

Supporting Information

Electroselective C(sp³)-H Deuteration of Isoindolinones

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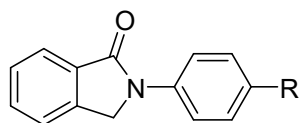
I. General Information

All reactions were performed under air atmosphere, using round bottom flasks. All substrates were obtained from the commercial sources or synthesized following literature procedures. All reagents were commercial and were used without further purification. The electrochemical reaction device follows our previous work.¹ The instrument for electrolysis is Single Output DC Power Supply (KRP-305DM) (made in China). Chromatography was carried on flash silica gel (300-400 mesh). All reactions were monitored by TLC, which was performed on percolated aluminum sheets of silica gel 60 (F254). Melting points were uncorrected. The ¹H and ¹³C NMR data were obtained on a 300 MHz NMR spectrometer with TMS as the internal standard and CDCl₃ or DMSO - *d*₆ as solvent. Multiplicities are indicated as it follows: s, singlet; d, doublet; t, triplet; q, quartet; m, multiplet; dd, doubled doublet; br, broad. Coupling constants (*J* values) where noted are quoted in Hertz. Highresolution mass spectra (HRMS) were obtained with a time-of-flight (TOF) mass spectrometer (ESI).

Electrode materials/dimensions:

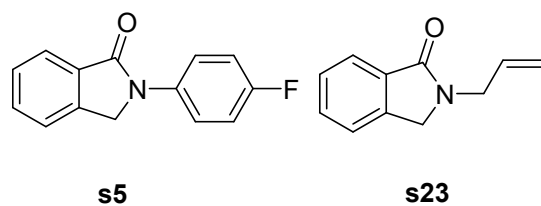
The graphite electrodes, aluminium electrodes and zinc electrodes are purchased from Tianjin Zhongnuotansu Technology Co., Ltd. The dimensions of the electrodes are 5 mm × 50 mm (the submerged height of the electrode is approximately 5 mm).

List of commercially available substrates:



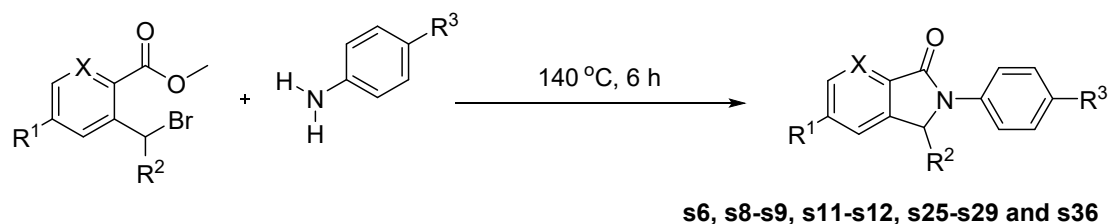
- 1**, R = H
- s3**, R = OMe
- s4**, R = Me
- s7**, R = Br
- s10**, R = CO₂Et

List of available substrates from our group's previous work²:



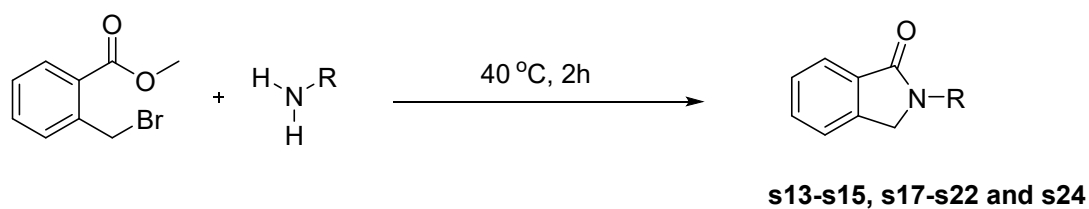
II. Substrates Synthesis and Characterization

Method A : Synthesis of compounds **s6**, **s8-s9**, **s11-s12**, **s25-s29** and **s36**.



A mixture of corresponding substituted methyl 2-(bromomethyl)benzoate (5 mmol, 1.0 eq.) and substituted amine (6 mmol, 1.2 eq.) in a 100 mL round bottom flask was stirred at 140 °C for 6 hours. The HBr tail gas absorption device was connected to this system throughout. After the completion of the reaction, the mixture was cooled to room temperature. The solution was diluted with DCM (30 mL) and quenched with 1N HCl aqueous or water (30 mL). The aqueous layer was extracted with DCM (3 × 30 mL). The combined DCM layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc as eluent afforded the desired product **s6**, **s8-s9**, **s11-s12**, **s25-s29** and **s36**.

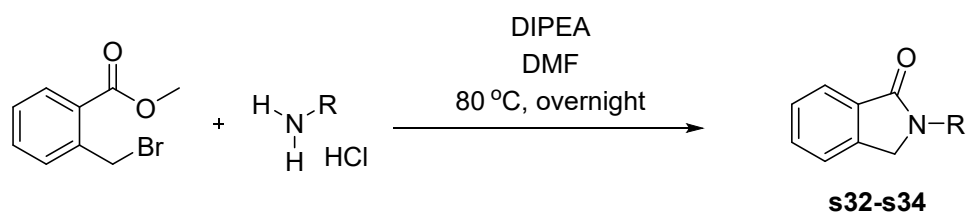
Method B : Synthesis of compounds **s13-s15**, **s17-s22** and **s24**.



A mixture of methyl 2-(bromomethyl)benzoate (5 mmol, 1.0 eq.) and substituted amine (6 mmol, 1.2 eq.) in a 100 mL round bottom flask was stirred at 40 °C for 2 hours. The HBr tail gas absorption device was connected to this system throughout. After the completion of the reaction, the mixture was cooled to room temperature. The solution was diluted with DCM (30 mL) and quenched with 1N HCl aqueous (30 mL). The

aqueous layer was extracted with DCM (3 × 30 mL). The combined DCM layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc as eluent afforded the desired product **s13-s15**, **s17-s22** and **s24**.

Method C : Synthesis of compounds **s32-s34**.



A mixture of corresponding substituted methyl 2-(bromomethyl)benzoate (5 mmol, 1.0 eq.), substituted amine hydrochloride (6 mmol, 1.2 eq.), DIPEA (15 mmol, 3 eq.) and DMF (15 mL) in a 100 mL round bottom flask was stirred at 80 °C for overnight. After the completion of the reaction, the mixture was cooled to room temperature. The solution was diluted with EtOAc (30 mL) and quenched with 1N HCl aqueous (30 mL). The aqueous layer was extracted with EtOAc (3 × 30 mL). The combined EtOAc layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc as eluent afforded the desired product **s32-s34**.

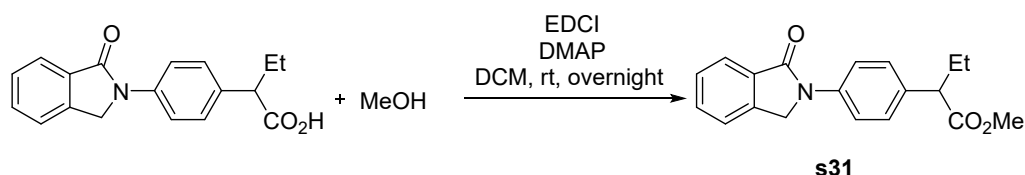
Method D : Synthesis of compounds **s30**.



To a solution of 2-benzoyl-N-phenylbenzamide (3.0 mmol, 1.0 eq.) in CH₃CN (5 mL) were added Et₃SiH (6.0 mmol, 2.0 eq.) and Al(OTf)₃ (1.5 mmol, 0.5 eq.) at room temperature. The reaction mixture was heated at reflux for 4 hours and then concentrated *in vacuo*. The solution was diluted with DCM (30 mL) and quenched with 1N HCl aqueous (30 mL). The aqueous layer was extracted with DCM (3 × 30 mL). The combined DCM layers were washed with brine (3 × 30 mL), dried over Na₂SO₄,

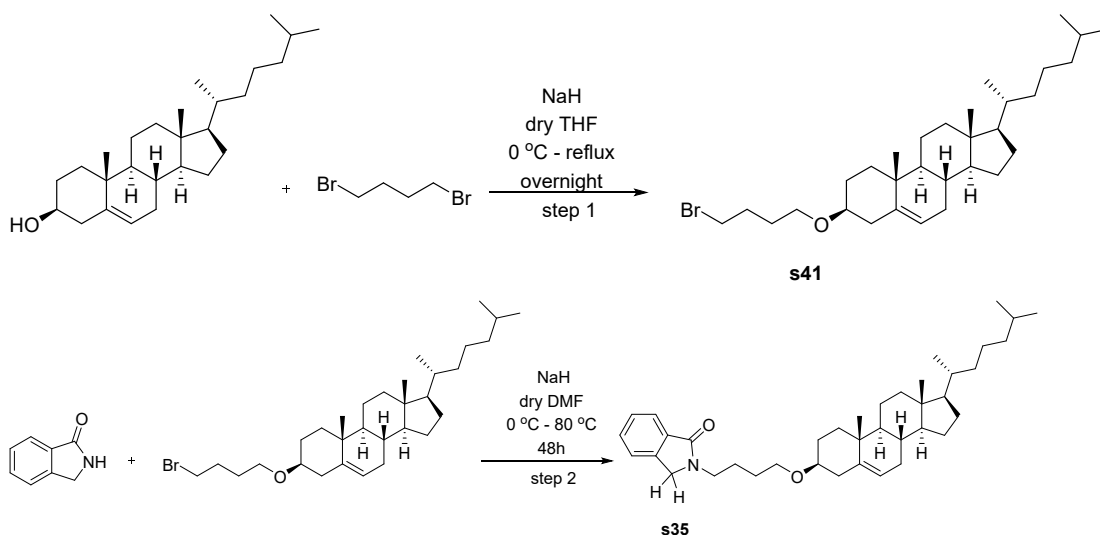
and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 3/1) as eluent afforded the desired product **s30**.

Method E : Synthesis of compounds **s31**.



At room temperature, indobufen (2 mmol, 1.0 eq.) was dissolved in DCM (20 mL) in a 50 mL round bottom flask, then EDCI (3 mmol, 1.5 eq.) and DMAP (2.4 mmol, 1.2 eq.) were added, and then the MeOH (4 mmol, 2 eq.) was added dropwise to the above solution, and the obtained solution was stirred at room temperature for overnight. The solution was diluted with DCM (30 mL) and quenched with 1N HCl aqueous (30 mL). The aqueous layer was extracted with DCM (3 × 30 mL). The combined DCM layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 10/1) as eluent afforded the desired product **s31**.

Method F : Synthesis of compounds **s35**.

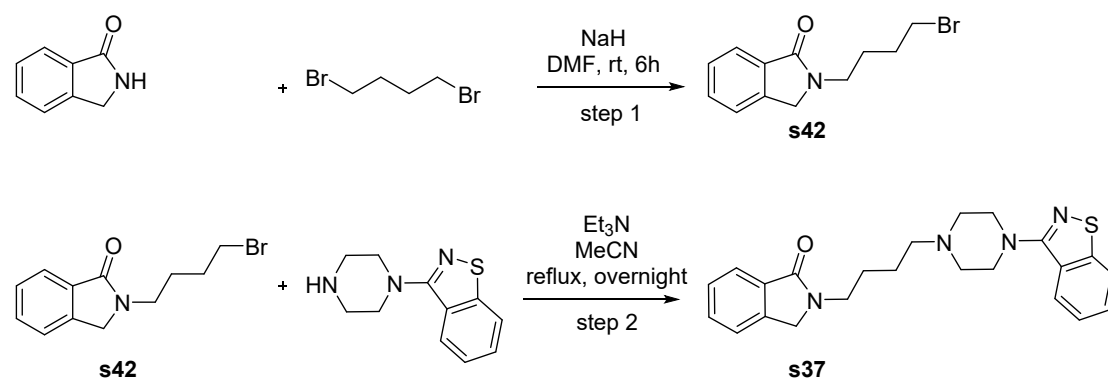


Step 1: At room temperature, cholesterol (10 mmol, 1.0 eq.) was dissolved in THF (10 mL) in a 50 mL round bottom flask, then at 0 °C NaH (20 mmol, 2 eq.) were added, then the 1,4-dibromobutane (50 mmol, 5 eq.) was added dropwise to the above solution,

and the obtained solution heated at reflux for overnight. After the completion of the reaction, the mixture was cooled to room temperature and added ice water into solution, then concentrated *in vacuo*. The solution was diluted with DCM (30 mL) and quenched with water (30 mL). The aqueous layer was extracted with DCM (3 × 30 mL). The combined DCM layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 10/1) as eluent afforded the desired product **s41**.

Step 2: At room temperature, isoindolin-1-one (2 mmol, 1.0 eq.) was dissolved in DMF (10 mL) in a 50 mL round bottom flask, then at 0 °C NaH (4 mmol, 2.0 eq.) were added, and then the **s41** (2.4 mmol, 1.2 eq.) was added dropwise to the above solution, and the obtained solution heated at 80 °C for 48 hours. After the completion of the reaction, the mixture was cooled to room temperature and added ice water into solution. The solution was diluted with Et₂O (30 mL) and quenched with water (30 mL). The aqueous layer was extracted with Et₂O (3 × 30 mL). The combined Et₂O layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 3/1) as eluent afforded the desired product **s35**.

Method G : Synthesis of compounds **s37**.

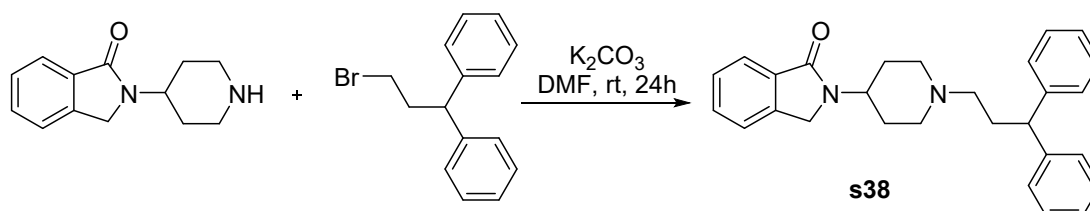


Step 1: At room temperature, isoindolin-1-one (10 mmol, 1.0 eq.) was dissolved in DMF (10 mL) in a 50 mL round bottom flask, then at 0 °C NaH (12 mmol, 1.2 eq.) were added, and then the 1,4-dibromobutane (50 mmol, 5 eq.) was added dropwise to the above solution, and the obtained solution was stirred at room temperature for 6

hours. After the completion of the reaction, the mixture was cooled to room temperature and added ice water into solution. The solution was diluted with EtOAc (30 mL) and quenched with water (30 mL). The aqueous layer was extracted with EtOAc (3 × 30 mL). The combined EtOAc layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc = (v/v = 3/1) as eluent afforded the desired product **s42**.

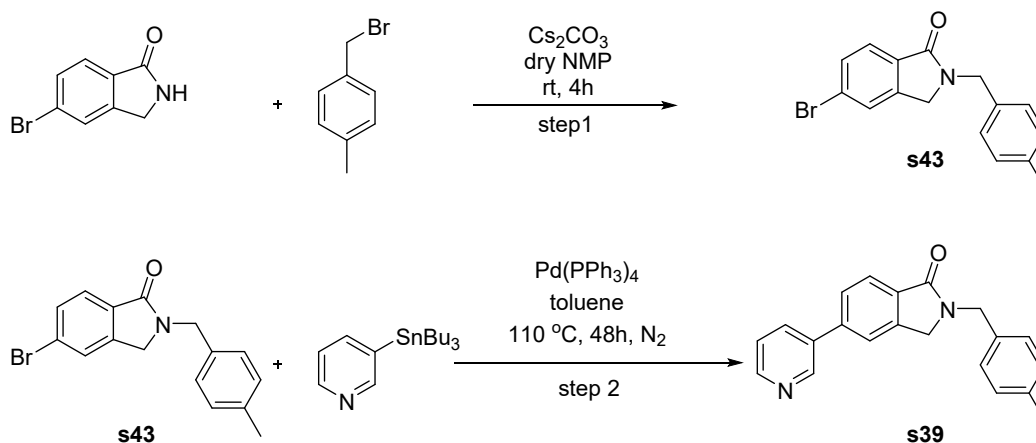
Step 2: A mixture of **s42** (5 mmol, 1.0 eq.), 3-(piperazin-1-yl)benzo[d]isothiazole (5.5 mmol, 1.1 eq.), Et₃N (6 mmol, 1.2 eq.) and MeCN (10 mL) in a 100 mL round bottom flask was heated at reflux for overnight. After the completion of the reaction, the mixture was cooled to room temperature and then concentrated *in vacuo*. The solution was diluted with DCM (30 mL) and quenched with 1N HCl aqueous (30 mL). The aqueous layer was extracted with DCM (3 × 30 mL). The combined DCM layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 1/3) as eluent afforded the desired product **s37**.

Method H : Synthesis of compounds **s38**.



A mixture of (3-bromopropyl)diphenylmethane (2 mmol, 1.0 eq.), 2-(piperidin-4-yl)isoindolin-1-one (2.2 mmol, 1.1 eq.), K₂CO₃ (4 mmol, 2.0 eq.) and DMF (10 mL) in a 100 mL round bottom flask was at room temperature for 24 hours. After the completion of the reaction, the solution was diluted with EtOAc (30 mL) and quenched with 1N HCl aqueous (30 mL). The aqueous layer was extracted with EtOAc (3 × 30 mL). The combined EtOAc layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 1/1) as eluent afforded the desired product **s38**.

Method I : Synthesis of compounds **s39**.

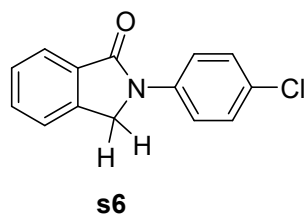


Step 1: A mixture of 5-bromoisoindolin-1-one (10 mmol, 1.0 eq.), 1-(bromomethyl)-4-methylbenzene (15 mmol, 1.5 eq.), Cs₂CO₃ (16 mmol, 2 eq.) and NMP (10 mL) in a 100 mL round bottom flask was at 60 °C for 4 hours. After the completion of the reaction, the solution was diluted with EtOAc (30 mL) and quenched with 1N HCl aqueous (30 mL). The aqueous layer was extracted with EtOAc (3 × 30 mL). The combined EtOAc layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 10/1) as eluent afforded the desired product **s43**.

Step 2: At room temperature, a mixture of **s43** (5 mmol, 1.0 eq.) and 3-(tributylstannyl)pyridine (7.5 mmol, 1.5 eq.) was dissolved in toluene (10 mL) in a 100 mL three-compartment cell, the reaction system was purged with N₂ three times, followed by the addition of Pd(PPh₃)₄ (1 mmol, 0.2 eq.). The mixture was refluxed at 110 °C for 48 h. The solution was diluted with EtOAc (30 mL) and quenched with water (30 mL). The aqueous layer was extracted with EtOAc (3 × 30 mL). The combined EtOAc layers were washed with brine (3 × 30 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 10/1) as eluent afforded the desired product **s39**.

2-(4-Chlorophenyl)isoindolin-1-one (s6)

The ^1H spectra data matched with values reported in the literature.³

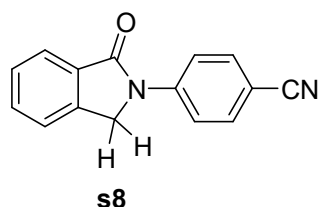


White solid (900 mg, 75% yield); m.p. 184 – 186 °C;

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.91 (d, $J = 8.3$ Hz, 1H), 7.86 – 7.79 (m, 2H), 7.65 – 7.56 (m, 1H), 7.54 – 7.47 (m, 2H), 7.41 – 7.32 (m, 2H), 4.82 (s, 2H).

4-(1-Oxoisoindolin-2-yl)benzonitrile (s8)

The ^1H spectra data matched with values reported in the literature.⁴

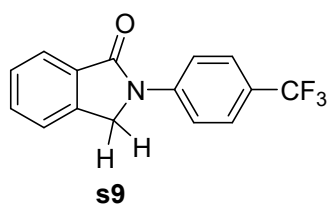


White solid (780 mg, 67% yield); m.p. 204 – 206 °C;

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 8.09 – 8.03 (m, 2H), 7.96 – 7.92 (m, 1H), 7.74 – 7.69 (m, 2H), 7.69 – 7.62 (m, 1H), 7.58 – 7.51 (m, 2H), 4.89 (s, 2H).

2-(4-(Trifluoromethyl)phenyl)isoindolin-1-one (s9)

The ^1H spectra data matched with values reported in the literature.⁴

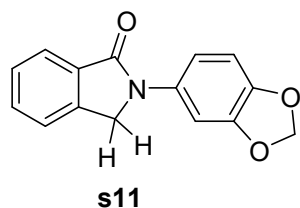


White solid (970 mg, 70% yield); m.p. 231 – 232 °C;

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 8.04 (d, $J = 8.4$ Hz, 2H), 7.94 (d, $J = 7.3$ Hz, 1H), 7.72 – 7.66 (m, 2H), 7.65 – 7.60 (m, 1H), 7.58 – 7.49 (m, 2H), 4.90 (s, 2H).

2-(Benzo[*d*][1,3]dioxol-5-yl)isoindolin-1-one (s11)

The ^1H spectra data matched with values reported in the literature.⁵

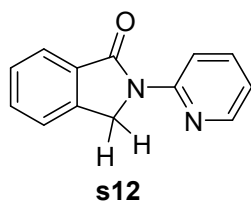


White solid (830 mg, 66% yield); m.p. 167 – 169 °C;

^1H NMR (300 MHz, CDCl_3): δ 7.94 – 7.88 (m, 1H), 7.63 – 7.55 (m, 2H), 7.53 – 7.45 (m, 2H), 7.10 (dd, $J = 8.4, 2.3$ Hz, 1H), 6.85 (dd, $J = 8.4, 0.4$ Hz, 1H), 5.99 (s, 2H), 4.80 (s, 2H).

2-(Pyridin-2-yl)isoindolin-1-one (s12)

The ^1H spectra data matched with values reported in the literature.⁴

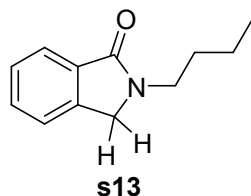


White solid (300 mg, 29% yield); m.p. 176 – 177 °C;

^1H NMR (300 MHz, CDCl_3): δ 8.67 (dt, $J = 8.5, 0.9$ Hz, 1H), 8.40 (ddd, $J = 5.0, 2.0, 0.9$ Hz, 1H), 7.96 – 7.90 (m, 1H), 7.81 – 7.72 (m, 1H), 7.62 (ddd, $J = 7.6, 7.1, 1.2$ Hz, 1H), 7.58 – 7.45 (m, 2H), 7.10 – 7.01 (m, 1H), 5.11 (s, 2H).

2-Butylisoindolin-1-one (s13)

The ^1H spectra data matched with values reported in the literature.²

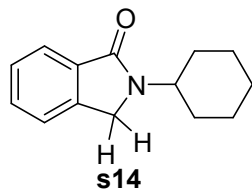


Colourless liquid (800 mg, 85% yield);

^1H NMR (300 MHz, CDCl_3): δ 7.86 – 7.78 (m, 1H), 7.54 – 7.47 (m, 1H), 7.46 – 7.40 (m, 2H), 4.36 (s, 2H), 3.64 (dt, $J = 19.2, 7.4$ Hz, 2H), 1.73 – 1.48 (m, 2H), 1.46 – 1.29 (m, 2H), 1.07 – 0.82 (m, 3H).

2-Cyclohexylisoindolin-1-one (s14)

The ^1H spectra data matched with values reported in the literature.³

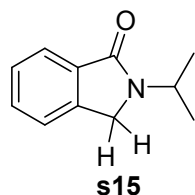


Colourless liquid (750 mg, 70% yield);

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.89 – 7.74 (m, 1H), 7.55 – 7.48 (m, 1H), 7.48 – 7.41 (m, 2H), 4.35 (s, 2H), 4.30 – 4.17 (m, 1H), 1.87 (s, 4H), 1.73 (d, $J = 13.1$ Hz, 1H), 1.57 – 1.38 (m, 4H), 1.27 – 1.11 (m, 1H).

2-Isopropylisoindolin-1-one (s15)

The ^1H spectra data matched with values reported in the literature.³

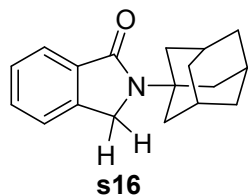


Colourless liquid (825 mg, 94% yield);

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.84 (dd, $J = 7.8, 1.5$ Hz, 1H), 7.55 – 7.49 (m, 1H), 7.44 (ddd, $J = 6.7, 4.0, 1.4$ Hz, 2H), 4.77 – 4.50 (m, 1H), 4.34 (s, 2H), 1.29 (d, $J = 6.8$ Hz, 6H).

2-(Adamantan-1-yl)isoindolin-1-one (s16)

The ^1H spectra data matched with values reported in the literature.⁶

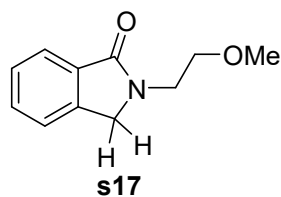


White solid (1.1 g, 82% yield); m.p. 202 – 204 °C;

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.81 – 7.75 (m, 1H), 7.49 (td, $J = 7.3, 1.4$ Hz, 1H), 7.45 – 7.37 (m, 2H), 4.46 (s, 2H), 2.31 (d, $J = 3.0$ Hz, 6H), 2.16 (s, 3H), 1.74 (dd, $J = 7.5, 4.2$ Hz, 6H).

2-(2-Methoxyethyl)isoindolin-1-one (s17)

The ^1H spectra data matched with values reported in the literature.²

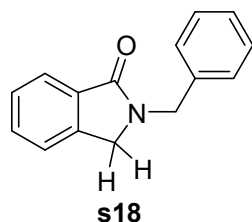


Colourless liquid (700 mg, 73% yield);

^1H NMR (300 MHz, CDCl_3): δ 7.85 – 7.77 (m, 1H), 7.54 – 7.46 (m, 1H), 7.44 – 7.36 (m, 2H), 4.49 (s, 2H), 3.80 – 3.72 (m, 2H), 3.63 – 3.57 (m, 2H), 3.32 (s, 3H).

2-Benzylisoindolin-1-one (s18)

The ^1H spectra data matched with values reported in the literature.²

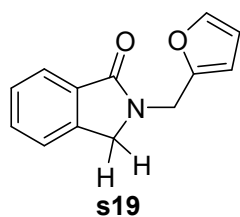


White solid (900 mg, 81% yield); m.p. 88 – 89 °C;

^1H NMR (300 MHz, CDCl_3): δ 7.95 – 7.85 (m, 1H), 7.56 – 7.42 (m, 2H), 7.38 (ddt, J = 7.3, 1.5, 0.8 Hz, 2H), 7.35 – 7.27 (m, 4H), 4.81 (s, 2H), 4.27 (s, 2H).

2-(Furan-2-ylmethyl)isoindolin-1-one (s19)

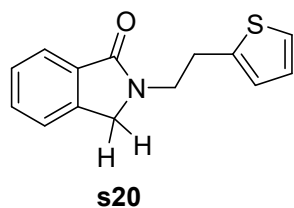
The ^1H spectra data matched with values reported in the literature.⁷



Yellow liquid (500 mg, 47% yield);

^1H NMR (300 MHz, CDCl_3): δ 7.84 (dt, J = 7.3, 1.1 Hz, 1H), 7.50 (td, J = 7.3, 1.7 Hz, 1H), 7.45 – 7.36 (m, 2H), 7.34 (dd, J = 1.7, 1.1 Hz, 1H), 6.34 – 6.25 (m, 2H), 4.77 (s, 2H), 4.35 (s, 2H).

2-(2-(Thiophen-2-yl)ethyl)isoindolin-1-one (s20)



White solid (970 mg, 80% yield); m.p. 102 – 104 °C;

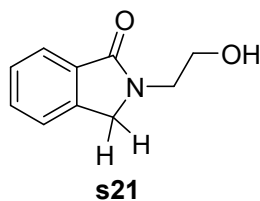
¹H NMR (300 MHz, CDCl₃): δ 7.89 – 7.78 (m, 1H), 7.54 – 7.48 (m, 1H), 7.46 (ddd, *J* = 7.3, 1.4, 0.6 Hz, 1H), 7.41 – 7.36 (m, 1H), 7.14 (dd, *J* = 5.1, 1.2 Hz, 1H), 6.91 (dd, *J* = 5.1, 3.5 Hz, 1H), 6.86 (dt, *J* = 3.5, 1.0 Hz, 1H), 4.25 (s, 2H), 3.90 (t, *J* = 7.0 Hz, 2H), 3.23 (t, *J* = 7.0, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 168.6, 141.3, 132.9, 131.3, 128.1, 127.2, 125.5, 124.1, 123.7, 122.7, 50.8, 44.4, 29.1.

HRMS (ESI): Calcd. for C₁₄H₁₄NOS⁺ [M + H]⁺: 244.0791, found: 244.0785.

2-(2-Hydroxyethyl)isoindolin-1-one (s21)

The ¹H spectra data matched with values reported in the literature.²

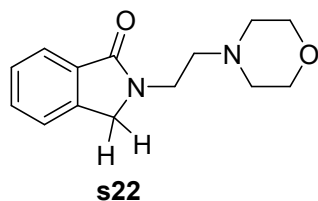


White solid (800 mg, 90% yield); m.p. 113 – 115 °C;

¹H NMR (300 MHz, CDCl₃): δ 7.82 (dt, *J* = 7.5, 1.1 Hz, 1H), 7.57 – 7.49 (m, 1H), 7.48 – 7.38 (m, 2H), 4.51 (s, 2H), 3.92 (dd, *J* = 5.6, 4.3 Hz, 2H), 3.77 (dd, *J* = 5.6, 4.3 Hz, 2H).

2-(2-Morpholinoethyl)isoindolin-1-one (s22)

The ¹H spectra data matched with values reported in the literature.⁸

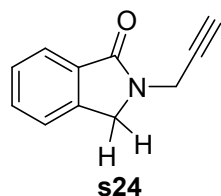


Colourless liquid (740 mg, 60% yield);

¹H NMR (300 MHz, CDCl₃): δ 7.89 – 7.82 (m, 1H), 7.58 – 7.50 (m, 1H), 7.49 – 7.41 (m, 2H), 4.50 (s, 2H), 3.75 (t, *J* = 6.4 Hz, 2H), 3.71 – 3.61 (m, 4H), 2.64 (t, *J* = 6.4 Hz, 2H), 2.57 – 2.43 (m, 4H).

2-(Prop-2-yn-1-yl)isoindolin-1-one (s24)

The ¹H spectra data matched with values reported in the literature.²

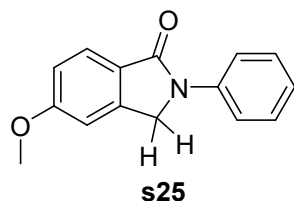


Colourless liquid (700 mg, 82% yield);

¹H NMR (300 MHz, CDCl₃): δ 7.90 – 7.81 (m, 1H), 7.60 – 7.51 (m, 1H), 7.49 – 7.43 (m, 2H), 4.50 (s, 2H), 4.45 (d, *J* = 2.6 Hz, 2H), 2.28 (t, *J* = 2.6 Hz, 1H).

5-Methoxy-2-phenylisoindolin-1-one (s25)

The ¹H spectra data matched with values reported in the literature.²

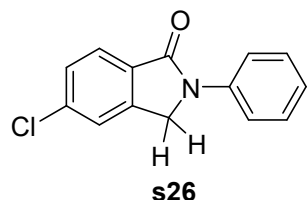


White solid (600 mg, 50% yield); m.p. 142 – 144 °C;

¹H NMR (300 MHz, CDCl₃): δ 7.88 – 7.75 (m, 3H), 7.48 – 7.34 (m, 2H), 7.18 – 7.08 (m, 1H), 7.04 – 6.90 (m, 2H), 4.74 (s, 2H), 3.87 (s, 3H).

5-Chloro-2-phenylisoindolin-1-one (s26)

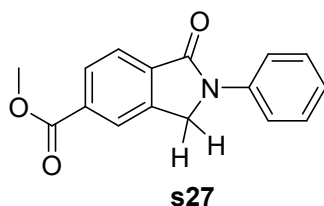
The ¹H spectra data matched with values reported in the literature.⁷



Yellow solid (400 mg, 30% yield); m.p. 155 – 156 °C;

¹H NMR (300 MHz, CDCl₃): δ 7.90 – 7.81 (m, 3H), 7.55 – 7.47 (m, 2H), 7.44 (dd, *J* = 8.7, 7.4 Hz, 2H), 7.21 (d, *J* = 7.4 Hz, 1H), 4.85 (s, 2H).

Methyl 1-oxo-2-phenylisoindoline-5-carboxylate (s27)



White solid (670 mg, 50% yield); m.p. 142 – 144 °C;

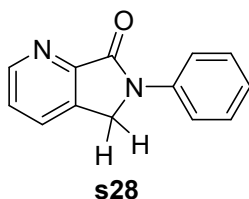
¹H NMR (300 MHz, CDCl₃): δ 8.27 – 8.15 (m, 2H), 8.02 – 7.95 (m, 1H), 7.90 – 7.82 (m, 2H), 7.45 (t, *J* = 7.9 Hz, 2H), 7.21 (t, *J* = 7.4 Hz, 1H), 4.91 (s, 2H), 3.98 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 166.5, 165.5, 140.1, 139.3, 137.2, 133.7, 130.0, 129.4, 125.1, 124.3, 124.2, 119.8, 52.7, 50.8.

HRMS (ESI): Calcd. for C₁₆H₁₄NO₃⁺ [M + H]⁺: 268.0968, found: 268.0971.

6-Phenyl-5,6-dihydro-7H-pyrrolo[3,4-*b*]pyridin-7-one (s28)

The ¹H spectra data matched with values reported in the literature.⁹

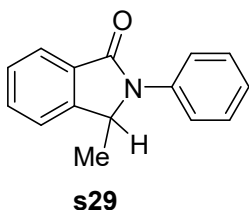


Yellow solid (400 mg, 64% yield); m.p. 192 – 194 °C;

¹H NMR (300 MHz, CDCl₃): δ 8.84 (d, *J* = 4.8 Hz, 1H), 7.92 – 7.77 (m, 3H), 7.60 – 7.38 (m, 3H), 7.25 – 7.18 (m, 1H), 4.89 (s, 2H).

3-Methyl-2-phenylisoindolin-1-one (s29)

The ¹H spectra data matched with values reported in the literature.³

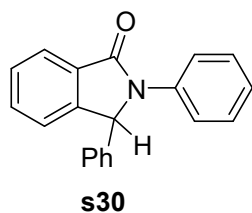


Colourless liquid (1.03 g, 92% yield);

¹H NMR (300 MHz, CDCl₃): δ 7.99 – 7.85 (m, 1H), 7.67 – 7.55 (m, 3H), 7.55 – 7.41 (m, 4H), 7.28 – 7.19 (m, 1H), 5.21 (q, *J* = 6.7 Hz, 1H), 1.46 (d, *J* = 6.7, 3H).

2,3-Diphenylisoindolin-1-one (s30)

The ^1H spectra data matched with values reported in the literature.¹⁰

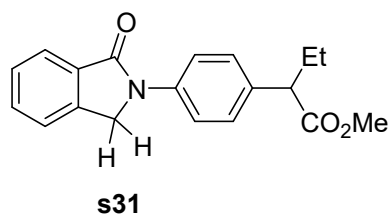


White solid (600 mg, 70% yield); m.p. 198 – 199 °C;

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 8.01 – 7.95 (m, 1H), 7.64 – 7.58 (m, 2H), 7.56 – 7.47 (m, 2H), 7.35 – 7.18 (m, 8H), 7.13 – 7.04 (m, 1H), 6.10 (s, 1H).

Methyl 2-(4-(1-oxoisoindolin-2-yl)phenyl)butanoate (s31)

The ^1H spectra data matched with values reported in the literature.¹¹

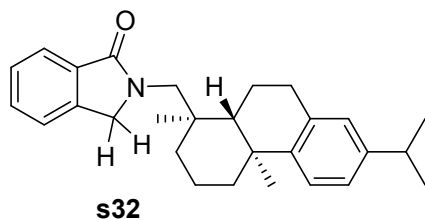


Colourless liquid (560 mg, 90% yield);

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.96 – 7.89 (m, 1H), 7.83 (d, $J = 8.6$ Hz, 2H), 7.63 – 7.56 (m, 1H), 7.52 (d, $J = 7.2$ Hz, 2H), 7.37 (d, $J = 8.6$ Hz, 2H), 4.85 (s, 2H), 3.67 (s, 3H), 3.47 (t, $J = 14.0$ Hz, 1H), 2.19 – 2.00 (m, 1H), 1.81 (dt, $J = 14.0, 7.4$ Hz, 1H), 0.91 (t, $J = 7.4$ Hz, 3H).

2-(((1*R*,4*aS*,10*aR*)-7-Isopropyl-1,4*a*-dimethyl-1,2,3,4,4*a*,9,10,10*a*-octahydrophenanthren-1-yl)methyl)isoindolin-1-one (s32)

The ^1H spectra data matched with values reported in the literature.¹²

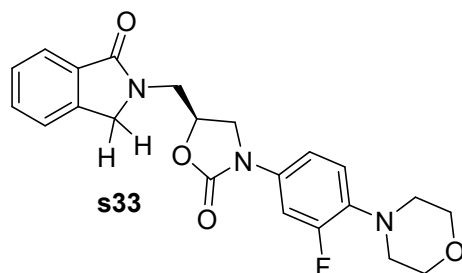


Yellow liquid (1.2 g, 60% yield);

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.85 (dt, $J = 7.4, 1.1$ Hz, 1H), 7.52 (td, $J = 7.4, 1.1$ Hz, 1H), 7.48 – 7.38 (m, 2H), 7.19 (d, $J = 8.1$ Hz, 1H), 7.01 (dd, $J = 8.1, 2.1$ Hz, 1H), 6.94 (d, $J = 2.1$ Hz, 1H), 4.51 (s, 2H), 3.50 (s, 2H), 3.06 – 2.94 (m, 2H), 2.85 (dt, $J = 13.8,$

6.4 Hz, 1H), 2.31 (dt, $J = 12.9, 3.4$ Hz, 1H), 2.17 (ddt, $J = 12.9, 6.4, 2.2$ Hz, 1H), 1.97 – 1.67 (m, 3H), 1.61 (q, $J = 2.7$ Hz, 2H), 1.56 (d, $J = 2.2$ Hz, 1H), 1.39 (td, $J = 12.7, 4.1$ Hz, 1H), 1.27 (d, $J = 2.5$ Hz, 6H), 1.24 (s, 3H), 1.09 (s, 3H).

(S)-3-(3-Fluoro-4-morpholinophenyl)-5-((1-oxoisindolin-2-yl)methyl)oxazolidin-2-one (s33)



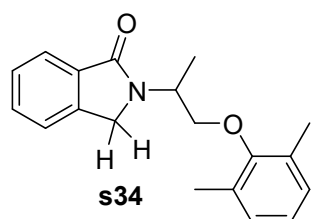
Yellow solid (800 mg, 40% yield); m.p. 233 – 236 °C;

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.88 – 7.80 (m, 1H), 7.61 – 7.53 (m, 1H), 7.50 – 7.42 (m, 2H), 7.39 (d, $J = 2.6$ Hz, 1H), 7.05 (dd, $J = 2.6, 1.1$ Hz, 1H), 6.92 (d, $J = 9.1$ Hz, 1H), 5.01 – 4.85 (m, 1H), 4.63 (d, $J = 1.8$ Hz, 2H), 4.10 (d, $J = 9.1$ Hz, 1H), 4.06 – 4.02 (m, 1H), 3.98 – 3.93 (m, 1H), 3.92 – 3.89 (m, 1H), 3.88 – 3.83 (m, 4H), 3.07 – 2.96 (m, 4H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 169.8, 157.2, 154.2, 153.9, 141.9, 136.6, 133.1, 132.1, 131.6, 128.3, 123.9, 123.0, 118.9, 114.1, 107.9, 107.5, 72.7, 67.0, 52.5, 51.0, 47.9, 45.6.

HRMS (ESI): Calcd. for $\text{C}_{22}\text{H}_{23}\text{FN}_3\text{O}_4^+$ $[\text{M} + \text{H}]^+$: 412.1667, found: 412.1677.

2-(1-(2,6-Dimethylphenoxy)propan-2-yl)isindolin-1-one (s34)



Colourless liquid (800 mg, 54% yield);

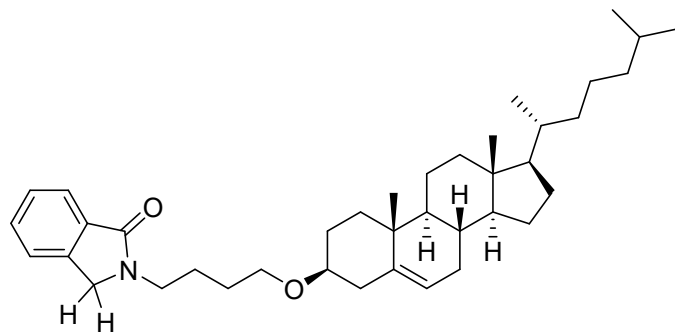
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.88 (m, 1H), 7.60 – 7.51 (m, 1H), 7.47 (m, 2H), 6.97 (m, 2H), 6.92 (s, 1H), 4.84 (tdd, $J = 7.1, 4.7, 2.4$ Hz, 1H), 4.72 (d, $J = 17.1$ Hz, 1H), 4.54 (d, $J = 17.1$ Hz, 1H), 4.03 – 3.89 (m, 2H), 2.26 – 2.15 (m, 6H), 1.56 (dd, $J = 7.1, 0.8$ Hz, 3H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 168.6, 155.3, 141.7, 133.0, 131.3, 130.8, 129.0, 128.0,

124.1, 123.7, 122.8, 74.0, 47.9, 47.5, 16.3, 15.5.

HRMS (ESI): Calcd. for $C_{19}H_{22}NO_2^+$ $[M + H]^+$: 296.1645, found: 296.1658.

2-((((3*S*,8*S*,9*S*,10*R*,13*R*,14*S*,17*R*)-10,13-Dimethyl-17-((*R*)-6-methylheptan-2-yl)-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[*a*]phenanthren-3-yl)oxy)methyl)isoindolin-1-one (s35)



s35

Colourless liquid (800 mg, 54% yield);

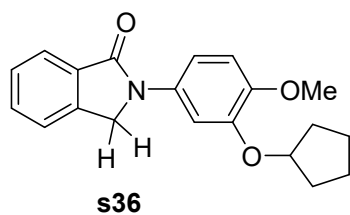
1H NMR (300 MHz, $CDCl_3$): δ 7.90 – 7.73 (m, 1H), 7.59 – 7.50 (m, 1H), 7.48 – 7.38 (m, 2H), 5.33 (s, 1H), 4.45 – 4.32 (m, 2H), 3.70 – 3.60 (m, 2H), 3.57 – 3.44 (m, 2H), 3.14 (m, 1H), 2.33 (m, 1H), 2.18 (d, $J = 12.1$ Hz, 1H), 2.08 – 1.94 (m, 2H), 1.91 – 1.62 (m, 5H), 1.62 – 1.06 (m, 21H), 1.00 (td, $J = 5.4, 4.9, 2.8$ Hz, 4H), 0.95 – 0.89 (m, 4H), 0.86 (m, 6H), 0.71 – 0.65 (m, 3H).

^{13}C NMR (75 MHz, $CDCl_3$): δ 168.6, 141.2, 141.1, 133.2, 131.2, 128.1, 123.7, 122.7, 121.6, 79.1, 67.4, 56.9, 56.3, 50.3, 50.1, 42.4, 42.2, 39.9, 39.6, 39.3, 37.3, 37.0, 36.3, 35.9, 32.0, 28.6, 28.3, 28.1, 27.5, 25.4, 24.4, 23.9, 22.9, 22.7, 21.2, 19.5, 18.8, 12.0.

HRMS (ESI): Calcd. for $C_{39}H_{60}NO_2^+$ $[M + H]^+$: 574.4619, found: 574.4612.

2-(3-(Cyclopentyloxy)-4-methoxyphenyl)isoindolin-1-one (s36)

The 1H spectra data matched with values reported in the literature.³



s36

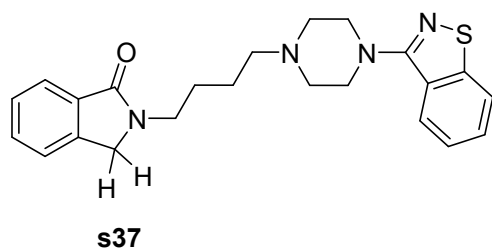
White solid (950 mg, 60% yield); m.p. 137 – 139 °C;

1H NMR (300 MHz, $CDCl_3$): δ 7.97 – 7.84 (m, 2H), 7.59 (ddd, $J = 7.8, 7.1, 1.2$ Hz,

1H), 7.54 – 7.44 (m, 2H), 7.06 – 6.98 (m, 1H), 6.94 – 6.84 (m, 1H), 4.92 – 4.85 (m, 1H), 4.82 (s, 2H), 3.86 (s, 3H), δ 2.04 (m, 2H), 1.98 – 1.88 (m, 2H), 1.85 (m, 2H), 1.61 (m, 2H).

2-(4-(4-(Benzo[d]isothiazol-3-yl)piperazin-1-yl)butyl)isoindolin-1-one (s37)

The ^1H spectra data matched with values reported in the literature.¹³

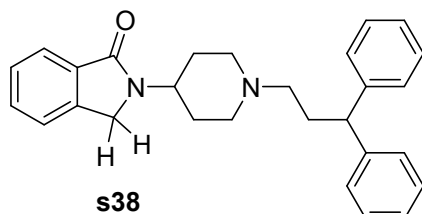


White solid (1.4 g, 69% yield); m.p. 233 – 234 °C;

^1H NMR (300 MHz, CDCl_3): δ 7.88 (d, $J = 8.1$ Hz, 1H), 7.85 – 7.81 (m, 1H), 7.78 (d, $J = 8.1$ Hz, 1H), 7.55 – 7.48 (m, 1H), 7.48 – 7.41 (m, 3H), 7.37 – 7.29 (m, 1H), 4.38 (s, 2H), 3.66 (t, $J = 7.0$ Hz, 2H), 3.54 (t, $J = 4.9$ Hz, 4H), 2.65 (t, $J = 4.9$ Hz, 4H), 2.47 (t, $J = 7.0$ Hz, 2H), 1.80 – 1.67 (m, 2H), 1.61 (m, 2H).

2-(1-(3,3-Diphenylpropyl)piperidin-4-yl)isoindolin-1-one (s38)

The ^1H spectra data matched with values reported in the literature.¹⁴

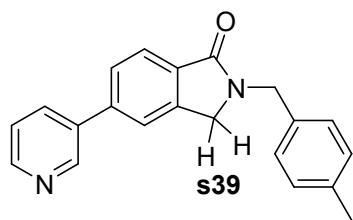


Colourless liquid (400 mg, 50% yield);

^1H NMR (300 MHz, CDCl_3): δ 7.84 (dd, $J = 8.2, 1.1$ Hz, 1H), 7.56 – 7.49 (m, 1H), 7.48 – 7.41 (m, 2H), 7.32 – 7.22 (m, 8H), 7.21 – 7.14 (m, 2H), 4.36 (s, 2H), 4.34 – 4.15 (m, 1H), 3.98 (t, $J = 7.0$ Hz, 1H), 3.00 (d, $J = 7.0$ Hz, 2H), 2.29 (m, 4H), 2.16 – 2.00 (m, 2H), 1.83 (p, $J = 4.0$ Hz, 4H).

2-(4-Methylbenzyl)-5-(pyridin-3-yl)isoindolin-1-one (s39)

The ^1H spectra data matched with values reported in the literature.¹⁵

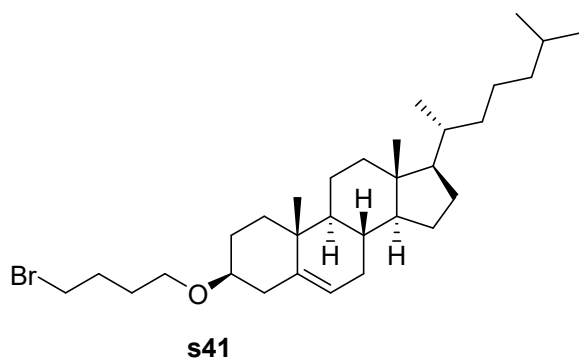


Yellow solid (800 mg, 40% yield); m.p. 145 – 147 °C;

¹H NMR (300 MHz, CDCl₃): δ 8.87 – 8.82 (m, 1H), 8.63 (dd, *J* = 4.9, 1.7 Hz, 1H), 7.99 (dd, *J* = 7.9, 0.8 Hz, 1H), 7.88 (ddd, *J* = 7.9, 2.4, 1.7 Hz, 1H), 7.67 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.56 (dd, *J* = 1.6, 0.8 Hz, 1H), 7.43 – 7.36 (m, 1H), 7.22 (d, *J* = 8.1 Hz, 2H), 7.18 – 7.13 (m, 2H), 4.79 (s, 2H), 4.33 (s, 2H), 2.33 (s, 3H).

(3*S*,8*S*,9*S*,10*R*,13*R*,14*S*,17*R*)-3-(4-Bromobutoxy)-10,13-dimethyl-17-((*R*)-6-methylheptan-2-yl)-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[*a*]phenanthrene (s41)

The ¹H spectra data matched with values reported in the literature.¹⁶

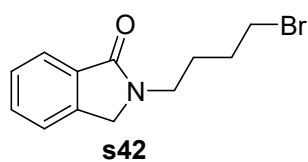


Colourless liquid (1.3 g, 25% yield);

¹H NMR (300 MHz, CDCl₃): δ 5.34 (d, *J* = 5.1 Hz, 1H), 3.56 – 3.38 (m, 4H), 3.12 (tt, *J* = 11.3, 4.4 Hz, 1H), 2.40 – 2.27 (m, 1H), 2.24 – 2.11 (m, 1H), 2.04 – 1.79 (m, 6H), 1.70 (dd, *J* = 8.8, 6.0 Hz, 2H), 1.54 – 0.82 (m, 34H), 0.68 (s, 3H).

2-(4-Bromobutyl)isoindolin-1-one (s42)

The ¹H spectra data matched with values reported in the literature.¹⁷



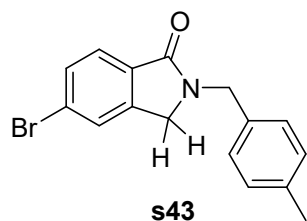
Colourless liquid (2.1 g, 80% yield);

¹H NMR (300 MHz, CDCl₃): δ 7.80 – 7.72 (m, 1H), 7.50 – 7.43 (m, 1H), 7.42 – 7.34

(m, 2H), 4.32 (s, 2H), 3.59 (t, $J = 6.6$ Hz, 2H), 3.45 – 3.34 (m, 3H), 2.02 – 1.91 (m, 1H), 1.89 – 1.69 (m, 4H).

5-Bromo-2-(4-methylbenzyl)isoindolin-1-one (s43)

The ^1H spectra data matched with values reported in the literature.¹⁵

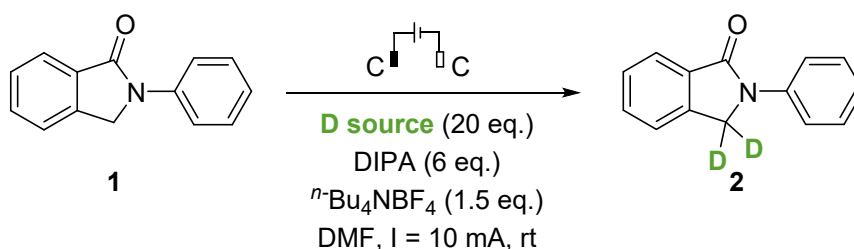


Yellow solid (1.4 g, 70% yield); m.p. 95 – 96 °C;

^1H NMR (300 MHz, CDCl_3): δ 7.71 (d, $J = 7.9$ Hz, 1H), 7.62 – 7.46 (m, 2H), 7.15 (m, 4H), 4.72 (s, 2H), 4.20 (s, 2H), 2.32 (s, 3H).

