = 1.11.



Figure S5. SEC trace for P5 and P5<sub>F</sub>.



**P6**: Prepared according to Polymerization General Procedure using **M6** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.98 – 7.61 (m, 63H), 7.36 – 7.29 (m, 63H), 6.66 (m, 1H), 6.35 (dd, J = 16.3, 7.3 Hz, 1H), 6.04 (dd, J = 15.8, 3.9 Hz, 18H), 5.84 – 5.70 (m, 14H), 5.65 – 5.28 (m, 61H), 5.11 – 4.90 (m, 2H), 4.46 (s, 31H), 4.22 (m, 63H), 2.42 (d, J = 7.9 Hz, 98H), 2.13 (q, J = 7.9 Hz, 61H), 1.88 – 1.69 (m, 61H), 1.54 – 1.35 (m, 61H).

**SEC** (CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 9.4 kg/mol, *Đ* = 1.39.



**P6**<sub>F:</sub> Prepared according to Polymerization General Procedure using **M6** and dihydrofuran with a targeted degree of polymerization of 30.

<sup>1</sup>**H NMR** (500 MHz, CDCl<sub>3</sub>): δ 7.78 – 7.65 (m, 64H), 7.42 – 7.29 (m, 64H), 6.79 – 6.52 (m, 1H), 6.41 – 6.18 (m, 1H), 6.12 – 5.98 (m, 20H), 5.84 – 5.63 (m, 12H), 5.57 – 5.23 (m, 61H), 4.46 (s, 31H), 4.35 – 4.03 (m, 62H), 2.46 – 2.34 (m, 97H), 2.24 – 2.03 (m, 61H), 1.89 – 1.72 (m, 63H), 1.56 – 1.31 (m, 62H).

**SEC** (CHCl<sub>3</sub>, RI): *M*<sub>n</sub> = 10 kg/mol, *Đ* = 1.31.



Figure S6. SEC trace for P6 and P6<sub>F</sub>.



Analysis of <sup>1</sup>H NMR signal assignment for polymer P2 and P2<sub>F</sub>.

**Figure S7**. <sup>1</sup>H NMR for polymers: a.**P2**, b. **P2**<sub>F</sub>. <sup>1</sup>H NMRs shows no terminal alkene could be observed in **P2**<sub>F</sub>, exhibiting high efficiency of **F**<sub>enyne</sub> termination and quantitative conversion of Ru chain-end into **Ru-S**.

#### Model metathesis reaction of Fenyne with Grubbs initiators



Grubbs 3 catalyst with desired amount was added to an 8 mL vial (Chemglass, CG-4909-03), placed under a nitrogen atmosphere, at room temperature. Then the desired amount of CDCl<sub>3</sub> was added to make a stock solution (0.001 M). In another 8 mL vial equipped with a stir bar, a solution of  $\mathbf{F}_{enyne}$  (1.5eq) in degassed CDCl<sub>3</sub> was prepared. Appropriate volume of Grubbs 3 solution is rapidly added to the vial containing monomers using a microliter syringe. The reaction mixture was stirred at RT and aliquots (0.1 mL each) were taken after desired time periods. Each aliquot analyzed by <sup>1</sup>H NMR.



Figure S8. <sup>1</sup>H NMR for analysis of model reaction of G3 with F<sub>enyne</sub>.

Grubbs 3 catalyst with desired amount was added to an 8 mL vial (Chemglass, CG-4909-03), placed under a nitrogen atmosphere, at room temperature. Then the desired amount of CDCl<sub>3</sub> was added to make a stock solution (0.015 M). In another 8 mL vial equipped with a stir bar, a solution of monomers

(10 eq) in degassed CDCl<sub>3</sub> was prepared. At the same time, a stock solution of  $\mathbf{F}_{enyne}$  (0.01M) in degassed CDCl<sub>3</sub> was prepared. Appropriate volume of Grubbs 3 solution is rapidly added to the vial containing monomers using a microliter syringe. After 10 mins, a desired volume of a stock solution of  $\mathbf{F}_{enyne}$  (1.5eq) was added into the polymerization mixture. The mixture was stirred at RT and aliquots (0.1 mL each) were taken after desired time periods. Each aliquot analyzed by <sup>1</sup>H NMR.





Figure S9. <sup>1</sup>H NMR analysis for the chain-end termination in polymerization of M2.





