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Self-Assembly of Single-Chain Nanoparticles from Block Copolymers into Inverse

Bicontinuous Structures

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Synthesis routes of P4VP-CTA, P4VP-b-PS, and P4VP(SCNP)-b-PS



Scheme S1. The synthesis route of P4VP-CTA.



Scheme S2. The synthesis route of P4VP-b-PS.



Scheme S3. The synthesis route of P4VP(SCNP)-b-PS.

Characterization.

NMR Spectroscopy. ¹H NMR spectra were obtained by Bruker DMX Spectrometer (400 MHz) using CDCl₃ as a solvent.

Gel Permeation Chromatography (GPC). The P4VP-*b*-PS and P4VP(SCNP)-*b*-PS were measured on Waters 2410 GPC. The eluent was THF with a flow rate of 1 mL/min. The samples were dissolved in THF, and filtered through 0.22 μ m syringe filters. The calibration curves were obtained from PS standards. Some samples were also measured on a system comsisted of Wyatt Technology detector, Agilent HPLC pump, and Agilent mixed columns (Plgel 20 μ m MIXED-A and PLgel 10 μ m MIXED-B). The eluent of DMF with LiBr (0.05 mol/L) was used and the measurements were conducted at 25 °C with a flow rate of 1 mL/min.

Dynamic Light Scattering (DLS). DLS measurements were conducted on the instrument of Shandong Naikete NKT-N9 with a 532 nm He-Ne laser.

Scanning Electron Microscopy (SEM). Images were taken using a Tescan Vega3 microscope with an accelerating voltage of 30 kV and a Zeiss SUPRA55 instrument at an accelerating voltage of 10

kV. The dilute dispersion solution was dropped onto a silicon wafer and then sputtered with gold.

Transmission Electron Microscopy (TEM). Images were taken using a JEM-2100Plus microscope with an accelerating voltage of 200 kV. The dilute dispersion solution was dropped onto a copper grid.

Small Angle X-ray Scattering (SAXS). SAXS measurements were performed on the synchrotron radiation at BL16B1 beamline provided by Shanghai Synchrotron Radiation Facility (SSRF) with a wavelength of 1.24 Å at room temperature.

Synthesis of P4VP-CTA via RAFT solution polymerization

P4VP macromolecular chain transfer agents (macro-CTAs) were synthesized by reversible additionfragmentation chain transfer (RAFT) solution polymerization (Scheme S1). Typically, 4-VP (0.63 g, 6.0 mmol), CPADB (40.5 mg, 0.1 mmol), and AIBN (1.64 mg, 0.01 mmol) dissolved in 3 mL ethanol were added into a Schlenk flask. After three freeze-pump-thaw cycles, the flask was sealed. The reaction was quenched in liquid nitrogen after 6 h at 70 °C and precipitated in petroleum ether three times. Then the product was dialyzed in ethanol and dried under vacuum. The degree of polymerization of P4VP (DP_{P4VP}) is calculated by ¹H NMR (Figure S1) as follows:

$$DP_{P4VP} = \frac{I_d}{I_b}$$
(S1)

where I_b represents the integrated value of the proton peaks originating from the phenyl groups of 4-Cyano-4-(phenylcarbonothioylthio) pentanoic acid (CPADB) (signal b, $\delta = 7.78-7.90$ ppm); I_d represents the integrated value of the proton peaks attributed to the pyridine groups of P4VP block (signal d, $\delta = 8.15-8.65$ ppm).



Figure S1 ¹H NMR spectra of P4VP₄₀-CTA and P4VP₅₃-CTA (from top to bottom).

Synthesis of P4VP-b-PS via RAFT dispersion polymerization

The degree of polymerization of PS (DP_{PS}) is calculated by ¹H NMR as follows:¹

$$\frac{I_{a}}{I_{b+c+d}} = \frac{2 \times DP_{P4VP}}{5 \times DP_{PS} + 2 \times DP_{P4VP}}$$
(S2)

where I_a represents the integrated value of the proton peaks at 8.20-8.60 ppm, originating from the pyridine groups of P4VP block (signal a); I_{b+c+d} represents the integrated value of the proton peaks at 6.20-7.20 ppm, originating from the phenyl groups of PS block and the pyridine groups of P4VP block. Accordingly, the DP_{PS} is calculated as follows:

$$DP_{PS} = \frac{2 \times DP_{P4VP} \times \left(\frac{I_{b+c+d+e}}{I_{a}} - 1\right)}{5}$$
(S3)

Taking P4VP₅₃-*b*-PS₄₉₉ as an example:

$$DP_{PS} = \frac{2 \times DP_{P4VP} \times (\frac{I_{b+c+d+e}}{I_{a}} - 1)}{5} = \frac{2 \times 53 \times (\frac{24.56}{1} - 1)}{5} \approx 499$$



Figure S2. ¹H NMR spectrum of P4VP₅₃-*b*-PS₄₉₉.