III. Investigation of Reaction Conditions

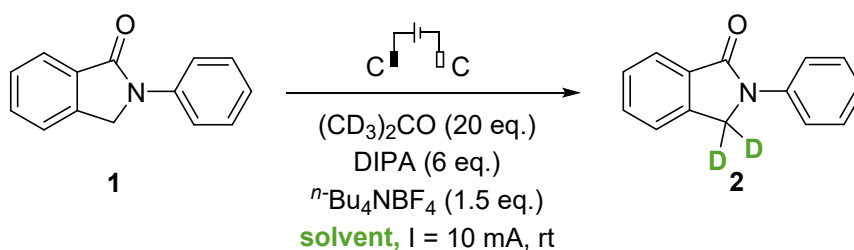
Table S1. Investigation of deuterium source^a:



| entry | variation of deuterium source | D % of 2 ^b |
|-------|------------------------------------|------------------------------|
| 1 | (CD ₃) ₂ SO | 30 |
| 2 | CD ₃ OD | 60 |
| 3 | EtOD | 70 |
| 4 | D ₂ O | 75 |
| 5 | CD ₃ CN | 77 |
| 6 | (CD ₃) ₂ CO | 82 (64%) ^c |

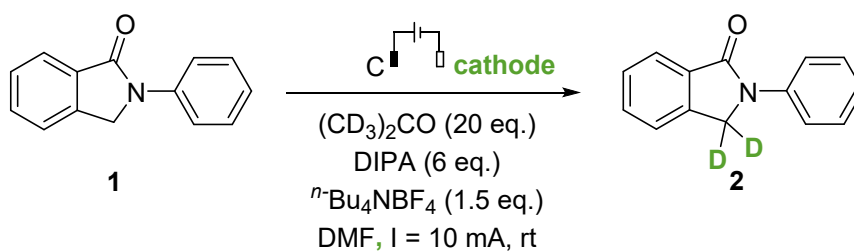
^aReaction conditions: graphite anode (d = 5 mm), graphite cathode (d = 5 mm), **1** (0.2 mmol), deuterium source (20 eq.), DIPA (6 eq.), $n\text{-Bu}_4\text{NBF}_4$ (1.5 eq.), DMF (3.0 mL), undivided cell, constant current = 10 mA, room temperature, 3 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

Table S2. Investigation of solvent^a:



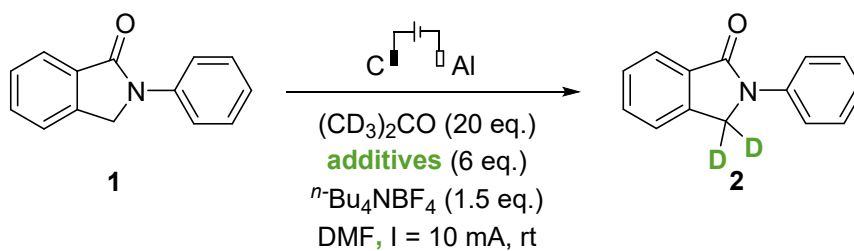
| entry | variation of solvent (3 mL) | D % of 2 ^b |
|-------|-----------------------------|------------------------------|
| 1 | MeCN | 10 |
| 2 | DCM | 15 |
| 3 | NMP | 70 |
| 4 | THF | 75 |
| 5 | DMA | 80 |

^aReaction conditions: graphite anode (d = 5 mm), graphite cathode (d = 5 mm), **1** (0.2 mmol), (CD₃)₂CO (20 eq.), DIPA (6 eq.), $n\text{-Bu}_4\text{NBF}_4$ (1.5 eq.), solvent (3.0 mL), undivided cell, constant current = 10 mA, room temperature, 3 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

Table S3. Investigation of cathode^a:

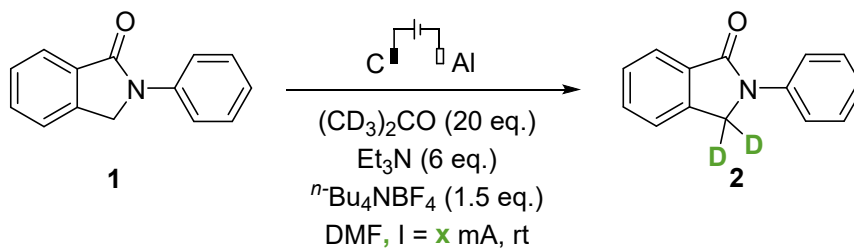
| entry | variation of cathode | D % of 2 ^b |
|-------|----------------------|------------------------------|
| 1 | Pt | 80 |
| 2 | Ni | 75 |
| 3 | Mo | 80 |
| 4 | steel | 80 |
| 5 | Zn | 85 |
| 6 | Mg | 85 |
| 7 | Al | 90 (66%) ^c |

^aReaction conditions: graphite anode ($d = 5 \text{ mm}$), cathode ($d = 5 \text{ mm}$), **1** (0.2 mmol), $(\text{CD}_3)_2\text{CO}$ (20 eq.), DIPA (6 eq.), $n\text{-Bu}_4\text{NBF}_4$ (1.5 eq.), DMF (3.0 mL), undivided cell, constant current = 10 mA, room temperature, 3 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

Table S4. Investigation of additives^a:

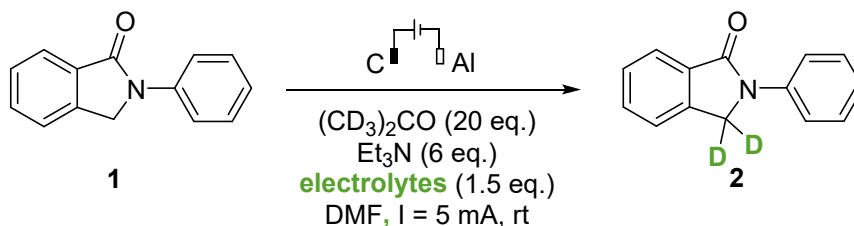
| entry | variation of additives | D % of 2 ^b |
|-------|------------------------|------------------------------|
| 1 | PivOH | 0 |
| 2 | DIPEA | 95 (36%) ^c |
| 3 | Et_3N | 96 (36%) ^c |
| 4 | PMP | 95 (30%) ^c |

^aReaction conditions: graphite anode ($d = 5 \text{ mm}$), aluminium cathode ($d = 5 \text{ mm}$), **1** (0.2 mmol), $(\text{CD}_3)_2\text{CO}$ (20 eq.), additives (6 eq.), $n\text{-Bu}_4\text{NBF}_4$ (1.5 eq.), DMF (3.0 mL), undivided cell, constant current = 10 mA, room temperature, 1.5 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

Table S5. Investigation of current^a:

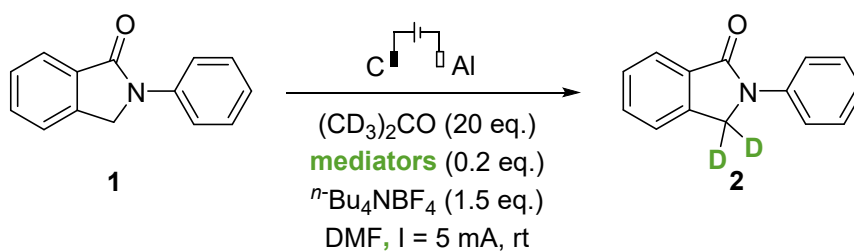
| entry | variation of current | D % of 2 ^b |
|-------|----------------------|------------------------------|
| 1 | 3 | 90 |
| 2 | 5 | 96 (60%) ^c |
| 3 | 20 | 96 (20%) ^c |

^aReaction conditions: graphite anode (d = 5 mm), aluminium cathode (d = 5 mm), **1** (0.2 mmol), $(\text{CD}_3)_2\text{CO}$ (20 eq.), Et_3N (6 eq.), $n\text{-Bu}_4\text{NBF}_4$ (1.5 eq.), DMF (3.0 mL), undivided cell, constant current = x mA, room temperature, 1.5 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

Table S6. Investigation of electrolytes^a:

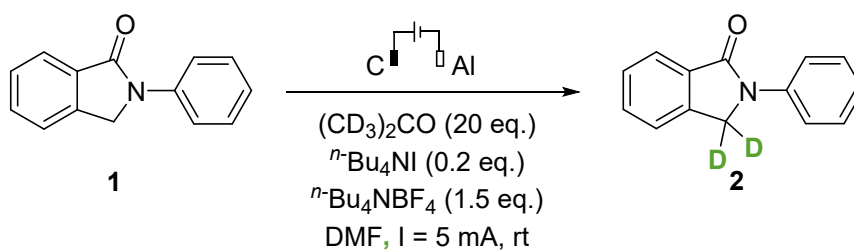
| entry | variation of electrolytes | D % of 2 ^b |
|-------|------------------------------|------------------------------|
| 1 | $n\text{-Bu}_4\text{NPF}_6$ | 95 (55%) ^c |
| 2 | $n\text{-Bu}_4\text{NClO}_4$ | 95 (47%) ^c |
| 3 | LiClO_4 | 0 |
| 4 | NaSbF_6 | 0 |
| 5 | $n\text{-Bu}_4\text{NBr}$ | 96 (53%) ^c |
| 6 | $n\text{-Bu}_4\text{NI}$ | 96 (55%) ^c |

^aReaction conditions: graphite anode (d = 5 mm), aluminium cathode (d = 5 mm), **1** (0.2 mmol), $(\text{CD}_3)_2\text{CO}$ (20 eq.), Et_3N (6 eq.), electrolytes (1.5 eq.), DMF (3.0 mL), undivided cell, constant current = 5 mA, room temperature, 1.5 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

Table S7. Investigation of mediators^a:

| entry | variation of mediators | D % of 2 ^b |
|-------|----------------------------------|------------------------------|
| 1 | KI | 90 |
| 2 | NHPI | 95 (47%) ^c |
| 3 | TEMPO | 94 (66%) ^c |
| 4 | ⁿ -Bu ₄ NI | 96 (81%) ^c |

^aReaction conditions: graphite anode (d = 5 mm), aluminium cathode (d = 5 mm), **1** (0.2 mmol), (CD₃)₂CO (20 eq.), mediators (0.2 eq.), ⁿ-Bu₄NBF₄ (1.5 eq.), DMF (3.0 mL), undivided cell, constant current = 5 mA, room temperature, 1.5 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

Table S8. Investigation of other conditions^a:

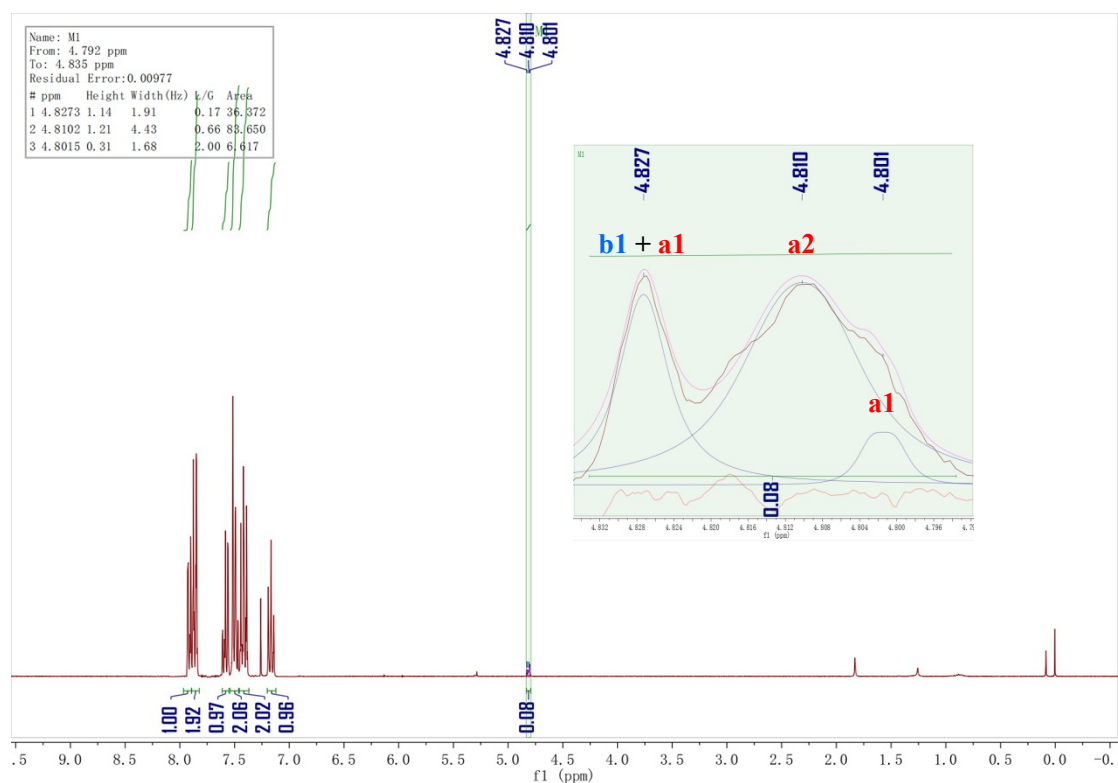
| entry | variation from standard conditions | D % of 2 ^b |
|-------|---|------------------------------|
| 1 | none | 96 (81%) ^c |
| 2 | no ⁿ -Bu ₄ NI | 95 (50%) ^c |
| 3 | no ⁿ -Bu ₄ NBF ₄ | 95 (62%) ^c |
| 4 | D ₂ O instead of (CD ₃) ₂ CO | 90 (71%) ^c |
| 5 | DMSO- <i>d</i> ₆ instead of (CD ₃) ₂ CO | 27 (60%) ^c |
| 6 | DCM instead of DMF | 92 (69%) ^c |
| 7 | EtOH instead of DMF | 0 |
| 8 | graphite instead of aluminium | 93 (71%) ^c |
| 9 | 10 mA, 45min instead of 5 mA, 1.5h | 95 (57%) ^c |
| 10 | no electric current | 0 |

^aReaction conditions: graphite anode (d = 5 mm), aluminium cathode (d = 5 mm), **1** (0.2 mmol), (CD₃)₂CO (20 eq.), ⁿ-Bu₄NI (0.2 eq.), ⁿ-Bu₄NBF₄ (1.5 eq.), DMF (3.0 mL), undivided cell, constant

current = 5 mA, room temperature, 1.5 h. ^bDegree of deuteration. ^cIsolated yield in parentheses.

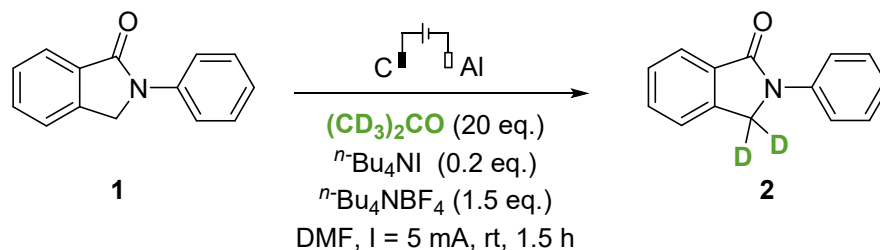
Scheme S1: Peak fitting function analyzing the ratio of CD₂/CDH/CH₂.

Peak fitting function in Mestrenova software was adopted to analyze the residue ¹H signal around 4.81 ppm. It was found that the residue benzylic hydrogen in product **2a** exists in the major form of Ar-CDH (triplet). Another existence is the Ar-CH₂ (single). By the analysis of the peak area of Ar-CDH (consisting 2*a1 + a2) and Ar-CH₂ (consisting b1), it was deduced the ratio to be 6.5:1. Take the overall peak area 0.08 (0 for Ar-CD₂, and 2.0 for Ar-CH₂) in account, the ratio of Ar-CD₂/Ar-CDH/Ar-CH₂ was 93:6:1.



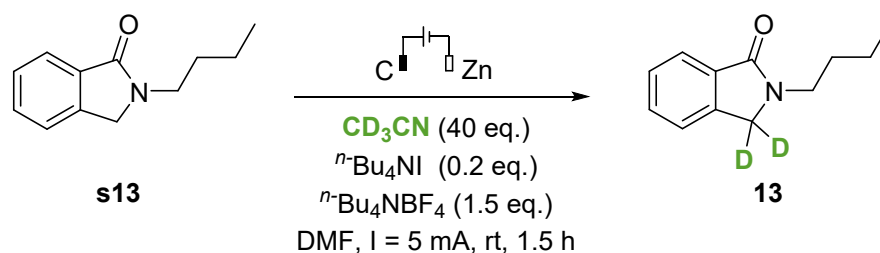
IV. Experimental Procedures and Compound Characterization

General procedure for electrochemical deuteration of isolindolines (Procedure A):



A 10 mL distillation flask equipped with a magnetic stir bar was charged with compound **1** (42 mg, 0.2 mmol, 1.0 eq.), $(\text{CD}_3)_2\text{CO}$ (0.3 mL, 20 eq.), $n\text{-Bu}_4\text{NI}$ (15 mg, 0.04 mmol), $n\text{-Bu}_4\text{NBF}_4$ (99 mg, 0.3 mmol, 1.5 eq.) and DMF (3.0 mL). The flask equipped with graphite rod anode ($d = 5$ mm) and aluminium rod cathode ($d = 5$ mm). The resulting solution was stirred and electrolyzed at a constant current of 5 mA (Single Output DC Power Supply: KRP-305DM) for 1.5 h at room temperature. The solution was diluted with EtOAc (5 mL) and brine (20 mL), and extracted with EtOAc (3×20 mL). The combined organic layers were washed with brine (3×20 mL), dried over Na_2SO_4 , and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc ($v/v = 10/1$) as eluent afforded the desired product **2**

(Procedure B):

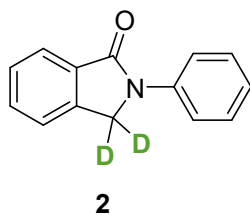


A 10 mL distillation flask equipped with a magnetic stir bar was charged with compound **s13** (38 mg, 0.2 mmol, 1.0 eq.), CD_3CN (0.43 mL, 40 eq.), $n\text{-Bu}_4\text{NI}$ (15 mg, 0.04 mmol), $n\text{-Bu}_4\text{NBF}_4$ (99 mg, 0.3 mmol, 1.5 eq.) and DMF (3.0 mL). The flask equipped with graphite rod anode ($d = 5$ mm) and zinc rod cathode ($d = 5$ mm). The resulting solution was stirred and electrolyzed at a constant current of 5 mA (Single Output DC Power Supply: KRP-305DM) for 1.5 h at room temperature. The solution

was diluted with EtOAc (5 mL) and brine (20 mL), and extracted with EtOAc (3 × 20 mL). The combined organic layers were washed with brine (3 × 20 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 10/1) as eluent afforded the desired product **13**.

2-Phenylisoindolin-1-one-3,3-*d*₂ (**2**)

Following Procedure A.



White solid (34 mg, 81% yield, 96% D); m.p. 161 – 163 °C;

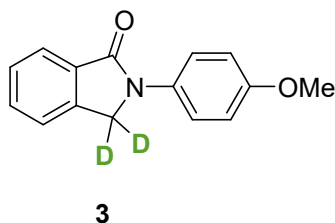
¹H NMR (300 MHz, CDCl₃): δ 7.95 – 7.89 (m, 1H), 7.89 – 7.82 (m, 2H), 7.63 – 7.55 (m, 1H), 7.54 – 7.47 (m, 2H), 7.46 – 7.38 (m, 2H), 7.22 – 7.13 (m, 1H), 4.85 – 4.82 (s, 0.08H).

¹³C NMR (75 MHz, CDCl₃): δ 167.4, 139.9, 139.4, 133.1, 131.9, 129.0, 128.2, 124.2, 123.9, 122.5, 119.2, 50.5 – 49.6 (m).

HRMS (ESI): Calcd. for C₁₄H₁₀D₂NO⁺ [M + H]⁺: 212.1039, found: 212.1037.

2-(4-Meoxyphenyl)isoindolin-1-one-3,3-*d*₂ (**3**)

Following Procedure A, using 2.0 h instead of 1.5 h.



White solid (35 mg, 73% yield, 95% D); m.p. 147 – 149 °C;

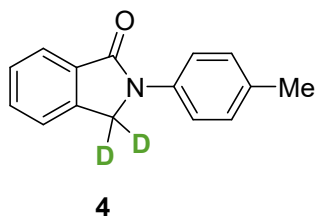
¹H NMR (300 MHz, CDCl₃): δ 7.94 – 7.84 (m, 1H), 7.78 – 7.66 (m, 2H), 7.62 – 7.49 (m, 1H), 7.54 – 7.42 (m, H), 6.99 – 6.87 (m, 2H), 4.75 (s, 0.10H), 3.80 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 167.0, 156.3, 139.9, 133.1, 132.5, 131.6, 128.0, 123.6, 122.4, 121.0, 114.0, 55.2, 51.2 – 49.9 (m).

HRMS (ESI): Calcd. for C₁₅H₁₂D₂NO₂⁺ [M + H]⁺: 242.1145, found: 242.1156.

2-(*p*-Tolyl)isoindolin-1-one-3,3-*d*₂ (4)

Following Procedure A.



White solid (36 mg, 80% yield, 95% D); m.p. 128 – 130 °C;

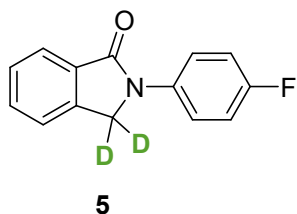
¹H NMR (300 MHz, CDCl₃): δ 7.96 – 7.88 (m, 1H), 7.77 – 7.70 (m, 2H), 7.59 (ddd, *J* = 7.7, 7.0, 1.3 Hz, 1H), 7.54 – 7.46 (m, 2H), 7.26 – 7.19 (m, 2H), 4.81 (s, 0.10H), 2.35 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 167.2, 139.9, 136.8, 133.8, 133.2, 131.7, 129.4, 128.1, 123.7, 122.4, 119.2, 50.6 – 50.0 (m), 20.7.

HRMS (ESI): Calcd. for C₁₅H₁₂D₂NO⁺ [M + H]⁺: 226.1195, found: 226.1190.

2-(4-Fluorophenyl)isoindolin-1-one-3,3-*d*₂ (5)

Following Procedure A.



White solid (38 mg, 82% yield, 96% D); m.p. 172 – 174 °C;

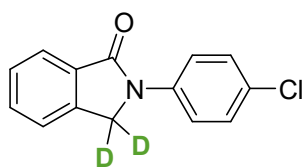
¹H NMR (300 MHz, CDCl₃): δ 7.95 – 7.87 (m, 1H), 7.86 – 7.75 (m, 2H), 7.63 – 7.55 (m, 1H), 7.54 – 7.42 (m, 2H), 7.15 – 6.95 (m, 2H), 4.80 (s, 0.08H).

¹³C NMR (75 MHz, CDCl₃): δ 167.3, 161.0, 157.7, 139.8, 135.5, 135.5, 132.9, 132.0, 128.3, 123.9, 122.6, 121.0, 115.8, 115.5, 50.8 – 50.2 (m).

HRMS (ESI): Calcd. for C₁₄H₉D₂FNO⁺ [M + H]⁺: 230.0945, found: 230.0942.

2-(4-Chlorophenyl)isoindolin-1-one-3,3-*d*₂ (6)

Following Procedure A.



6

White solid (37 mg, 75% yield, 95% D); m.p. 184 – 186 °C;

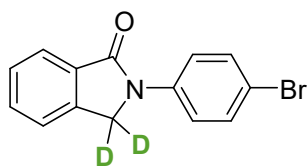
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.91 – 7.85 (m, 1H), 7.84 – 7.76 (m, 2H), 7.64 – 7.55 (m, 1H), 7.53 – 7.44 (m, 2H), 7.37 – 7.30 (m, 2H), 4.78 (s, 0.10H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 167.4, 139.7, 138.0, 132.8, 132.2, 129.3, 129.0, 128.4, 124.0, 122.6, 120.2, 50.5 – 49.4 (m).

HRMS (ESI): Calcd. for $\text{C}_{14}\text{H}_9\text{D}_2\text{ClNO}^+ [\text{M} + \text{H}]^+$: 246.0649, found: 246.0650.

2-(4-Bromophenyl)isoindolin-1-one-3,3- d_2 (7)

Following Procedure A, using $n\text{-Bu}_4\text{NI}$ (3.0 eq.) instead of $n\text{-Bu}_4\text{NI}$ (0.2 eq.), using 1.0 h instead of 1.5 h.



7

White solid (41 mg, 70% yield, 95% D); m.p. 188 – 189 °C;

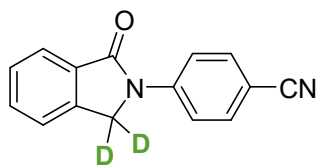
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.94 – 7.86 (m, 1H), 7.81 – 7.73 (m, 2H), 7.60 (m, 1H), 7.54 – 7.46 (m, 4H), 4.79 (s, 0.10H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 167.4, 139.7, 138.5, 132.8, 132.2, 132.0, 128.4, 124.0, 122.6, 120.4, 117.0, 50.5 – 49.8 (m).

HRMS (ESI): Calcd. for $\text{C}_{14}\text{H}_9\text{D}_2\text{BrNO}^+ [\text{M} + \text{H}]^+$: 290.0144, found: 290.0133.

4-(1-Oxoisindolin-2-yl-3,3- d_2)benzonitrile (8)

Following Procedure A.



8

White solid (33 mg, 70% yield, 95% D); m.p. 207 – 209 °C;

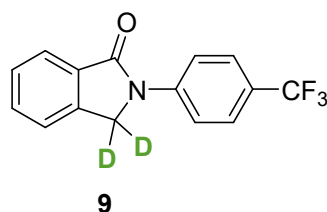
¹H NMR (300 MHz, CDCl₃): δ 8.06 – 7.97 (m, 2H), 7.93 – 7.85 (m, 1H), 7.70 – 7.58 (m, 3H), 7.56 – 7.47 (m, 2H), 4.83 (s, 0.10H).

¹³C NMR (75 MHz, DMSO-*d*₆): δ 167.3, 143.3, 140.9, 133.2, 132.9, 131.8, 128.3, 123.5, 123.4, 118.9, 118.7, 105.5, 50.2 – 48.6 (m).

HRMS (ESI): Calcd. for C₁₅H₉D₂N₂O⁺ [M + H]⁺: 237.0991, found: 237.0983.

2-(4-(Trifluoromethyl)phenyl)isoindolin-1-one-3,3-*d*₂ (9)

Following Procedure A, using 3.5 h instead of 1.5 h.



White solid (34 mg, 61% yield, 91% D); m.p. 233 – 235 °C;

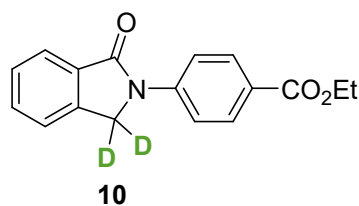
¹H NMR (300 MHz, CDCl₃): δ 8.03 (d, *J* = 8.5 Hz, 2H), 7.92 (d, *J* = 7.5 Hz, 1H), 7.73 – 7.59 (m, 3H), 7.54 (d, *J* = 7.5 Hz, 2H), 4.88 (s, 0.18H).

¹³C NMR (75 MHz, (CD₃)₂CO): δ 174.3, 145.7, 141.0, 133.5, 129.2, 129.0, 127.2 – 127.0 (m), 126.9, 124.5, 124.2 – 124.0 (m), 122.7, 120.0, 60.6 – 60.4 (m).

HRMS (ESI): Calcd. for C₁₅H₉D₂F₃NO⁺ [M + H]⁺: 280.0913, found: 280.0918.

Ethyl 4-(1-oxoisoindolin-2-yl)benzoate-3,3-*d*₂ (10)

Following Procedure A.



White solid (40 mg, 71% yield, 95% D); m.p. 180 – 181 °C;

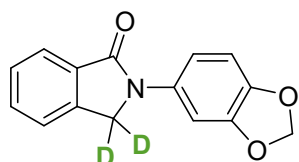
¹H NMR (300 MHz, CDCl₃): δ 8.09 – 8.01 (m, 2H), 7.98 – 7.91 (m, 2H), 7.90 – 7.85 (m, 1H), 7.63 – 7.54 (m, 1H), 7.52 – 7.44 (m, 2H), 4.81 (s, 0.10H), 4.36 (t, *J* = 7.1 Hz, 2H), 1.39 (t, *J* = 7.1 Hz, 3H).

^{13}C NMR (75 MHz, CDCl_3): δ 167.7, 166.0, 143.3, 139.8, 132.7, 132.5, 131.4, 130.6, 128.4, 125.6, 124.1, 122.6, 117.7, 113.6, 60.7, 58.2 – 57.51 (m), 14.3.

HRMS (ESI): Calcd. for $\text{C}_{17}\text{H}_{14}\text{D}_2\text{NO}_3^+ [\text{M} + \text{H}]^+$: 284.1250, found: 284.1244.

2-(Benzo[d][1,3]dioxol-5-yl)isoindolin-1-one-3,3- d_2 (11)

Following Procedure A.



11

White solid (26 mg, 50% yield, 92% D); m.p. 167 – 169 °C;

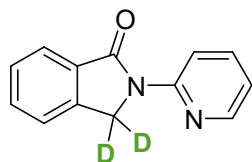
^1H NMR (300 MHz, CDCl_3): δ 7.94 – 7.79 (m, 1H), 7.63 – 7.54 (m, 2H), 7.52 – 7.46 (m, 2H), 7.08 (dd, $J = 8.4, 2.3$ Hz, 1H), 6.83 (dd, $J = 8.4, 0.4$ Hz, 1H), 5.97 (s, 2H), 4.78 (s, 0.10H).

^{13}C NMR (75 MHz, CDCl_3): δ 167.2, 147.9, 144.3, 139.8, 133.8, 133.1, 131.8, 128.2, 123.8, 122.5, 112.5, 108.0, 102.3, 101.3, 51.3 – 50.6 (m).

HRMS (ESI): Calcd. for $\text{C}_{15}\text{H}_{10}\text{D}_2\text{NO}_3^+ [\text{M} + \text{H}]^+$: 256.0937, found: 256.0936.

2-(Pyridin-2-yl)isoindolin-1-one-3,3- d_2 (12)

Following Procedure A, using 3.5 h instead of 1.5 h.



12

White solid (35 mg, 83% yield, 94% D); m.p. 177 – 179 °C;

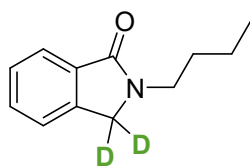
^1H NMR (300 MHz, CDCl_3): δ 8.65 (dt, $J = 8.5, 1.0$ Hz, 1H), 8.37 (ddd, $J = 4.9, 2.0, 1.0$ Hz, 1H), 7.91 (dt, $J = 7.6, 1.0$ Hz, 1H), 7.73 (ddd, $J = 8.5, 7.3, 2.0$ Hz, 1H), 7.65 – 7.43 (m, 3H), 7.04 (ddd, $J = 7.3, 4.9, 1.0$ Hz, 1H), 5.06 (s, 0.12H).

^{13}C NMR (75 MHz, CDCl_3): δ 167.7, 151.8, 147.6, 140.8, 137.8, 132.9, 132.4, 128.1, 124.0, 122.9, 119.3, 114.0, 49.7 – 49.2 (m).

HRMS (ESI): Calcd. for $\text{C}_{13}\text{H}_9\text{D}_2\text{N}_2\text{O}^+ [\text{M} + \text{H}]^+$: 213.0991, found: 213.0996.

2-Butylisindolin-1-one-3,3-*d*₂ (13)

Following Procedure B.



13

Colourless liquid (34 mg, 89% yield, 96% D);

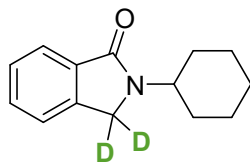
¹H NMR (300 MHz, CDCl₃): δ 7.85 – 7.74 (m, 1H), 7.55 – 7.35 (m, 3H), 4.32 (s, 0.08H), 3.73 – 3.47 (m, 2H), 1.69 – 1.51 (m, 2H), 1.47 – 1.18 (m, 2H), 0.93 (t, *J* = 7.3 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 168.3, 140.8, 132.9, 130.9, 127.7, 123.3, 122.5, 49.7 – 49.1 (m), 41.8, 30.3, 19.8, 13.6.

HRMS (ESI): Calcd. for C₁₂H₁₄D₂NO⁺ [M + H]⁺: 192.1352, found: 192.1351.

2-Cyclohexylisindolin-1-one-3,3-*d*₂ (14)

Following Procedure B, using 3.0 h instead of 1.5 h.



14

Colourless liquid (38 mg, 88% yield, 95% D);

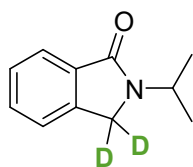
¹H NMR (300 MHz, CDCl₃): δ 7.90 – 7.72 (m, 1H), 7.57 – 7.38 (m, 3H), 4.31 (s, 0.10H), 4.23 (tt, *J* = 11.8, 3.3 Hz, 1H), 1.91 – 1.79 (m, 4H), 1.71 (m, 1H), 1.56 – 1.33 (m, 4H), 1.28 – 1.04 (m, 1H).

¹³C NMR (75 MHz, CDCl₃): δ 167.5, 140.9, 133.0, 130.6, 127.5, 123.0, 122.4, 50.15, 45.6 – 45.1 (m), 31.0, 25.2, 25.1.

HRMS (ESI): Calcd. for C₁₄H₁₆D₂NO⁺ [M + H]⁺: 218.1508, found: 218.1509.

2-Isopropylisindolin-1-one-3,3-*d*₂ (15)

Following Procedure B.



15

Colourless liquid (27 mg, 77% yield, 97% D);

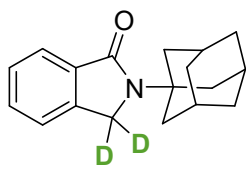
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.88 – 7.76 (m, 1H), 7.61 – 7.34 (m, 3H), 4.66 (pd, J = 6.8, 0.6 Hz, 1H), 4.30 (s, 0.06H), 1.27 (dd, J = 6.8, 0.6 Hz, 6H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 167.5, 140.8, 133.1, 130.7, 127.6, 123.1, 122.5, 44.4 – 43.8 (m), 42.3, 20.5.

HRMS (ESI): Calcd. for $\text{C}_{11}\text{H}_{12}\text{D}_2\text{NO}^+$ [$\text{M} + \text{H}$] $^+$: 178.1195, found: 178.1195.

2-(Adamantan-1-yl)isoindolin-1-one-3,3- d_2 (16)

Following Procedure B, using 0.1 mmol substrate instead of 0.2 mmol, using 1.25 h instead of 1.5 h.



16

White solid (20 mg, 74% yield, 95% D); m.p. 202.5 – 205 °C;

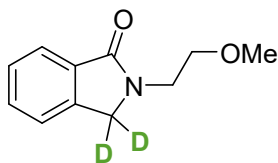
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.78 (dt, J = 7.3, 1.2 Hz, 1H), 7.53 – 7.35 (m, 3H), 4.46 (s, 0.10H), 2.30 (d, J = 3.0 Hz, 6H), 2.21 – 2.11 (m, 3H), 1.83 – 1.63 (m, 6H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 168.6, 140.7, 134.6, 130.6, 127.6, 122.9, 122.2, 55.29, 47.5 – 46.7 (m), 39.9, 36.2, 29.5.

HRMS (ESI): Calcd. for $\text{C}_{18}\text{H}_{20}\text{D}_2\text{NO}^+$ [$\text{M} + \text{H}$] $^+$: 270.1821, found: 270.1824.

2-(2-Methoxyethyl)isoindolin-1-one-3,3- d_2 (17)

Following Procedure B.



17

Colourless liquid (30 mg, 78% yield, 97% D);

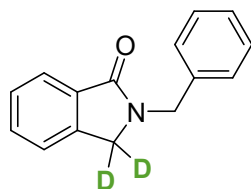
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.83 (dtd, $J = 7.4, 1.3, 0.5$ Hz, 1H), 7.55 – 7.47 (m, 1H), 7.46 – 7.38 (m, 2H), 4.48 (s, 0.06H), 3.81 – 3.72 (m, 2H), 3.62 (dd, $J = 5.6, 4.5$ Hz, 2H), 3.34 (s, 3H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 168.4, 141.4, 132.6, 131.0, 127.6, 123.3, 122.5, 77.4, 58.51, 51.4 – 50.2 (m), 42.1.

HRMS (ESI): Calcd. for $\text{C}_{11}\text{H}_{12}\text{D}_2\text{NO}_2^+ [\text{M} + \text{H}]^+$: 194.1145, found: 194.1145.

2-Benzylisoindolin-1-one-3,3- d_2 (18)

Following Procedure B, using 3.0 h instead of 1.5 h.



18

White solid (34 mg, 76% yield, 96% D); m.p. 88 – 89 °C;

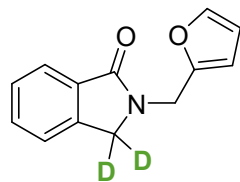
$^1\text{H NMR}$ (300 MHz, CDCl_3): 7.90 – 7.82 (m, 1H), 7.53 – 7.38 (m, 2H), 7.37 – 7.21 (m, 6H), 4.77 (s, 2H), 4.22 (s, 0.08H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 168.3, 141.0, 136.9, 132.5, 131.2, 128.6, 127.9, 127.9, 127.5, 123.6, 122.6, 49.9 – 48.2 (m), 46.2.

HRMS (ESI): Calcd. for $\text{C}_{15}\text{H}_{12}\text{D}_2\text{NO}^+ [\text{M} + \text{H}]^+$: 226.1195, found: 226.1200.

2-(Furan-2-ylmethyl)isoindolin-1-one-3,3- d_2 (19)

Following Procedure B.



19

Yellow liquid (35 mg, 81% yield, 95% D);

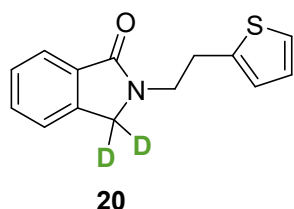
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.85 (ddd, $J = 7.3, 1.4, 1.0$ Hz, 1H), 7.56 – 7.37 (m, 3H), 7.35 (dd, $J = 1.7, 1.0$ Hz, 1H), 6.31 (t, $J = 1.5$ Hz, 2H), 4.78 (s, 2H), 4.35 (s, 0.10H).

^{13}C NMR (75 MHz, CDCl_3): δ 168.0, 150.2, 142.3, 141.0, 132.3, 131.2, 127.8, 123.6, 122.6, 110.3, 108.3, 50.3 – 47.8 (m), 38.8.

HRMS (ESI): Calcd. for $\text{C}_{13}\text{H}_{10}\text{D}_2\text{NO}_2^+ [\text{M} + \text{H}]^+$: 216.0988, found: 216.0993.

2-(2-(Thiophen-2-yl)ethyl)isoindolin-1-one-3,3- d_2 (20)

Following Procedure B, using 1.0 h instead of 1.5 h.



White solid (43.1 mg, 88% yield, 95% D); m.p. 102 – 104 °C;

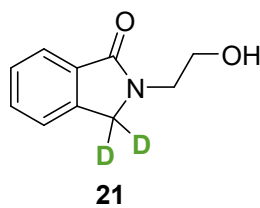
^1H NMR (300 MHz, CDCl_3): δ 7.84 (d, $J = 9$ Hz, 1H), 7.56 – 7.33 (m, 3H), 7.13 (dt, $J = 5.1, 1.0$ Hz, 1H), 6.94 – 6.87 (m, 1H), 6.84 (dt, $J = 3.1, 1.0$ Hz, 1H), 4.22 (s, 0.10H), 4.02 – 3.65 (m, 2H), 3.21 (td, $J = 7.0, 0.9$ Hz, 2H).

^{13}C NMR (75 MHz, CDCl_3): δ 168.4, 141.0, 140.9, 132.6, 131.1, 127.8, 126.9, 125.2, 123.8, 123.4, 122.5, 52.1 – 48.7 (m), 44.1, 28.8.

HRMS (ESI): Calcd. for $\text{C}_{14}\text{H}_{12}\text{D}_2\text{NOS}^+ [\text{M} + \text{H}]^+$: 246.0916, found: 246.0922.

2-(2-Hydroxyethyl)isoindolin-1-one-3,3- d_2 (21)

Following Procedure B, using 3.0 h instead of 1.5 h.



White solid (25 mg, 70% yield, 95% D); m.p. 114 – 115 °C;

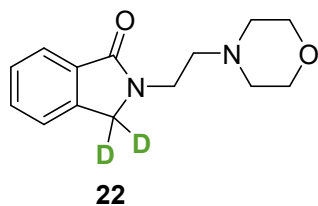
^1H NMR (300 MHz, CDCl_3): δ 7.79 (dq, $J = 7.4, 1.1$ Hz, 1H), 7.56 – 7.47 (m, 1H), 7.45 – 7.37 (m, 2H), 4.48 (s, 0.10H), 3.94 – 3.84 (m, 2H), 3.80 – 3.69 (m, 2H), 3.45 (s, 1H).

^{13}C NMR (75 MHz, CDCl_3): δ 169.5, 141.5, 132.5, 131.3, 127.9, 123.4, 122.7, 61.1, 51.8 – 50.7 (m), 45.7.

HRMS (ESI): Calcd. for $\text{C}_{10}\text{H}_{10}\text{D}_2\text{NO}_2^+ [\text{M} + \text{H}]^+$: 180.0988, found: 180.0982.

2-(2-Morpholinoethyl)isoindolin-1-one-3,3-*d*₂ (22)

Following Procedure B, using 1.25 h instead of 1.5 h.



Colourless liquid (37 mg, 75% yield, 97% D);

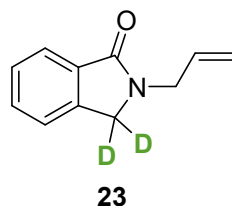
¹H NMR (300 MHz, CDCl₃): δ 7.97 – 7.71 (m, 1H), 7.59 – 7.42 (m, 3H), 4.48 (s, 0.06H), 3.87 – 3.57 (m, 6H), 2.63 (t, *J* = 6.4 Hz, 2H), 2.51 (dd, *J* = 5.7, 3.7 Hz, 4H).

¹³C NMR (75 MHz, CDCl₃): δ 168.4, 141.2, 132.8, 131.1, 127.8, 123.5, 122.6, 66.8, 56.9, 53.5, 50.7 – 49.7 (m), 39.0.

HRMS (ESI): Calcd. for C₁₄H₁₇D₂N₂O₂⁺ [M + H]⁺: 249.1567, found: 249.1575.

2-Allylisoindolin-1-one-3,3-*d*₂ (23)

Following Procedure B, using 2.0 h instead of 1.5 h.



Colourless liquid (25 mg, 70% yield, 94% D);

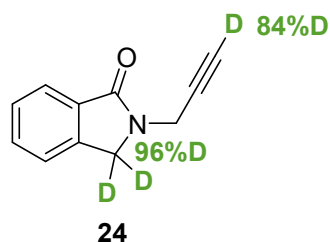
¹H NMR (300 MHz, CDCl₃): δ 7.84 (dt, *J* = 7.3, 1.2 Hz, 1H), 7.57 – 7.38 (m, 3H), 5.85 (ddt, *J* = 17.4, 9.8, 6.0 Hz, 1H), 5.24 (m, 1H), 5.19 (t, *J* = 1.4 Hz, 1H), 4.33 (s, 0.12H), 4.22 (dt, *J* = 6.0, 1.4 Hz, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 168.0, 140.9, 132.8, 132.5, 131.1, 127.7, 123.4, 122.6, 117.6, 49.30 – 48.7 (m), 44.7.

HRMS (ESI): Calcd. for C₁₁H₁₀D₂NO⁺ [M + H]⁺: 176.1039, found: 176.1041.

2-(Prop-2-yn-1-yl-3-*d*)isoindolin-1-one-3,3-*d*₂ (24)

Following Procedure B.



Colourless liquid (25 mg, 29% yield);

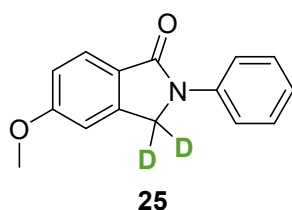
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 8.02 – 7.64 (m, 1H), 7.61 – 7.41 (m, 3H), 4.80 (s, 0.08H), 4.45 (s, 2H), 2.29 (s, 0.16H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 168.1, 141.2, 132.2, 131.7, 128.2, 123.9, 123.0, 77.4, 72.7-72.5 (m), 49.3 – 48.4 (m), 31.8.

HRMS (ESI): Calcd. for $\text{C}_{11}\text{H}_7\text{D}_3\text{NO}^+$ $[\text{M} + \text{H}]^+$: 175.0945, found: 175.0945.

5-Methoxy-2-phenylisoindolin-1-one-3,3- d_2 (25)

Following Procedure A, using 2.5 h instead of 1.5 h.



White solid (34 mg, 70% yield, 95% D); m.p. 144 – 146 °C;

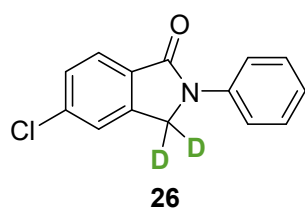
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.88 – 7.75 (m, 3H), 7.42 (dd, $J = 8.7, 7.4$ Hz, 2H), 7.20 – 7.10 (m, 1H), 7.07 – 6.95 (m, 2H), 4.80 (s, 0.10H), 3.90 (s, 3H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 167.5, 163.3, 142.4, 139.8, 129.2, 125.9, 125.6, 124.1, 119.2, 115.2, 107.4, 55.79, 50.5 – 49.0 (m).

HRMS (ESI): Calcd. for $\text{C}_{15}\text{H}_{12}\text{D}_2\text{NO}_2^+$ $[\text{M} + \text{H}]^+$: 242.1145, found: 242.1145.

5-Chloro-2-phenylisoindolin-1-one-3,3- d_2 (26)

Following Procedure A, using 3.0 h instead of 1.5 h.



yellow solid (35 mg, 71% yield, 93% D); m.p. 156 – 158 °C;

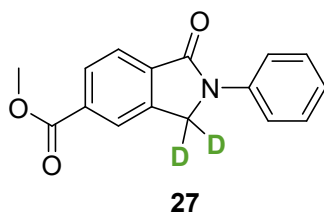
¹H NMR (300 MHz, CDCl₃): δ 7.95 – 7.72 (m, 3H), 7.54 – 7.47 (m, 2H), 7.43 (dd, *J* = 8.7, 7.3 Hz, 2H), 7.23 – 7.16 (m, 1H), 4.84 (s, 0.14H).

¹³C NMR (75 MHz, DMSO - *d*₆): δ 165.7, 143.0, 139.2, 137.0, 129.0, 128.6, 128.2, 125.0, 124.3, 123.6, 119.3, 50.2 – 49.2(m).

HRMS (ESI): Calcd. for C₁₄H₉D₂ClNO⁺ [M + H]⁺: 246.0649, found: 246.0654.

3-(Benzo[*d*][1,3]dioxol-5-yl)-*N*-ethyl-*N*-phenylpropanamide-2,2-*d*₂ (27)

Following Procedure A, using 0.1 mmol instead of 0.2 mmol, using 50 °C instead of rt.



White solid (27 mg, 50% yield, 95% D); m.p. 142 – 145 °C;

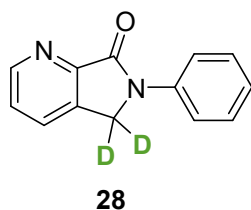
¹H NMR (300 MHz, CDCl₃): δ 8.20 (d, *J* = 7.8 Hz, 2H), 7.99 (d, *J* = 8.3 Hz, 1H), 7.92 – 7.82 (d, *J* = 8.3 Hz, 2H), 7.45 (t, *J* = 7.8 Hz, 2H), 7.30 – 7.11 (m, 1H), 4.91 (s, 0.10H), 3.98 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 166.6, 166.5, 140.0, 139.3, 137.3, 135.1, 133.7, 130.0, 129.4, 125.1, 124.3, 119.7, 52.7, 52.3 – 51.9 (m).

HRMS (ESI): Calcd. for C₁₆H₁₂D₂NO₃⁺ [M + H]⁺: 270.1094, found: 270.1088.

6-Phenyl-5,6-dihydro-7*H*-pyrrolo[3,4-*b*]pyridin-7-one-5,5-*d*₂ (28)

Following Procedure A.



Yellow solid (25 mg, 60% yield, 95% D); m.p. 193 – 195 °C;

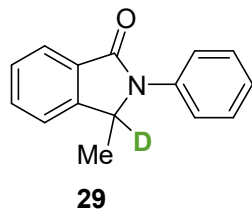
¹H NMR (300 MHz, CDCl₃): δ 8.79 (dd, *J* = 4.8, 1.5 Hz, 1H), 7.99 – 7.83 (m, 3H), 7.61 – 7.38 (m, 3H), 7.19 (ddt, *J* = 8.7, 7.5, 1.2 Hz, 1H), 4.85 (s, 0.10H).

¹³C NMR (75 MHz, CDCl₃): δ 165.2, 151.1, 150.5, 138.9, 134.1, 131.3, 129.1, 125.7, 124.9, 119.3, 48.7 – 47.4 (m).

HRMS (ESI): Calcd. for $C_{13}H_9D_2N_2O^+$ $[M + H]^+$: 213.0991, found: 213.0994.

3-Methyl-2-phenylisoindolin-1-one-3-*d* (29)

Following Procedure A, using 6.0 h instead of 1.5 h.



Yellow liquid (36 mg, 80% yield, 93% D);

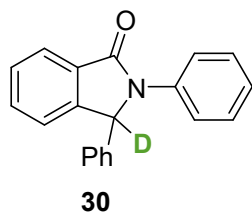
1H NMR (300 MHz, $CDCl_3$): δ 7.93 (dt, $J = 7.5, 1.1$ Hz, 1H), 7.66 – 7.56 (m, 3H), 7.55 – 7.40 (m, 4H), 7.27 – 7.20 (m, 1H), 5.21 (q, $J = 6.7$ Hz, 0.10H), 1.45 (s, 3H).

^{13}C NMR (75 MHz, $CDCl_3$): δ 166.9, 146.2, 137.0, 132.0, 131.7, 129.0, 128.3, 125.3, 124.0, 123.3, 122.0, 56.8 – 56.2 (m), 18.6.

HRMS (ESI): Calcd. for $C_{15}H_{13}DNO^+$ $[M + H]^+$: 225.1133, found: 225.1142.

2,3-Diphenylisoindolin-1-one-3-*d* (30)

Following Procedure A, using 3.0 h instead of 1.5 h.



White solid (44 mg, 77% yield, 95% D); m.p. 198 – 199 °C;

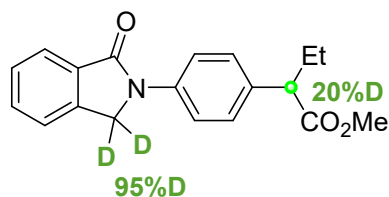
1H NMR (300 MHz, $CDCl_3$): δ 8.02 – 7.93 (m, 1H), 7.68 – 7.59 (m, 2H), 7.57 – 7.45 (m, 2H), 7.36 – 7.18 (m, 8H), 7.10 (td, $J = 7.3, 1.1$ Hz, 1H), 6.10 (s, 0.05H).

^{13}C NMR (75 MHz, $CDCl_3$): δ 167.9, 145.6, 137.6, 137.5, 132.4, 131.1, 129.1, 128.8, 128.5, 128.3, 126.8, 124.9, 124.0, 123.0, 122.4, 65.5 – 64.9 (m).

HRMS (ESI): Calcd. for $C_{20}H_{15}DNO^+$ $[M + H]^+$: 287.1289, found: 287.1300.

Methyl 2-(4-(1-oxoisoindolin-2-yl-3,3-*d*)phenyl)butanoate (31)

Following Procedure A.