19.1 19.0 18.9 18.8 18.7 18.6 18.5 18.4 18.3 18.2 18.1 18.0 17.9 17.8 17.7 17.6 17.5 17.4 17.3 17.2 17.1 17.0 16.9 16.8 16.7 16.6 16.5 16.4 ppm







3.00 17.95 17.90 17.85 17.80 17.75 17.50 17.55 17.50 17.55 17.40 17.35 17.40 17.35 17.40 17.25 17.20 17.15 17.10 17.05 16.90 16.95 16.90 16

**Figure S11.** <sup>1</sup>H NMR analysis for the chain-end termination in polymerization of **M5**.



**Figure S12.** <sup>1</sup>H NMR analysis for the chain-end termination in polymerization of **M6**.

### Ruthenium removal protocols and analysis of ruthenium content of polymer samples

**MeOH wash:** After termination via addition of desired amount of EVE or  $F_{enyne}$ , 15-fold volume of methanol was added into the mixture, followed by stirring for 5 minutes. After centrifugation at 4300 rpm for 20 minutes, the solvent was decanted, and polymers were collected. Obtained polymer parts were dried under vacuum for analysis using GPC, NMR, and ICP-MS.

**Fluorous Solid-phase Extraction (F-SPE)**: Following termination via addition of desired amount of  $F_{enyne}$ , the solvent of mixture was evaporated. Then, the desired amount of DCM was added, and the mixture was loaded onto the pre-activated fluorous cartridge, followed by elution using DCM/DMSO (v/v=3/7). Obtained polymer parts were dried under vacuum for analysis using GPC, NMR, and ICP-MS.

**Digestion method for ICP-MS**: Samples ranging in mass from 10 to 15 mg were digested in Parr instrument A280AC vessels placed in a heat oven (Fisherbrand<sup>™</sup> 180L Gravity Oven) with 4 mL of 65% HNO<sub>3</sub> (Suprapur<sup>™</sup>, MilliporeSigma<sup>™</sup>) and 0.15 mL of 70% HClO<sub>4</sub> (99.999% trace metals basis, Sigma-Aldrich). Samples were pre-digested before heat by aging the sample mixed with acids for 16 h at room temperature. Then, the oven system was set at 130 °C for 8 hours. Final digests were diluted with deionized water to 10 ml stock solution in volumetric flasks and measured within 24 h.

**ICP-MS:** Calibration curve was obtained by measuring a series of standard solutions with concentrations ranging from 0.001 mg/L to 0.200 mg/L (prepared by diluting a standard solution containing 10 mg/L of ruthenium in a matrix of 10% HCl and 1% HNO<sub>3</sub>, PerkinElmer TruQms). The measurement was obtained as a count of ions with mass-to-charge ratio of 102. The apparatus was purged with 1% HNO<sub>3</sub> for 90 s after each sample and the signal from each sample was allowed to stabilize for 30 s before each measurement. Three measurements were obtained for each sample and averaged. The limit of quantification calculated from obtained results for blank solutions was 0.03 ppm.

Entrya	Monomor	[M]	Solvent	קט	Tominator	Mothod <sup>b</sup>	Ru level <sup>c</sup>	Ru Removal
Entry	WONUTIER	(mol/L)	Solvent	DF	Terrinator	Method	(ppm)	(%)
1	M1	1	DCM	30	EVE	MeOH	634±8	98.40
2	M1	1	DCM	30	F <sub>enyne</sub>	MeOH	182±3	99.54
3	M1	1	DCM	30	F <sub>enyne</sub>	CF <sub>3</sub> -OH	132±7	99.67
4	M1	1	DCM	30	F <sub>enyne</sub>	F-SPE	14±3	99.96
5	M2	0.2	DCM	30	EVE	MeOH	506±9	97.70
6	M2	0.2	DCM	30	Fenyne	MeOH	179±4	99.19
7	M2	0.2	DCM	30	Fenyne	F-SPE	70±4	99.68
8	M2	0.2	DCM	60	F <sub>enyne</sub>	MeOH	98±3	99.12
9	M2	0.2	DCM	60	Fenyne	F-SPE	36±4	99.67
10	M2	0.2	DCM	100	Fenyne	F-SPE	16±4	99.76
11	M2	0.2	DCM	500	F <sub>enyne</sub>	F-SPE	4±1	99.89
12	M3	0.2	DCM	30	EVE	MeOH	508±14	97.69
13	M3	0.2	DCM	30	Fenyne	MeOH	180±5	99.18
14	M3	0.2	DCM	30	F <sub>enyne</sub>	F-SPE	75±3	99.66
15	M4	0.2	DCM	30	EVE	MeOH	802±14	96.33

Table S2. Ruthenium residue content of polymers (P1 - P6,  $P1_F - P6_F$ ).