The volume fraction of the PS block (f_{PS}) is calculated as follows:²

$$f_{\rm PS} = \frac{V_{\rm PS}}{V_{\rm P4VP} + V_{\rm PS}} = \frac{M_{\rm PS}/\rho_{\rm PS}}{M_{\rm P4VP}/\rho_{\rm P4VP} + M_{\rm PS}/\rho_{\rm PS}}$$
(S4)

In equation S4, V_{P4VP} and V_{PS} represent the volumes of P4VP and PS chains respectively; M_{P4VP} and M_{PS} denote the molecular weights of the P4VP and PS blocks, respectively; ρ_{P4VP} represents the density of P4VP, which is about 1.15 g cm⁻³; ρ_{PS} represents the density of PS, which is about 1.05 g cm⁻³.

Taking P4VP₅₃-*b*-PS₄₉₉ as an example:

$$f_{\rm PS} = \frac{V_{\rm PS}}{V_{\rm P4VP} + V_{\rm PS}} = \frac{M_{\rm PS}/\rho_{\rm PS}}{M_{\rm P4VP}/\rho_{\rm P4VP} + M_{\rm PS}/\rho_{\rm PS}} = \frac{499 \times 104/1.05}{53 \times 105/1.15 + 499 \times 104/1.05} \approx 91.1\%$$

Preparation and characterization of tadpole-like SCNPs.

The number of DIB (x) linked to P4VP is calculated by 1 H NMR as follows:

$$x = \frac{1}{4} \times \frac{I_f \times 2 \times DP_{P4VP}}{I_a}$$
(S5)

Therefore, the actual CD of SCNPs is calculated by the following equation:

$$CD = \frac{2x}{DP_{P4VP}} \times 100\% = \frac{I_{f}}{I_{a}} \times 100\%$$
(S6)

Taking CD_{27%}-P4VP(SCNP)₅₃-*b*-PS₅₆₉as an example (Figure S3):

$$x = \frac{1}{4} \times \frac{I_{\rm f} \times 2 \times \rm{DP}_{\rm{P4VP}}}{I_{\rm a}} = \frac{1}{4} \times \frac{0.27 \times 2 \times 53}{1} = 7.155$$

CD =
$$\frac{2 \times 7.155}{53} \times 100\% = \frac{I_f}{I_a} \times 100\% = \frac{0.27}{1} \times 100\% = 27\%$$

So, this sample can be named CD_{27%}-P4VP(SCNP)₅₃-b-PS₅₆₉.



Figure S3. ¹H NMR spectrum of CD_{27%}-P4VP(SCNP)₅₃-b-PS₅₆₉ prepared in dilute DCM solution.



Figure S4. GPC traces of P4VP(SCNP)₄₀-*b*-PS₅₁₂ using DMF as the eluent.



Figure S5. (a) DLS curves of P4VP₅₃-*b*-PS₅₆₉ and P4VP(SCNP)₅₃-*b*-PS₅₆₉ at CD of 0%, 18%, and 27%. (b) Comparison curves between 27% and 40%.



Figure S6. TEM image of tadpole-like SCNPs (CD_{27%}-P4VP(SCNP)₅₃-*b*-PS₅₆₉) prepared in dilute DCM solution.



Self-assembly behavior of linear BCPs and tadpole-like SCNPs in solution.

Figure S7. Cubosomes obtained by linear P4VP-*b*-PS using DMF as co-solvent. (a, b) SEM image and SAXS pattern of P4VP₅₃-*b*-PS₈₂₀ (f_{PS} =94.4%) ; (c,d) TEM image and SAXS pattern of P4VP₄₀-*b*-PS₇₃₁ (f_{PS} =95.2%).



Figure S8. (a) SAXS profile of cubosomes formed by $CD_{10\%}$ -P4VP(SCNP)₄₀-*b*-PS₅₄₄. SEM images of (b) irregular cubosomes formed by $CD_{16\%}$ -P4VP(SCNP)₄₀-*b*-PS₅₄₄ and (c) LCMs formed by $CD_{28\%}$ -P4VP(SCNP)₄₀-*b*-PS₅₄₄ using DMF as co-solvent.