31

Colourless liquid (50 mg, 80% yield);

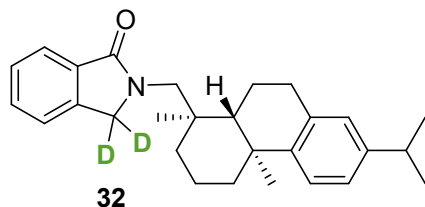
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.89 (dt, $J = 7.4, 1.2$ Hz, 1H), 7.85 – 7.78 (m, 2H), 7.61 – 7.53 (m, 1H), 7.52 – 7.43 (m, 2H), 7.39 – 7.31 (m, 2H), 4.81 (s, 0.10H), 3.66 (s, 3H), 3.46 (t, $J = 7.7$ Hz, 0.8H), 2.10 (ddd, $J = 13.6, 7.7, 7.4$ Hz, 1H), 1.82 (dq, $J = 13.6, 7.4$ Hz, 1H), 0.90 (t, $J = 7.4$ Hz, 3H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 174.2, 167.3, 139.8, 138.4, 134.9, 133.0, 131.9, 128.5, 128.2, 123.8, 122.5, 119.3, 52.6, 51.7, 50.5 – 49.9 (m), 26.5, 12.0.

HRMS (ESI): Calcd. for $\text{C}_{19}\text{H}_{18}\text{D}_2\text{NO}_3^+ [\text{M} + \text{H}]^+$: 312.1563, found: 312.1552.

2-(((1*R*,4*aS*,10*aR*)-7-Isopropyl-1,4*a*-dimethyl-1,2,3,4,4*a*,9,10,10*a*-octahydrophenanthren-1-yl)methyl)isoindolin-1-one-3,3-*d*₂ (32)

Following Procedure B, using 1.25 h instead of 1.5 h.



32

Yellow liquid (65 mg, 81% yield, 96% D);

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.88 – 7.78 (m, 1H), 7.56 – 7.33 (m, 3H), 7.16 (d, $J = 8.2$ Hz, 1H), 6.99 (dd, $J = 8.2, 2.1$ Hz, 1H), 6.91 (d, $J = 2.1$ Hz, 1H), 4.66 – 4.37 (s, 0.08H), 3.49 (d, $J = 2.8$ Hz, 2H), 3.09 – 2.70 (m, 3H), 2.29 (d, $J = 13.0$ Hz, 1H), 2.13 (d, $J = 6.5$ Hz, 1H), 1.97 – 1.65 (m, 3H), 1.59 (q, $J = 2.8, 2.3$ Hz, 2H), 1.56 – 1.52 (m, 1H), 1.40 (dd, $J = 17.6, 5.0$ Hz, 1H), 1.24 (d, $J = 4.4$ Hz, 6H), 1.21 (s, 3H), 1.07 (s, 3H).

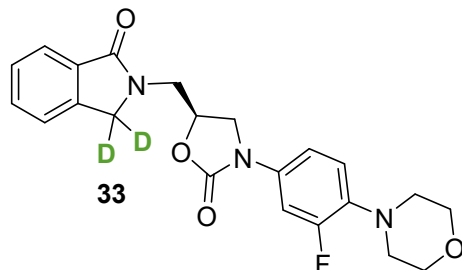
$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 169.9, 147.1, 145.4, 141.2, 134.4, 132.4, 131.1, 127.8, 126.8, 126.7, 123.9, 123.7, 123.5, 122.3, 54.8, 53.0 – 53.8 (m), 45.3, 39.4, 38.1, 37.4, 37.3, 33.3, 30.0, 25.5, 23.8, 23.8, 19.1, 19.0, 18.6.

HRMS (ESI): Calcd. for $\text{C}_{28}\text{H}_{34}\text{D}_2\text{NO}^+ [\text{M} + \text{H}]^+$: 404.2917, found: 404.2928.

(*S*)-3-(3-Fluoro-4-morpholinophenyl)-5-((1-oxoisoindolin-2-yl)-3,3-

***d*₂methyl)oxazolidin-2-one (33)**

Following Procedure A, using 0.1 mmol instead of 0.2 mmol, using 50 °C instead of rt, using 3.0 h instead of 1.5 h.



White solid (25 mg, 60% yield, 95% D); m.p. 235 – 237 °C;

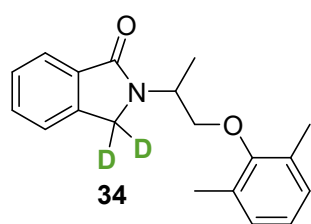
¹H NMR (300 MHz, CDCl₃): δ 7.87 – 7.81 (m, 1H), 7.61 – 7.53 (m, 1H), 7.51 – 7.45 (m, 2H), 7.41 (dd, *J* = 14.3, 2.6 Hz, 1H), 7.06 (ddd, *J* = 8.8, 2.6, 1.1 Hz, 1H), 6.90 (d, *J* = 8.8 Hz, 1H), 5.05 – 4.84 (m, 1H), 4.60 (s, 0.10H), 4.16 – 4.00 (m, 2H), 3.97 – 3.77 (m, 6H), 3.07 – 2.89 (m, 4H).

¹³C NMR (75 MHz, CDCl₃): δ 169.4, 162.4, 156.8, 154.0, 153.5, 141.5, 136.1, 132.9, 131.7, 131.4, 127.9, 123.4, 122.7, 118.6, 113.8, 72.1, 66.6, 50.7, 49.8 – 49.1 (m), 47.7, 45.3.

HRMS (ESI): Calcd. for C₂₂H₂₁D₂FN₃O₄⁺ [M + H]⁺: 414.1793, found: 414.1784.

2-(1-(2,6-Dimethylphenoxy)propan-2-yl)isoindolin-1-one-3,3-*d*₂ (34)

Following Procedure B, using 10 mA instead of 5 mA.



Colourless liquid (48.1 mg, 81% yield, 95% D);

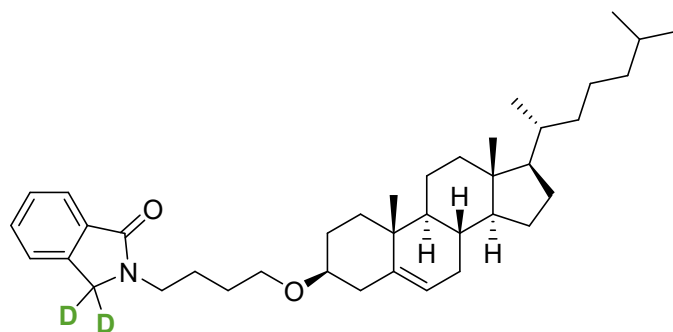
¹H NMR (300 MHz, CDCl₃): δ 7.94 – 7.84 (m, 1H), 7.58 – 7.51 (m, 1H), 7.47 (m, 2H), 6.97 (ddd, *J* = 7.0, 1.6, 0.6 Hz, 2H), 6.92 – 6.86 (m, 1H), 4.83 (tdd, *J* = 7.1, 4.6, 2.5 Hz, 1H), 4.53 (s, 0.10H), 3.95 (h, *J* = 4.6 Hz, 2H), 2.20 – 2.13 (m, 6H), 1.56 (d, *J* = 7.1 Hz, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 168.5, 155.1, 141.5, 133.0, 131.2, 130.6, 128.9, 127.9, 124.0, 124.0, 123.5, 122.7, 73.9, 47.4, 46.9 – 46.3 (m), 16.1, 15.4.

HRMS (ESI): Calcd. for $C_{19}H_{20}D_2NO_2^+ [M + H]^+$: 298.1771, found: 298.1777.

2-(4-(((3*S*,8*S*,9*S*,10*R*,13*R*,14*S*,17*R*)-10,13-Dimethyl-17-((*R*)-6-methylheptan-2-yl)-2,3,4,7,8,9,10,11,12,13,14,15,16,17-tetradecahydro-1*H*-cyclopenta[*a*]phenanthren-3-yl)oxy)butyl)isoindolin-1-one-3,3-*d*₂ (35)

Following Procedure B, using 2.0 h instead of 1.5 h.



35

Colourless liquid (95 mg, 82% yield, 97% D);

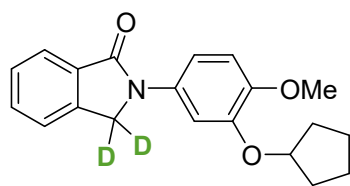
¹H NMR (300 MHz, CDCl₃): δ 7.85 – 7.76 (m, 1H), 7.52 – 7.45 (m, 1H), 7.41 (m, 2H), 5.30 (m, 1H), 4.36 (s, 0.06H), 3.62 (t, *J* = 7.2 Hz, 2H), 3.48 (t, *J* = 7.2 Hz, 2H), 3.10 (m, 1H), 2.32 (m, 1H), 2.24 – 2.08 (m, 1H), 1.98 (m, 2H), 1.93 – 1.67 (m, 5H), 1.65 – 1.05 (m, 21H), 0.96 (s, 4H), 0.89 (d, *J* = 6.4 Hz, 4H), 0.84 (dd, *J* = 6.4, 1.3 Hz, 6H), 0.65 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 168.6, 141.2, 141.1, 133.2, 131.2, 128.1, 123.7, 122.7, 121.6, 79.1, 67.4, 56.9, 56.3, 50.3, 50.1 – 49.9 (m), 42.4, 42.2, 39.9, 39.6, 39.3, 37.3, 37.0, 36.3, 35.9, 32.0, 28.6, 28.3, 28.1, 27.5, 25.4, 24.4, 23.9, 22.9, 22.7, 21.2, 19.5, 18.8, 12.0.

HRMS (ESI): Calcd. for $C_{39}H_{58}D_2NO_2^+ [M + H]^+$: 576.4744, found: 576.4736.

2-(3-(Cyclopentyloxy)-4-methoxyphenyl)isoindolin-1-one-3,3-*d*₂ (36)

Following Procedure B.



36

White solid (41 mg, 62% yield, 95% D); mp: 137 – 140 °C;

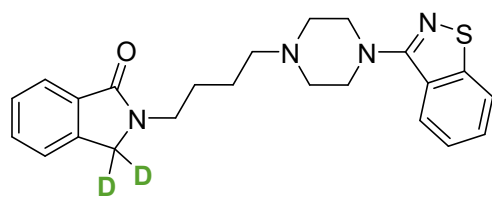
¹H NMR (300 MHz, CDCl₃): δ 7.94 – 7.78 (m, 2H), 7.62 – 7.53 (m, 1H), 7.52 – 7.44 (m, 1H), 6.99 (dd, *J* = 8.7, 2.6 Hz, 1H), 6.87 (d, *J* = 8.7 Hz, 1H), 4.86 (dt, *J* = 6.3, 3.1 Hz, 1H), 4.78 (s, 0.10H), 3.84 (s, 3H), 2.14 – 1.74 (m, 6H), 1.70 – 1.53 (m, 2H).

¹³C NMR (75 MHz, CDCl₃): δ 167.1, 147.8, 146.8, 139.8, 133.2, 133.0, 131.7, 128.1, 123.6, 122.4, 111.9, 110.6, 107.7, 107.6, 80.4, 56.1, 50.6 – 49.9 (m), 32.7, 24.0.

HRMS (ESI): Calcd. for C₂₀H₂₀D₂NO₃⁺ [M + H]⁺: 326.1720, found: 326.1717.

2-(4-(4-(Benzo[*d*]isothiazol-3-yl)piperazin-1-yl)butyl)isoindolin-1-one-3,3-*d*₂ (37)

Following Procedure B, using 4.0 h instead of 1.5 h.



37

White solid (50 mg, 61% yield, 96% D); mp: 233.2 – 234.8 °C;

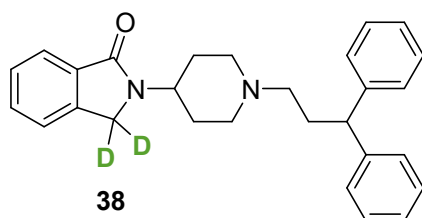
¹H NMR (300 MHz, CDCl₃): δ 7.87 (dt, *J* = 8.1, 1.0 Hz, 1H), 7.79 (ddt, *J* = 12.9, 8.1, 1.1 Hz, 2H), 7.55 – 7.38 (m, 4H), 7.36 – 7.29 (m, 1H), 4.36 (s, 0.08H), 3.64 (t, *J* = 7.0 Hz, 2H), 3.57 – 3.35 (m, 4H), 2.63 (t, *J* = 4.9 Hz, 4H), 2.45 (t, *J* = 7.0 Hz, 2H), 1.78 – 1.50 (m, 4H).

¹³C NMR (75 MHz, CDCl₃): δ 168.4, 163.7, 152.5, 140.8, 132.8, 131.0, 127.8, 127.3, 123.7, 123.4, 122.5, 120.3, 52.7, 49.7, 49.3 – 48.7 (m), 41.9, 26.1, 23.8.

HRMS (ESI): Calcd. for C₂₃H₂₅D₂N₄OS⁺ [M + H]⁺: 409.2026, found: 409.2037.

2-(1-(3,3-Diphenylpropyl)piperidin-4-yl)isoindolin-1-one-3,3-*d*₂ (38)

Following Procedure B, using 4.0 h instead of 1.5 h.



38

Colourless liquid (41 mg, 50% yield, 97% D);

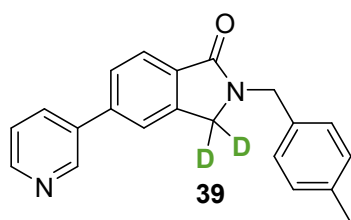
¹H NMR (300 MHz, CDCl₃): δ 7.88 – 7.81 (m, 1H), 7.62 – 7.41 (m, 4H), 7.35 – 7.23 (m, 8H), 7.18 (m, 2H), 4.45 (s, 0.10H), 4.38 – 4.20 (m, 1H), 3.98 (t, *J* = 7.1 Hz, 1H), 3.01 (m, 2H), 2.46 – 2.23 (m, 4H), 2.19 – 2.07 (m, 2H), 1.89 – 1.72 (m, 4H).

¹³C NMR (75 MHz, CDCl₃): δ 168.2, 144.7, 141.2, 133.2, 131.2, 128.5, 128.0, 127.8, 126.2, 123.6, 122.8, 56.9, 53.0, 49.3, 48.9, 45.9 – 45.1 (m), 33.0, 30.3.

HRMS (ESI): Calcd. for C₂₈H₂₉D₂N₂O⁺ [M + H]⁺: 413.2556, found: 413.2558.

2-(4-Methylbenzyl)-5-(pyridin-3-yl)isoindolin-1-one-3,3-*d*₂ (39)

Following Procedure B, using 2.0 h instead of 1.5 h.



Yellow solid (53 mg, 84% yield, 94% D); mp: 145 – 147 °C;

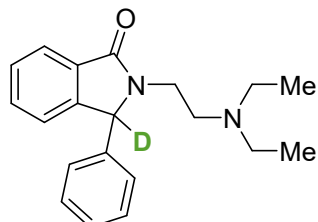
¹H NMR (300 MHz, CDCl₃): δ 8.81 (d, *J* = 2.4 Hz, 1H), 8.60 (dd, *J* = 4.8, 1.6 Hz, 1H), 7.95 (dd, *J* = 7.9, 0.8 Hz, 1H), 7.85 (dt, *J* = 7.9, 2.0 Hz, 1H), 7.63 (dd, *J* = 7.9, 1.6 Hz, 1H), 7.54 – 7.51 (m, 1H), 7.36 (dd, *J* = 8.1, 4.8 Hz, 1H), 7.19 (d, *J* = 8.1 Hz, 2H), 7.12 (d, *J* = 7.9 Hz, 2H), 4.76 (s, 2H), 4.29 (s, 0.12H), 2.30 (s, 3H).

¹³C NMR (75 MHz, CDCl₃): δ 167.9, 149.0, 148.3, 142.1, 141.0, 137.4, 135.9, 134.6, 133.8, 132.5, 129.4, 128.1, 127.3, 124.4, 123.7, 121.6, 49.3 – 48.1 (m), 46.1, 21.0.

HRMS (ESI): Calcd. for C₂₁H₁₇D₂N₂O⁺ [M + H]⁺: 317.1617, found: 317.1624.

2-(2-(Diethylamino)ethyl)-3-phenylisoindolin-1-one-3-*d* (40)

Following Procedure A, using *n*-Bu₄NI (1.5 eq.) instead of *n*-Bu₄NI (0.2 eq.), using 20 mA instead of 5 mA, using 3.0 h instead of 1.5 h.



40

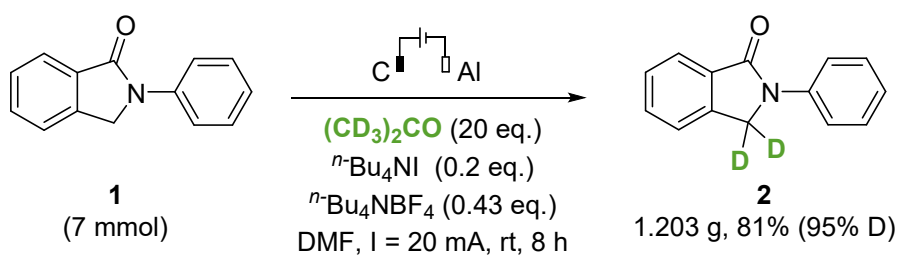
Colourless liquid (19 mg, 30% yield, 90% D);

¹H NMR (300 MHz, CDCl₃): δ 7.97 – 7.81 (m, 1H), 7.53 – 7.42 (m, 2H), 7.35 (m, 3H), 7.20 – 7.09 (m, 3H), 5.68 (s, 0.10H), 3.95 (m, 1H), 3.17 (m, 1H), 2.85 (dt, *J* = 13.6, 7.1 Hz, 1H), 2.69 (qd, *J* = 7.1, 2.8 Hz, 5H), 1.05 (t, *J* = 7.1 Hz, 6H).

¹³C NMR (75 MHz, CDCl₃): δ 169.3, 146.6, 136.8, 132.1, 131.3, 129.3, 128.9, 128.4, 127.8, 123.5, 123.3, 65.6 – 65.0 (m), 50.8, 47.4, 37.8, 10.8.

HRMS (ESI): Calcd. for C₂₀H₂₄DN₂O⁺ [M + H]⁺: 310.2024, found: 310.2039.

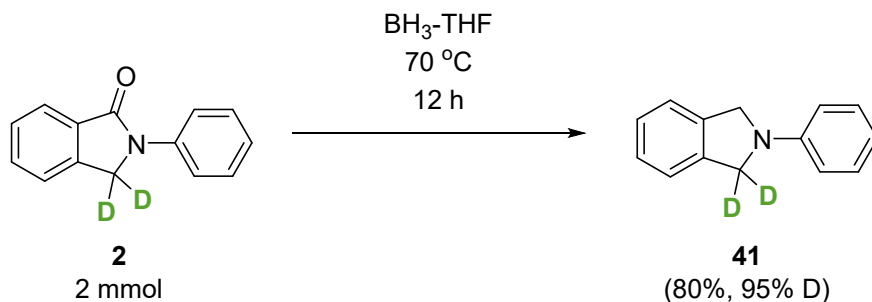
Procedure for gram-scale experiment (Procedure C):



A 50 mL distillation flask equipped with a magnetic stir bar was charged with compound **1** (1.463 g, 7 mmol, 1.0 eq.), (CD₃)₂CO (10.5 mL, 20 eq.), *n*-Bu₄NBF₄ (0.988 g, 3 mmol, 0.43 eq.), *n*-Bu₄NI (0.517 g, 1.4 mmol, 0.2 eq.) and DMF (30 mL). The flask equipped with graphite rod anode (*d* = 5 mm) and aluminium rod cathode (*d* = 5 mm) (the submerged height of the electrode is approximately 2 cm). The resulting solution was stirred and electrolyzed at a constant current of 20 mA (Single Output DC Power Supply: KRP-305DM) for 8 hours at room temperature. The solution was diluted with EtOAc (20 mL) and brine (20 mL), and extracted with EtOAc (3 × 30 mL). The combined organic layers were washed with brine (3 × 50 mL), dried over Na₂SO₄, and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (*v/v* = 10/1) as eluent afforded the desired product **2** (1.203 g, 81%, 95%D).

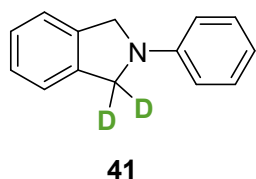
¹H NMR (300 MHz, CDCl₃): δ 7.95 – 7.89 (m, 1H), 7.89 – 7.82 (m, 2H), 7.63 – 7.55 (m, 1H), 7.54 – 7.47 (m, 2H), 7.46 – 7.38 (m, 2H), 7.22 – 7.13 (m, 1H), 4.85 – 4.82 (s, 0.10 H).

Procedure for transformation of isoindolinones:



A dry 50 mL three-necked bottle flask was charged with $\text{BH}_3\text{-THF}$ (12 mL, 12 mmol, 6 eq., 1 mol/L in anhydrous THF) and compound **2** (422 mg, 2 mmol, 1.0 eq.) under nitrogen atmosphere. The reaction mixture was heated to $70\text{ }^\circ\text{C}$ and refluxed for 12 h. The solution was cooled to room temperature under nitrogen, then concentrated *in vacuo*. The solution was diluted with DCM (5 mL) and brine (20 mL), and extracted with DCM ($3 \times 20\text{ mL}$). The combined organic layers were washed with brine ($3 \times 20\text{ mL}$), dried over Na_2SO_4 , and concentrated *in vacuo*. Purification by flash column chromatography using PE as eluent afforded the desired product **41**.

2-Phenylisoindoline-1,1- d_2 (**41**)



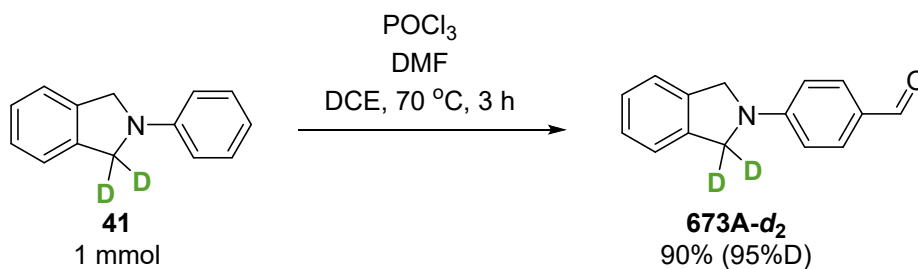
White solid (315 mg, 80% yield, 95% D); mp: $171 - 172\text{ }^\circ\text{C}$;

$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 7.38 – 7.28 (m, 6H), 6.76 (tt, $J = 7.3, 0.9\text{ Hz}$, 1H), 6.69 (dq, $J = 7.3, 0.9\text{ Hz}$, 2H), 4.66 (s, 2.10H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 147.1, 137.9, 137.7, 129.2, 127.0, 122.5, 116.0, 111.4, 53.6, 53.5 – 53.0 (m).

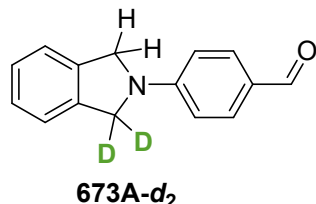
HRMS (ESI): Calcd. for $\text{C}_{14}\text{H}_{12}\text{D}_2\text{N}^+ [\text{M} + \text{H}]^+$: 198.1246, found: 198.1251.

Synthesis of compound 673A- d_2 :



A dry 50 mL three-necked bottle flask was charged with DMF (2 mmol, 2 eq.) in DCE (5 mL) and POCl_3 (2 mmol, 2.0 eq.) was added dropwise to the above solution at 0 °C under nitrogen atmosphere. The **41** (1 mmol, 1.0 eq.) was added after stir 20 minutes. The reaction mixture was at 70 °C for 3 h. The solution was cooled to room temperature and pour into ice water and saturated Na_2CO_3 solution. The solution was diluted with DCM (5 mL) and brine (20 mL), and extracted with DCM (3 × 20 mL). The combined organic layers were washed with brine (3 × 20 mL), dried over Na_2SO_4 , and concentrated *in vacuo*. Purification by flash column chromatography using PE/EtOAc (v/v = 3/1) as eluent afforded the desired product **673A-*d*₂**.

4-(Isoindolin-2-yl-1,1-*d*₂)benzaldehyde (**673A-*d*₂**)



White solid (202 mg, 90% yield, 95% D); mp: 174 – 176 °C;

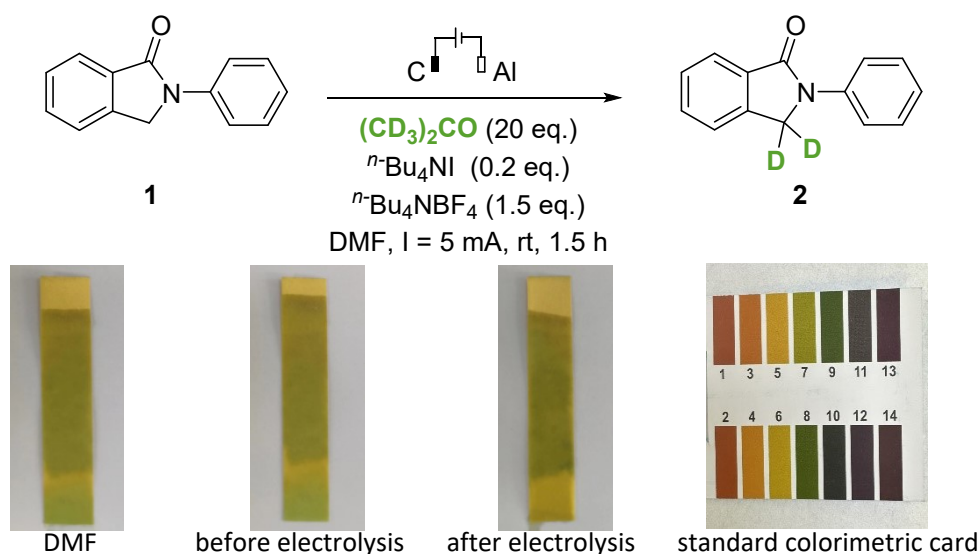
$^1\text{H NMR}$ (300 MHz, CDCl_3): δ 9.77 (s, 1H), 7.87 – 7.74 (m, 2H), 7.34 (d, $J = 1.7$ Hz, 4H), 6.75 – 6.65 (m, 2H), 4.73 (s, 2.10H).

$^{13}\text{C NMR}$ (75 MHz, CDCl_3): δ 190.1, 151.1, 136.5, 136.3, 132.0, 127.5, 125.4, 122.5, 111.0, 53.5 – 52.8 (m).

HRMS (ESI): Calcd. for $\text{C}_{15}\text{H}_{12}\text{D}_2\text{NO}^+$ [$\text{M} + \text{H}$] $^+$: 226.1195, found: 226.1198.

V Mechanism Research

Scheme S2: pH Detection.



(1) The pH test paper was moistened with water and anhydrous DMF was dropped on it. The color did not change obviously.

(2) The pH test paper was moistened with water and the reaction mixture before electrification was dropped on the pH test paper. The color did not change obviously.

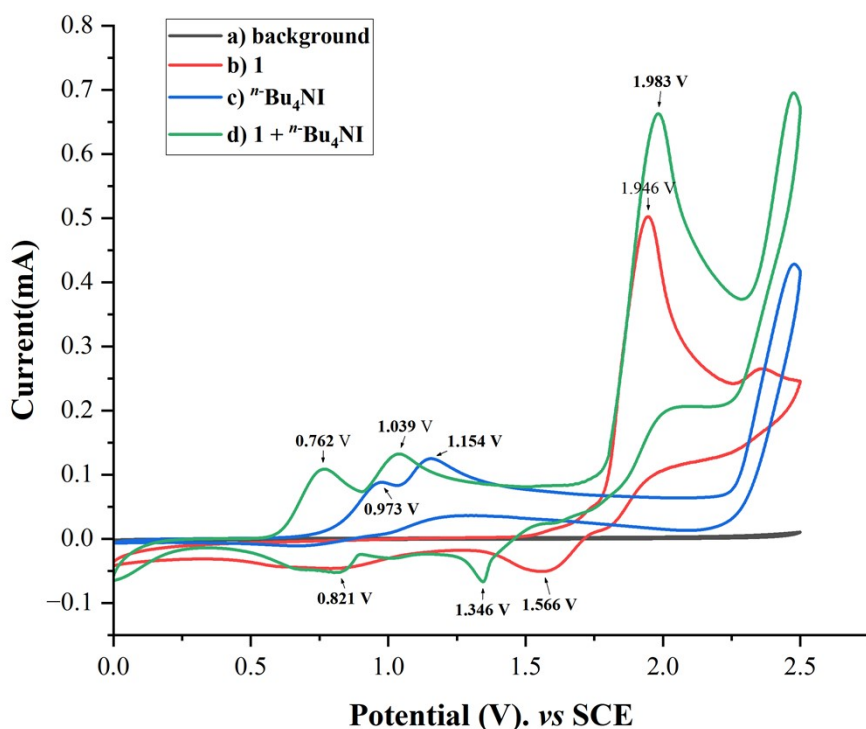
(3) After the reaction was completed under standard conditions, the pH test paper was moistened with water and the mixture was dropped on it. The color did not change obviously.

Scheme S3: Cyclic Voltammetry Experiment

Cyclic voltammetry was carried out in a glass cell with CHI760E electrochemical workstation. Cyclic voltammetry of compound **1** at a scan rate of $100 \text{ mV}\cdot\text{s}^{-1}$. The electrochemical experiments were performed in a three-compartment cell fitted with a glassy carbon as the working electrode, a saturated calomel electrode (SCE) as the reference electrode, and a graphite rod (diameter is 5 mm) as the counter electrode.

Electrolyte: $0.1 \text{ M } n\text{-Bu}_4\text{NBF}_4$ in MeCN; Concentration of a sample: 0.01 M .

In the mixture of **1** with $n\text{-Bu}_4\text{NI}$, the oxidation peak of $n\text{-Bu}_4\text{NI}$ dramatically shifted from 0.973 V to 0.762 V and from 1.154 V to 1.039 V . In addition, the reduction peak of **1** shifted from 1.566 V to 1.346 V and the another reduction peak remained at 0.821 V . (curve d)



Cyclic voltammograms obtained in $0.1 \text{ M } n\text{-Bu}_4\text{NBF}_4/\text{MeCN}$ using glass carbon (diameter, 3 mm) as the working electrode, graphite rod, and saturated calomel electrode (SCE) as the auxiliary and reference electrode, respectively, at a scan rate of $100 \text{ mV}\cdot\text{s}^{-1}$: (a) background, (b) $10 \text{ mM } \mathbf{1}$, (c) $10 \text{ mM } n\text{-Bu}_4\text{NI}$ and (d) $10 \text{ mM } \mathbf{1} + 10 \text{ mM } n\text{-Bu}_4\text{NI}$.

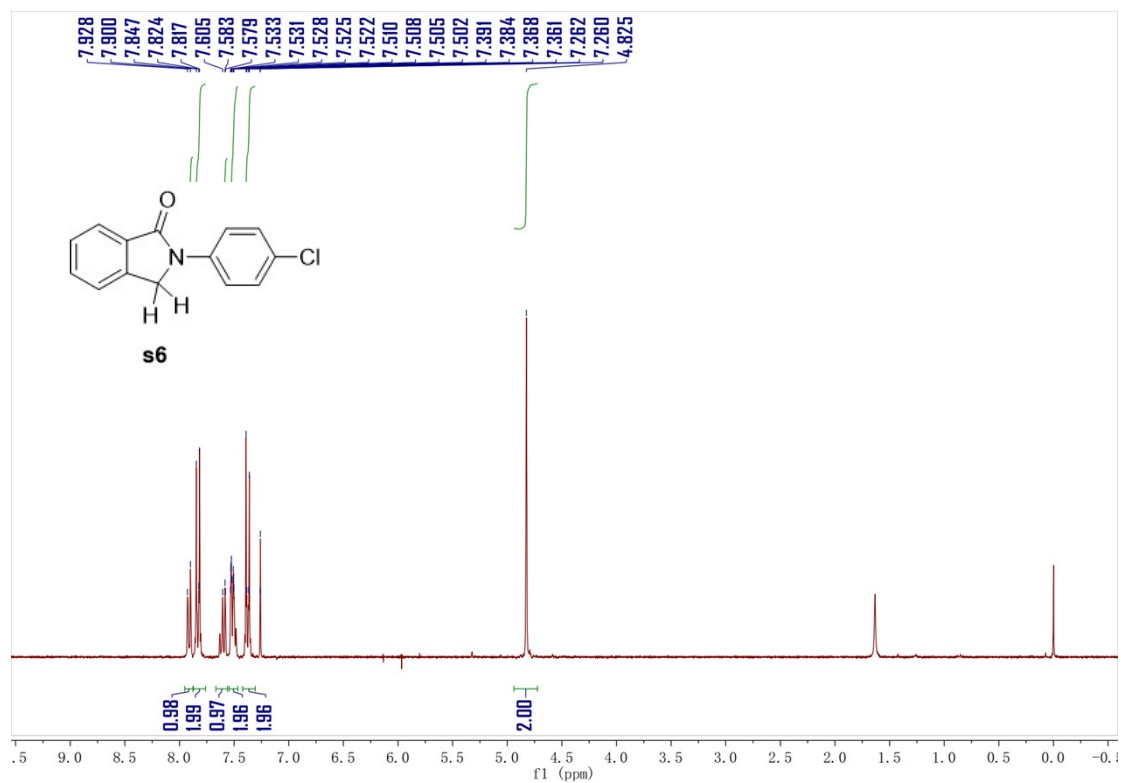
VI References

- 1 S. Ning, L. Zheng, Y. Bai, S. Wang, S. Wang, L. Shi, Q. Gao, X. Che, Z. Zhang and J. Xiang, Highly selective electroreductive linear dimerization of electron-deficient vinylarenes, *Tetrahedron*, 2021, **102**, 132535.
- 2 Y. Bai, L. Shi, L. Zheng, S. Ning, X. Che, Z. Zhang and J. Xiang, Electroselective and controlled reduction of cyclic imides to hydroxylactams and lactams, *Org. Lett.*, 2021, **23**, 2298–2302.
- 3 H. Wang, Z. Xie, B. Lu, K. Zhong, J. Lu and J. Liu, One-pot method to construct isoindolinones and its application to the synthesis of DWP205109 and intermediate of Lenalidomide, *Tetrahedron Lett.*, 2021, **74**, 153152–153157.
- 4 Y. Tian, J. Wei, M. Wang, G. Li and F. Xu, Hantzsch ester triggered metal-free cascade approach to isoindolinones, *Tetrahedron Lett.*, 2018, **59**, 1866–1870.
- 5 Y. Zhou, P. Chen, X. Lv, J. Niu, Y. Wang, M. Lei and L. Hu, A facile and efficient method for the synthesis of N-substituted isoindolin-1-one derivatives under Pd(OAc)₂/HCOOH system, *Tetrahedron Lett.*, 2017, **58**, 2232–2235.
- 6 Y. M. Zhu, Y. Fang, H. Li, X. P. Xu and S. J. Ji, Divergent reaction of isocyanides with o-bromobenzaldehydes: Synthesis of ketenimines and lactams with isoindolinone cores, *Org. Lett.*, 2021, **23**, 7342–7347.
- 7 Z. Zou, G. Cai, W. Chen, C. Zou, Y. Li, H. Wu, L. Chen, J. Hu, Y. Li and Y. Huang, Metal-free cascade formation of intermolecular C-N bonds accessing substituted isoindolinones under cathodic reduction, *J. Org. Chem.*, 2021, **86**, 15777–15784.
- 8 R. E. Dolle, C. MacLeod, B. Martinez-Teipel, W. Barker, P. R. Seida and T. Herbertz, Solid/solution-phase annulation reagents: single-step synthesis of cyclic amine derivatives, *Angew. Chem. Int. Ed.*, 2005, **44**, 5830–5833.
- 9 B. Fan, Z. Liu, M. Tang, Y. Xu, X. Tang and Z. Mao, Novel synthesis of azaphthalimidine hydroxylactams, *Synthetic Commun.*, 2008, **38**, 3231–3242.
- 10 T. Liu, W. Jia, Q. Xi, Y. Chen, X. Wang and D. Yin, Diversity-oriented synthesis of heterocycles: Al(OTf)₃-promoted cascade cyclization and ionic hydrogenation, *J. Org. Chem.*, 2018, **83**, 1387–1393.

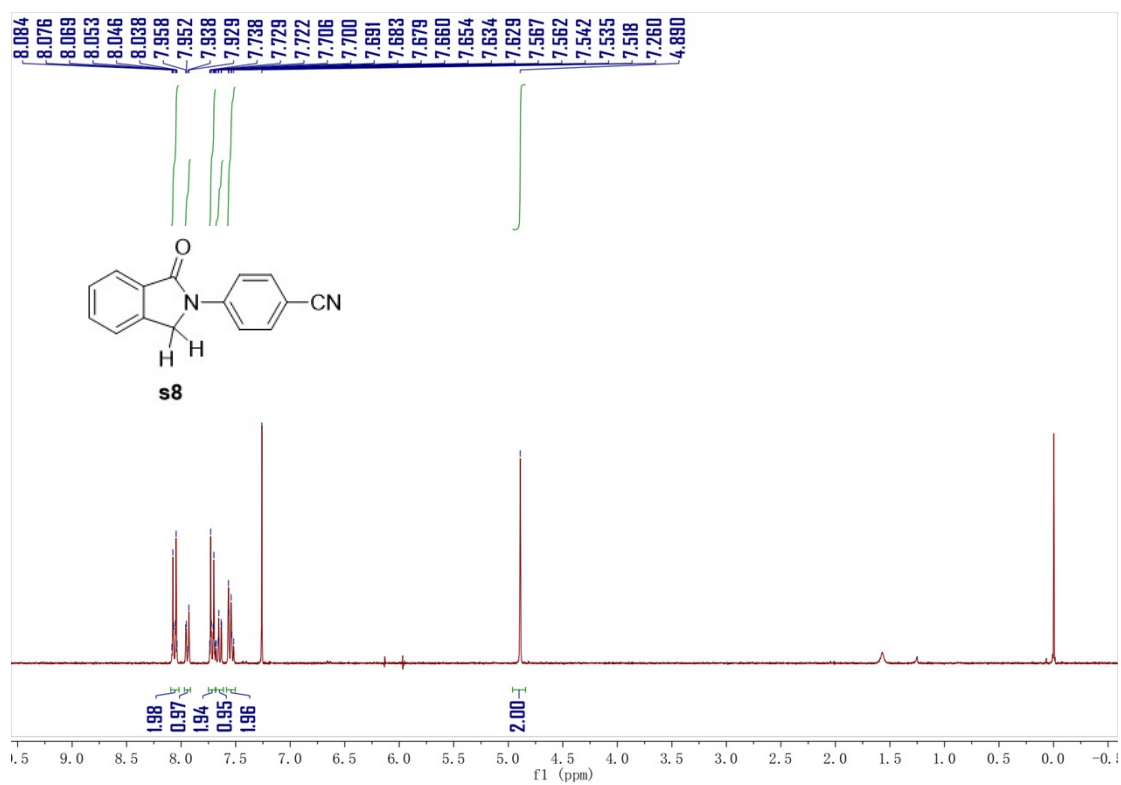
- 11 H. P. Deng, Q. Zhou and J. Wu, Microtubing-reactor-assisted aliphatic C-H functionalization with HCl as a hydrogen-atom-transfer catalyst precursor in conjunction with an organic photoredox catalyst, *Angew. Chem. Int. Ed.*, 2018, **57**, 12661–12665.
- 12 K. S. Kovaleva, O. I. Yarovaya, A. V. Shernyukov, V. V. Zarubaev, A. A. Shtro, Y. R. Orshanskaya and N. F. Salakhutdinov, Synthesis of new heterocyclic dehydroabietylamine derivatives and their biological activity, *Chem. Heterocycl. Comp.*, 2017, **53**, 364–370.
- 13 M. H. Norman, G. C. Rigdon, F. Navas III and B. R. Cooper, Cyclic benzamides as mixed dopamine D₂/serotonin 5-HT₂ receptor antagonists: potential atypical antipsychotic agents, *J. Med. Chem.*, 1994, **37**, 2552–2563.
- 14 H. Jamil, D. A. Gordon, D. C. Eustice, C. M. Brooks, J. K. Dickson, Y. Chen, B. Ricci, C. H. Chu, T. W. Harrity, C. P. Ciosek, S. A. Biller, R. E. Gregg and J. R. Wetterau, An inhibitor of the microsomal triglyceride transfer protein inhibits apoB secretion from HepG2 cells, *Proc. Natl. Acad. Sci.*, 1996, **93**, 11991–11995.
- 15 J. Clayton, F. Ma, B. V. Wagenen, R. Ukkiramapandian, L. Egle, J. Empfield, M. Isaac, A. Slassi, G. Steelman, R. Urbanek and S. Walsh, Preparation of isoindolones as metabotropic glutamate receptor potentiators, *PCT Int. Appl.*, WO 2006/020879 A1, 2006.
- 16 L. He, Y. Jiang, K. Liu, V. Gomez-Murcia, X. Ma, A. Torrecillas, Q. Chen, X. Zhu, E. Lesnefsky, J. C. Gomez-Fernandez, B. Xu and S. Zhang, Insights into the impact of a membrane-anchoring moiety on the biological activities of bivalent compounds As potential neuroprotectants for alzheimer's disease, *J. Med. Chem.*, 2018, **61**, 777–790.
- 17 M. H. Norman, D. J. Minick and G. C. Rigdon, Effect of linking bridge modifications on the antipsychotic profile of some phthalimide and isoindolinone derivatives, *J. Med. Chem.*, 1996, **39**, 149–157.

VII. Spectra

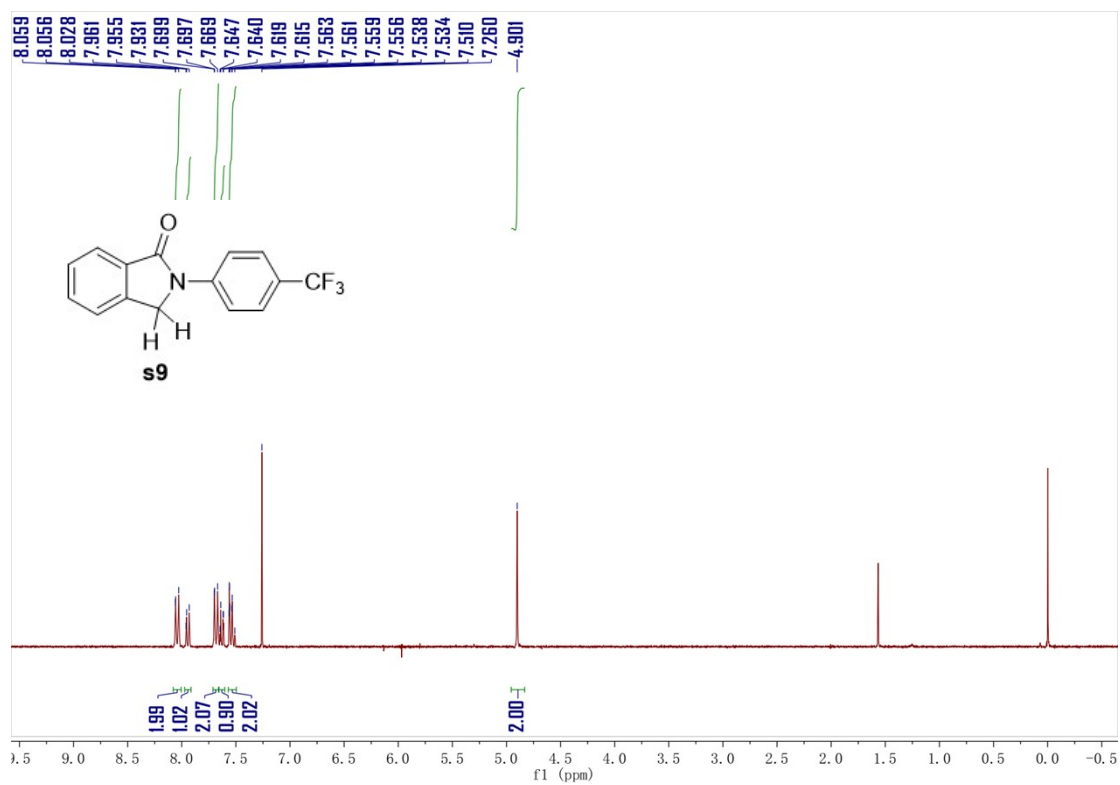
^1H NMR (300 MHz, CDCl_3):



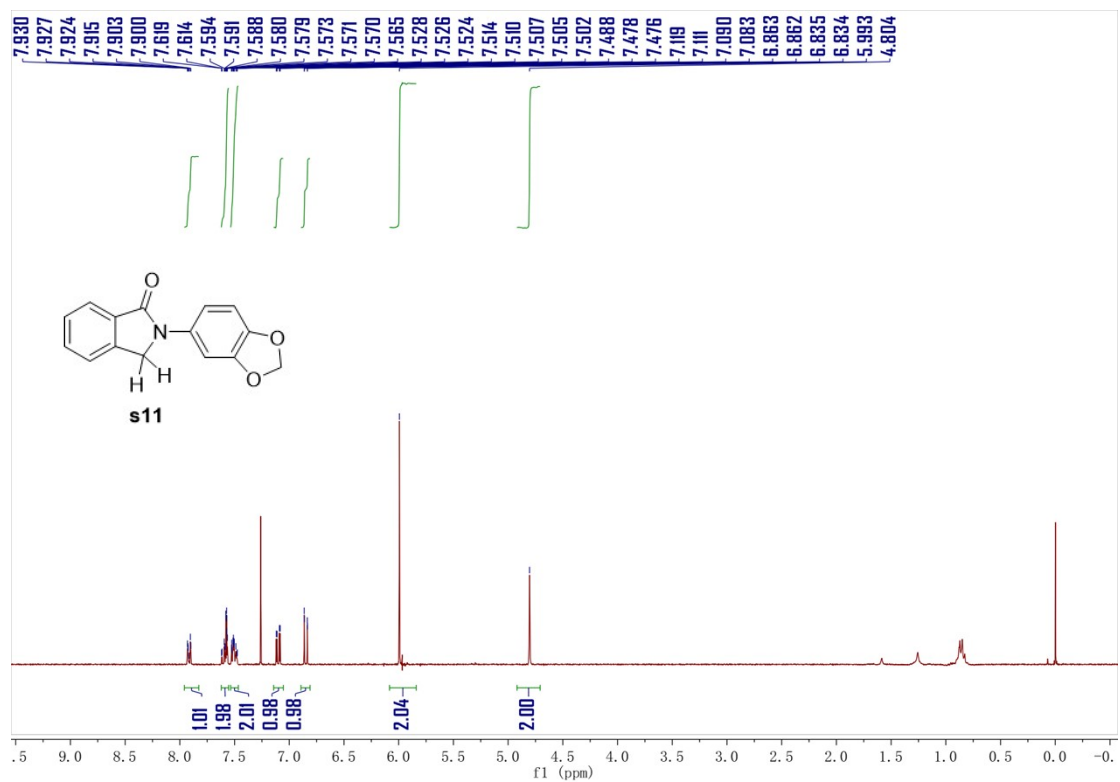
^1H NMR (300 MHz, CDCl_3):



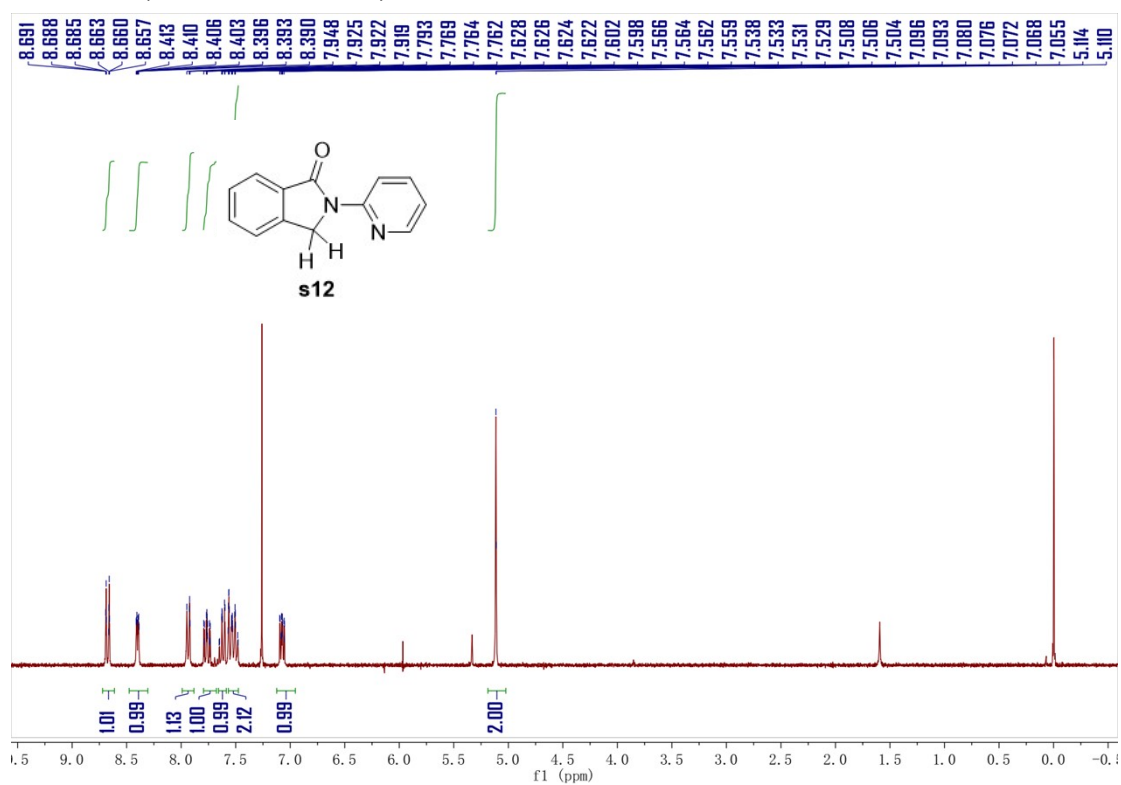
¹H NMR (300 MHz, CDCl₃):



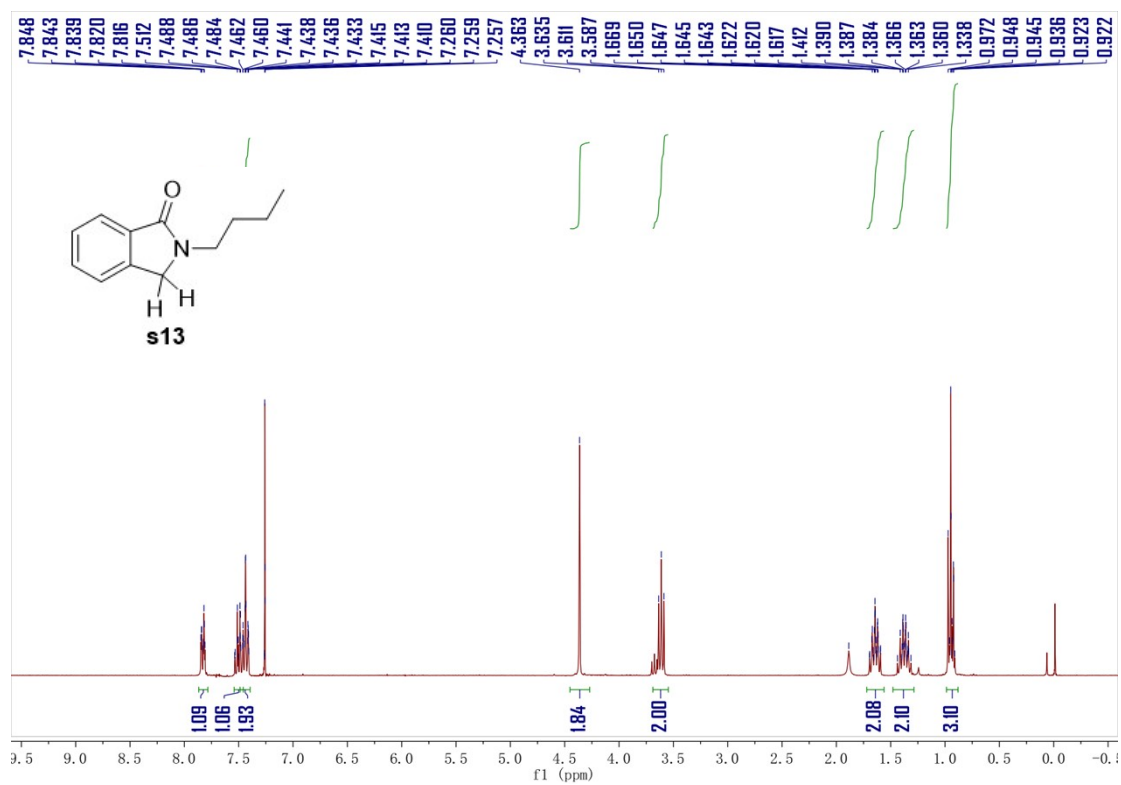
¹H NMR (300 MHz, CDCl₃):