16	M4	0.2	DCM	30	F <sub>enyne</sub>	MeOH	687±24	96.85
17	M4	0.2	DCM	30	F <sub>enyne</sub>	F-SPE	544±16	97.51
18	M5	0.2	DCM	30	EVE	MeOH	686±8	96.42
19 <sup>d</sup>	M5	0.2	DCM	30	F <sub>enyne</sub>	MeOH	254±5	98.67
20 <sup>d</sup>	M5	0.2	DCM	30	F <sub>enyne</sub>	F-SPE	59±4	99.69
21	M6	0.2	THF	30	EVE	MeOH	633±19	96.72
22	M6	0.2	THF	30	F <sub>enyne</sub>	MeOH	385±8	98.01
23	M6	0.2	THF	30	F <sub>enyne</sub>	F-SPE	167±5	99.13
24	M2	0.2	THF	30	F <sub>enyne</sub>	MeOH	179±7	99.19
25	M2	0.2	THF	30	F <sub>enyne</sub>	F-SPE	72±2	99.67
26	M2	0.2	EtOAc	30	F <sub>enyne</sub>	MeOH	21783±211	1.03
27	M2	0.2	Tol	30	F <sub>enyne</sub>	MeOH	21685±198	1.47

<sup>*a*</sup>Polymerization Condition: 3 µmol **G3** catalyst, termination using 0.2 ml **EVE** or 1.5 eq  $\mathbf{F}_{enyne}$ . <sup>*b*</sup>MeOH or CF3-OH: precipitation using 6 ml of MeOH or CF<sub>3</sub>CH<sub>2</sub>OH, F-SPE: Flourous Solid Phase Extraction. <sup>*c*</sup>Determined by ICP-MS, and results were obtained from 3 runs. <sup>*d*</sup>Termination using 5 eq  $\mathbf{F}_{enyne}$ .

Entry <sup>a</sup>	Monomer	[M] (mol/L)	DP	Method <sup>b</sup>	Elution solvent <sup>b</sup>	Ru level <sup>c</sup> (ppm)
1	M4	0.2	30	MeOH	DCM/DMSO (v/v=3/7)	532±21
2	M4	0.2	30	MeOH	THF/DMSO (v/v=1/5)	574±27
3	M4	0.2	30	F-SPE	DCM/MeOH (v/v=1/1)	593±22

Table S3. Ruthenium residue content of polymers (P4<sub>F</sub>).

<sup>*a*</sup>Polymerization Condition: 3  $\mu$ mol **G3** catalyst in DCM, termination using 1.5 eq **F**<sub>enyne</sub>. <sup>*b*</sup> MeOH: precipitation once using 6 mL methanol, F-SPE: Flourous Solid Phase Extraction, Elution solvent: hydrophobic solvent used for polymers purification. <sup>*c*</sup>Determined by ICP-MS, and results were obtained from 3 runs.



Figure S13. Residual Ruthenium content in P1 and P1<sub>F</sub> purified by different methods.



Figure S14. Residual Ruthenium content in P3 and  $P3_F$  purified by different methods.



Figure S15. Residual Ruthenium content in P4 and P4<sub>F</sub> purified by different methods.



Figure S16. Residual Ruthenium content in P5 and P5<sub>F</sub> purified by different methods.



Figure S17. Residual Ruthenium content in P6 and P6<sub>F</sub> purified by different methods.

Entry <sup>a</sup>	Monomer	[M] (mol/L)	DP	Ru level <sup>♭</sup> (ppm)
1	M1	1	30	49741±531
2	M2	0.2	30	22009±241
3	M2	0.2	60	11077±135
4	M2	0.2	100	6664±100
5	M2	0.2	500	4005±74
6	M3	0.2	30	22046±211
7	M4	0.2	30	21842±198
8	M5	0.2	30	19168±173
9	M6	0.2	30	19296±187

|--|

<sup>*a*</sup>Polymerization Condition: 3 µmol **G3** catalyst in DCM or THF, termination using EVE. <sup>*b*</sup>Determined by ICP-MS, and results were obtained from 3 runs.



**Figure S18.** TGA curves of **P2**<sub>F</sub> in N<sub>2</sub>. For **P2**<sub>F</sub> without purification, 92% loss at 300 °C, for **P2**<sub>F</sub> with methanol precipitation, 92% at 403 °C, and for **P2**<sub>F</sub> with F-SPE, 88% loss at 410 °C in the N<sub>2</sub>.





### <sup>1</sup>H and <sup>13</sup>C NMR Spectra











8.4 8.2 8.0 7.8 7.6 7.4 7.2 7.0 6.8 6.6 6.4 6.2 6.0 5.8 5.6 5.4 5.2 5.0 4.8 4.6 4.4 4.2 4.0 3.8 3.6 3.4 3.2 3.0 2.8 2.6 2.4 2.2 2.0 1.8 ppm







S31







S33









![](_page_37_Figure_0.jpeg)

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