Figure S9. (a) SAXS profile of cubosomes formed by $CD_{39\%}$ -P4VP(SCNP)₅₃-*b*-PS₄₉₉ and (b) TEM image of LCMs with irregular pores and elongated vesicles formed by $CD_{45\%}$ -P4VP(SCNP)₅₃-*b*-PS₄₉₉ using DMF as co-solvent.



Figure S10. (a, b) SEM images of LCVs formed by linear P4VP₅₃-*b*-PS₅₆₉; (c, e) SEM images and (d, f) SAXS patterns of cubosomes formed by $CD_{18\%}$ -P4VP(SCNP)₅₃-*b*-PS₅₆₉ and $CD_{27\%}$ -P4VP(SCNP)₅₃-*b*-PS₅₆₉, respectively; (g) SEM image and (h) TEM image of cubosomes with uneven surface formed by $CD_{40\%}$ -P4VP(SCNP)₅₃-*b*-PS₅₆₉. All the self-assembles were obtained by using DMF as co-solvent.



Figure S11. (a) TEM image and (b) SAXS pattern of sponges formed by linear $P4VP_{40}-b-PS_{512}$; (c, e) SEM images (inserts were TEM and SEM image at magnification) and (d, f) SAXS patterns of cubosomes formed by $CD_{11\%}$ -P4VP(SCNP)₄₀-b-PS₅₁₂ and $CD_{17\%}$ -P4VP(SCNP)₄₀-b-PS₅₁₂, respectively; (g) SEM images and (h) SAXS pattern of LCMs formed by $CD_{29\%}$ -P4VP(SCNP)₄₀-b-PS₅₁₂. All the self-assembles were obtained by using DMF as co-solvent.



Figure S12. SEM images of (a) sponges obtained by linear P4VP₅₃-*b*-PS₇₅₀, cubosomes obtained by (b) CD_{14%}-P4VP(SCNP)₅₃-*b*-PS₇₅₀ and (c) CD_{19%}-P4VP(SCNP)₅₃-*b*-PS₇₅₀, and (d) LCMs obtained by CD_{26%}-P4VP(SCNP)₅₃-*b*-PS₇₅₀ using DMF as co-solvent.



Figure S13. SEM images of cubosomes formed by (a) CD_{18%}-P4VP(SCNP)₅₃-b-PS₈₂₀ and (b) CD_{25%}-

P4VP(SCNP)₅₃-*b*-PS₈₂₀, and (c) LCMs formed by CD_{45%}-P4VP(SCNP)₅₃-*b*-PS₈₂₀ using DMF as co-solvent.



Figure S14. SEM images of (a) micelles formed by linear P4VP₆₇-*b*-PS₃₆₇, (b) sponges formed by CD_{5%}-P4VP(SCNP)₆₇-*b*-PS₃₆₇, and (c) nanowires formed by CD_{12%}-P4VP(SCNP)₆₇-*b*-PS₃₆₇ using DMF as co-solvent. SEM images of (d) sponges formed by linear P4VP₆₇-*b*-PS₉₀₁ and (e, f) nanowires formed by CD_{12%}-P4VP(SCNP)₆₇-*b*-PS₉₀₁ and CD_{40%}-P4VP(SCNP)₆₇-*b*-PS₉₀₁ using DMF as co-solvent.



Figure S15. SEM and TEM images of (a, c) sponges formed by CD_{6%}-P4VP(SCNP)₄₀-*b*-PS₅₄₄ and (b, d) LCMs with uneven surface formed by CD_{13%}-P4VP(SCNP)₄₀-*b*-PS₅₄₄ using DMF as co-solvent. The

interchain cross-linking nanopartices were prepared in the dilute dioxane solution.



Figure S16. GPC traces of P4VP(SCNP)₅₃-*b*-PS₅₆₉ prepared in dilute DCM solution when CD was 27% and 40%, using (a) DMF and (b) THF as the eluent, respectively.



Figure S17. SEM images of (a) LCVs formed by linear $P4VP_{40}$ -*b*- PS_{375} , (b) sponges formed by $CD_{12\%}$ -P4VP(SCNP)₄₀-*b*- PS_{375} , and LCMs with uneven surface formed by (c) $CD_{25\%}$ -P4VP(SCNP)₄₀-*b*- PS_{375} and (d) $CD_{31\%}$ -P4VP(SCNP)₄₀-*b*- PS_{375} using DMF as co-solvent.



Figure S18. SEM and TEM images of LCVs formed by (a, b) linear P4VP₄₀-*b*-PS₅₄₄ and (c, d) CD_{16%}-P4VP(SCNP)₄₀-*b*-PS₅₄₄ using THF as co-solvent.

Reference:

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