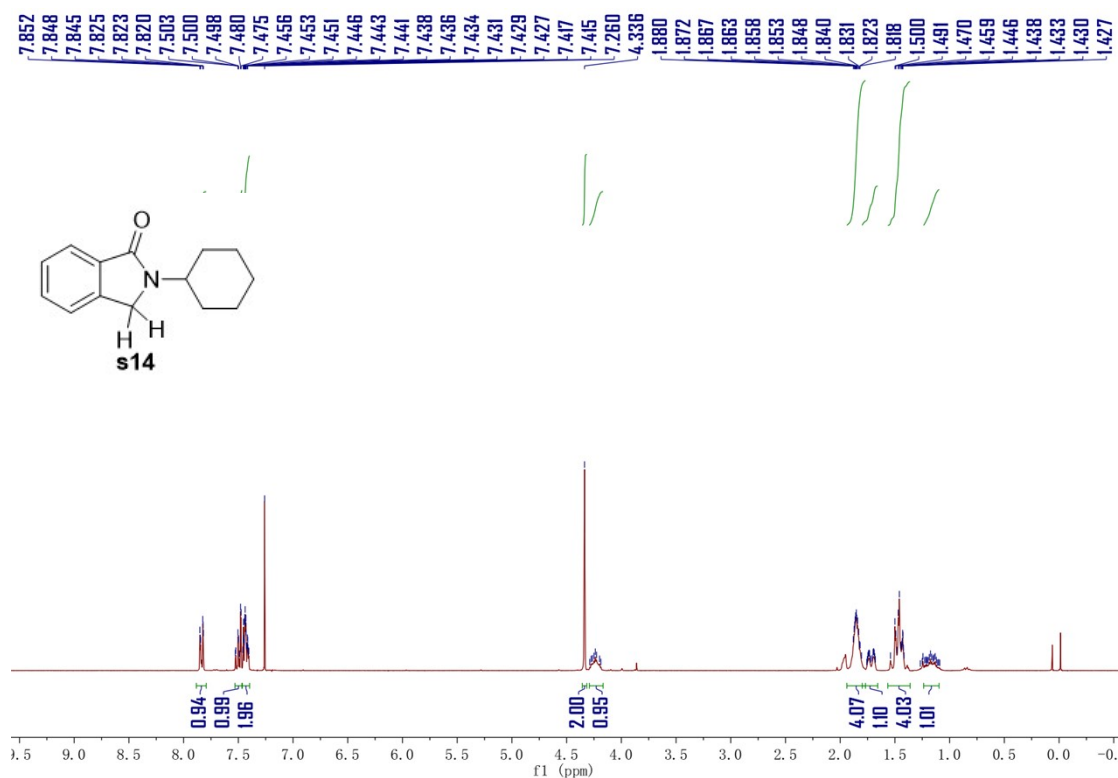
¹H NMR (300 MHz, CDCl₃):



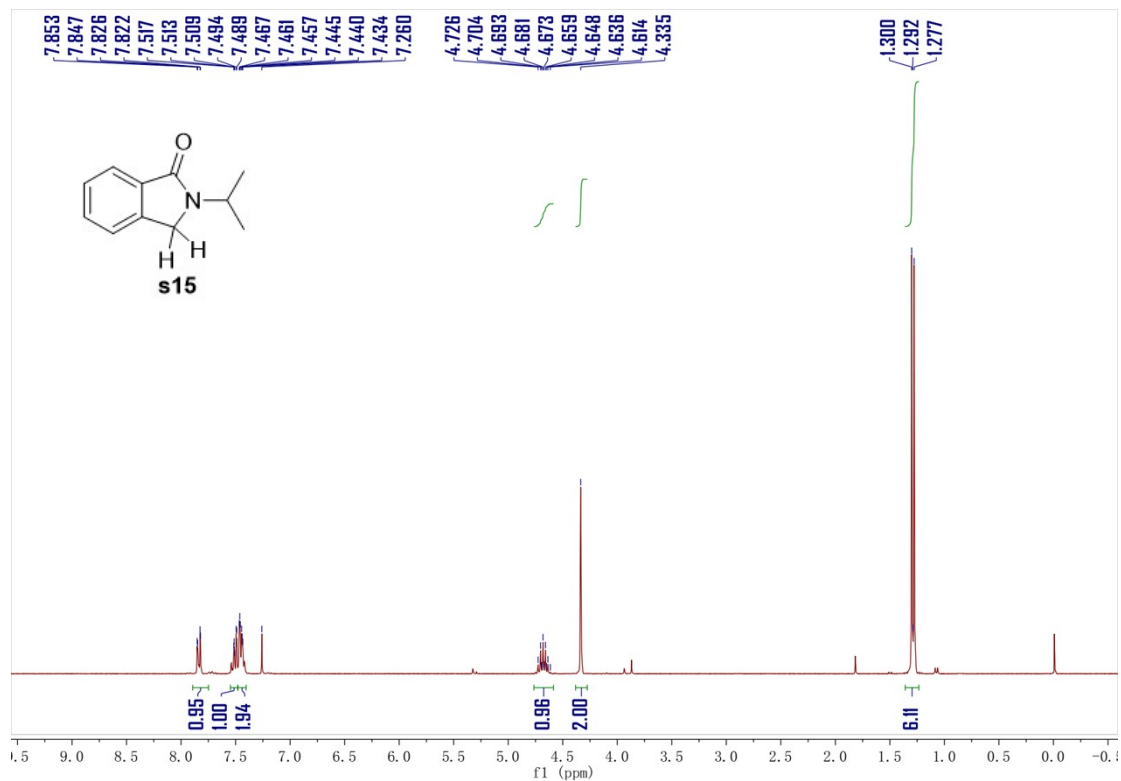
¹H NMR (300 MHz, CDCl₃):



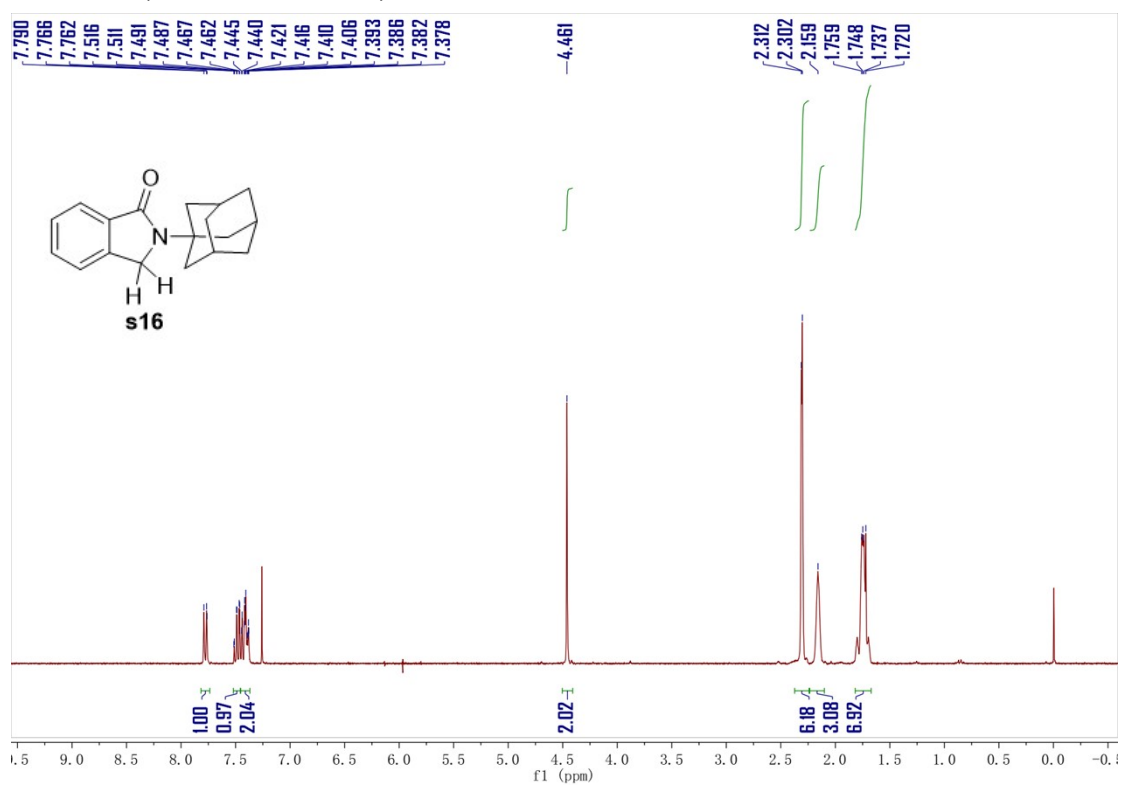
¹H NMR (300 MHz, CDCl₃):



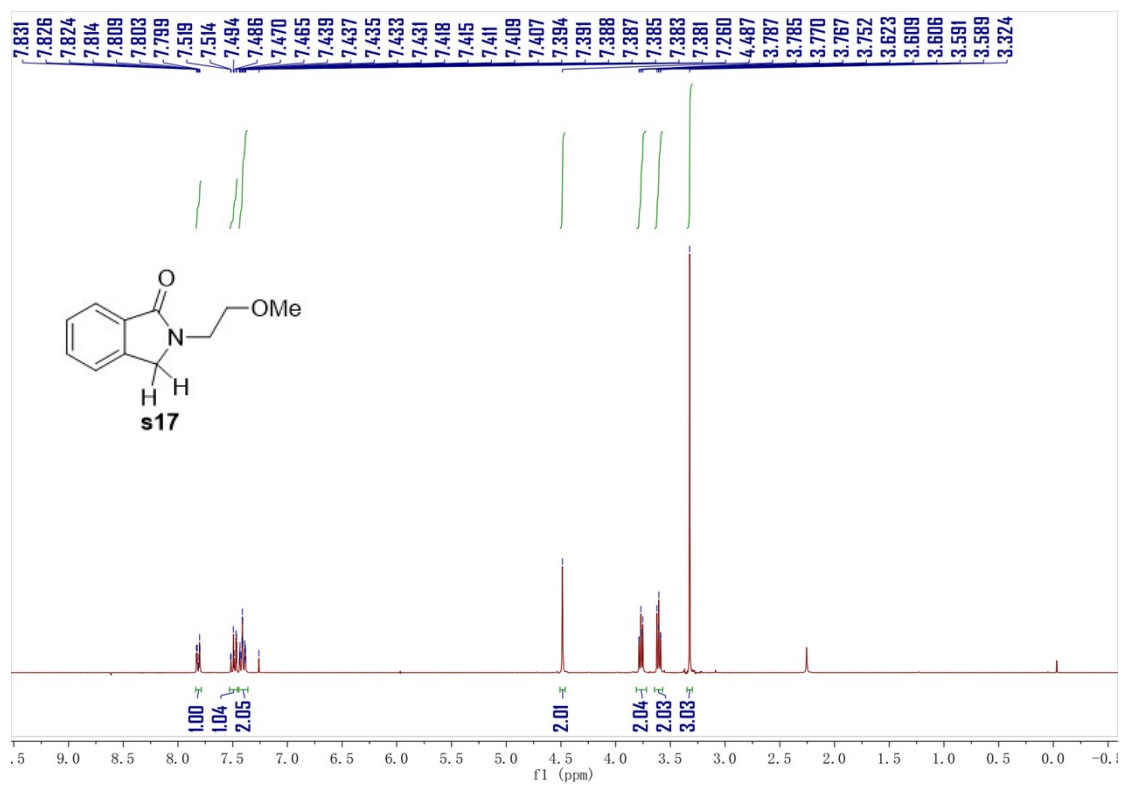
¹H NMR (300 MHz, CDCl₃):



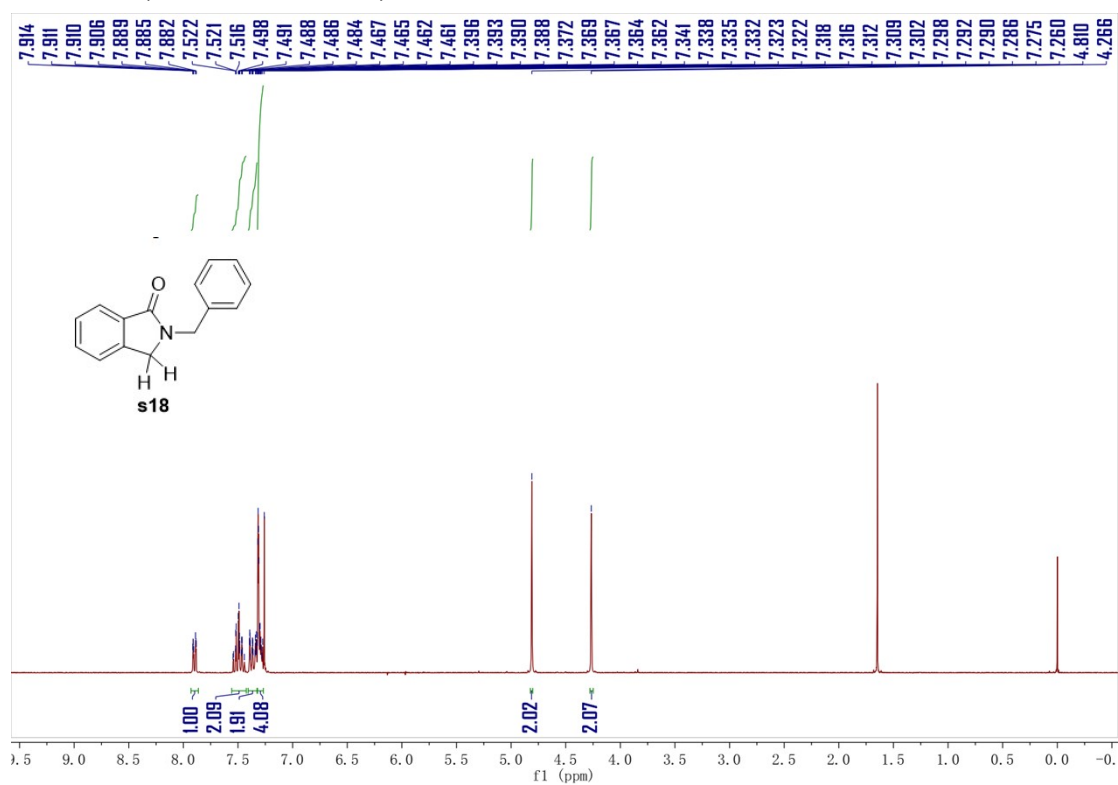
¹H NMR (300 MHz, CDCl₃):



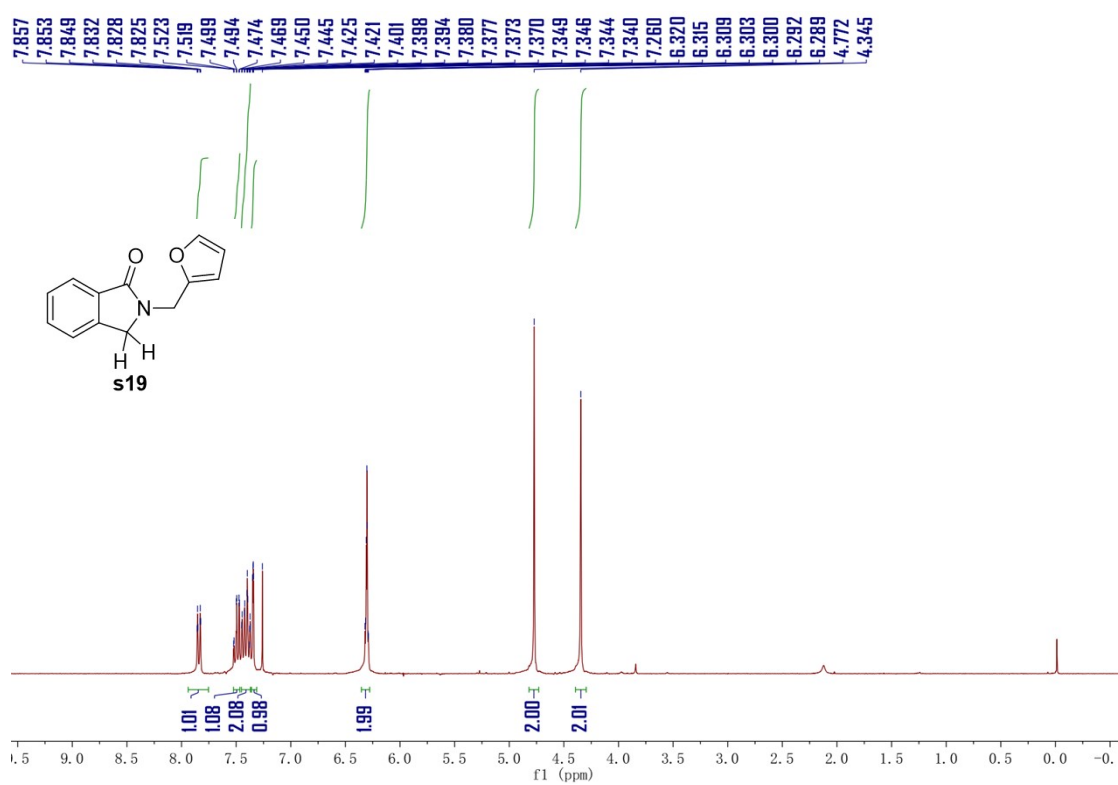
¹H NMR (300 MHz, CDCl₃):



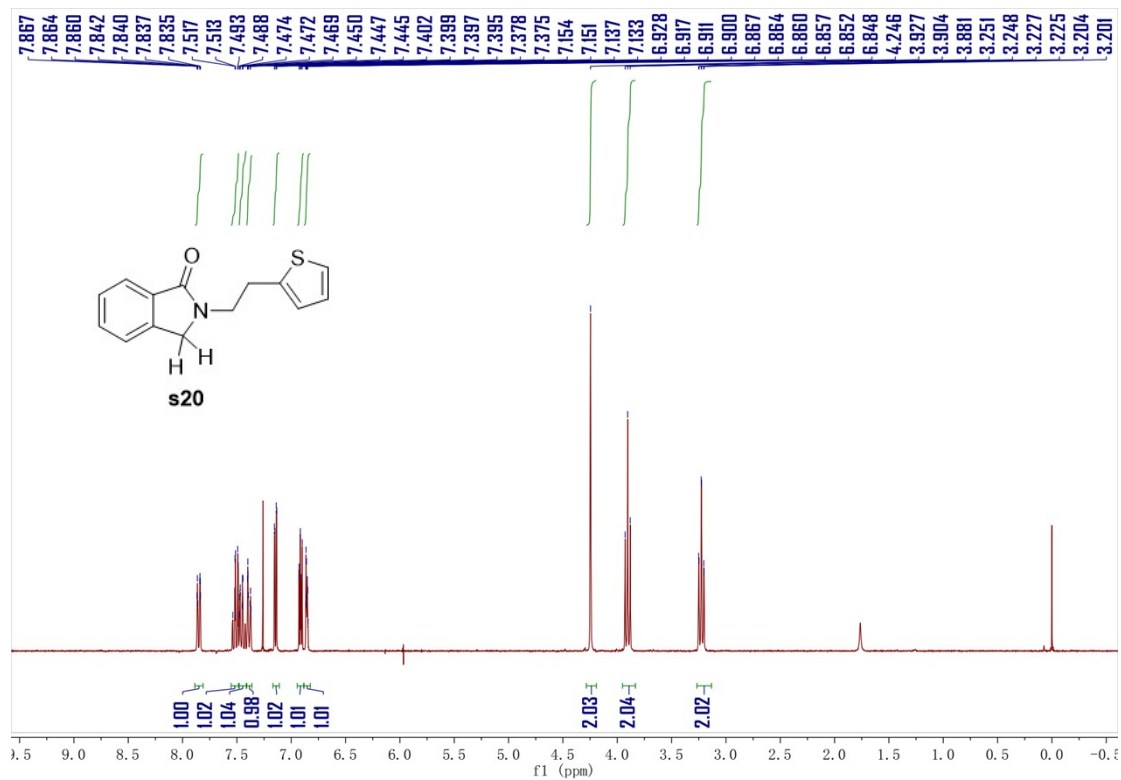
¹H NMR (300 MHz, CDCl₃):



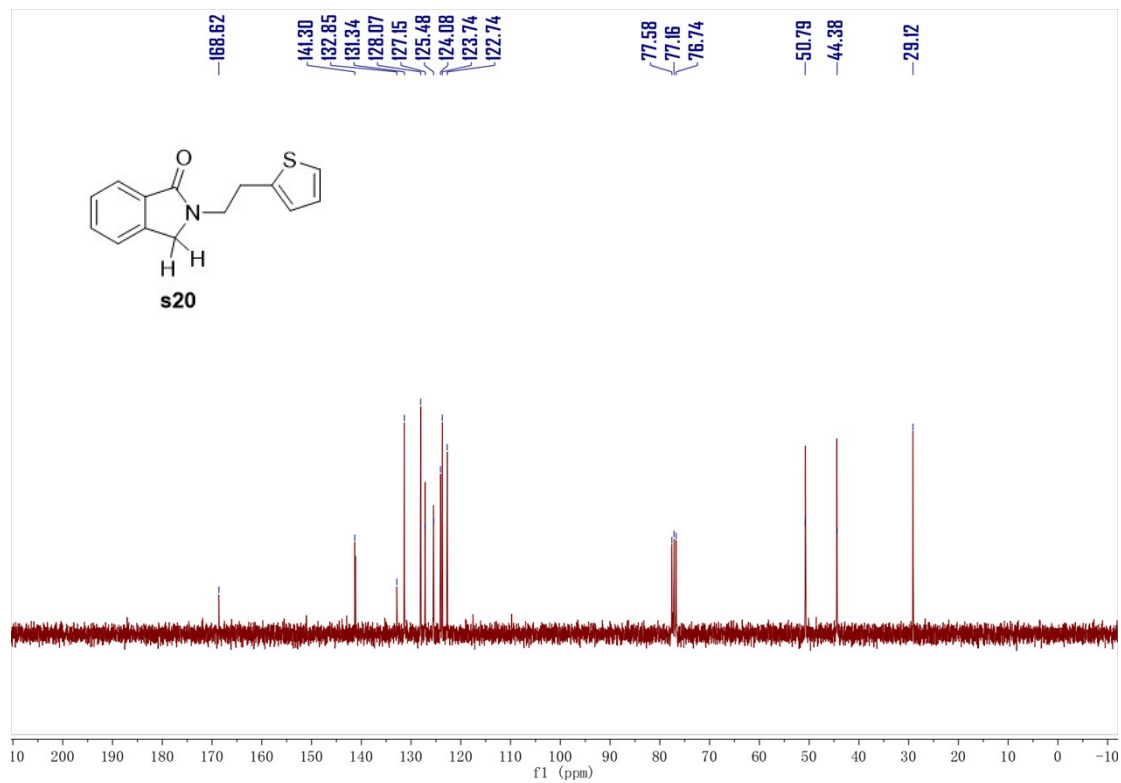
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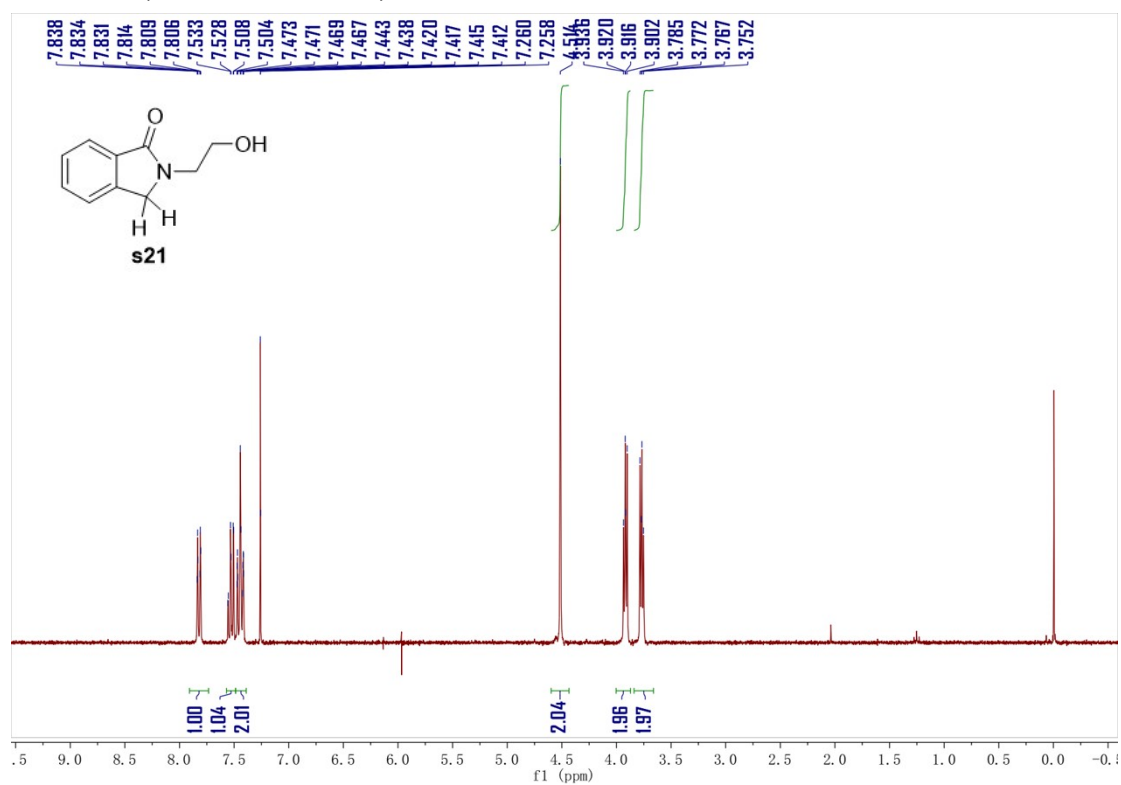
¹H NMR (300 MHz, CDCl₃):



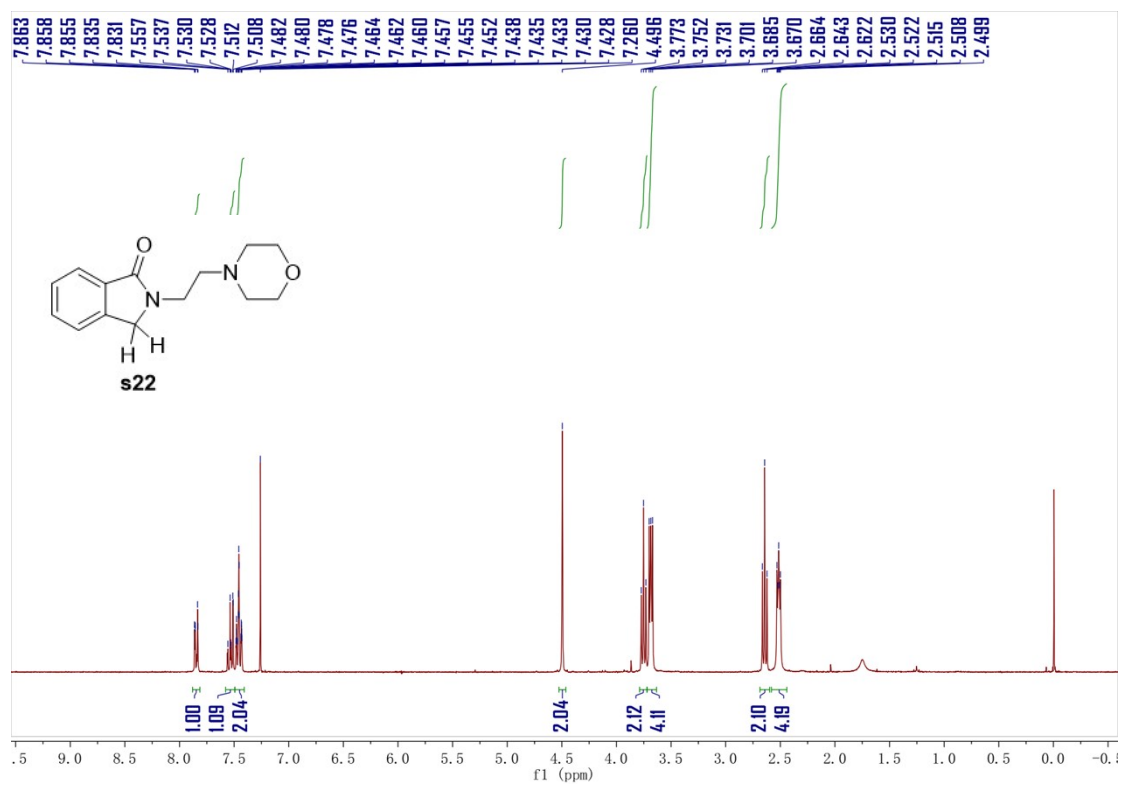
¹³C NMR (75 MHz, CDCl₃):



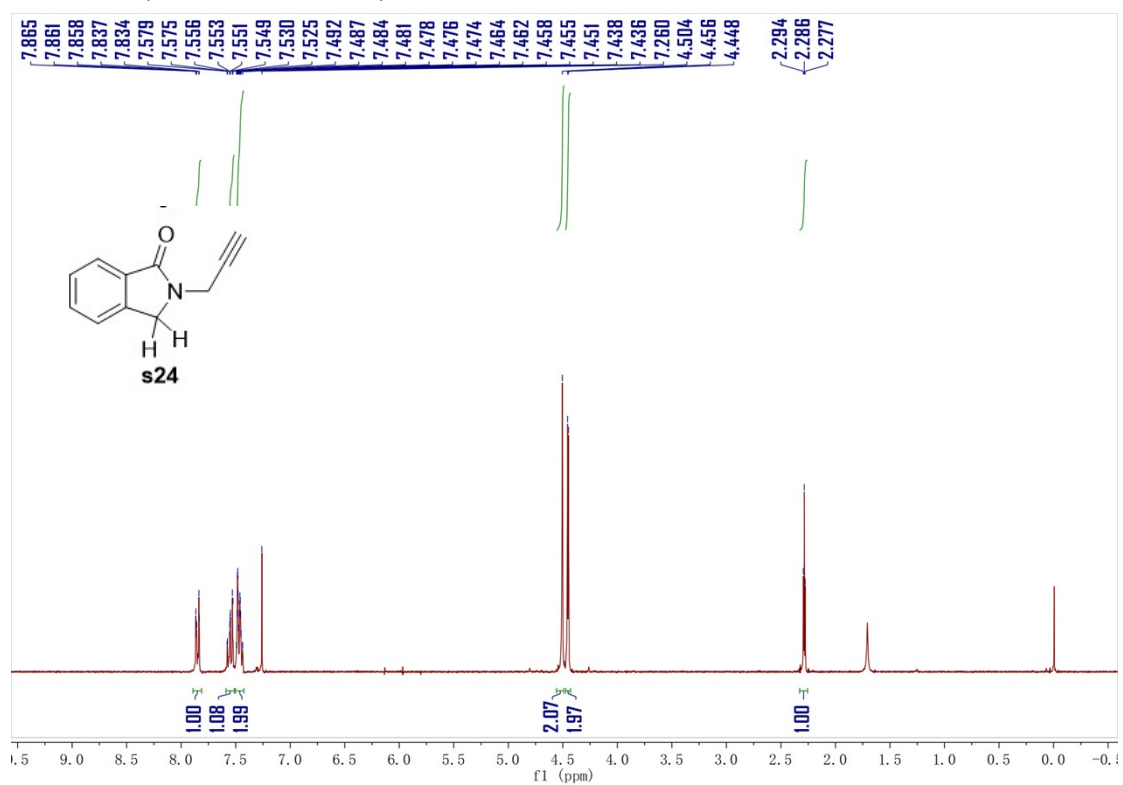
¹H NMR (300 MHz, CDCl₃):



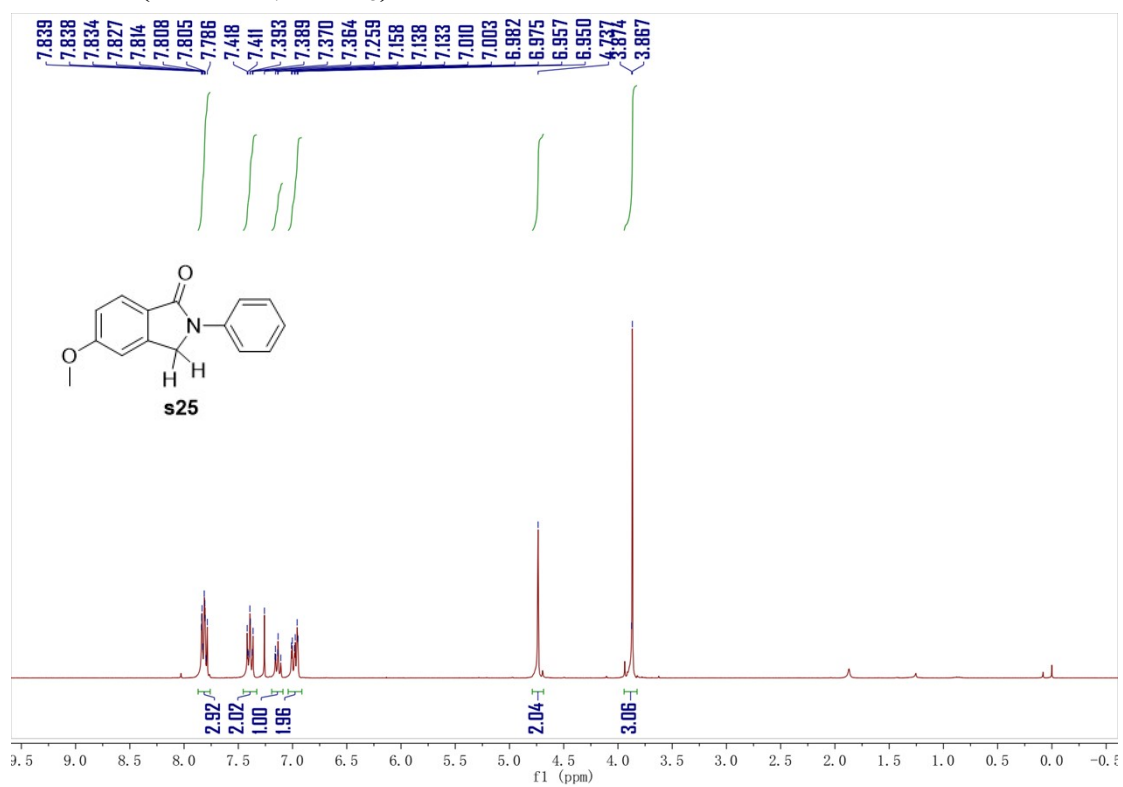
¹H NMR (300 MHz, CDCl₃):



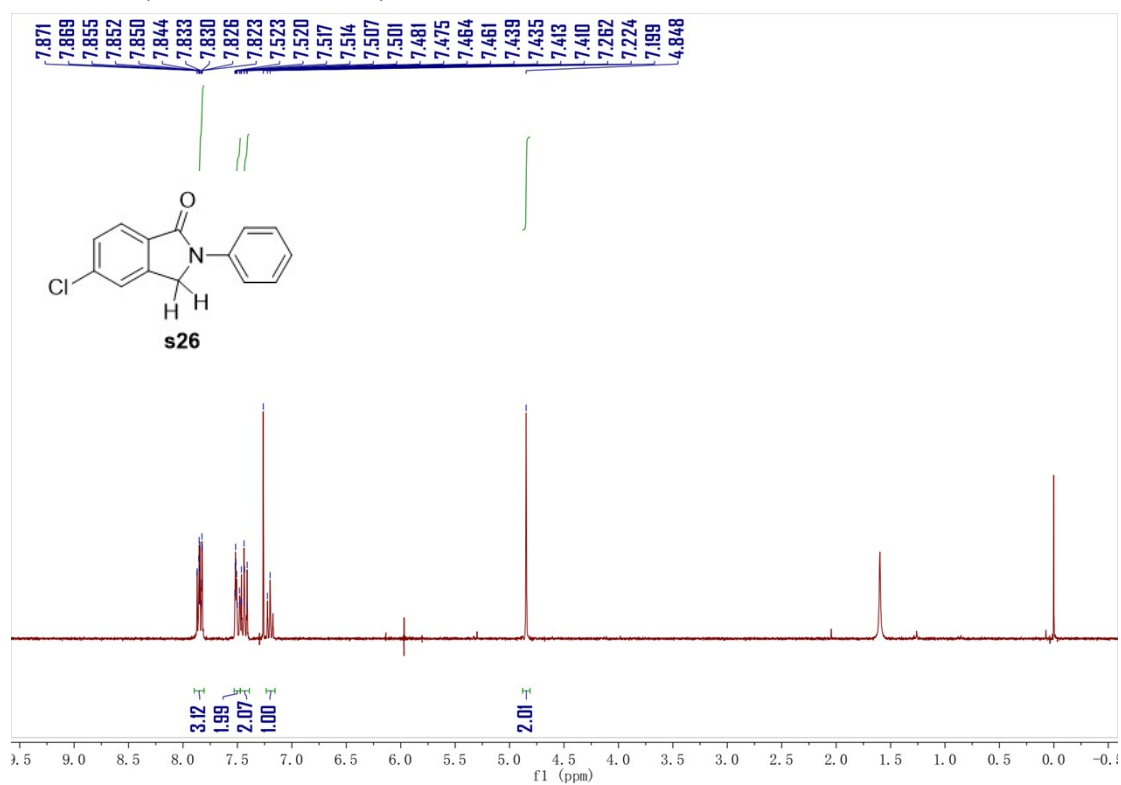
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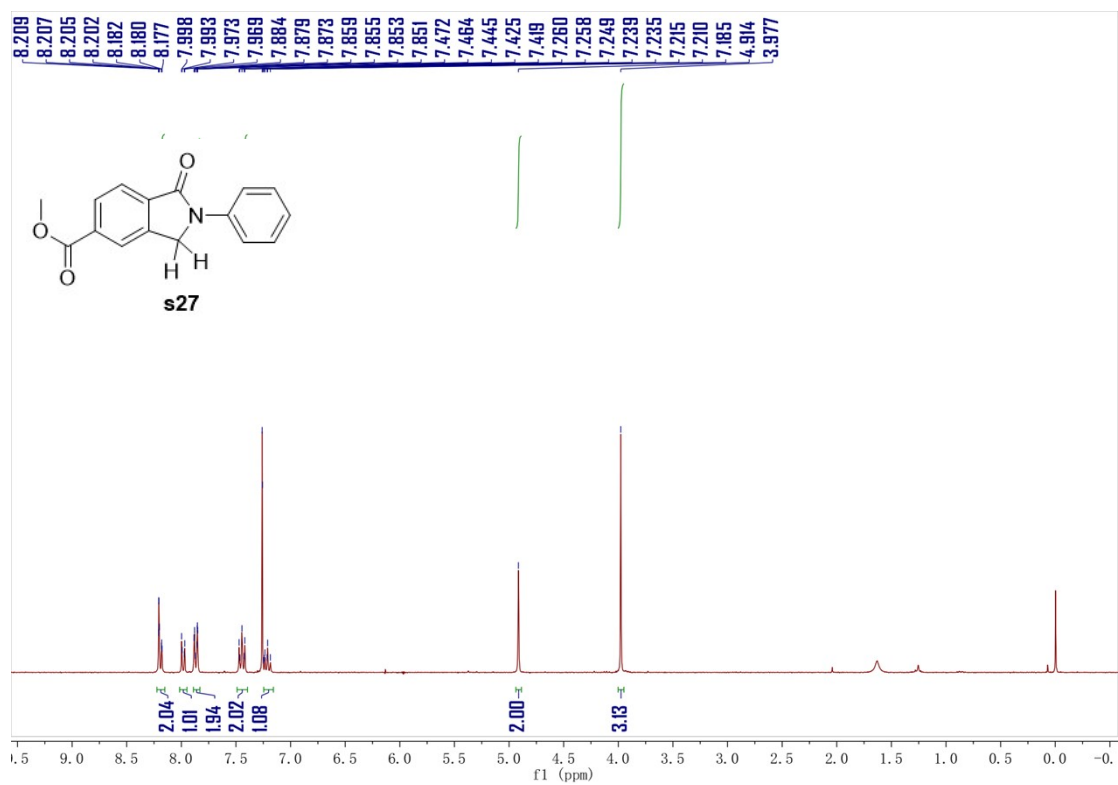
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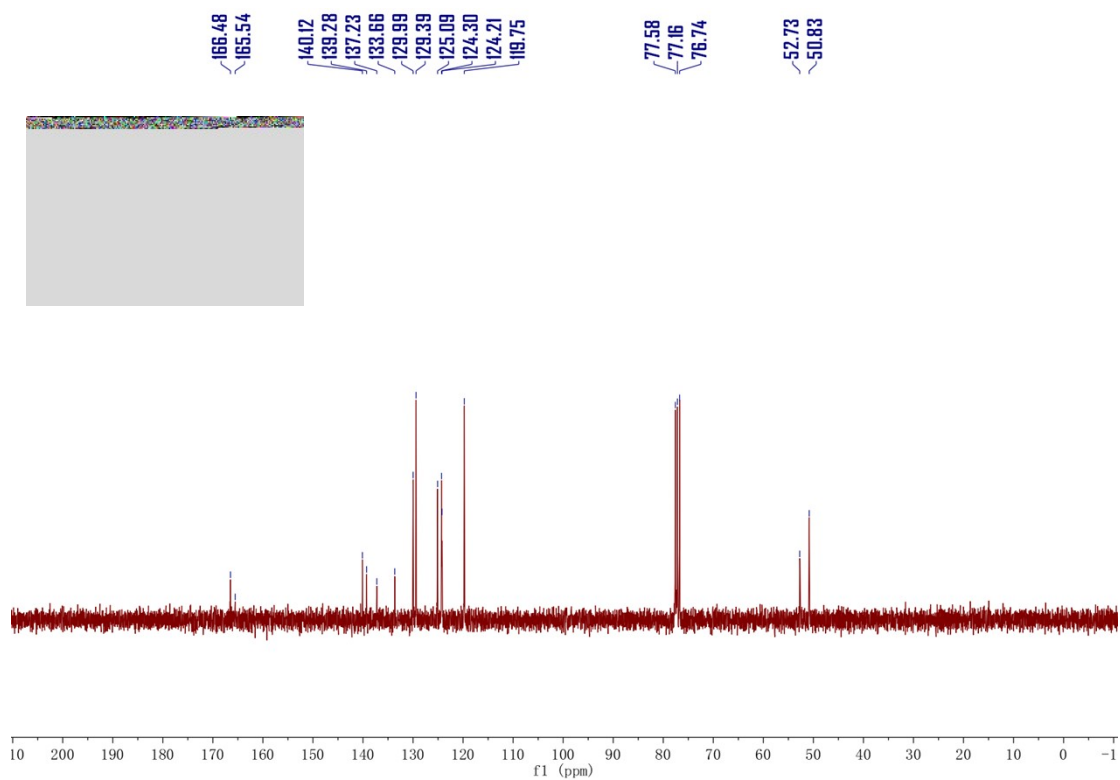
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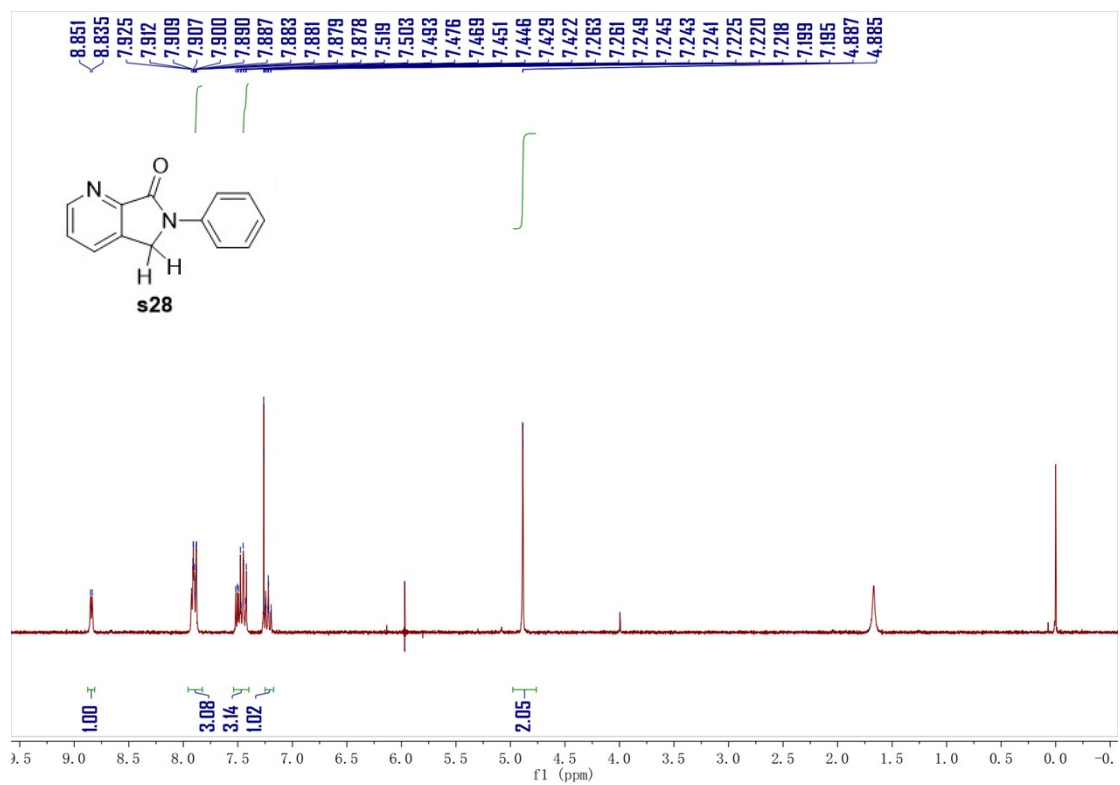
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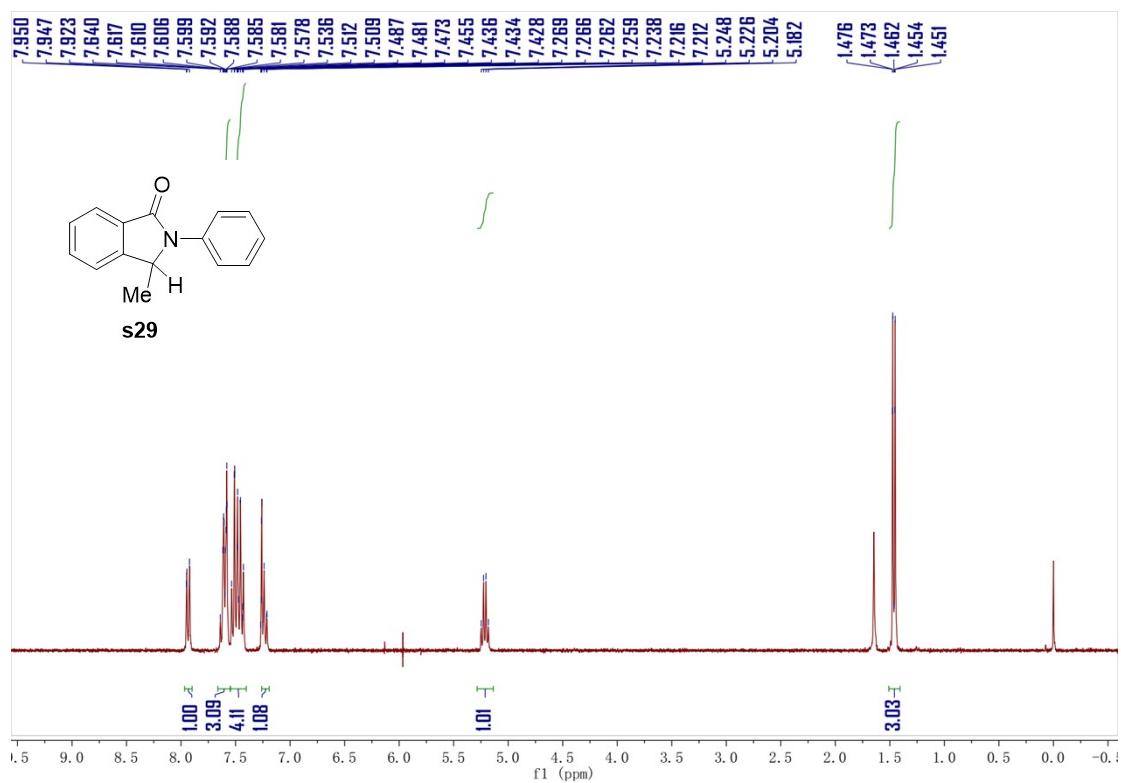
¹³C NMR (75 MHz, CDCl₃):



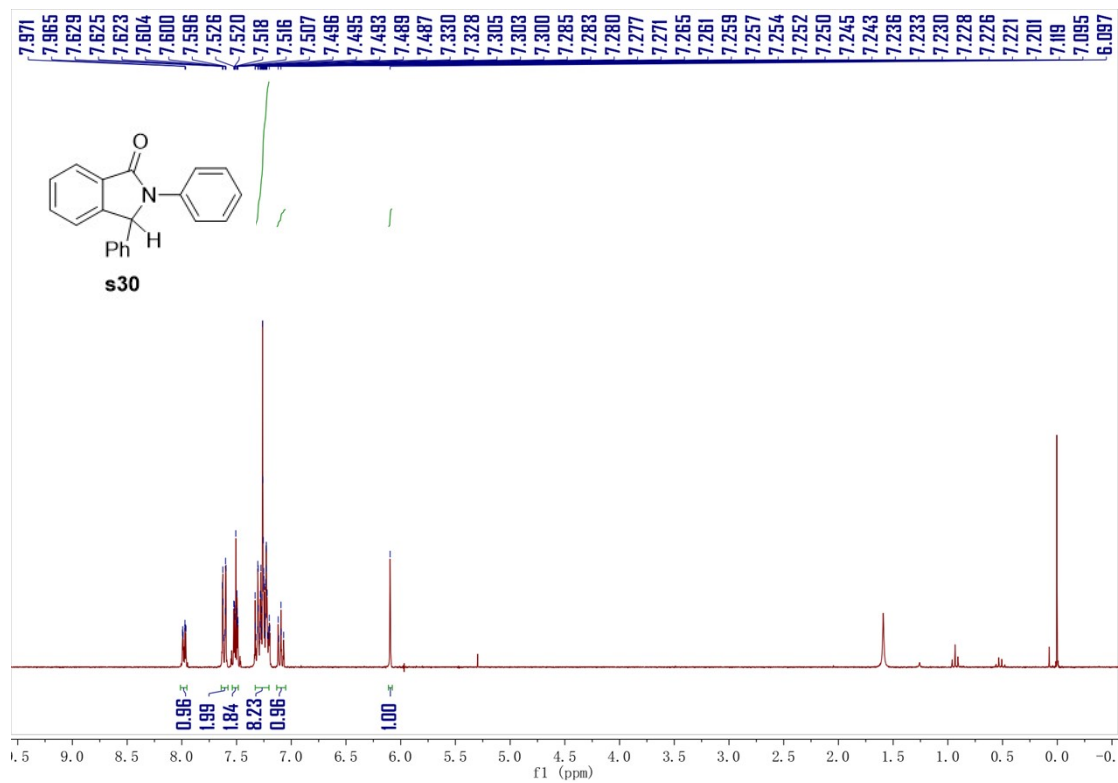
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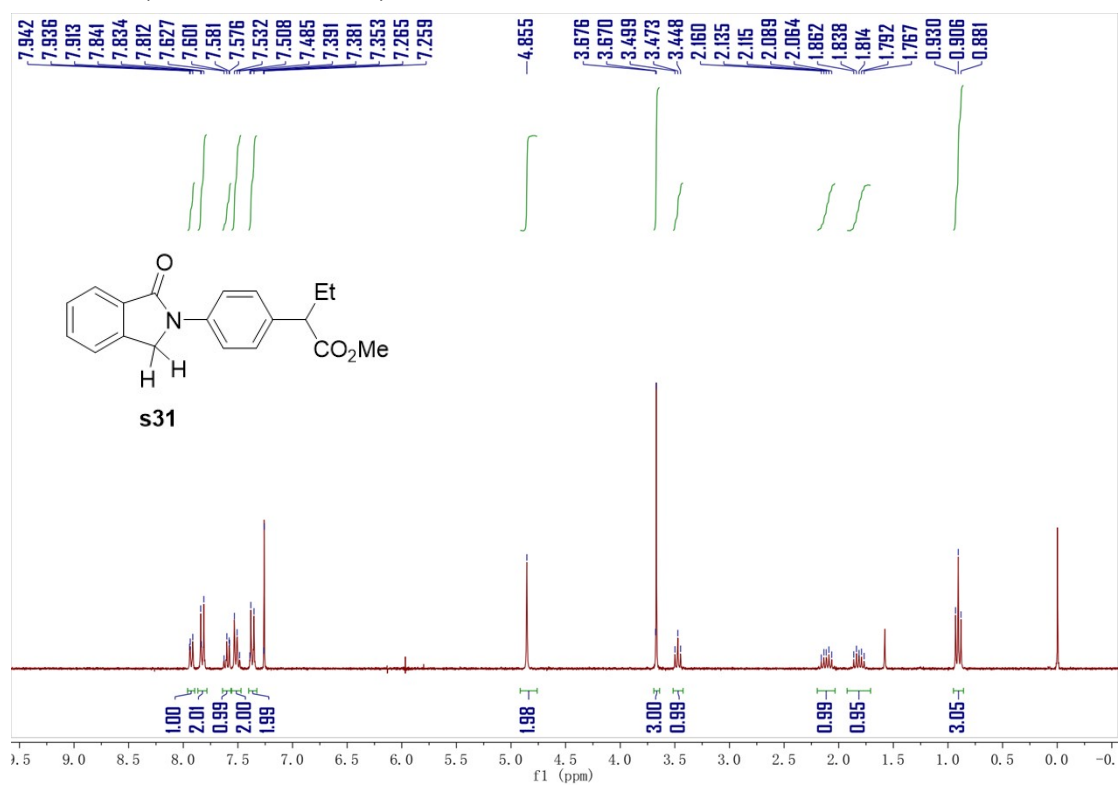
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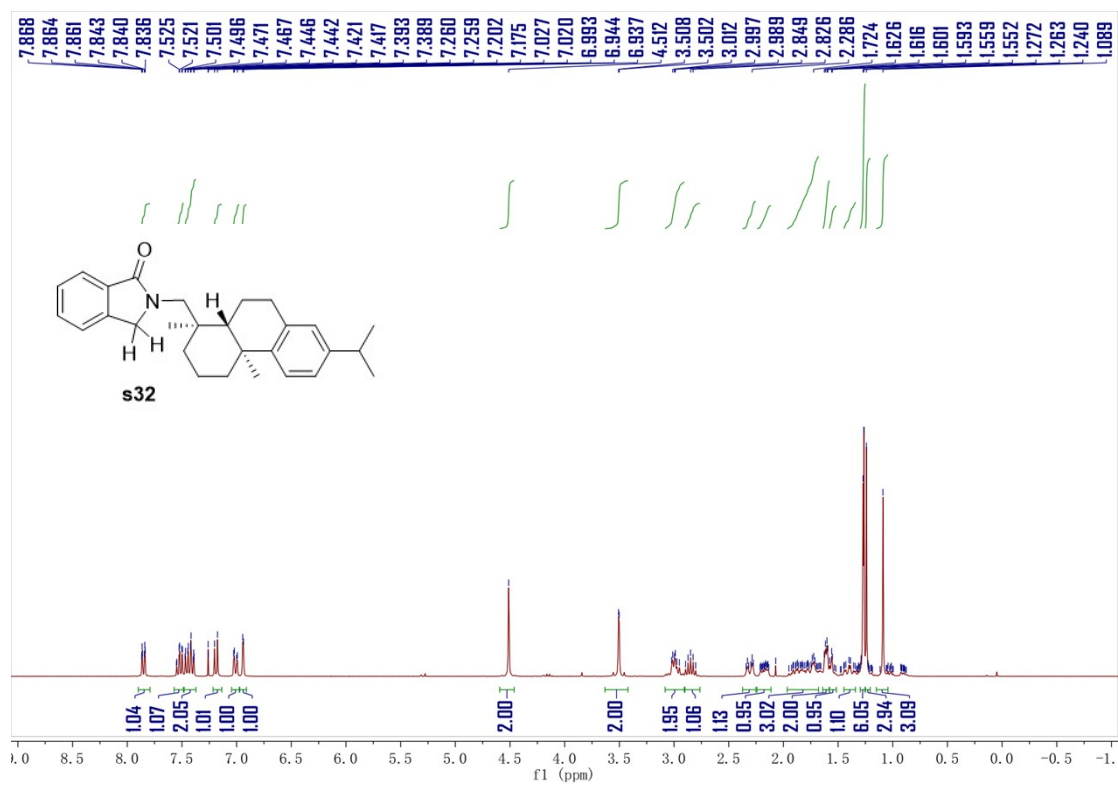
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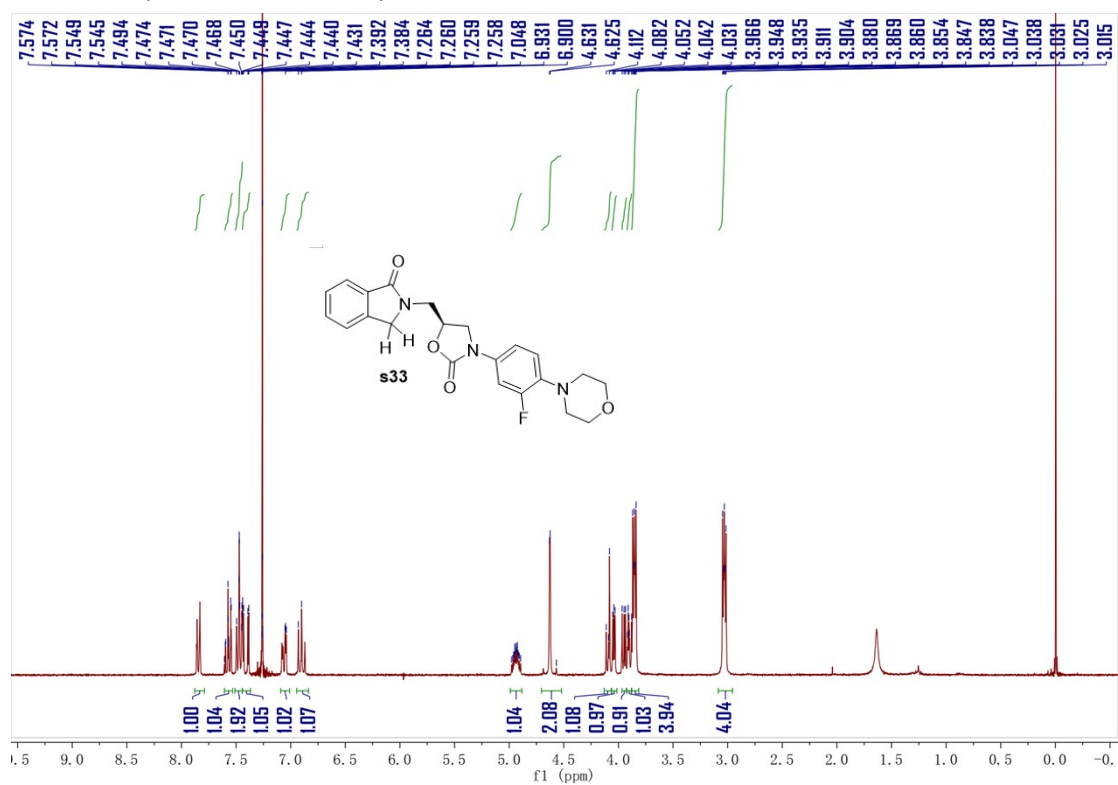
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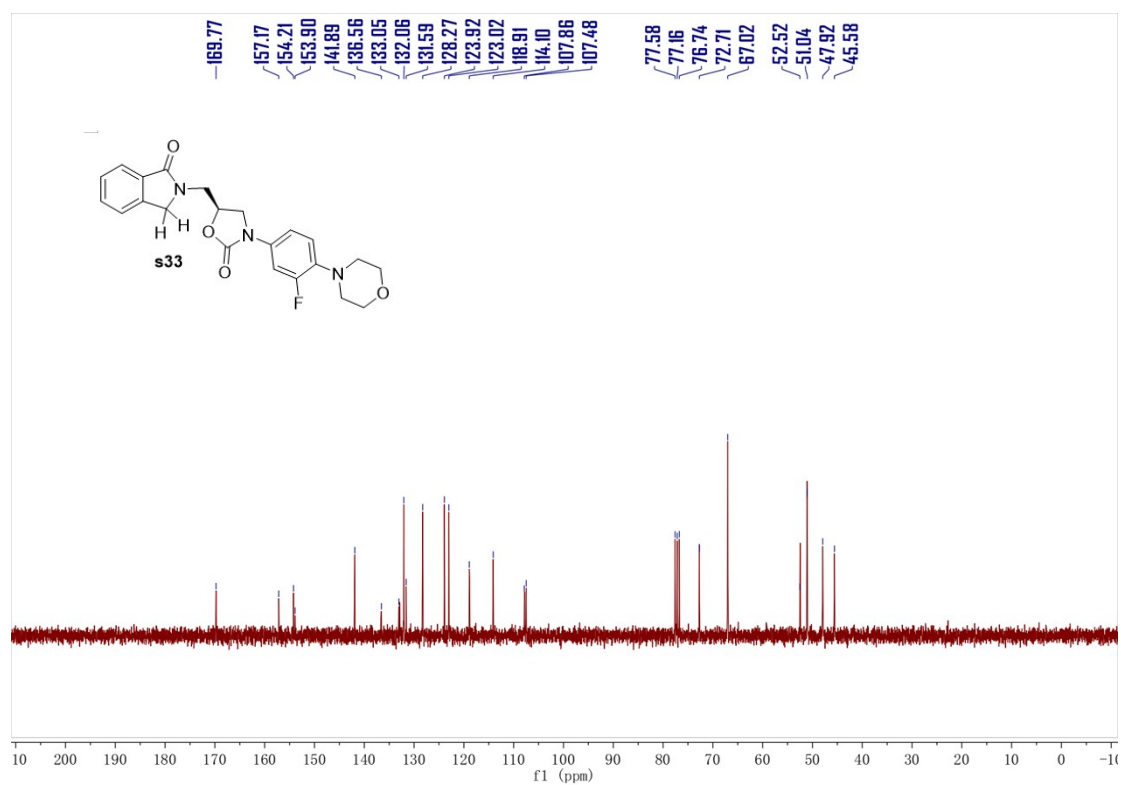
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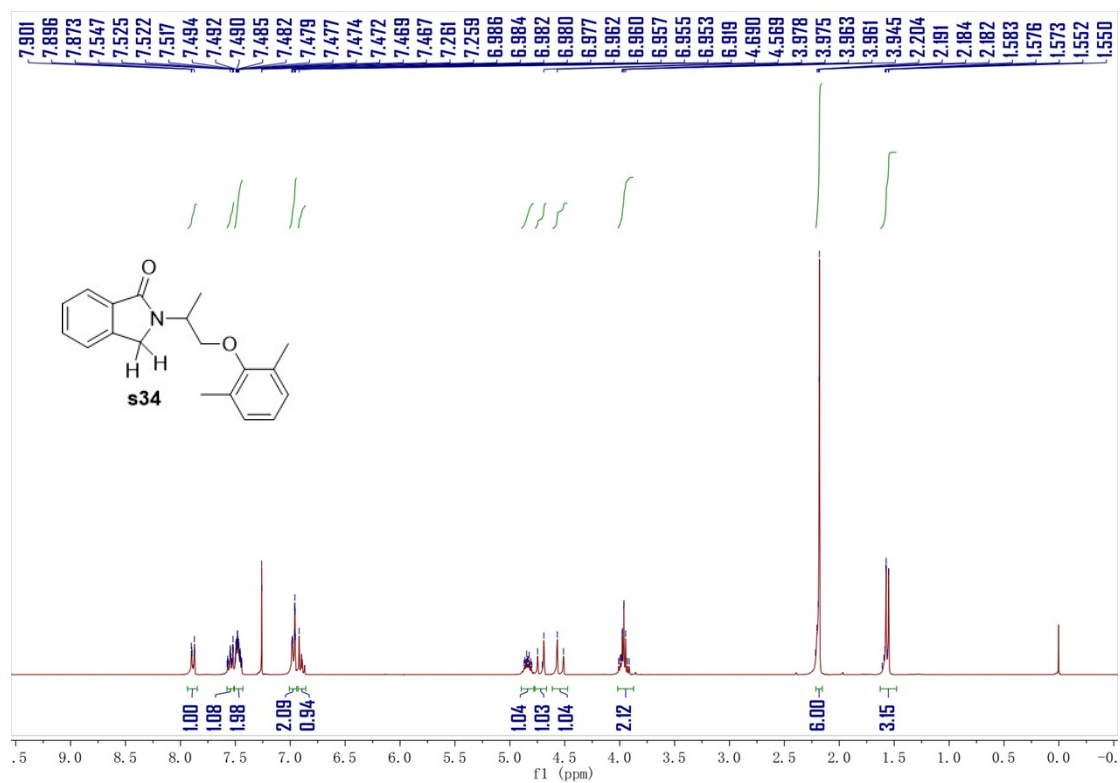
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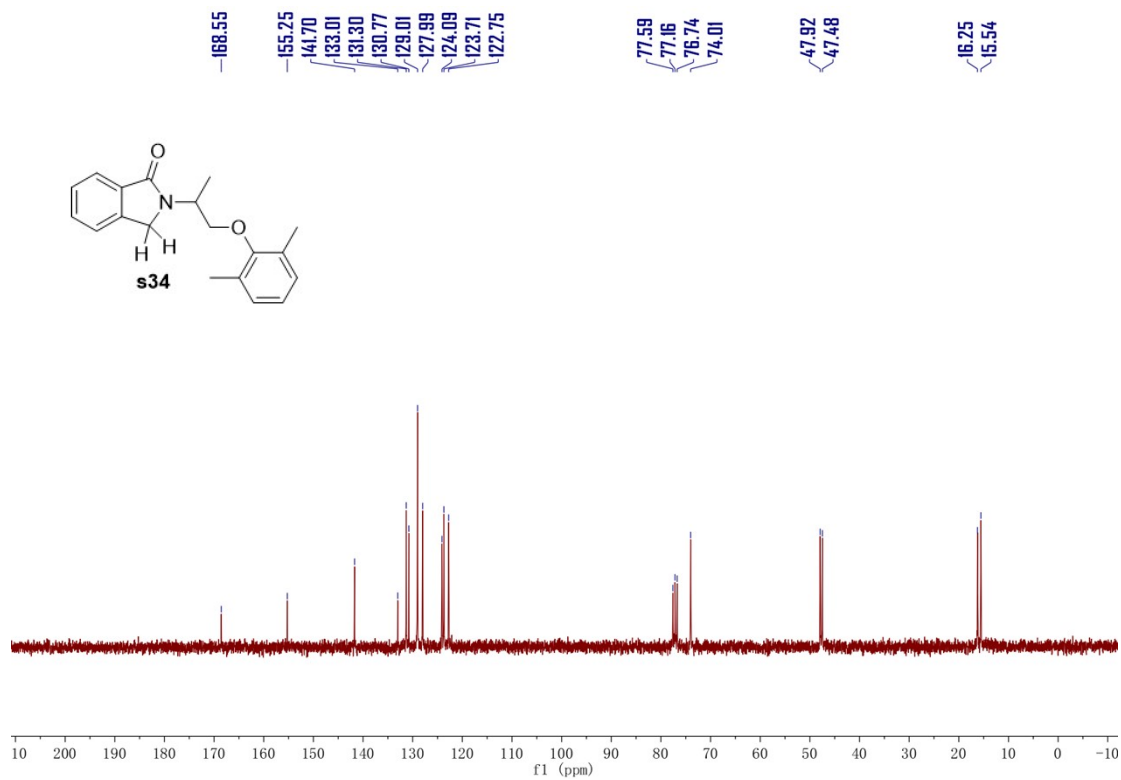
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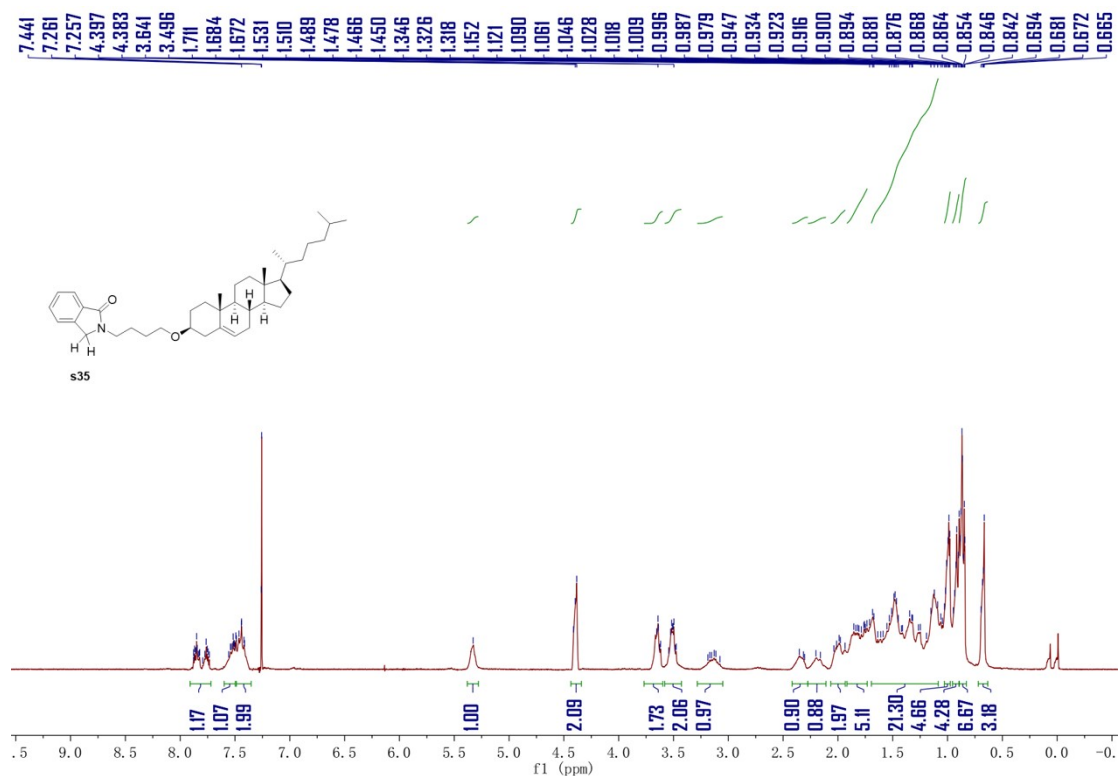
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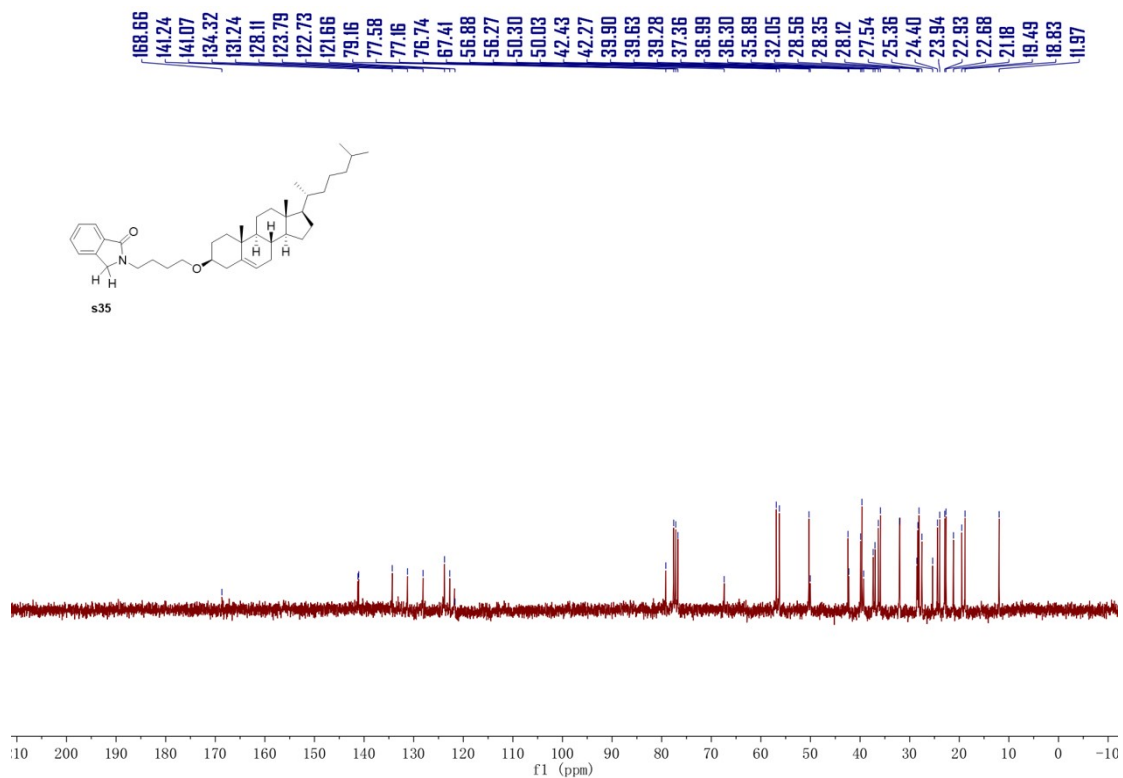
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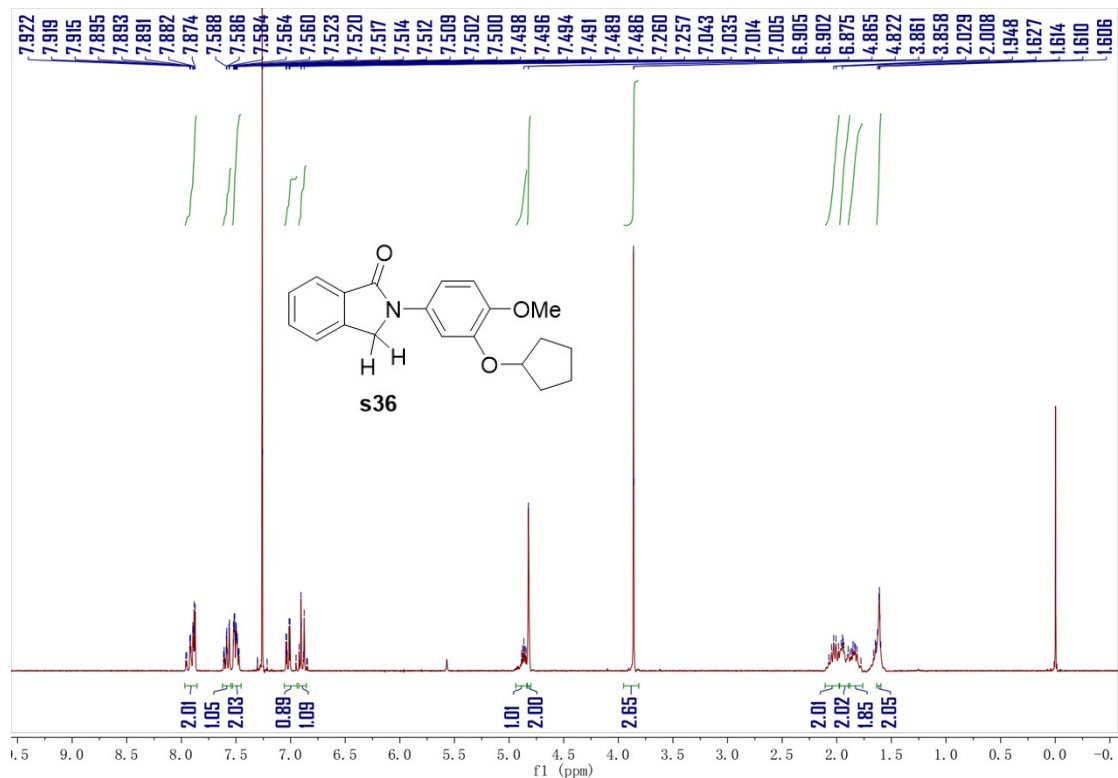
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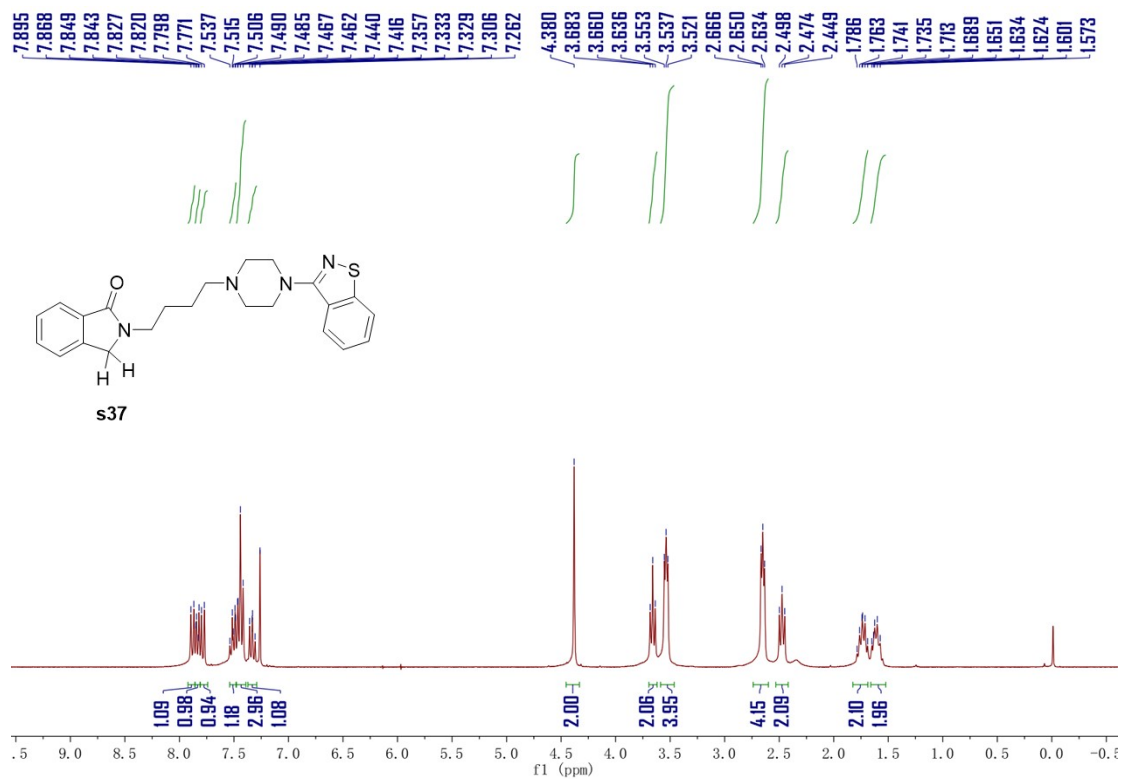
¹³C NMR (75 MHz, CDCl₃):



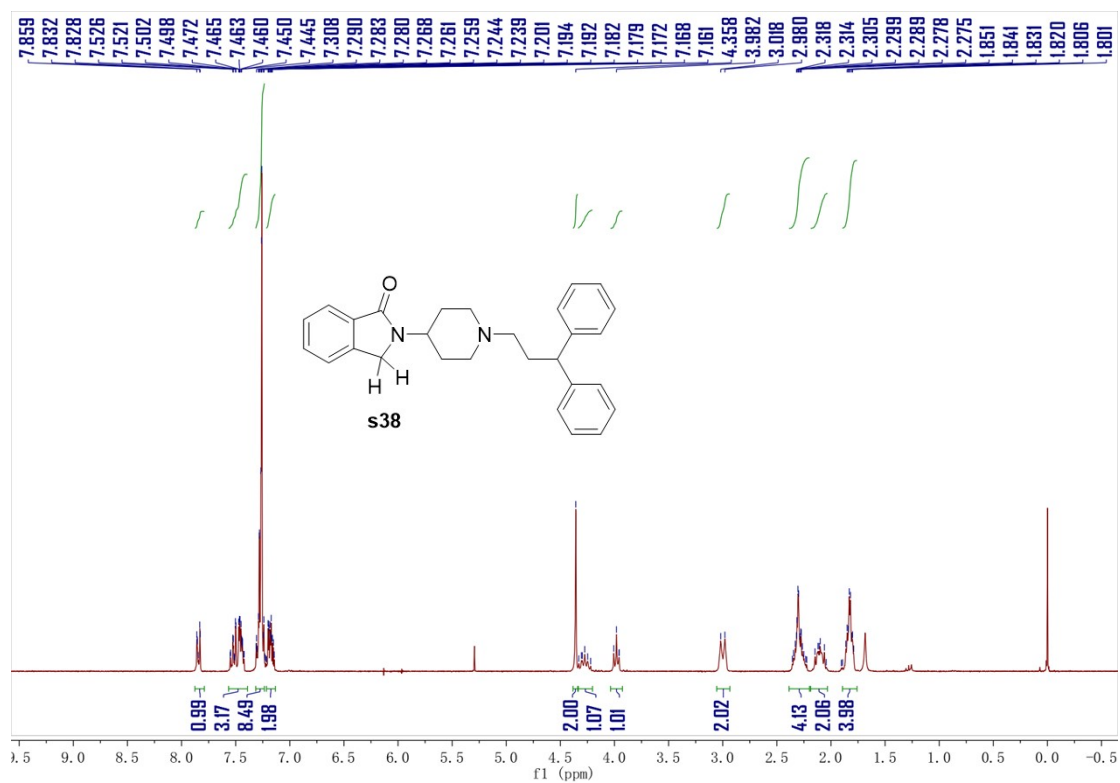
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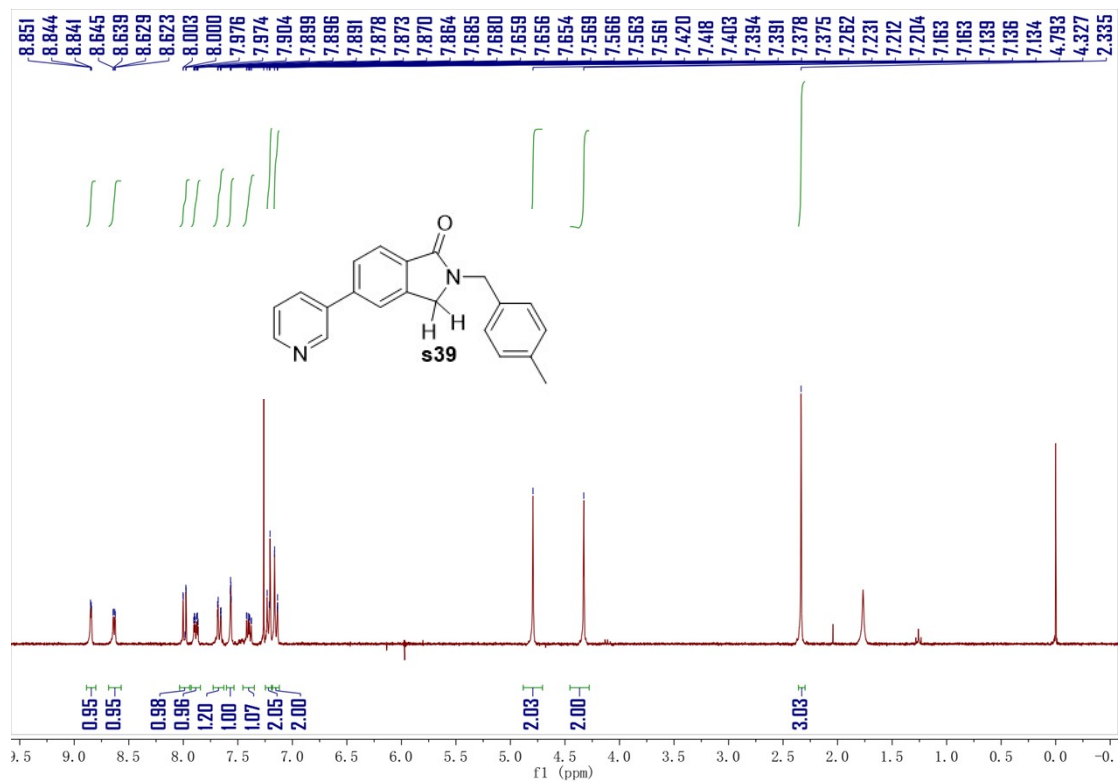
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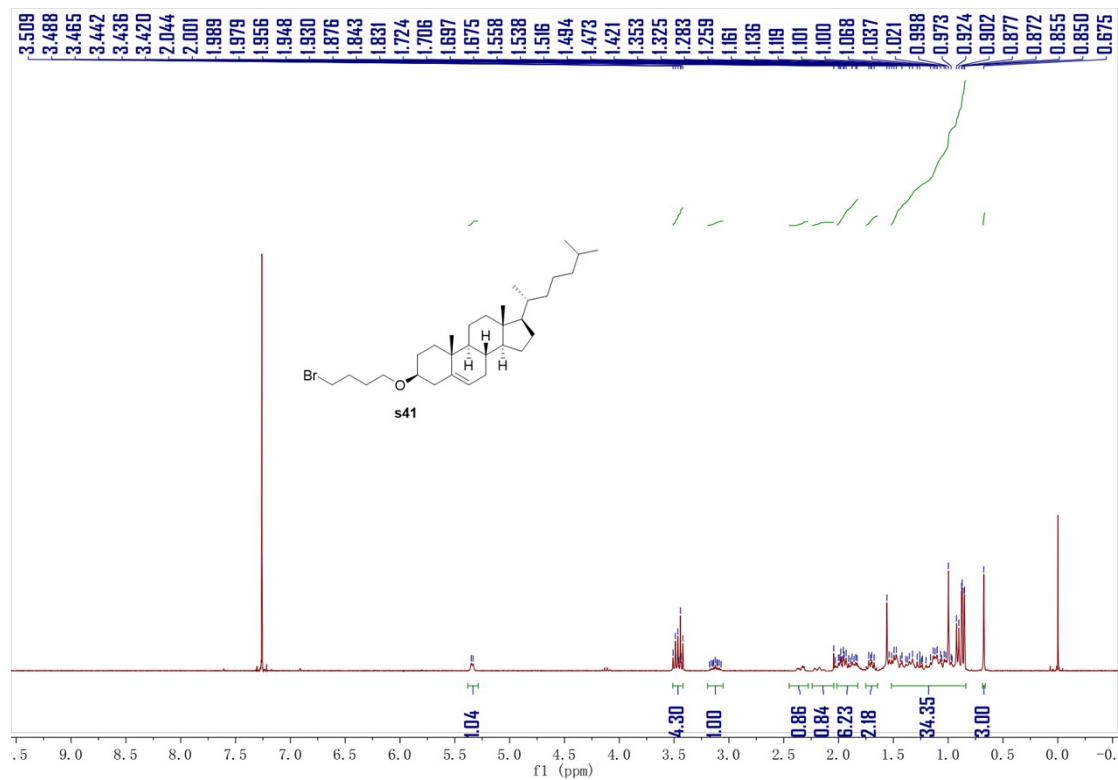
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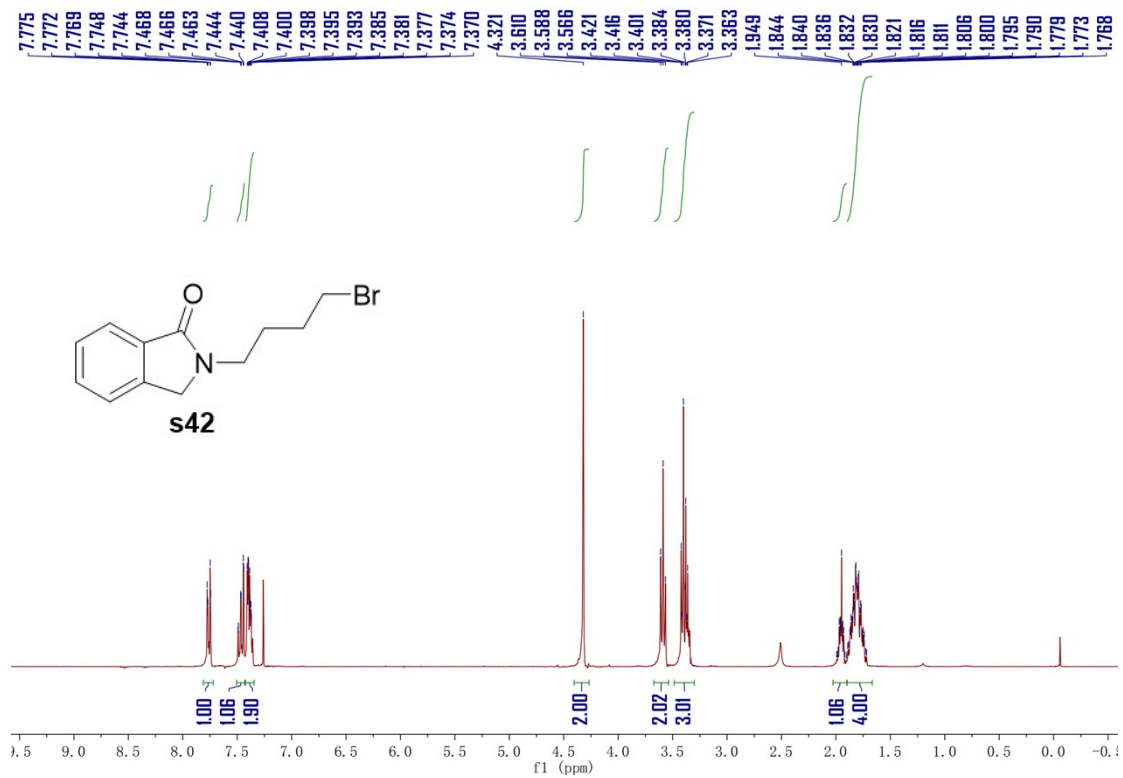
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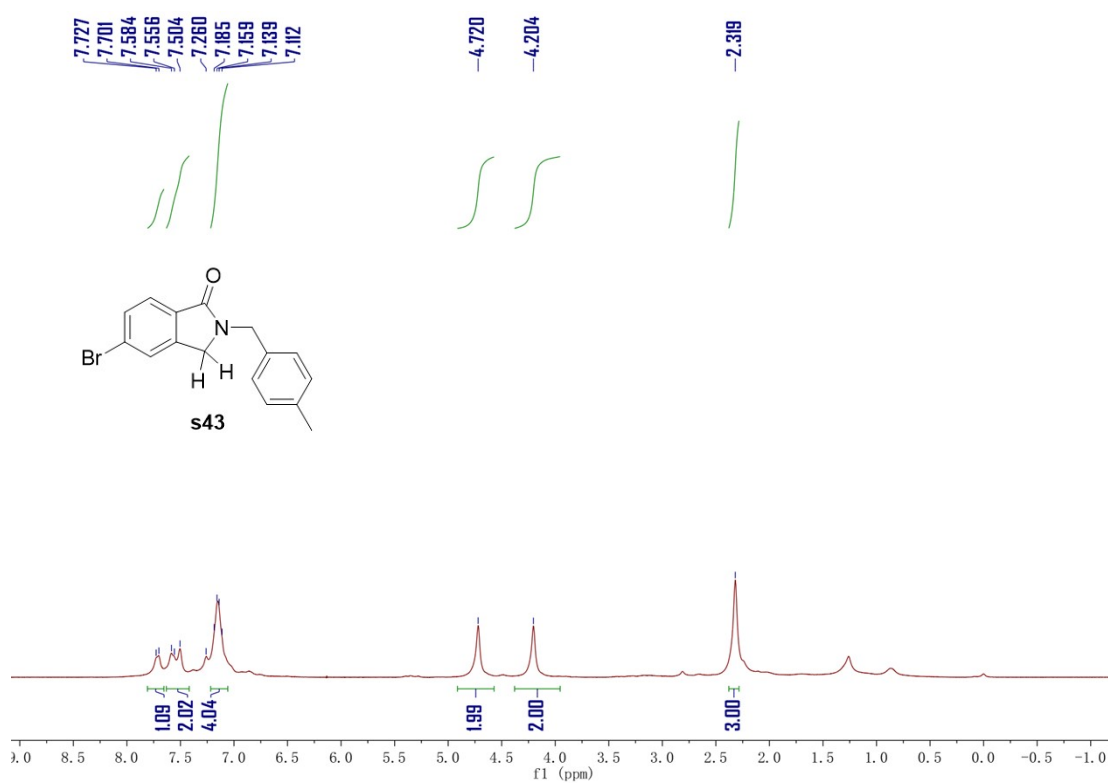
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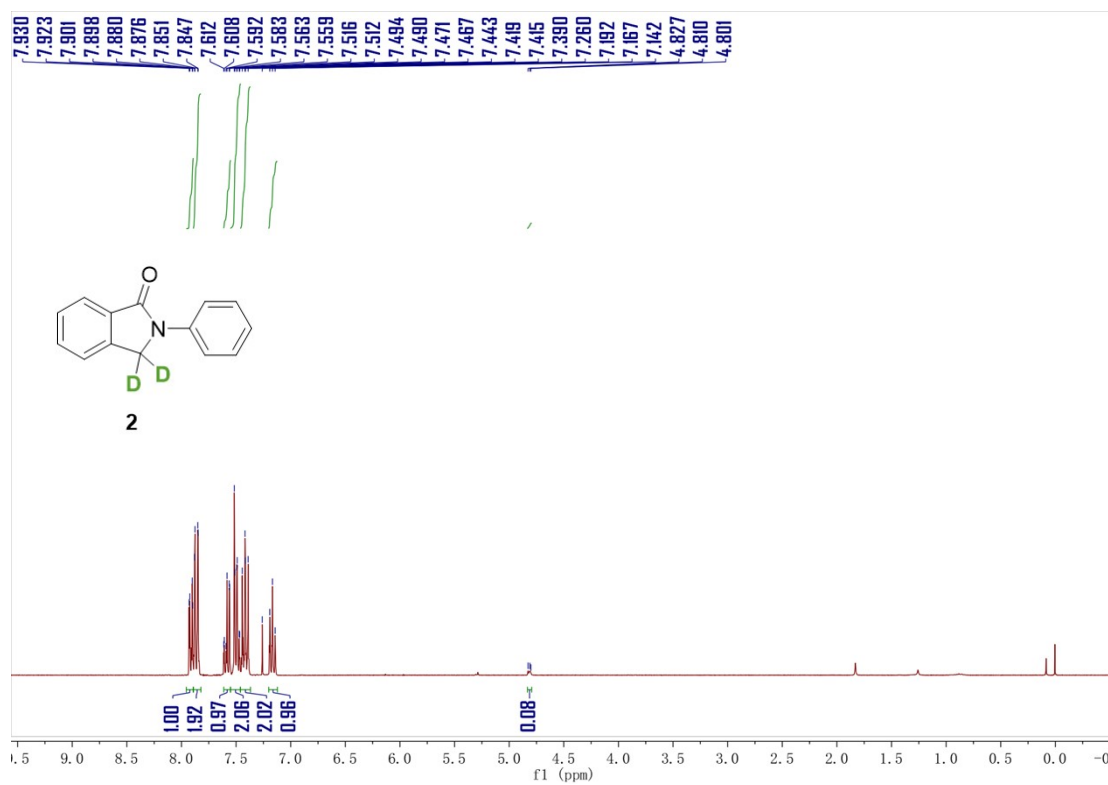
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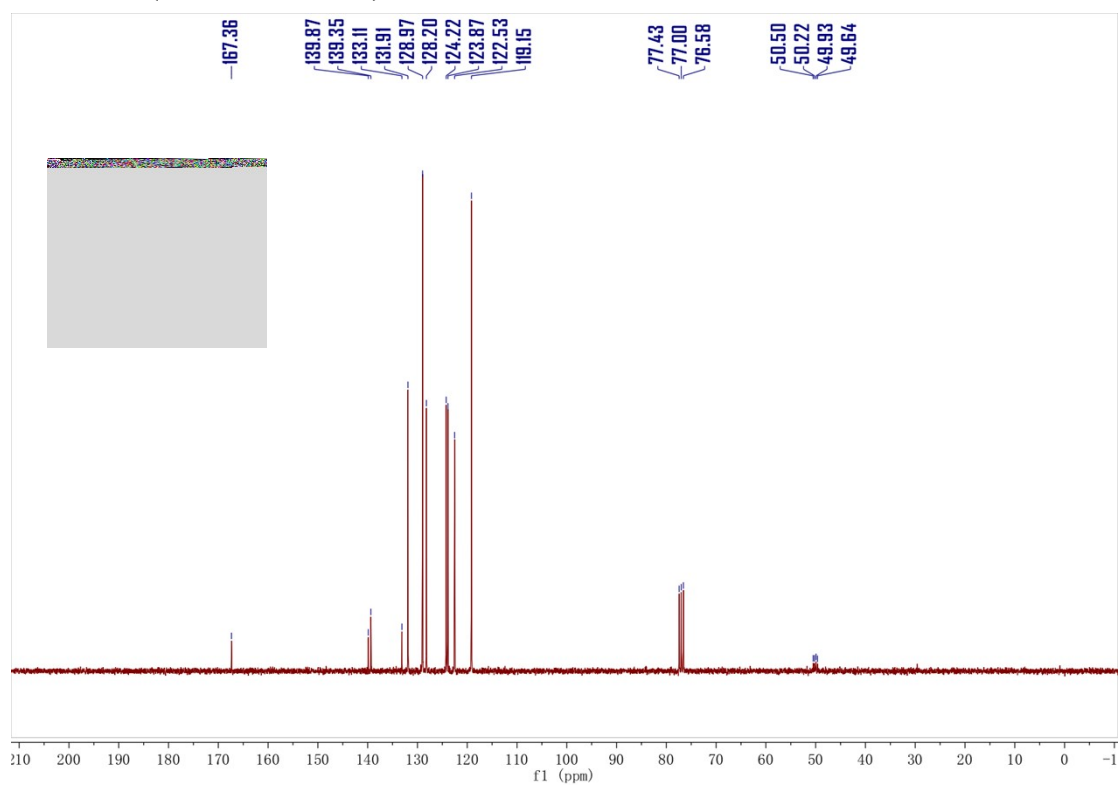
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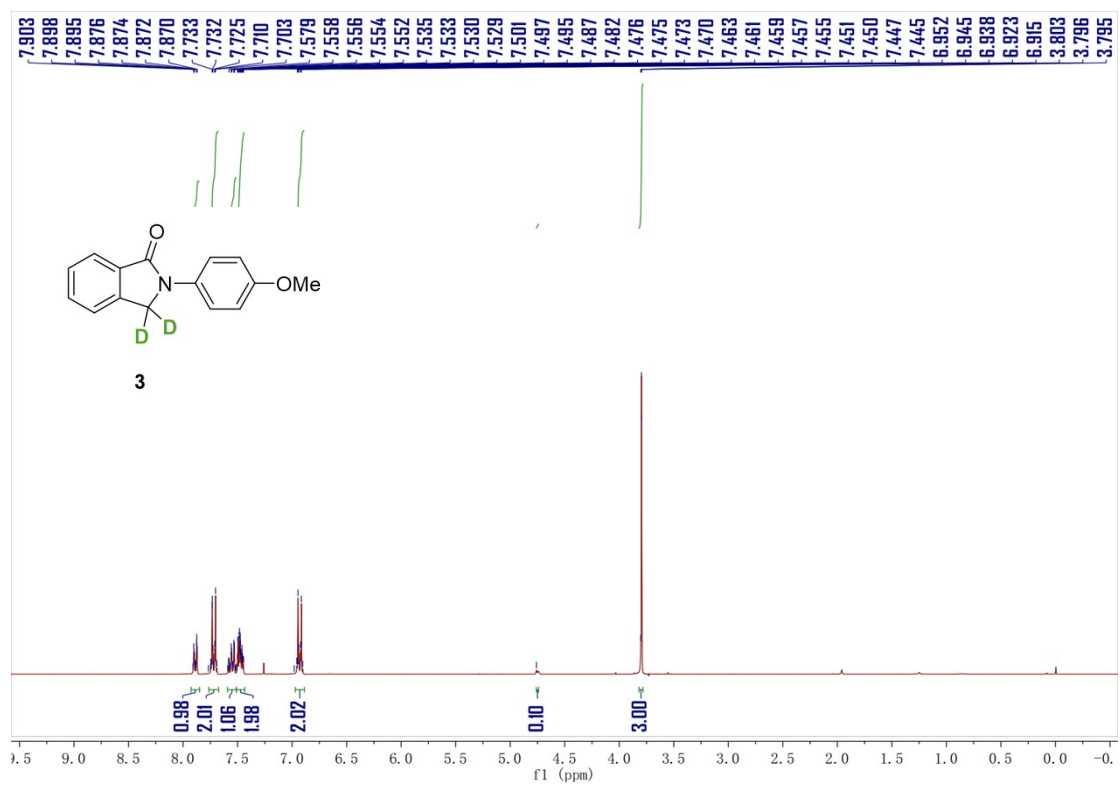
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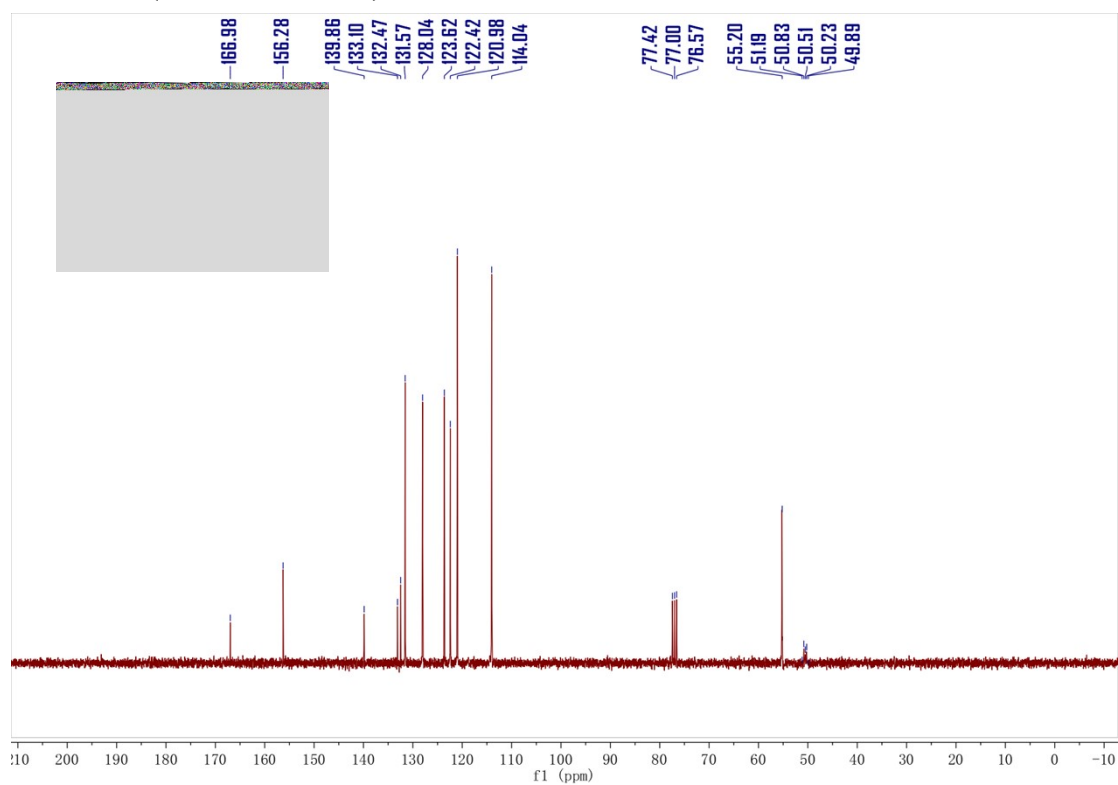
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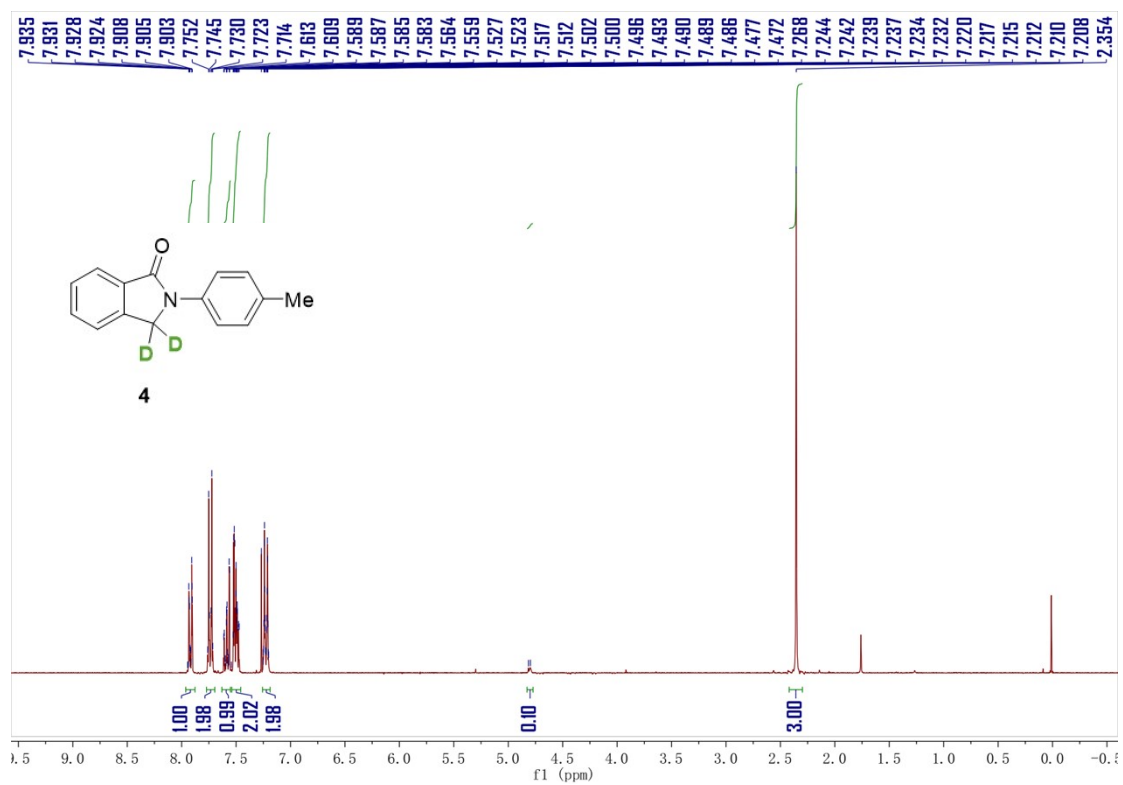
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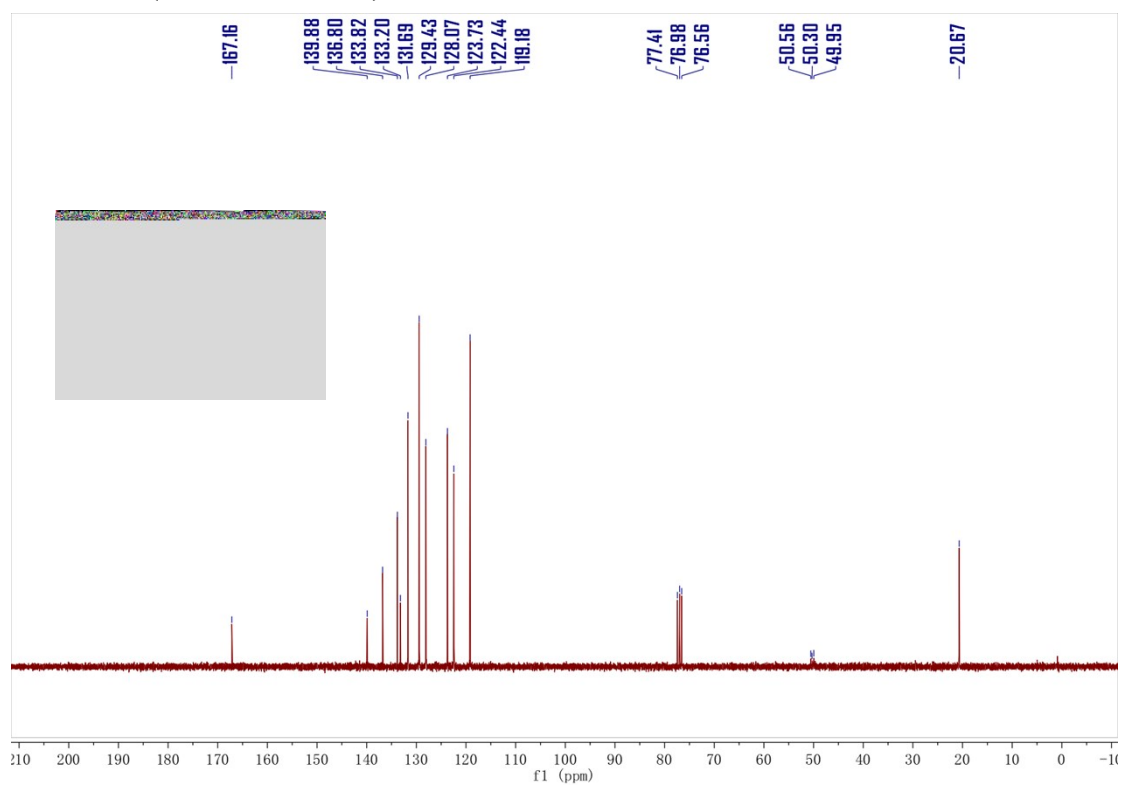
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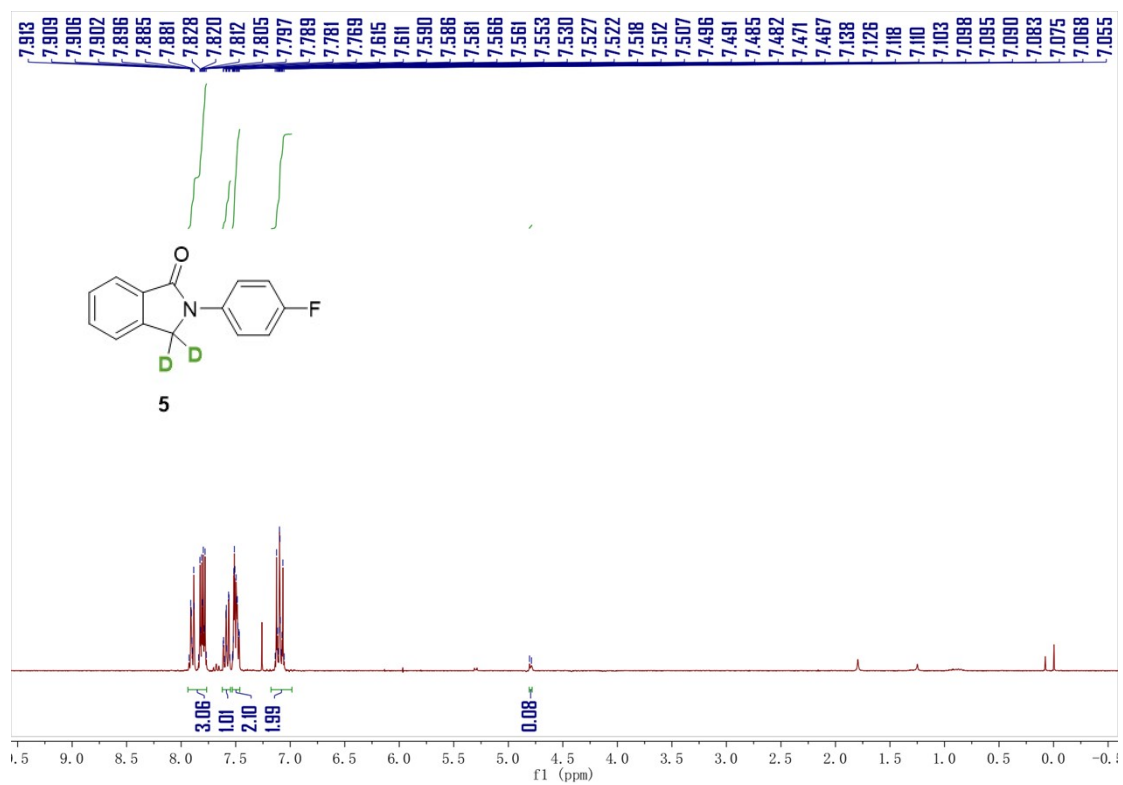
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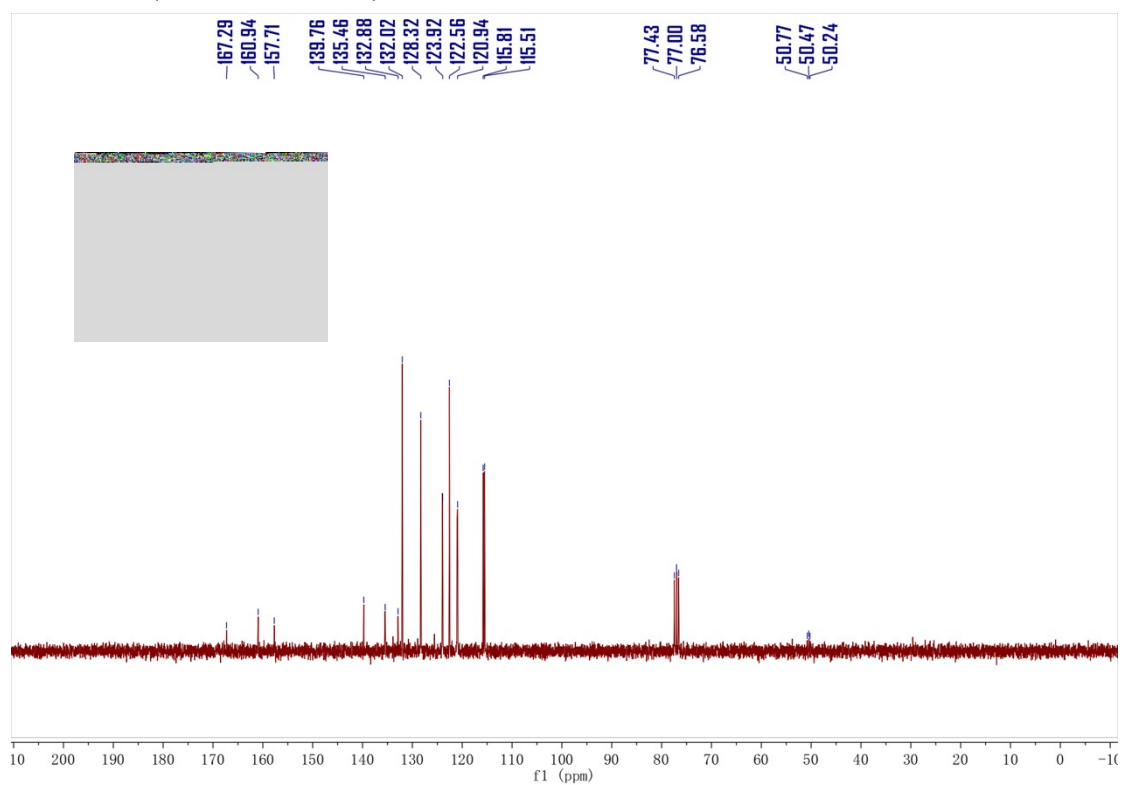
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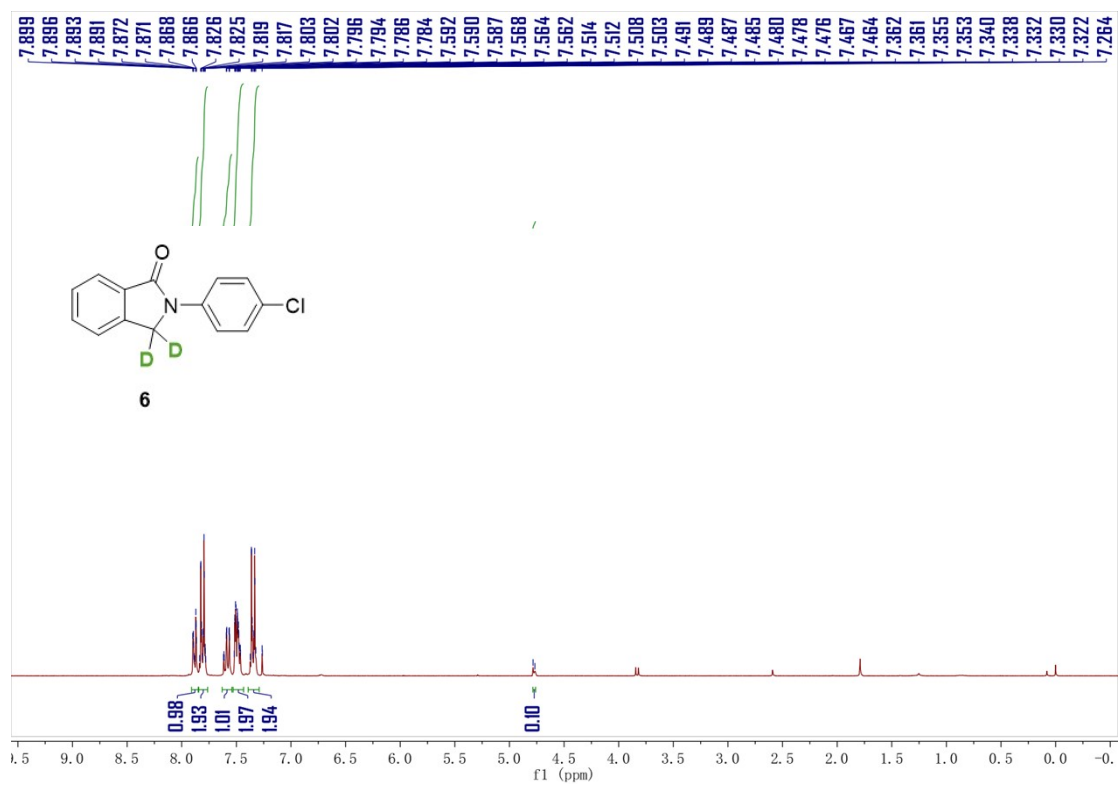
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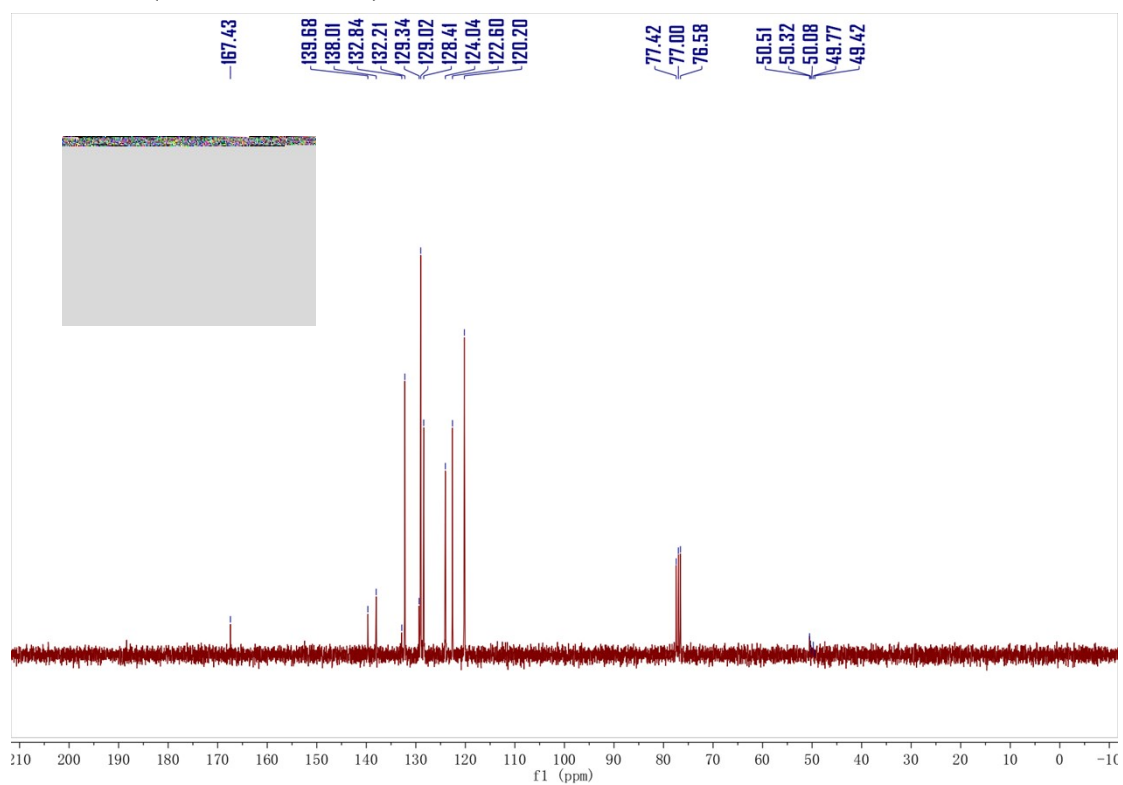
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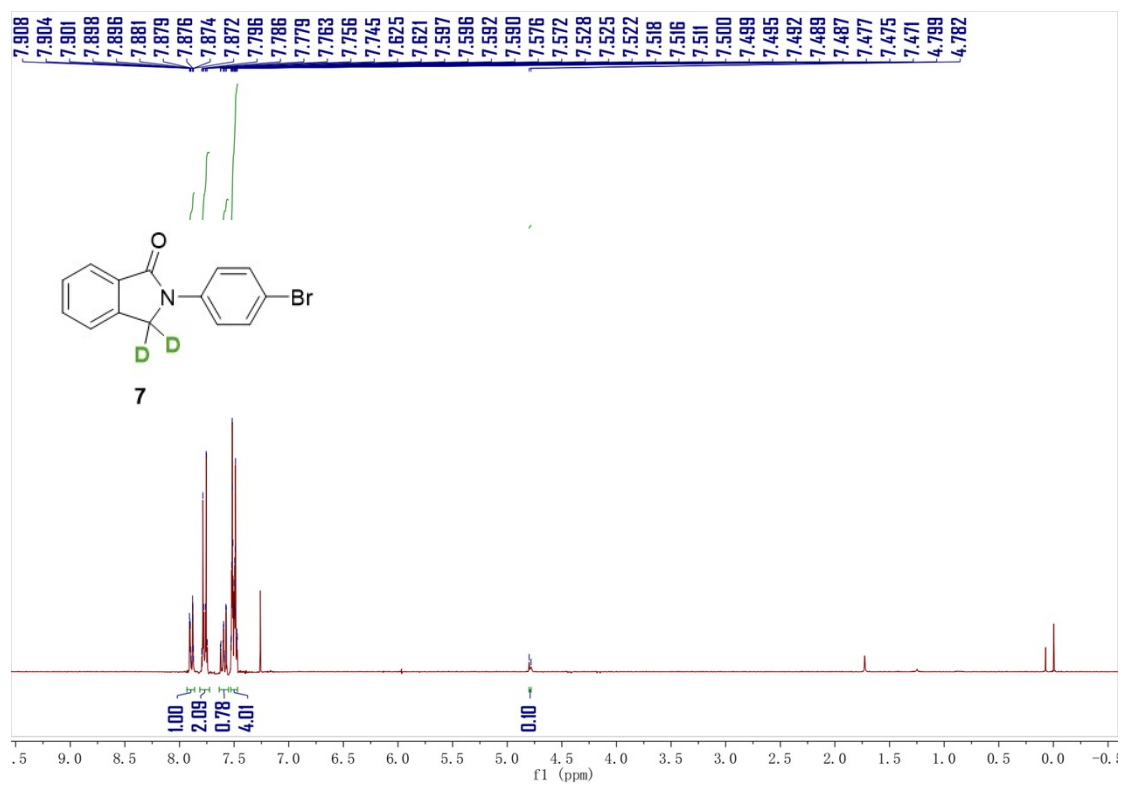
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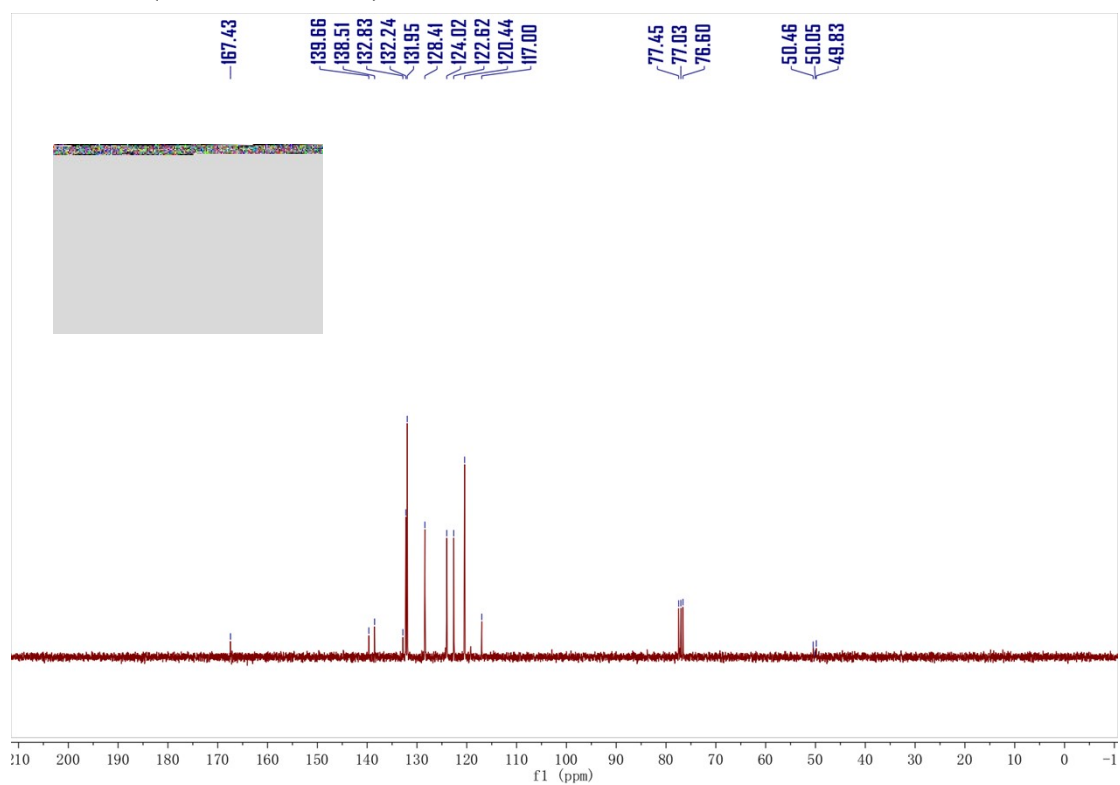
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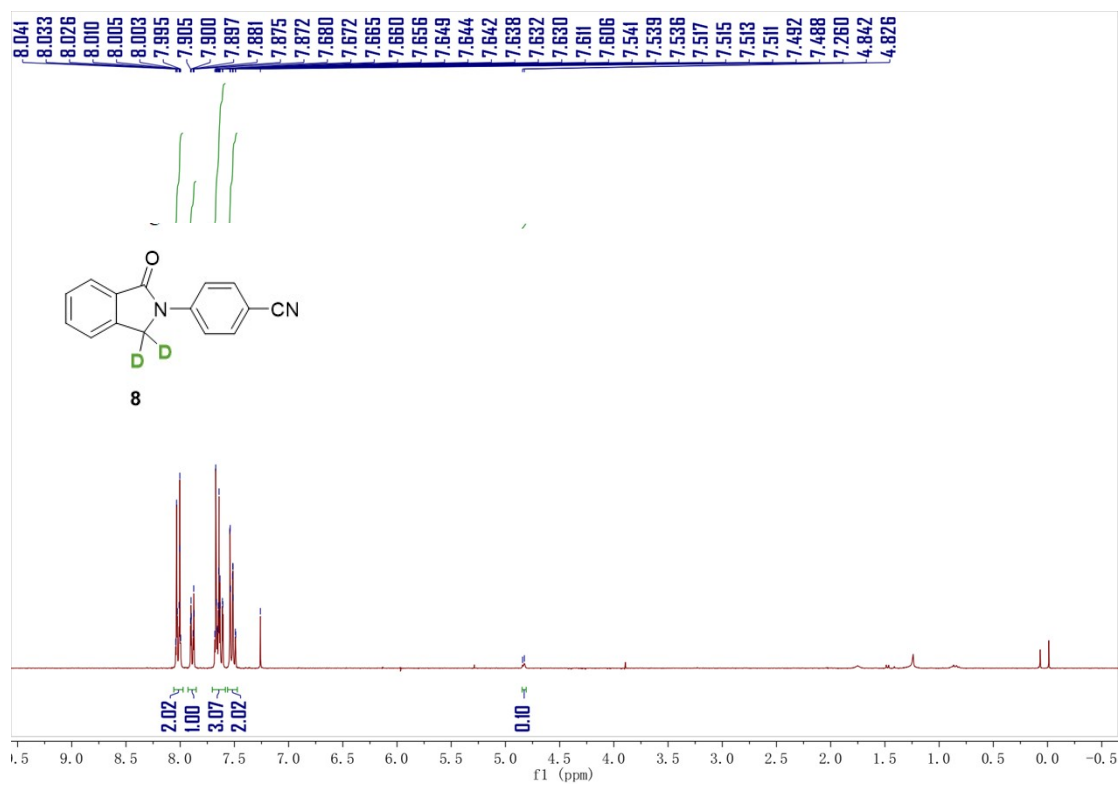
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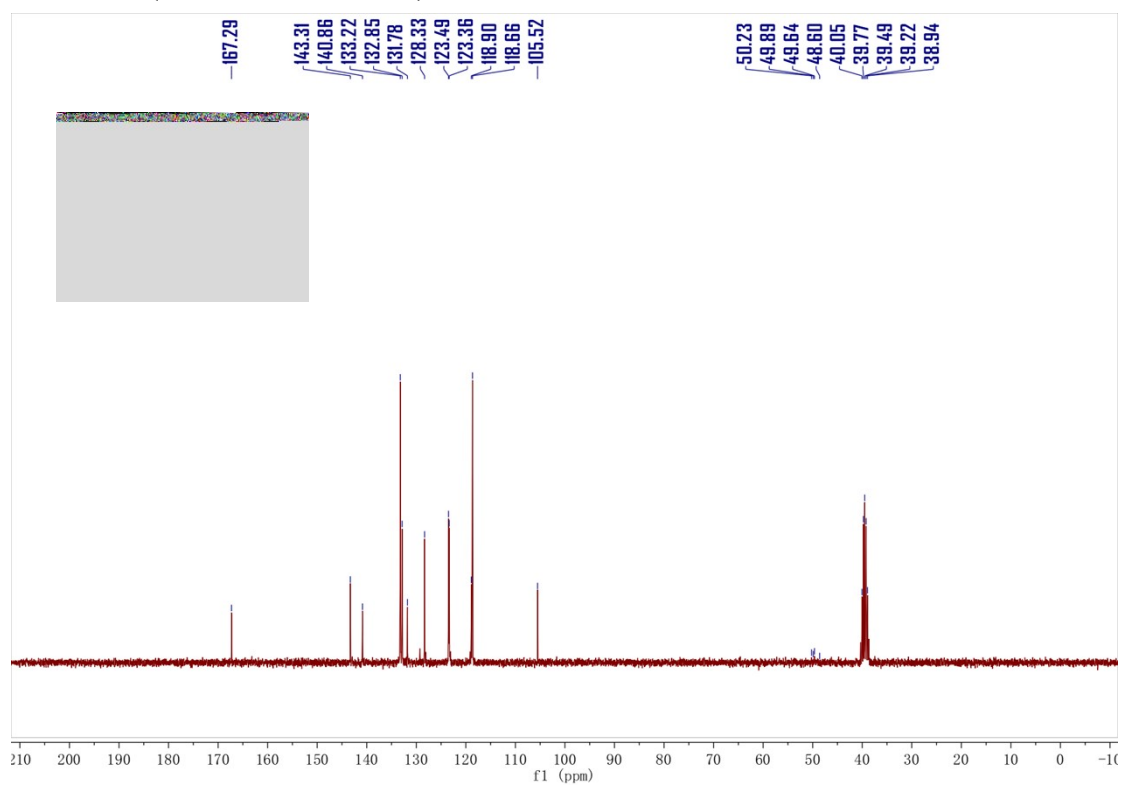
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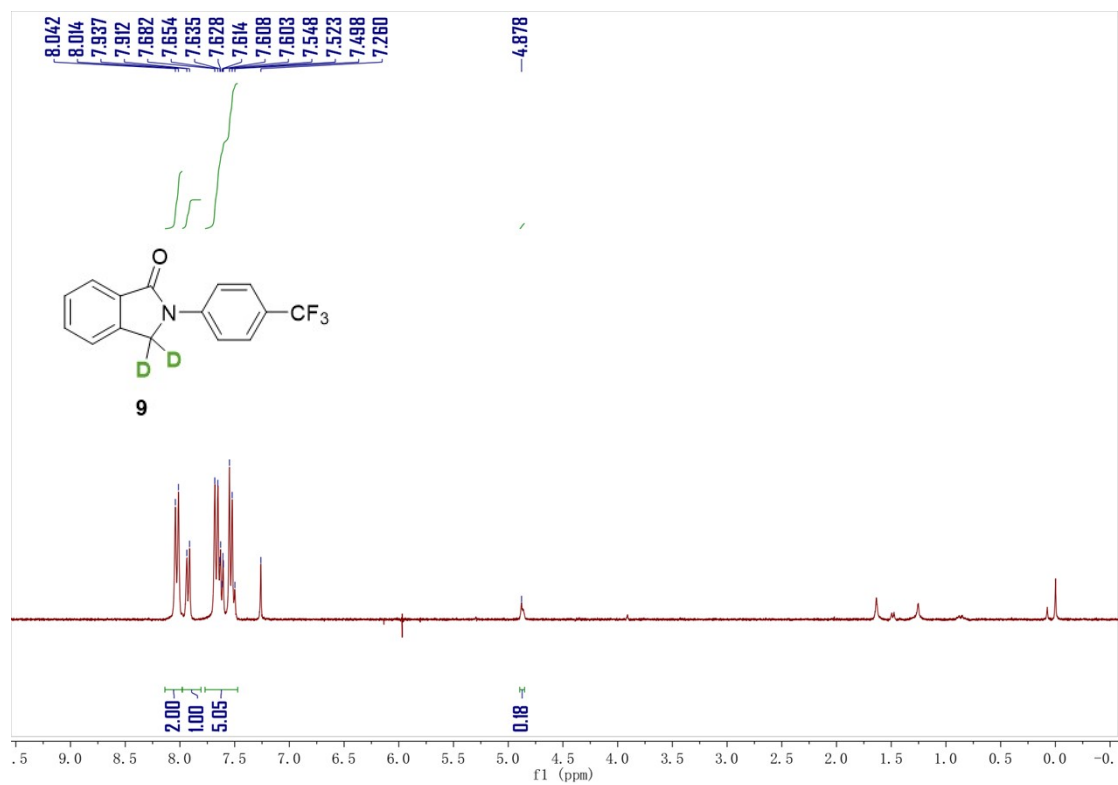
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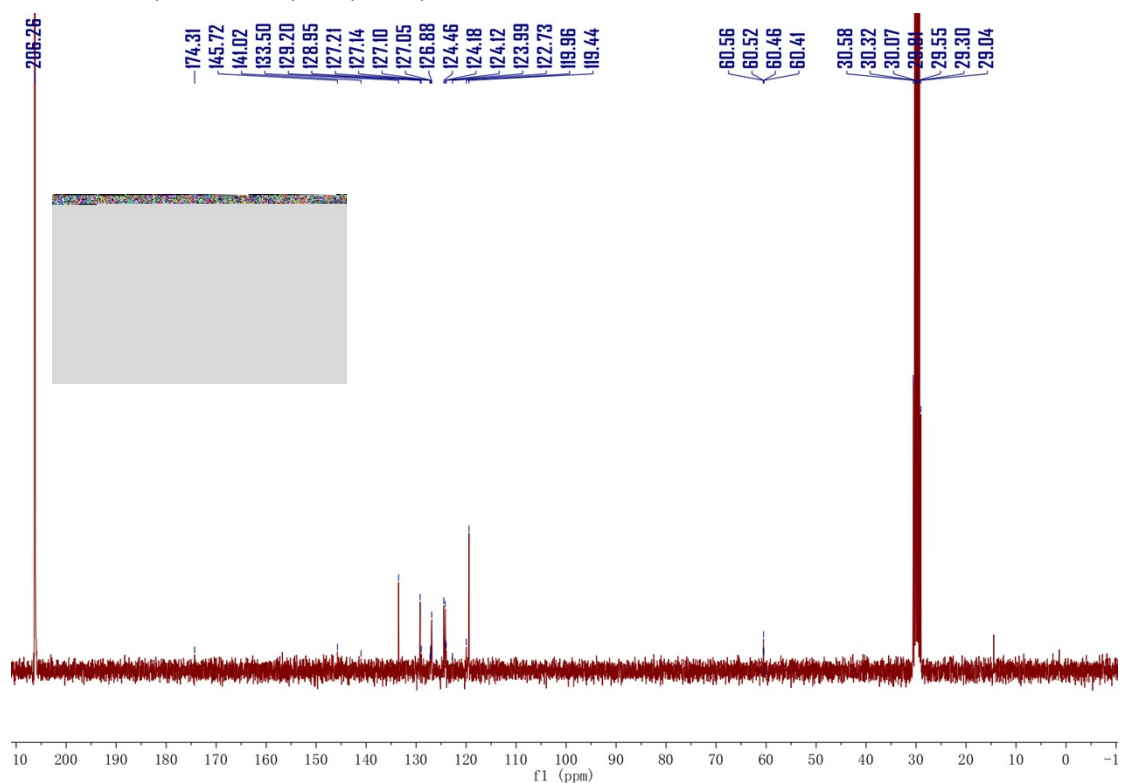
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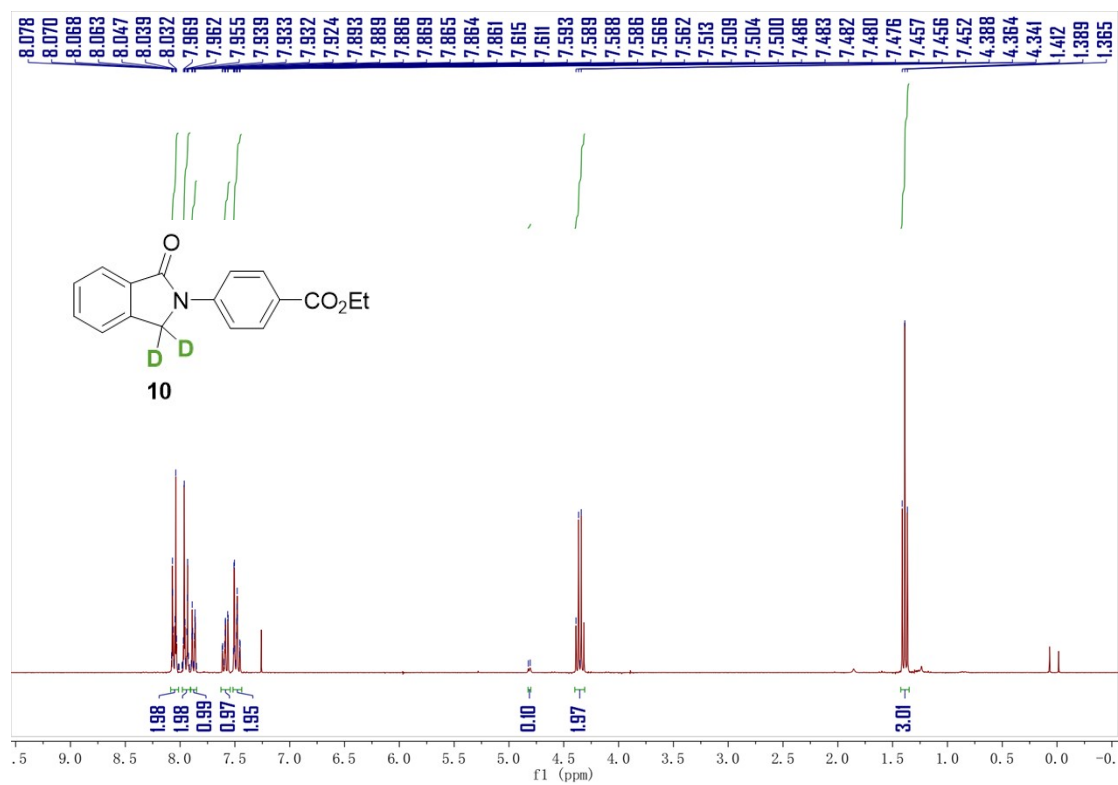
¹H NMR (300 MHz, CDCl₃):



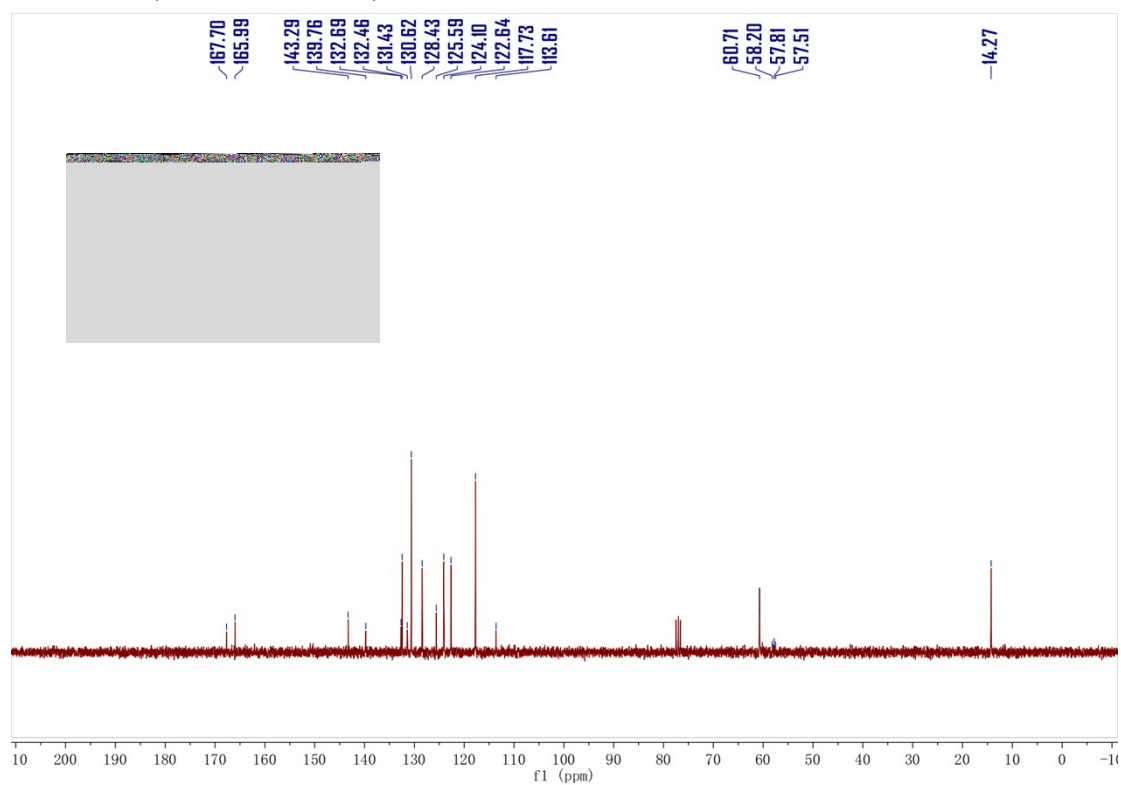
¹³C NMR (75 MHz, (CD₃)₂CO):



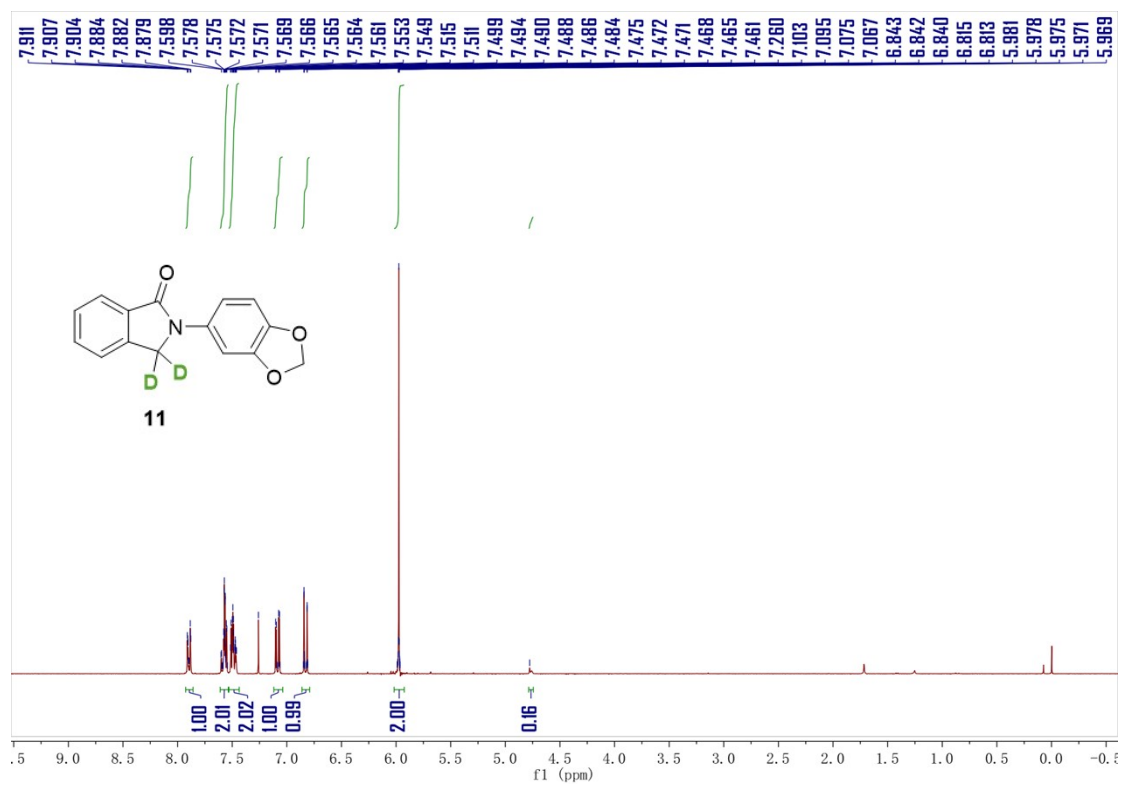
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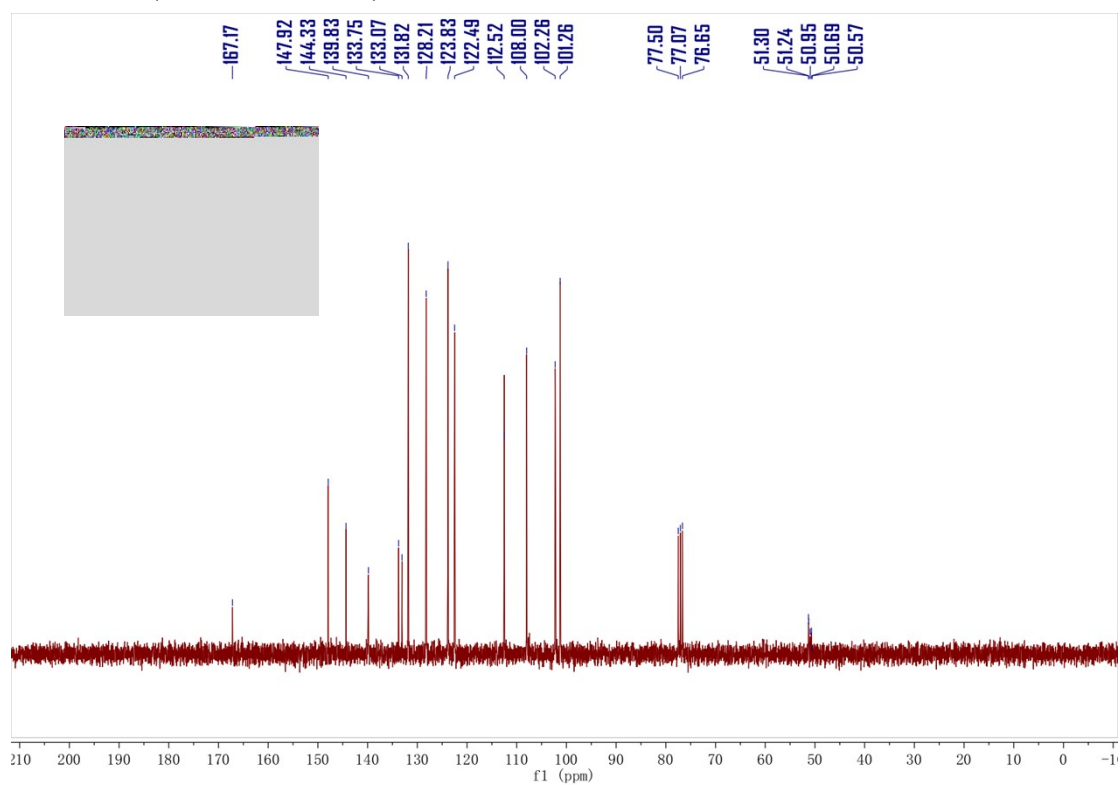
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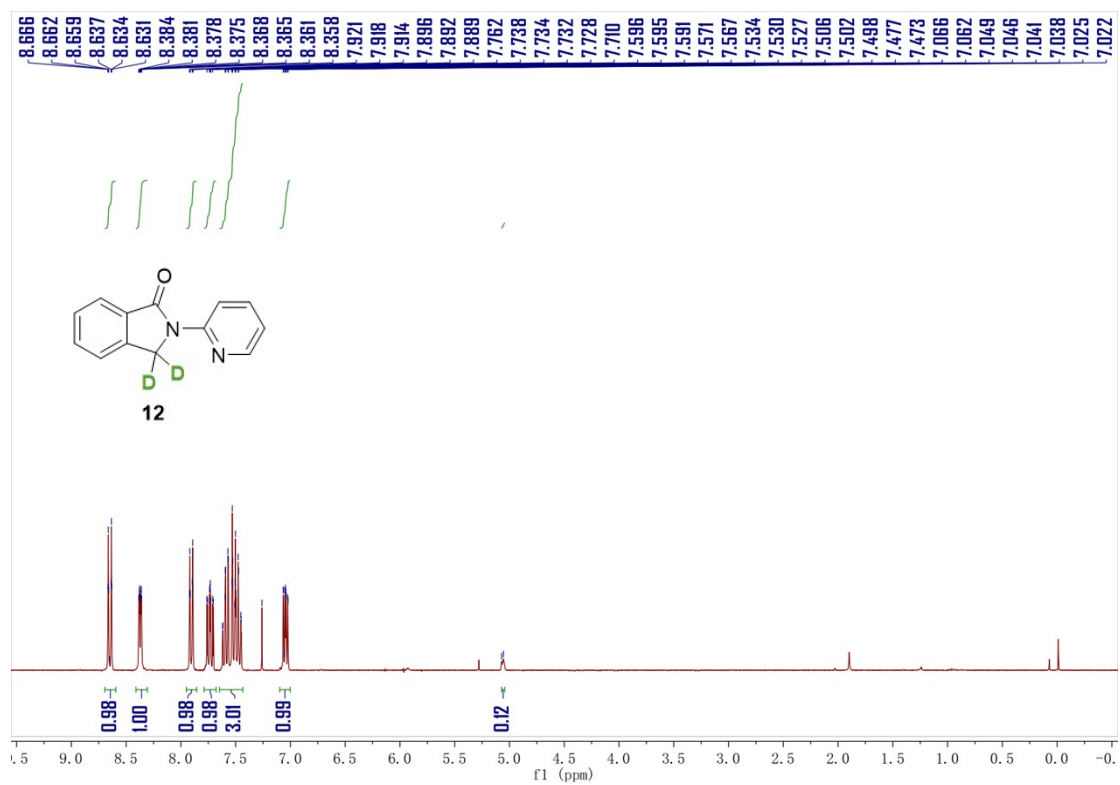
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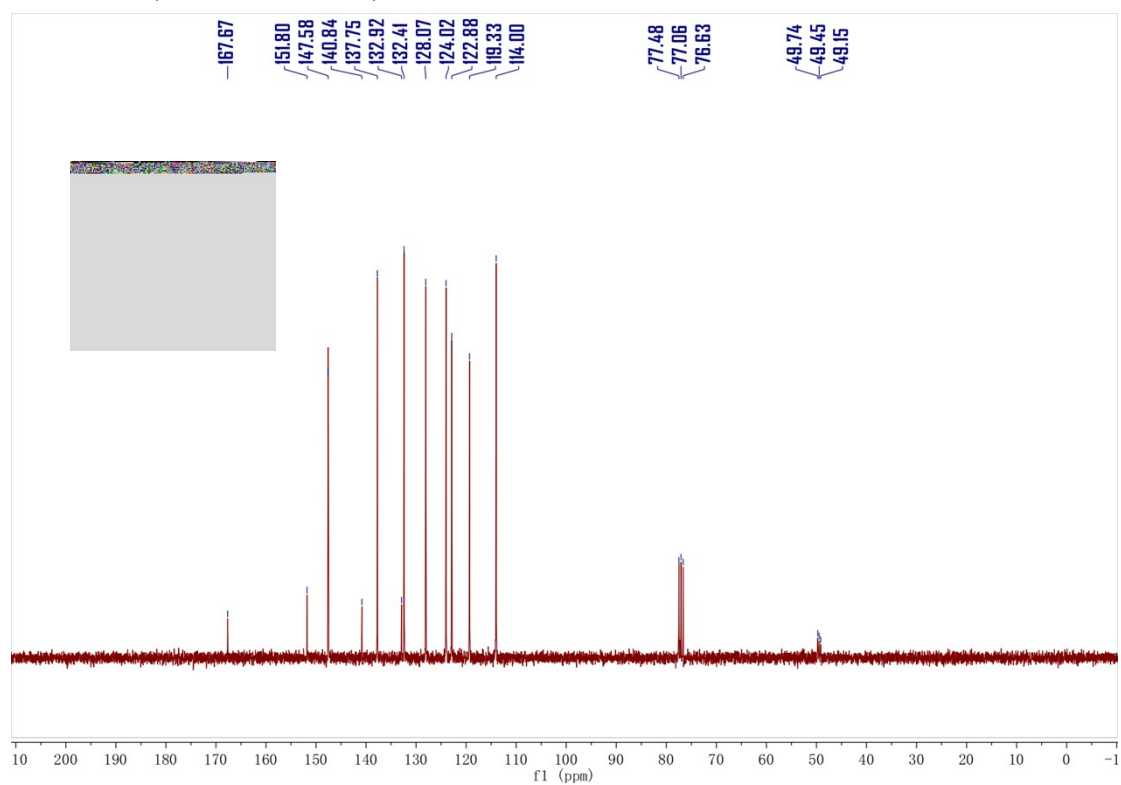
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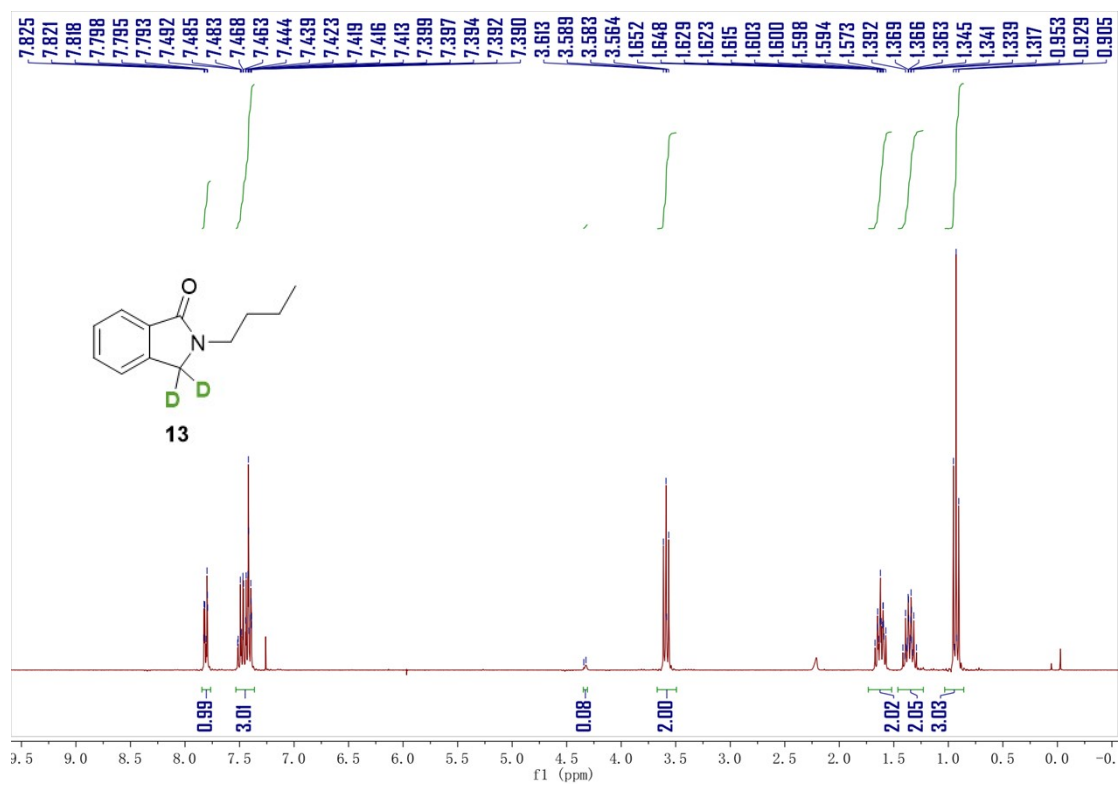
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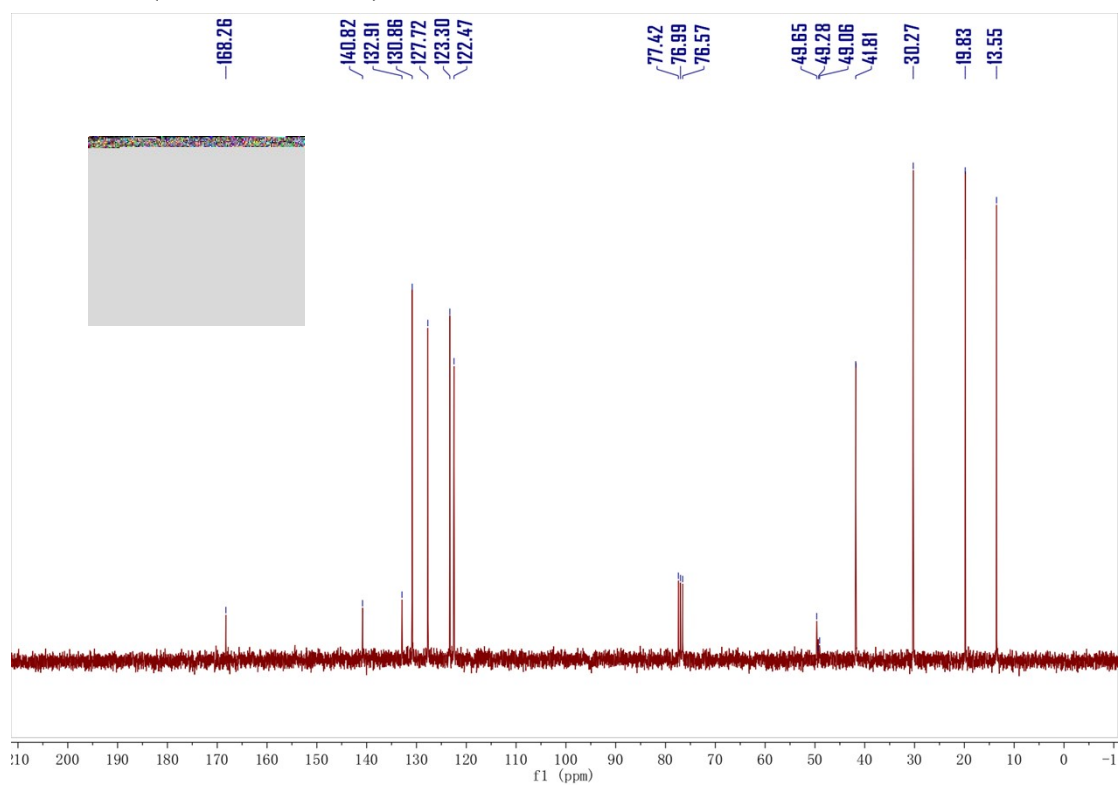
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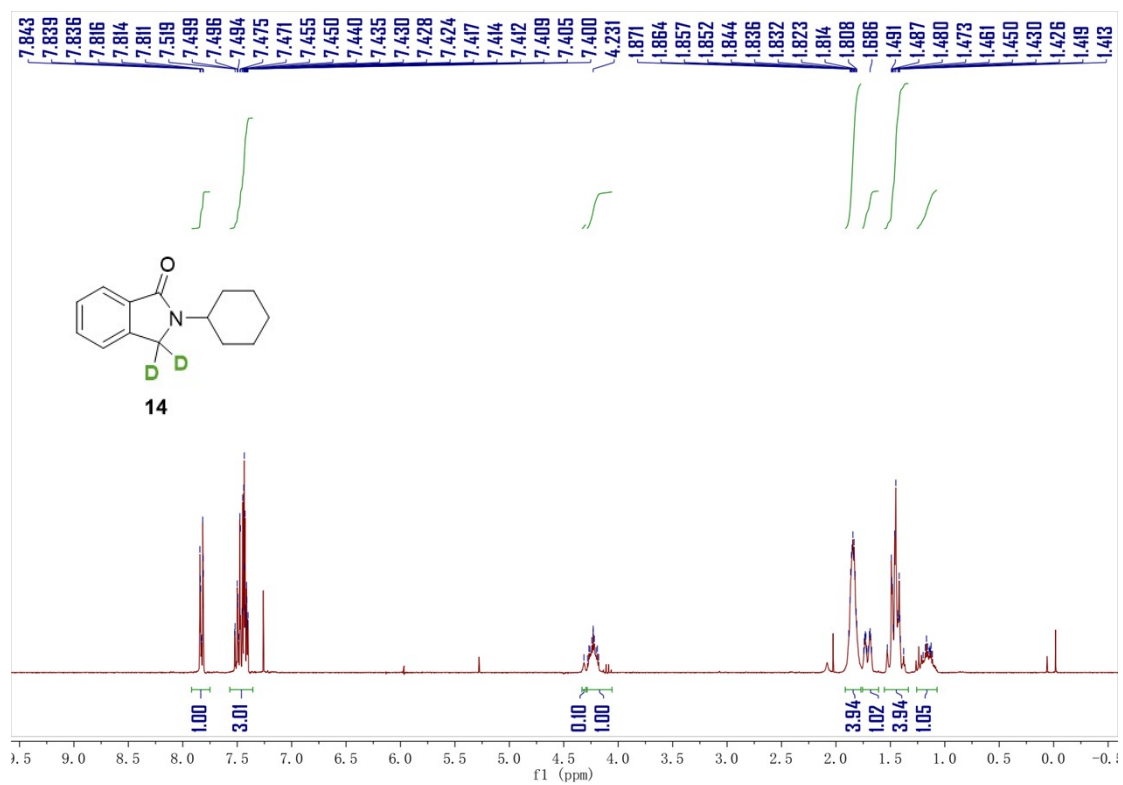
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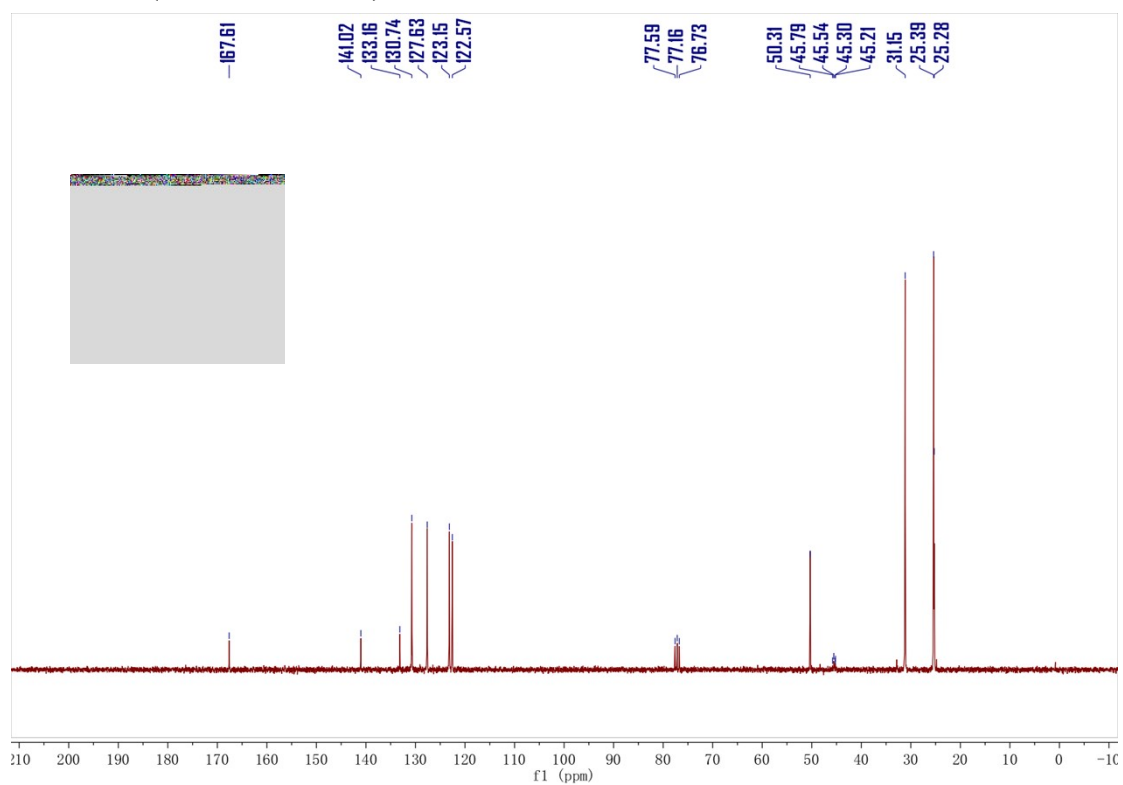
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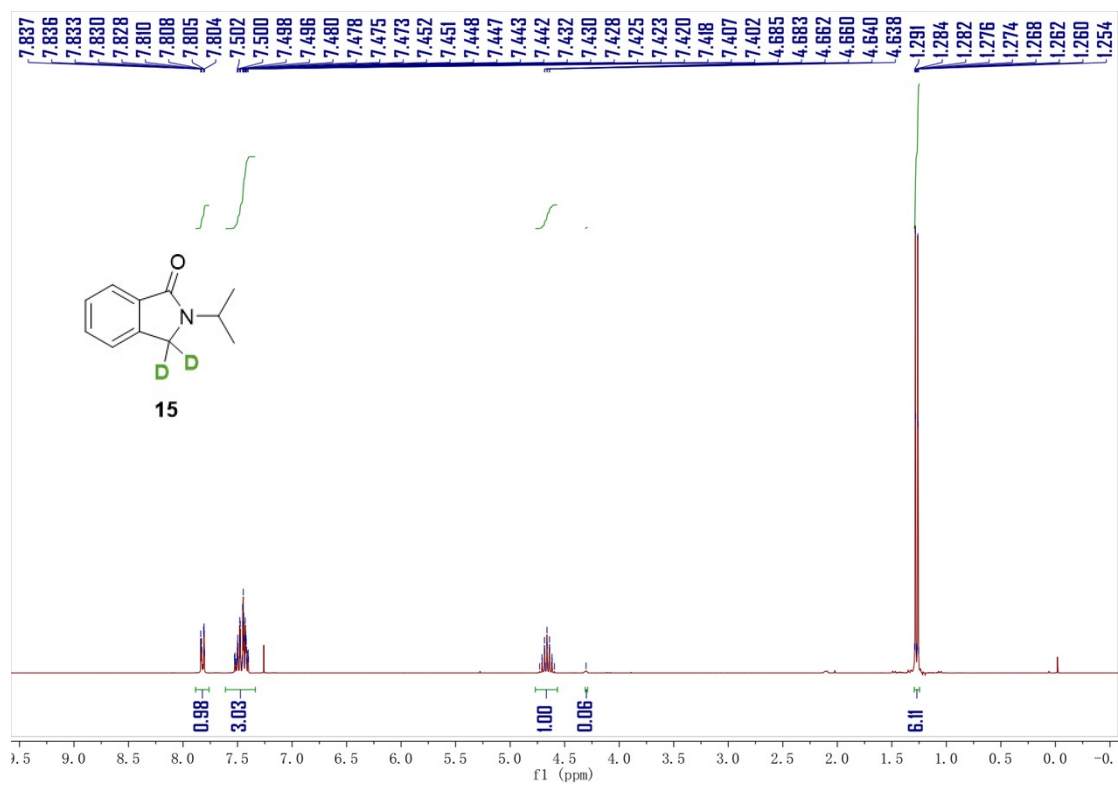
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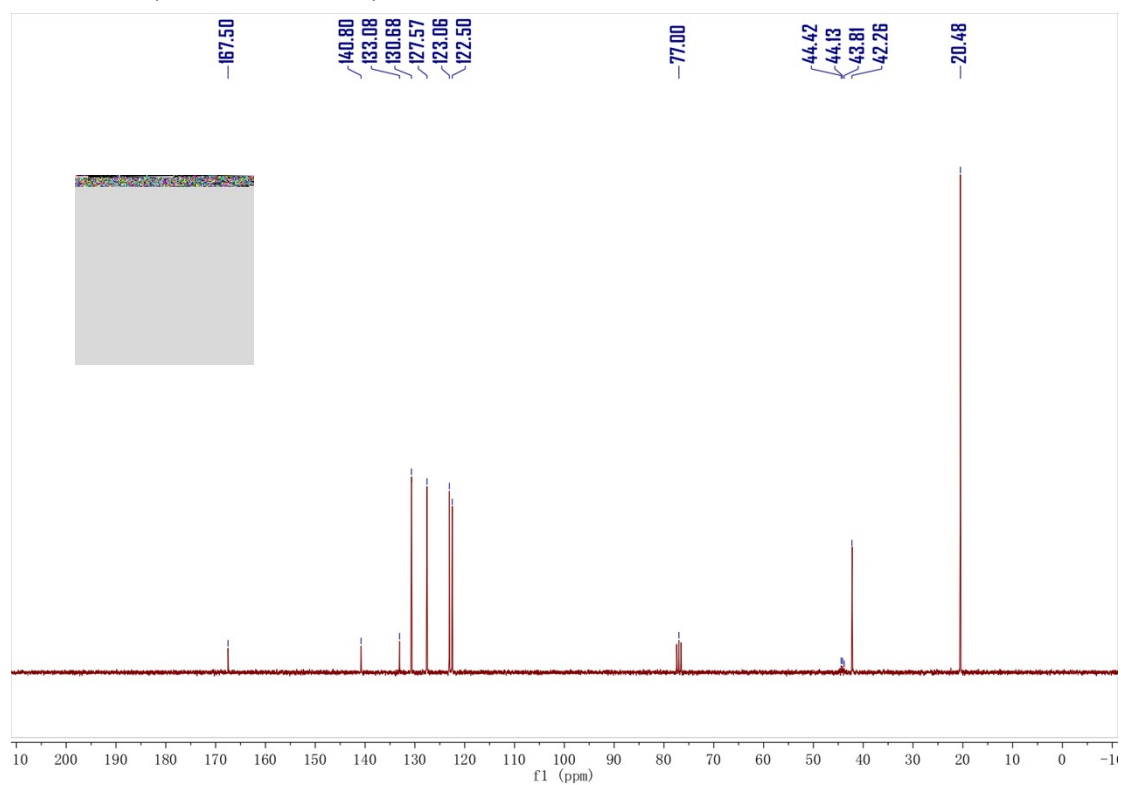
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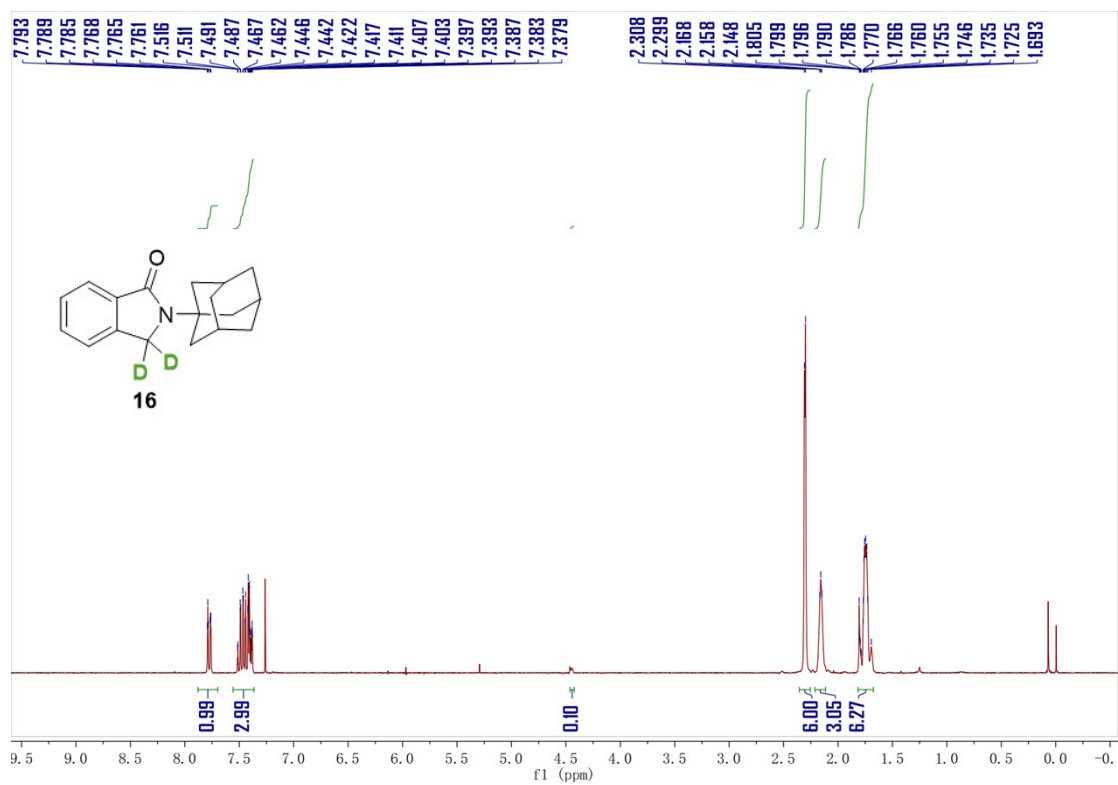
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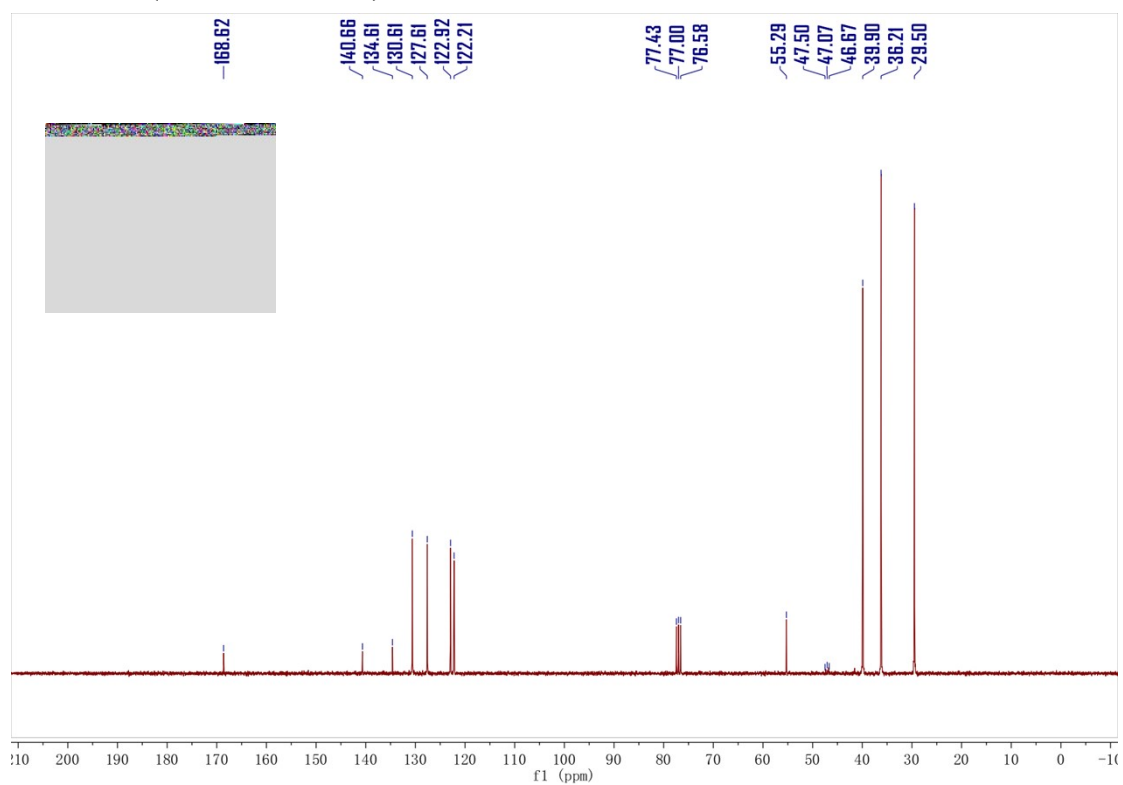
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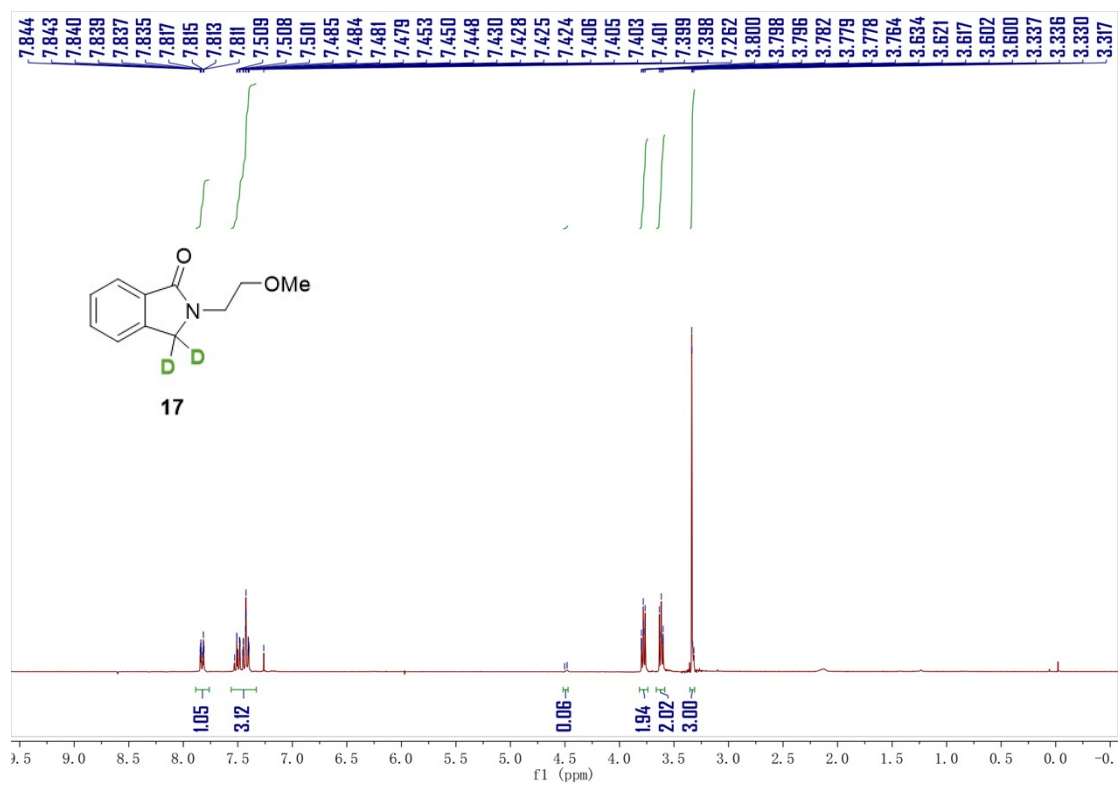
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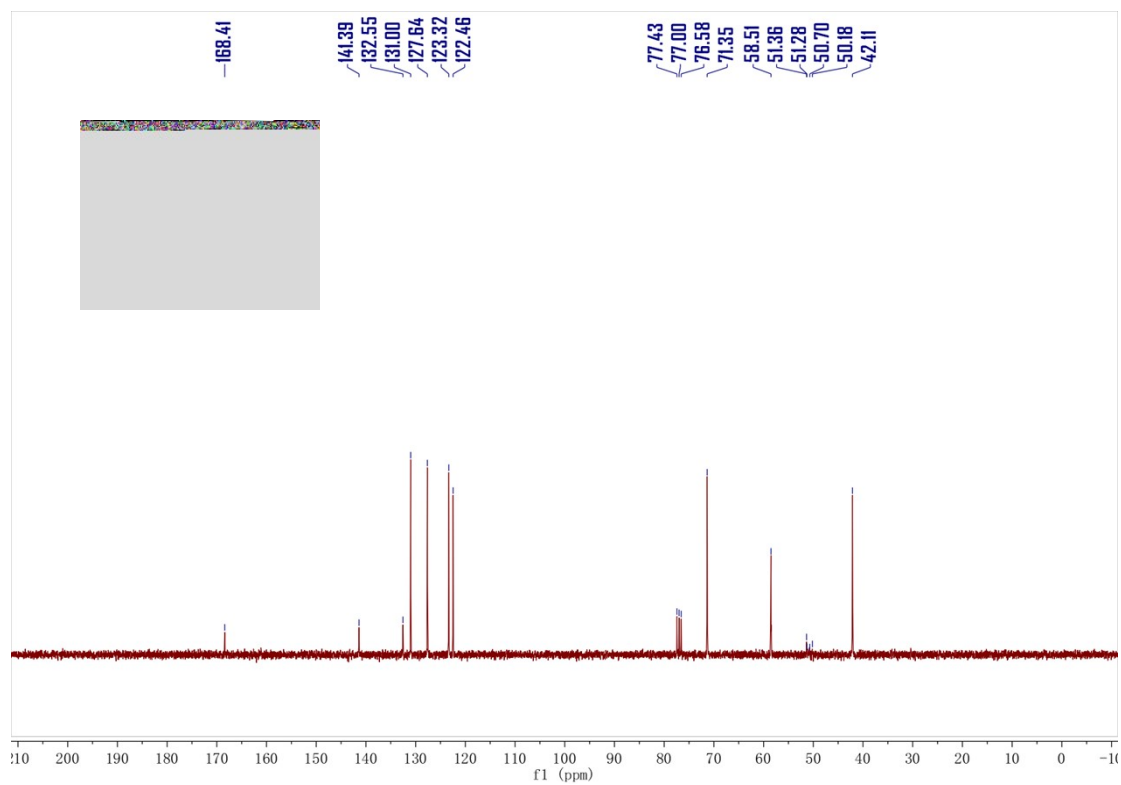
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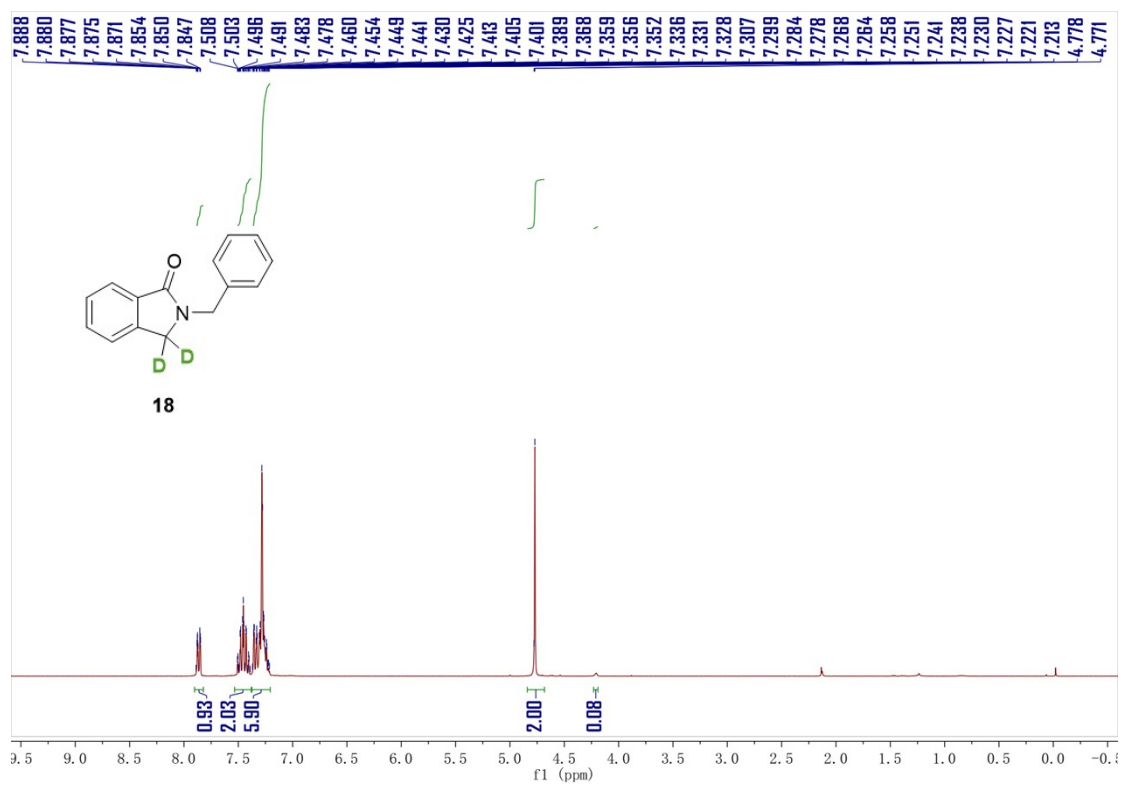
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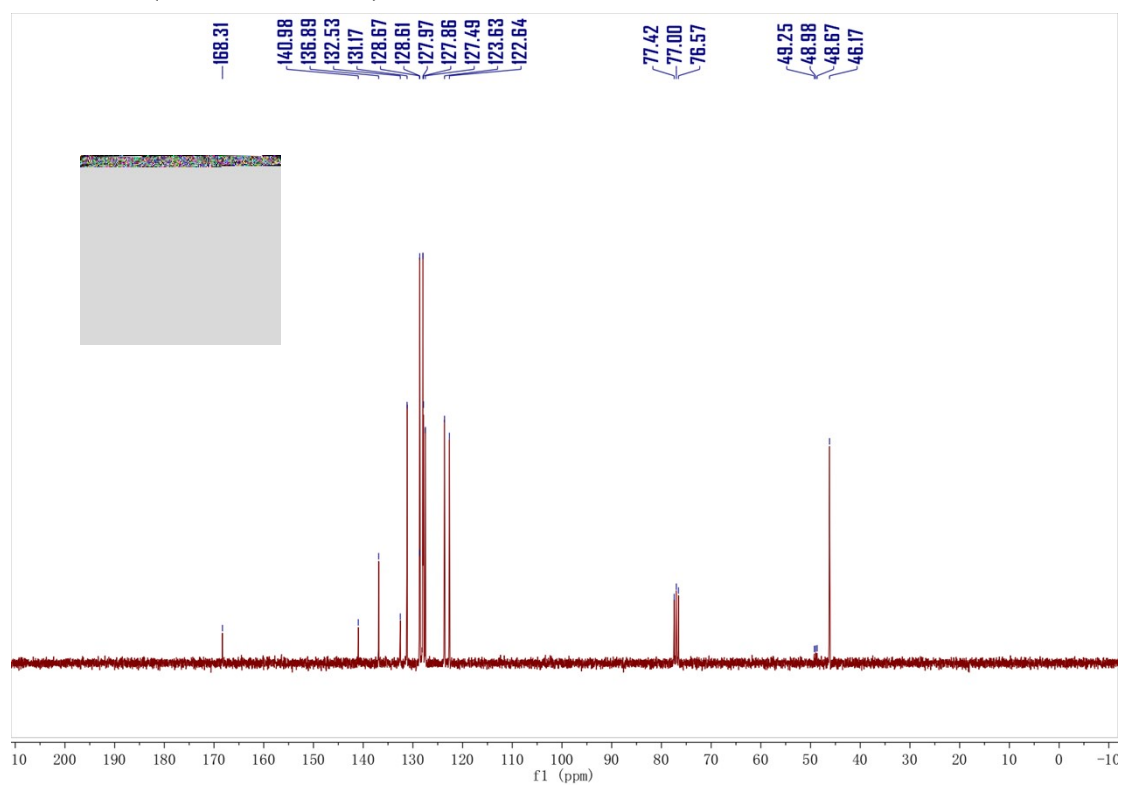
^{13}C NMR (75 MHz, CDCl_3):



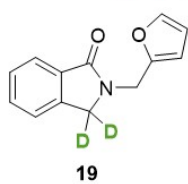
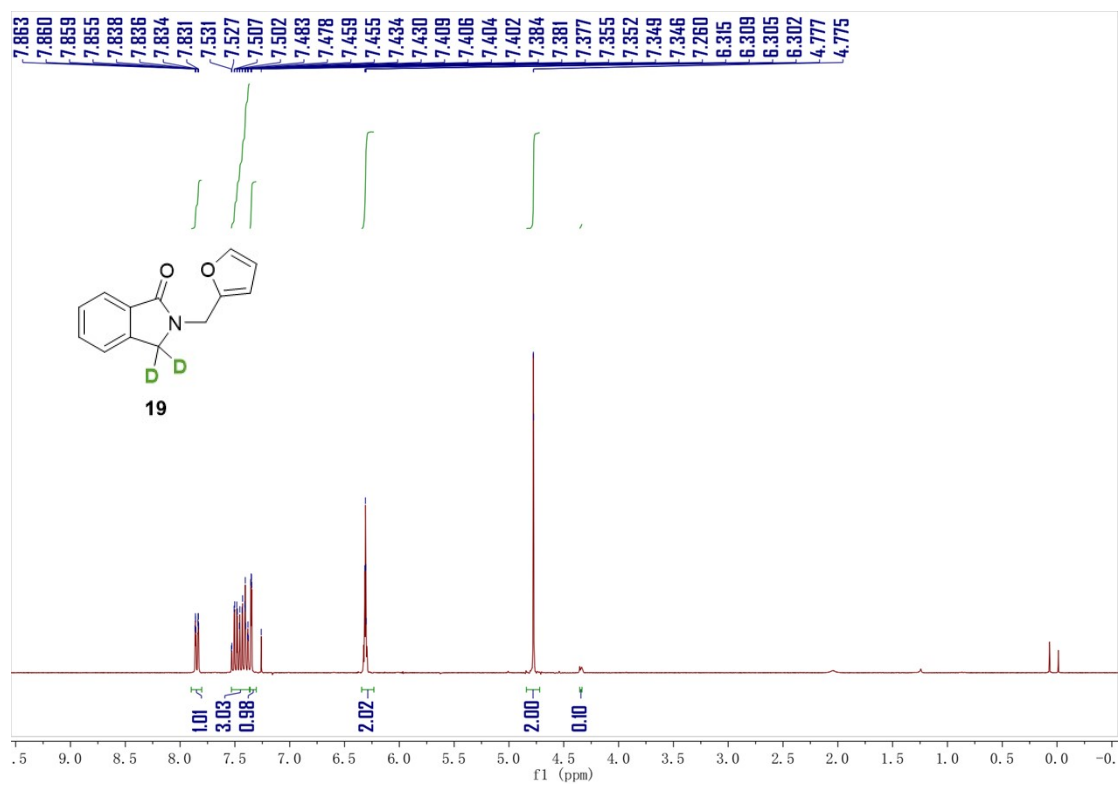
^1H NMR (300 MHz, CDCl_3):



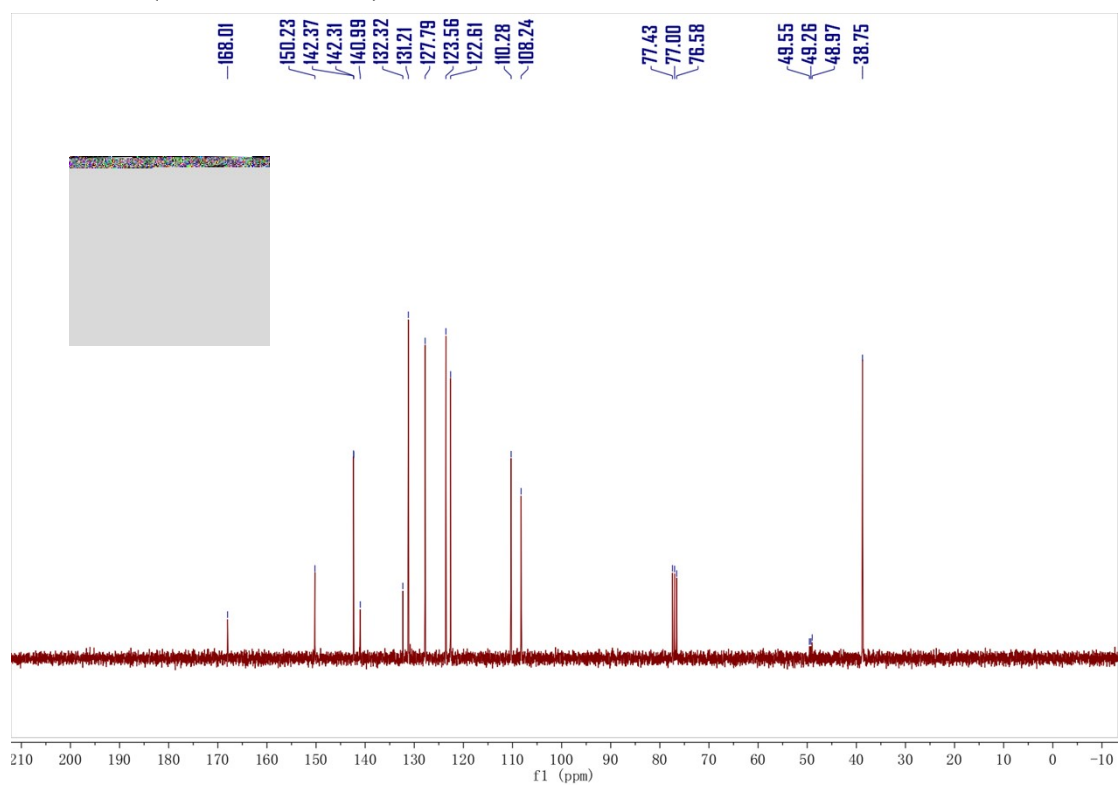
¹³C NMR (75 MHz, CDCl₃):



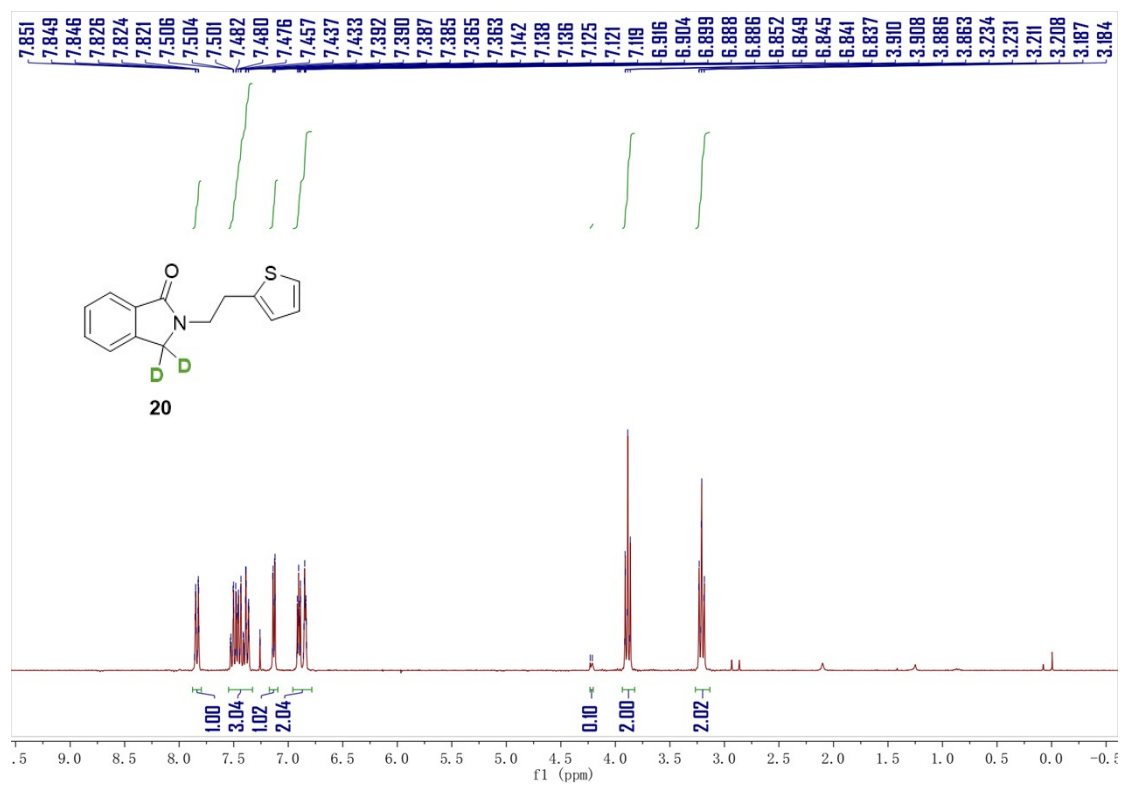
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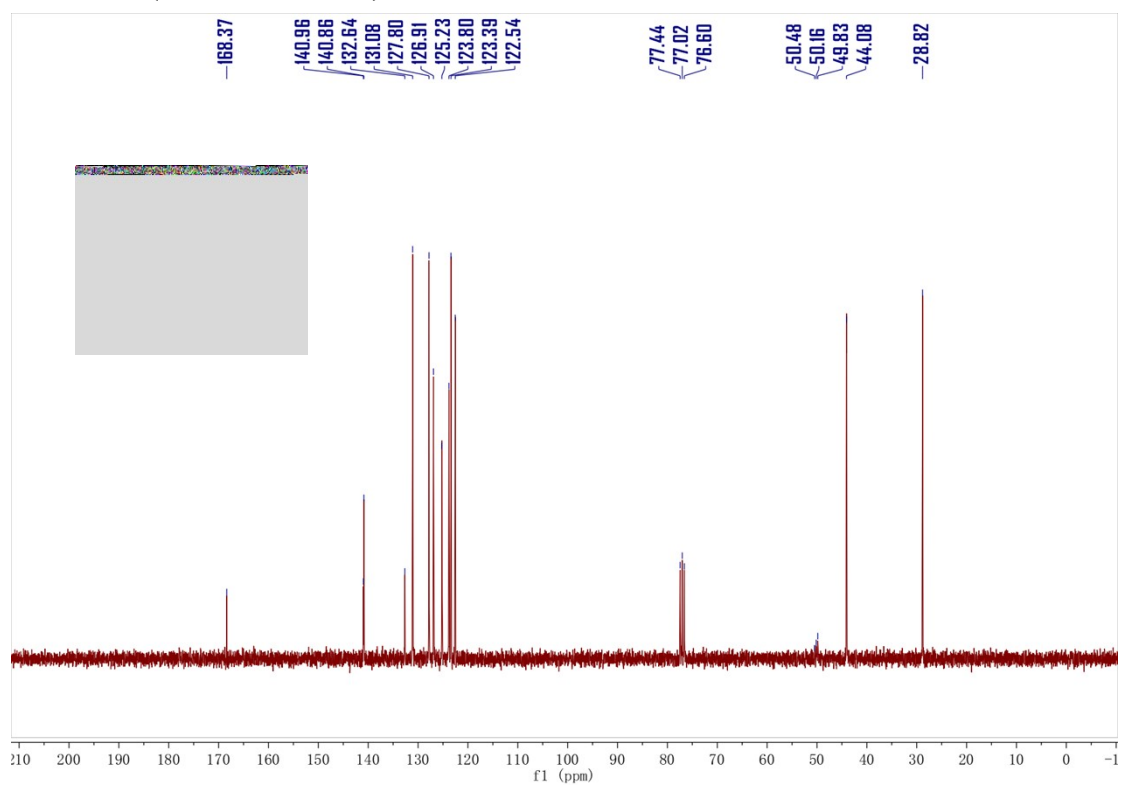
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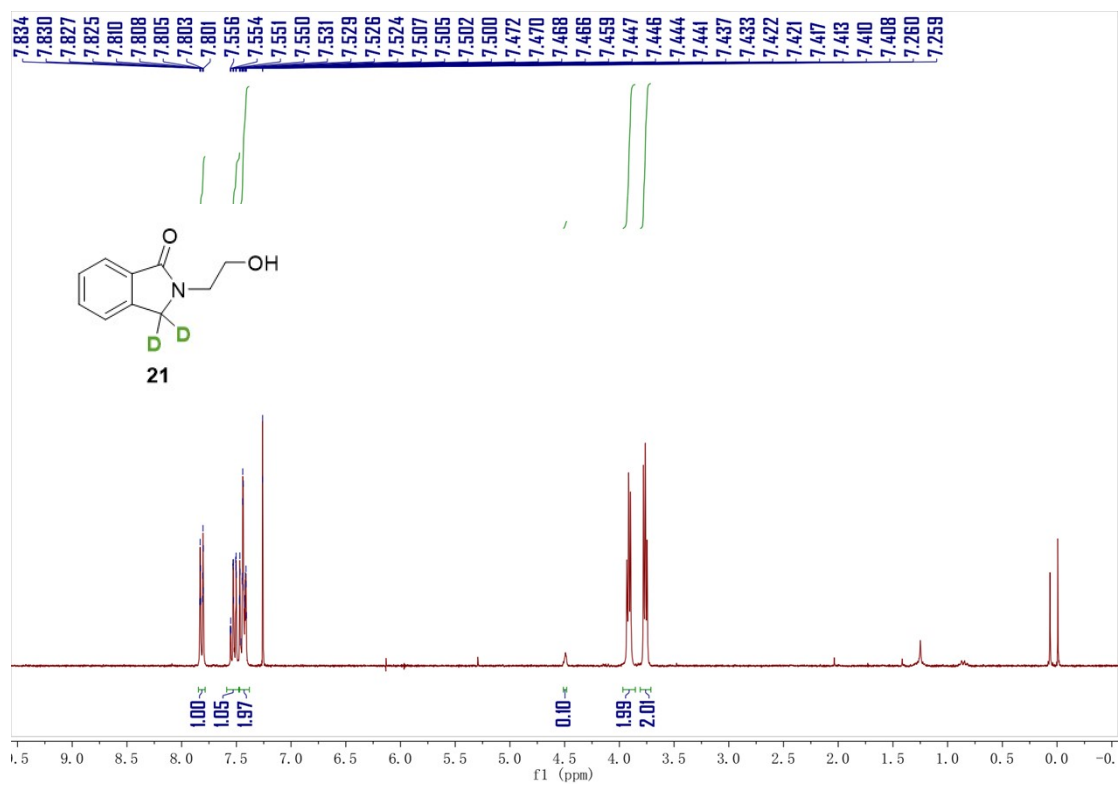
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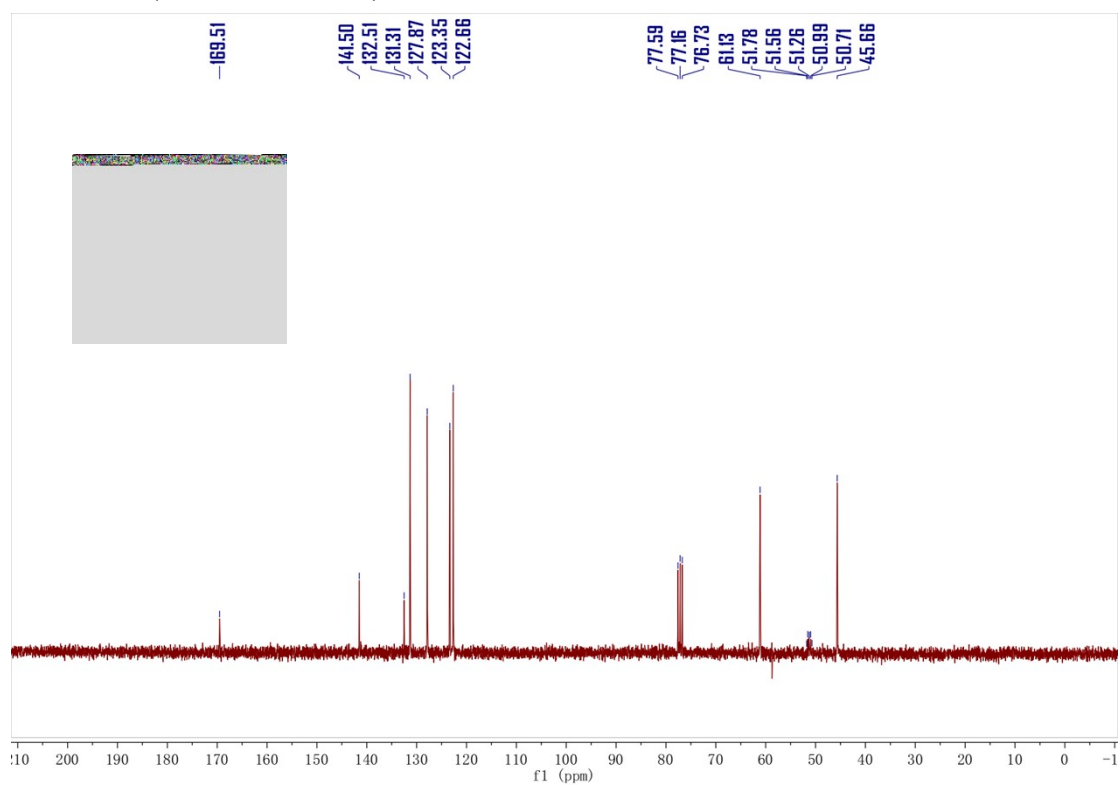
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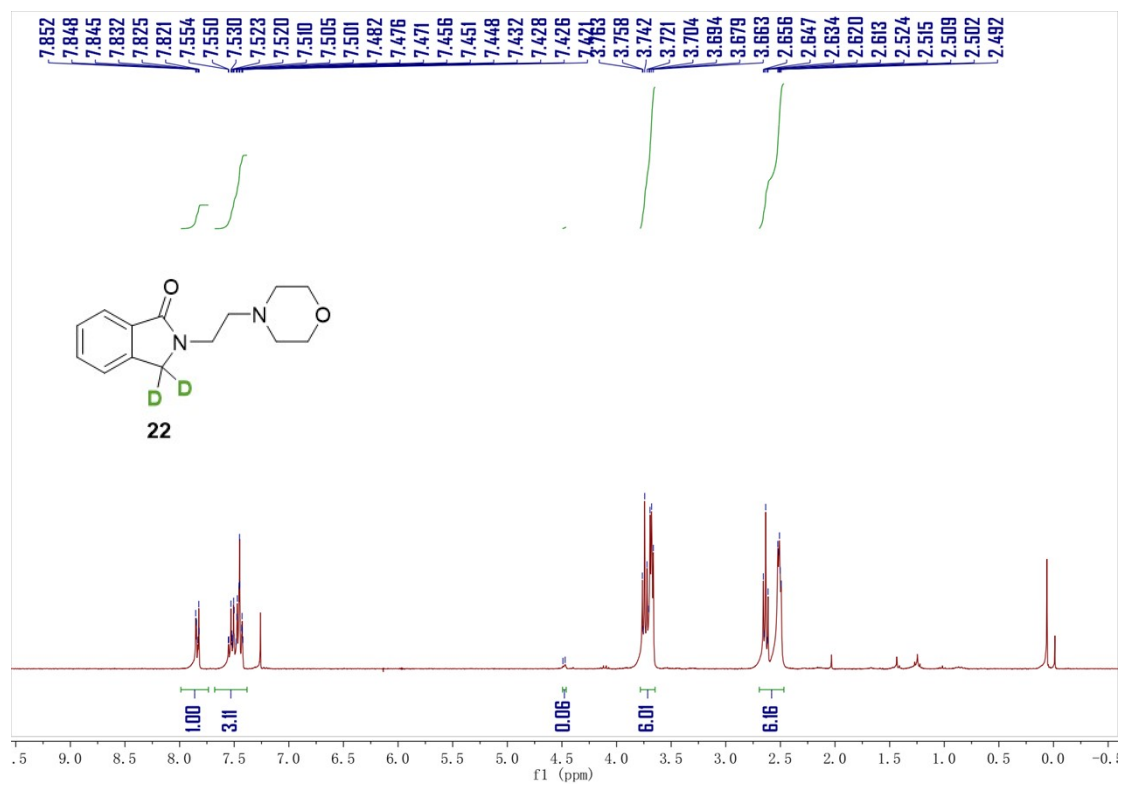
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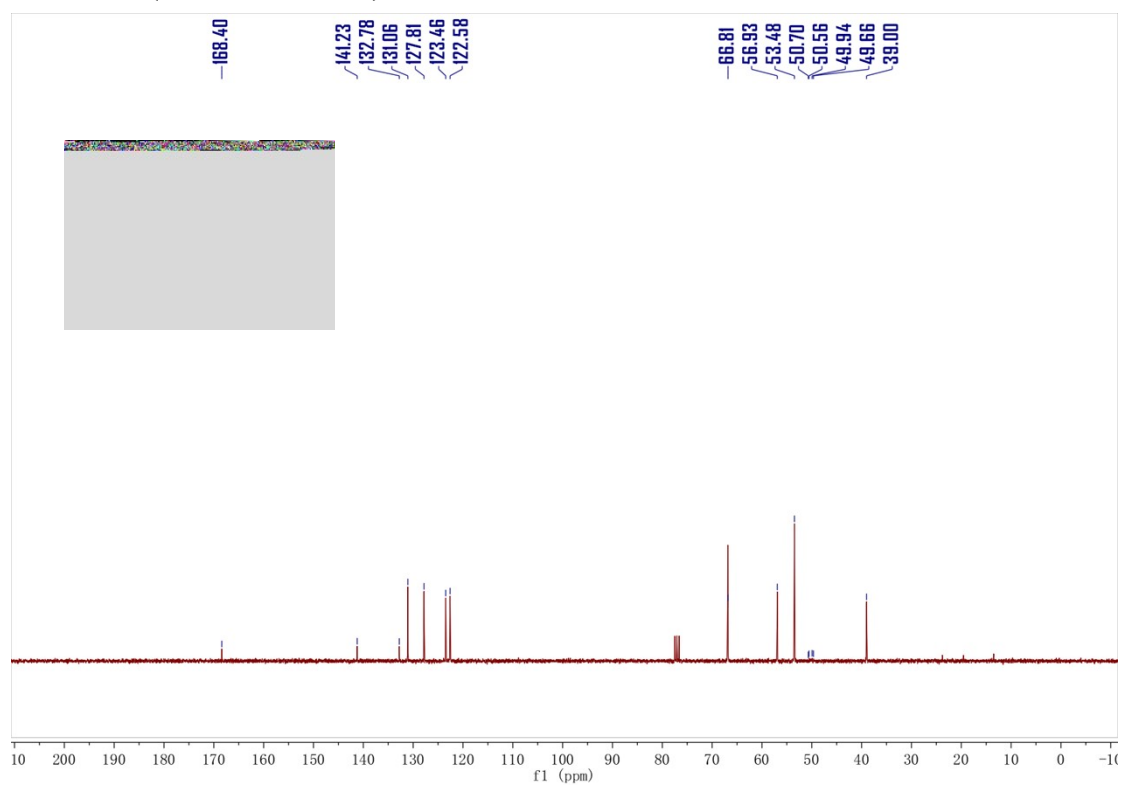
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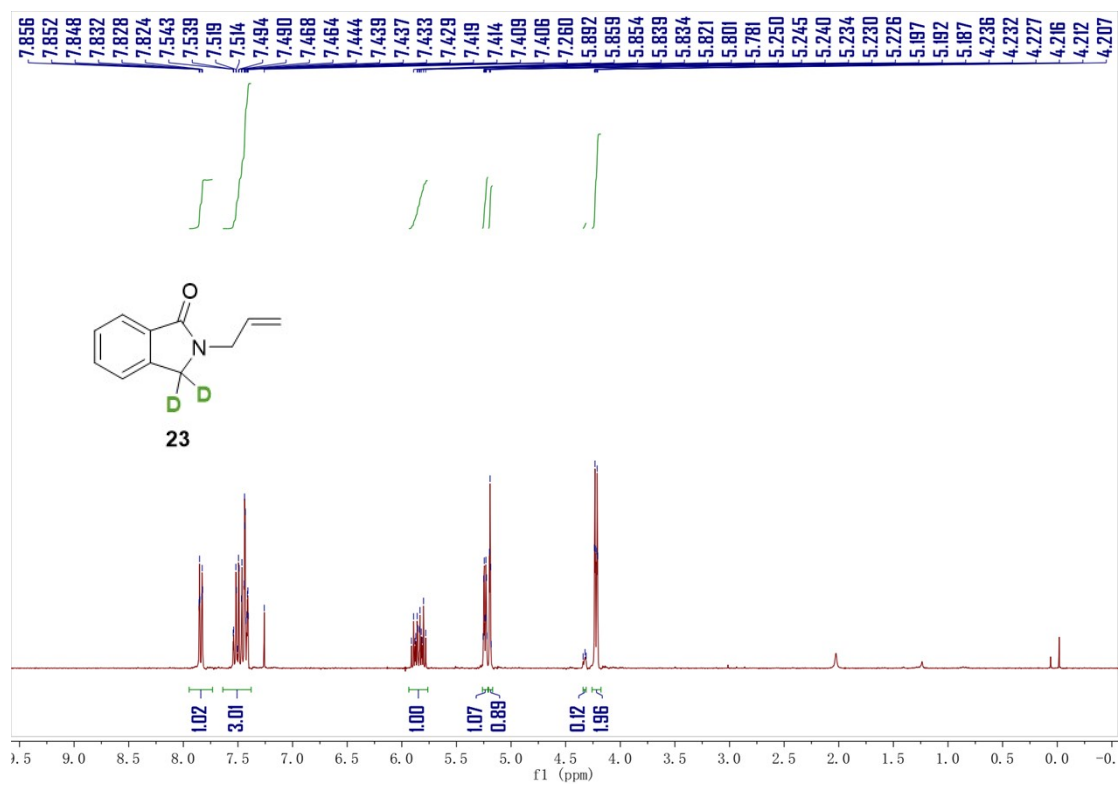
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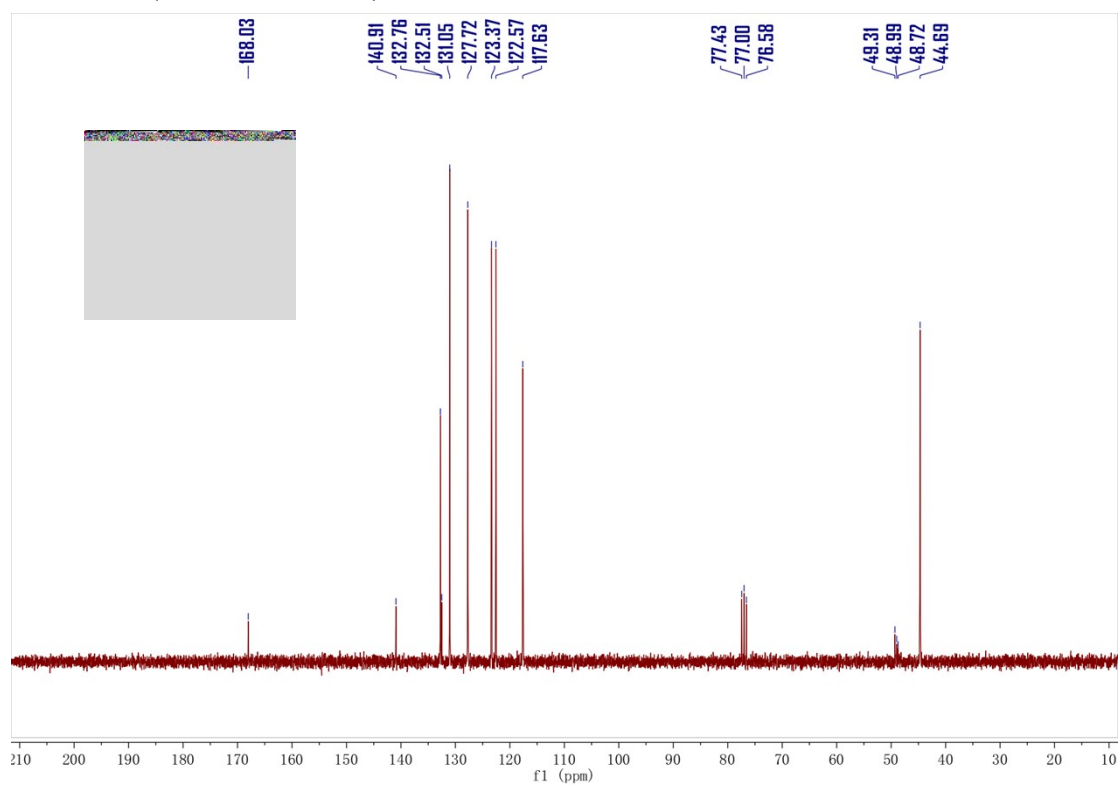
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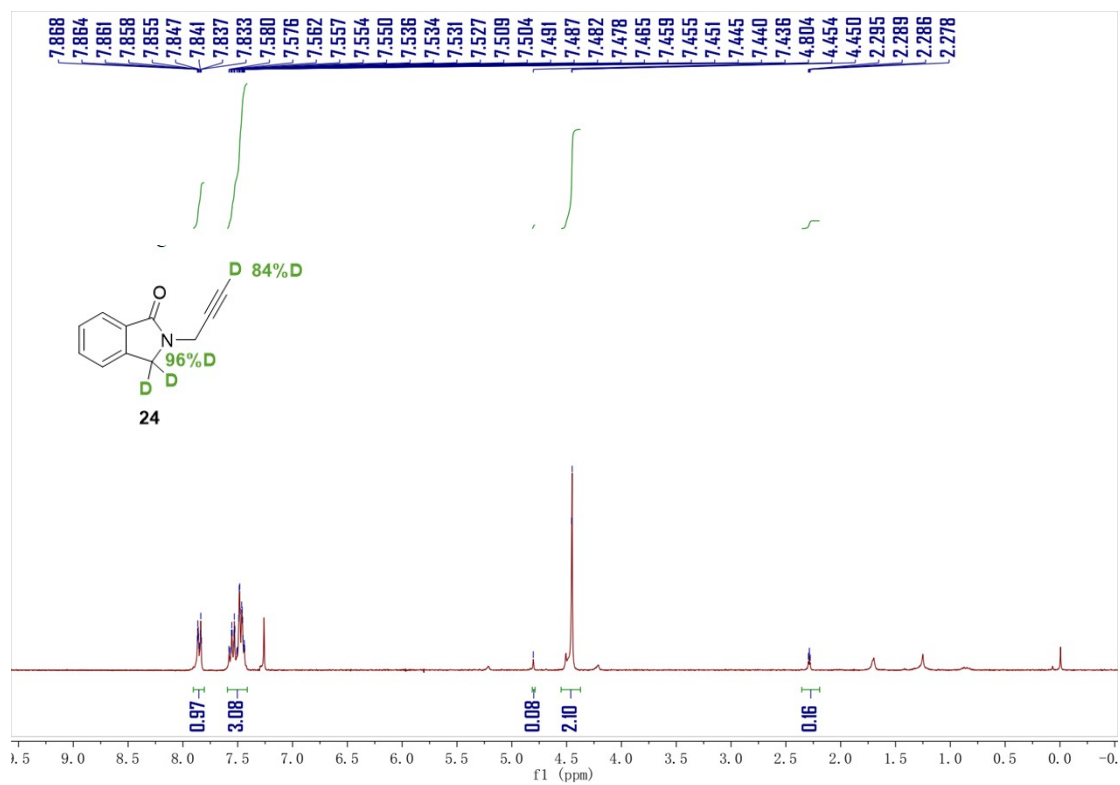
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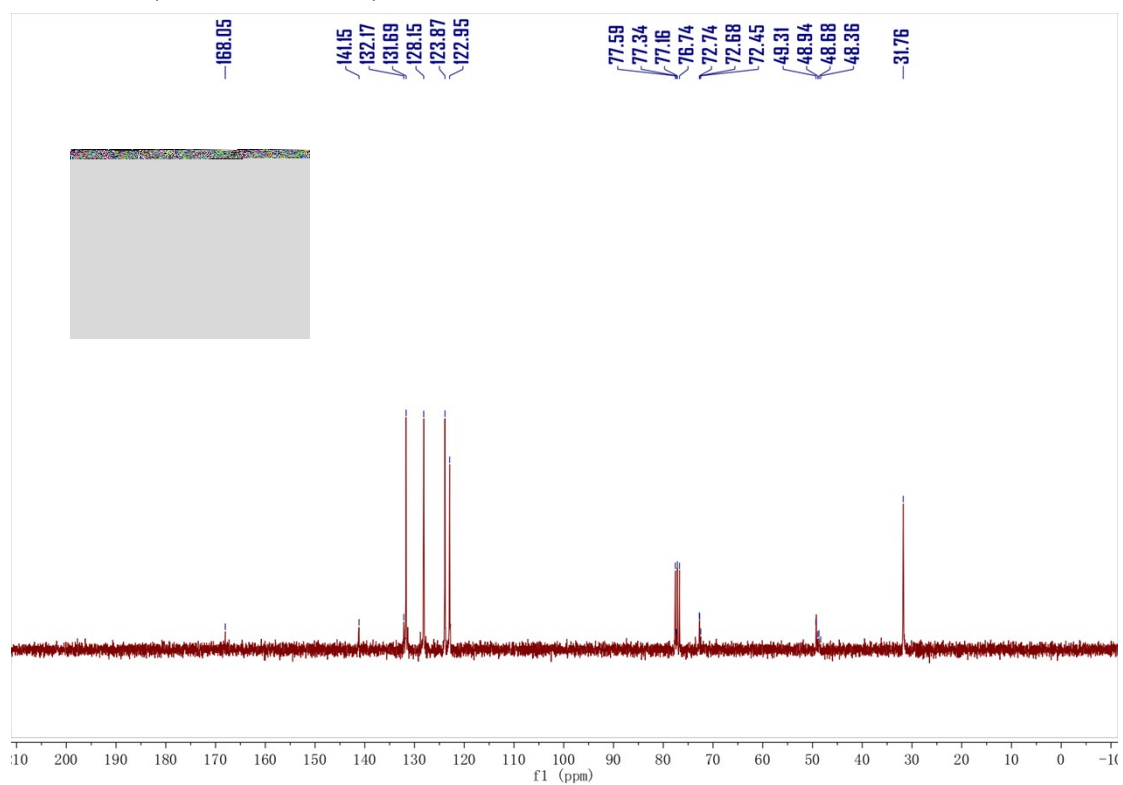
^{13}C NMR (75 MHz, CDCl_3):



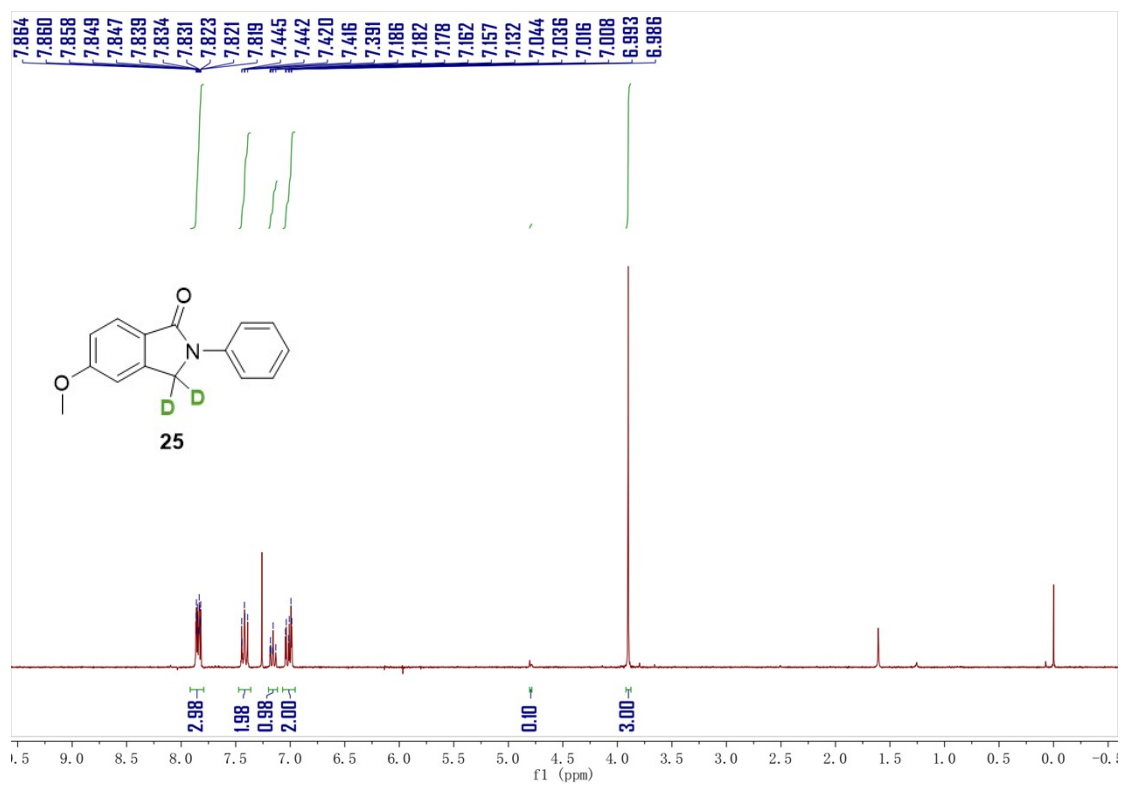
^1H NMR (300 MHz, CDCl_3):



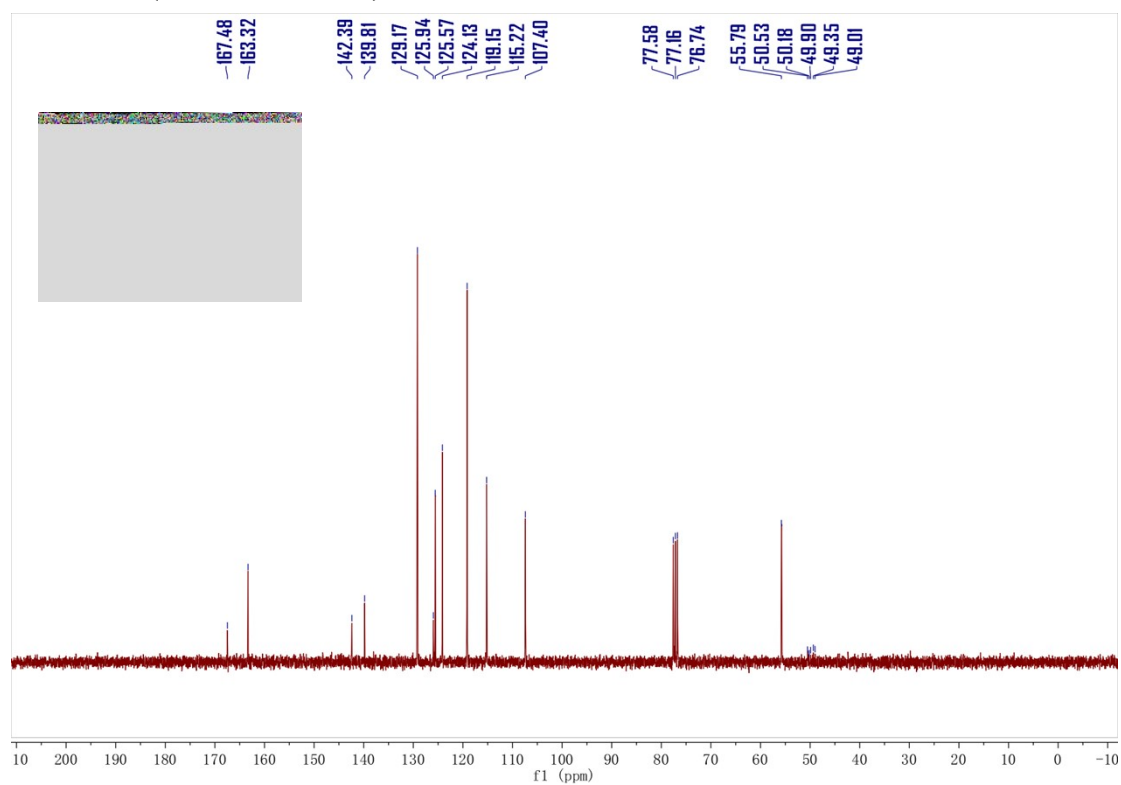
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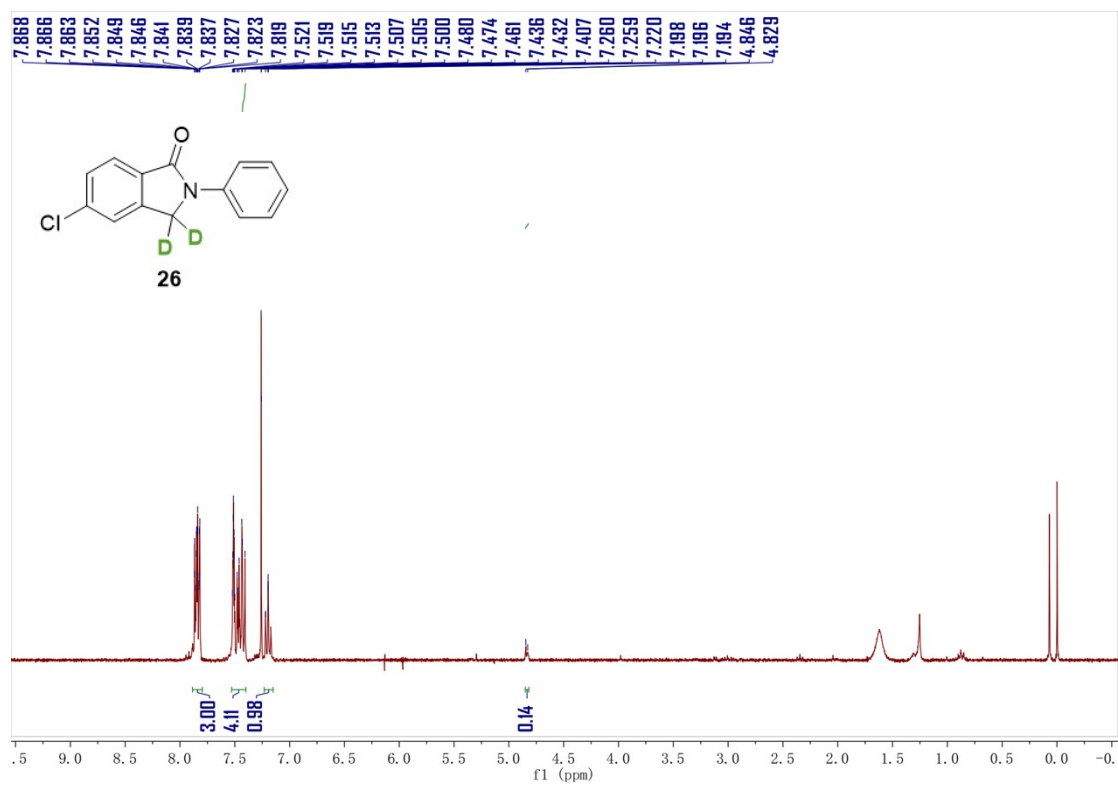
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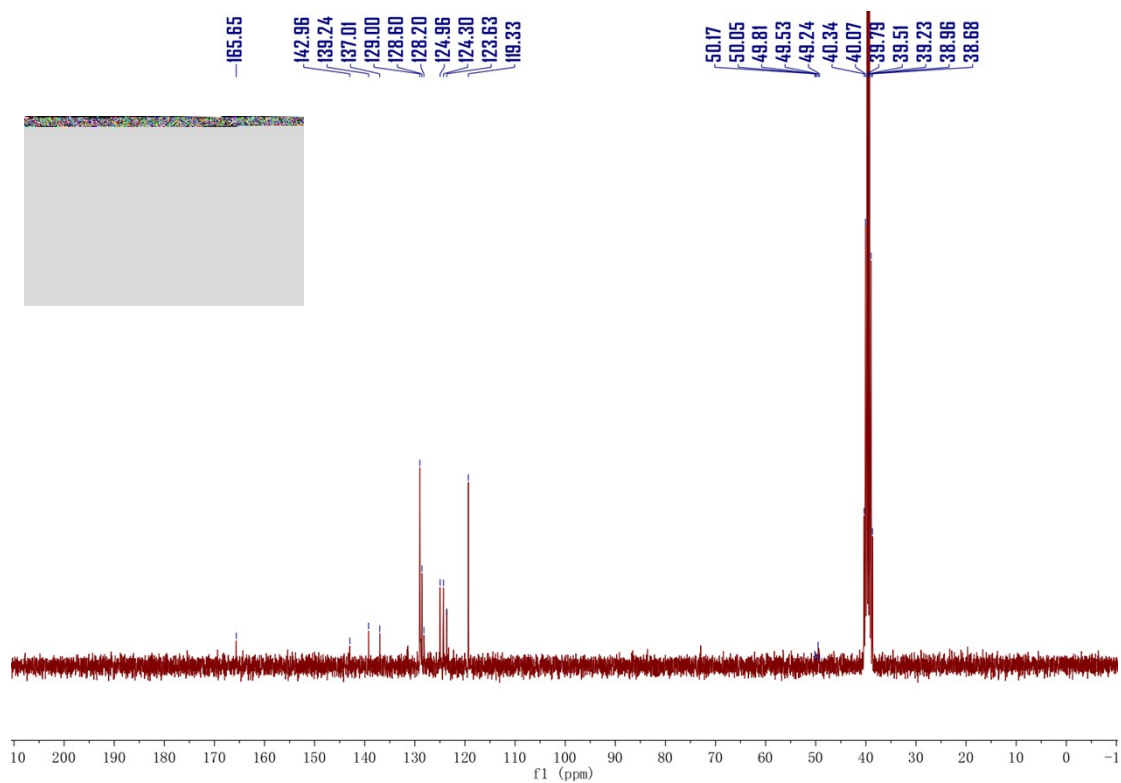
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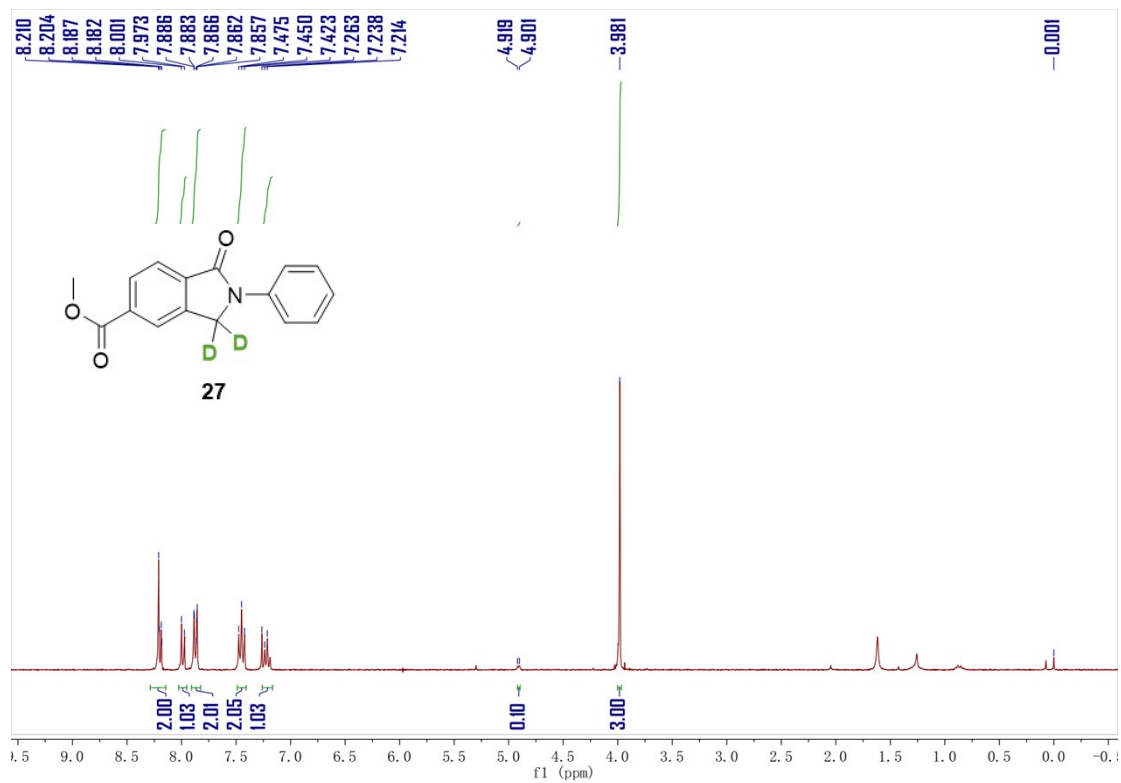
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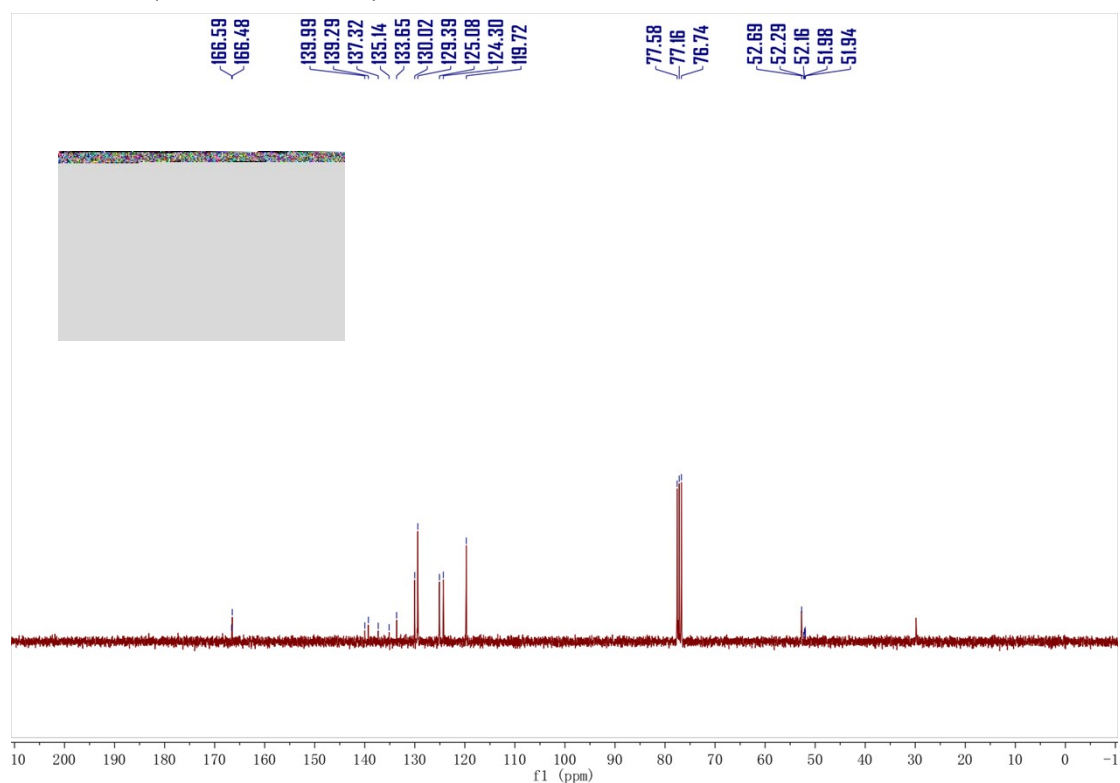
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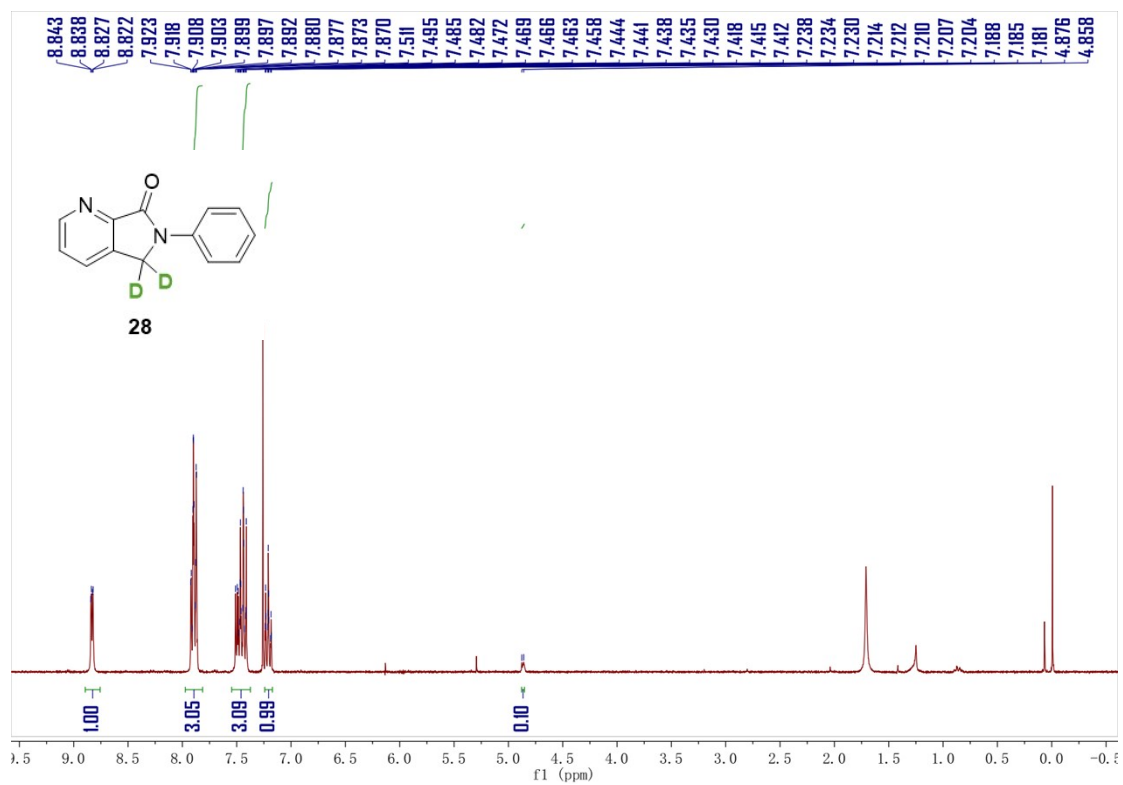
¹H NMR (300 MHz, CDCl₃):



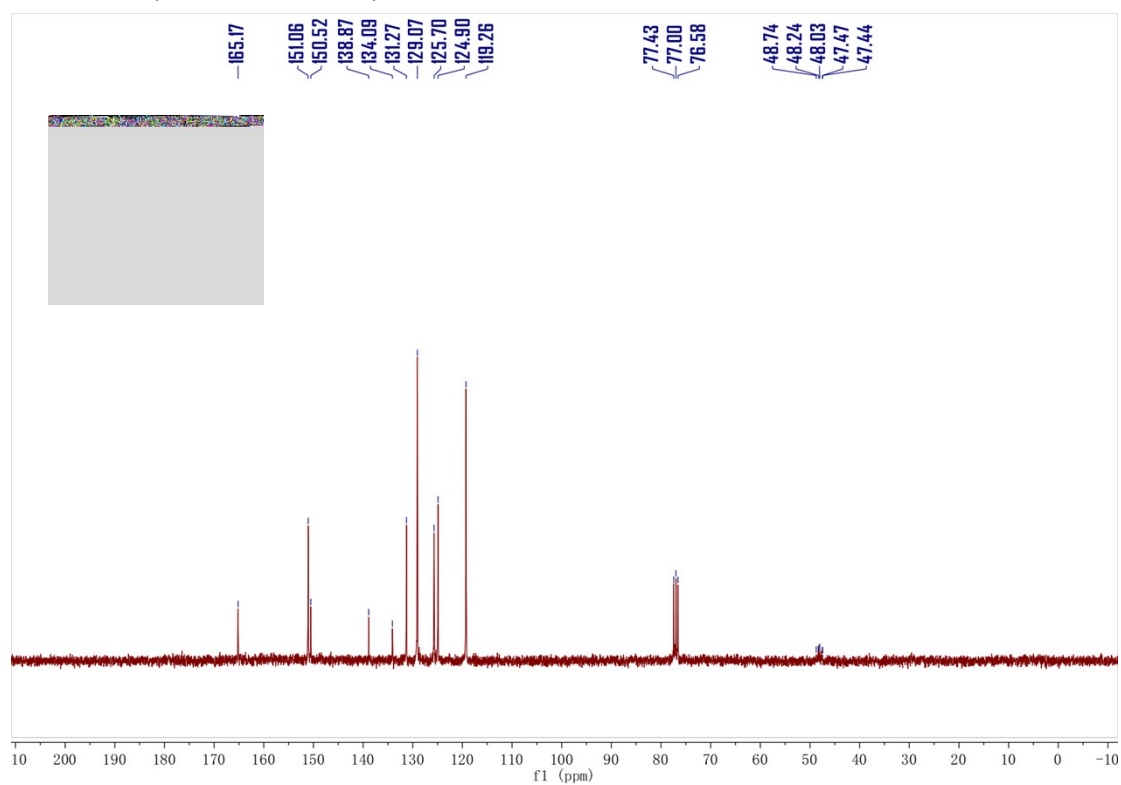
¹³C NMR (75 MHz, CDCl₃):



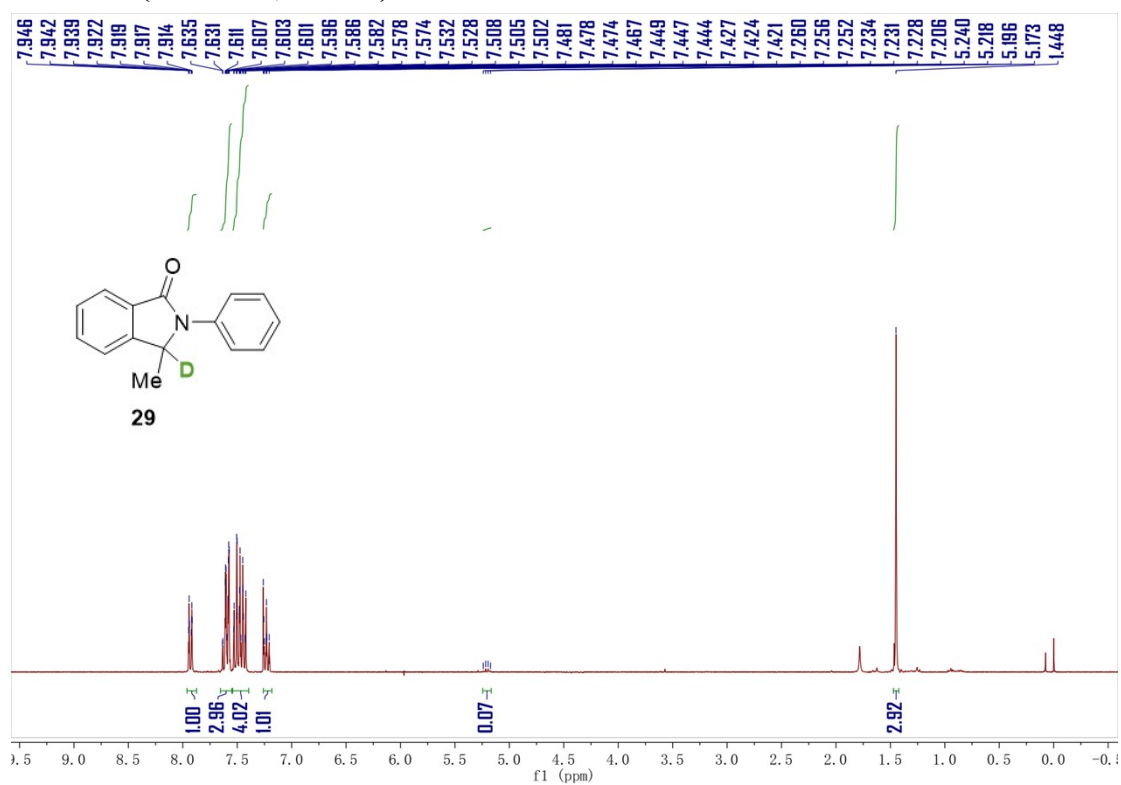
¹H NMR (300 MHz, CDCl₃):



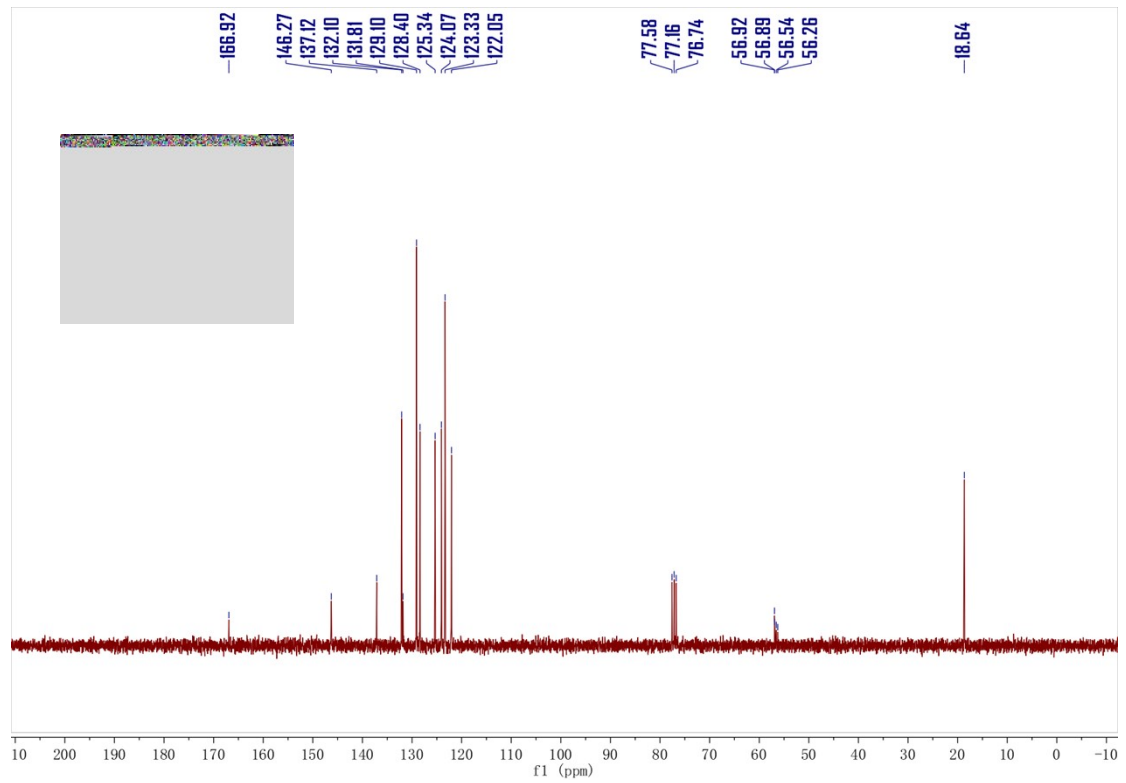
^{13}C NMR (75 MHz, CDCl_3):



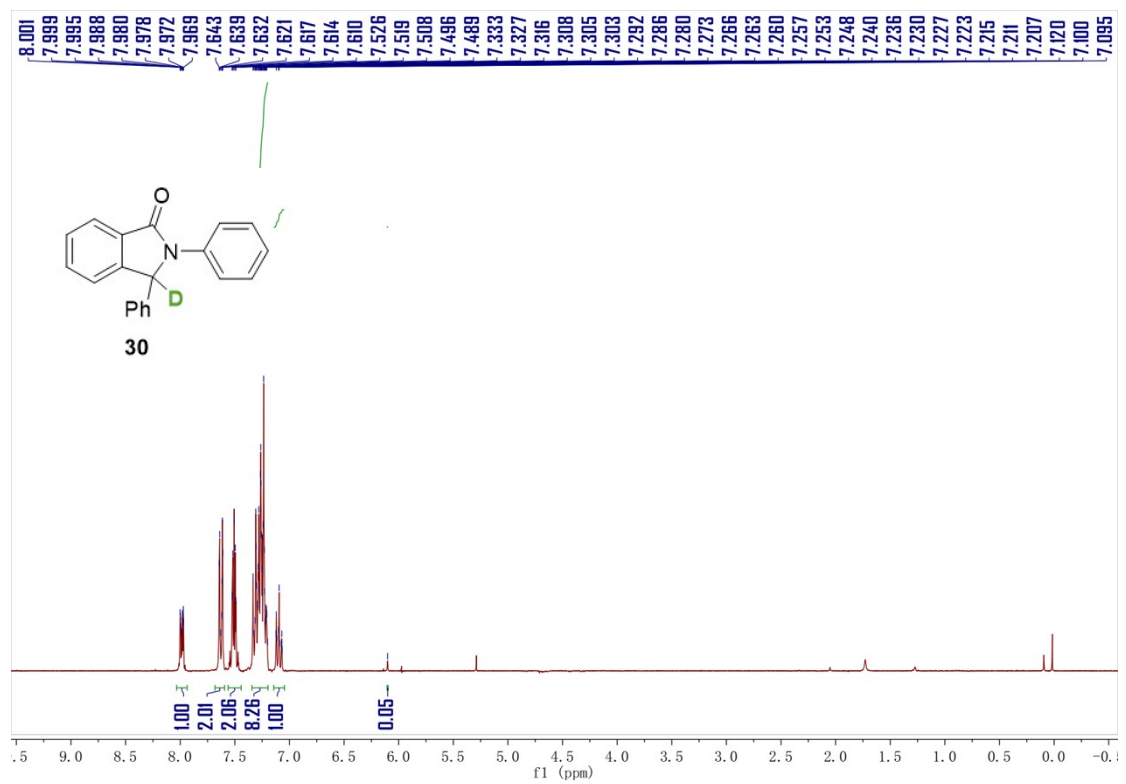
^1H NMR (300 MHz, CDCl_3):



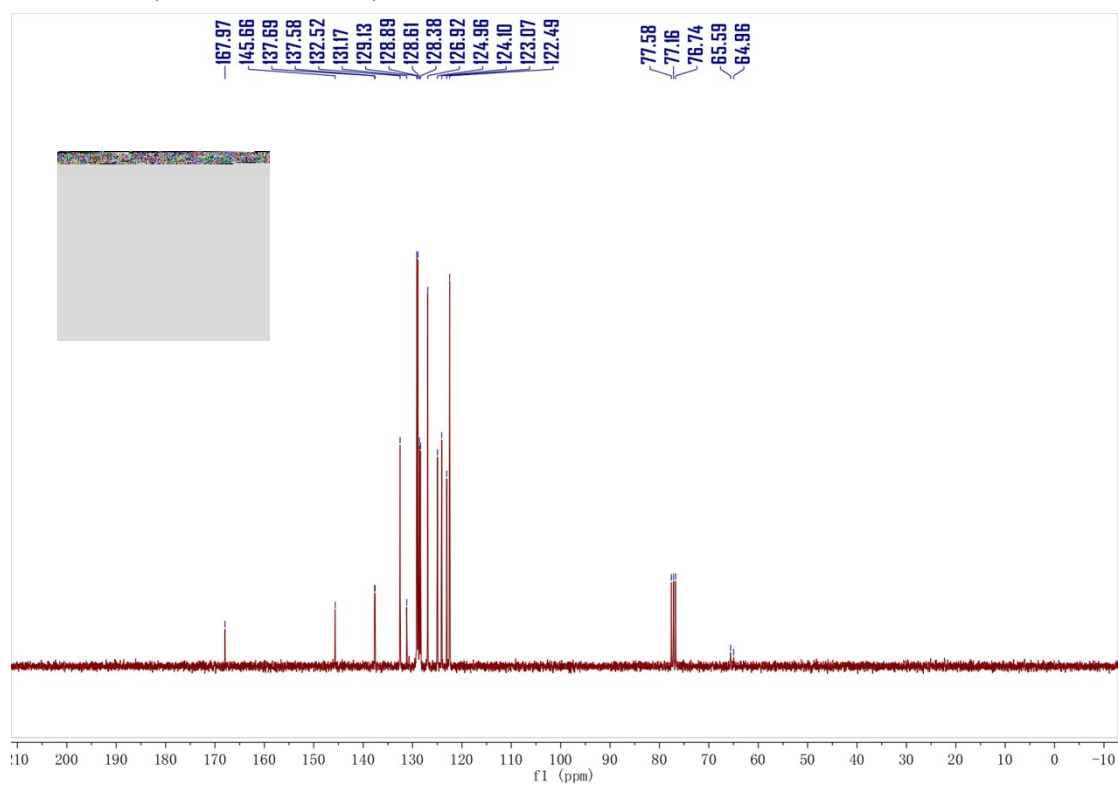
^{13}C NMR (75 MHz, CDCl_3):



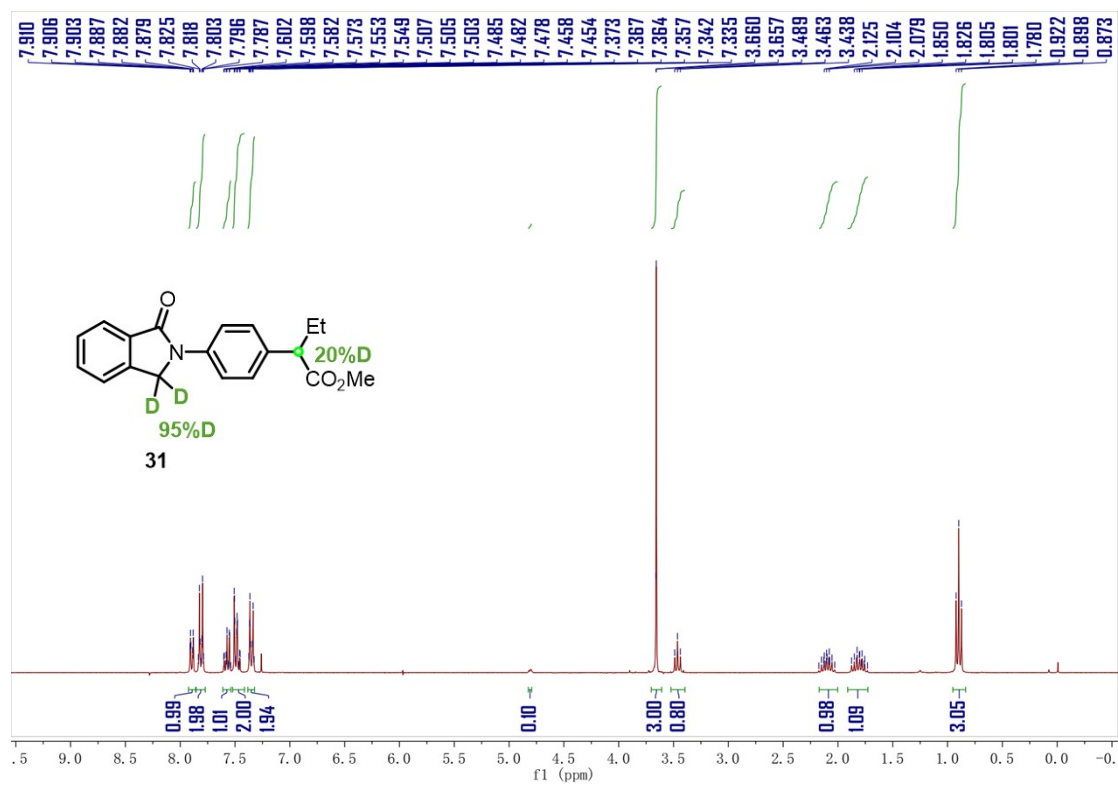
^1H NMR (300 MHz, CDCl_3):



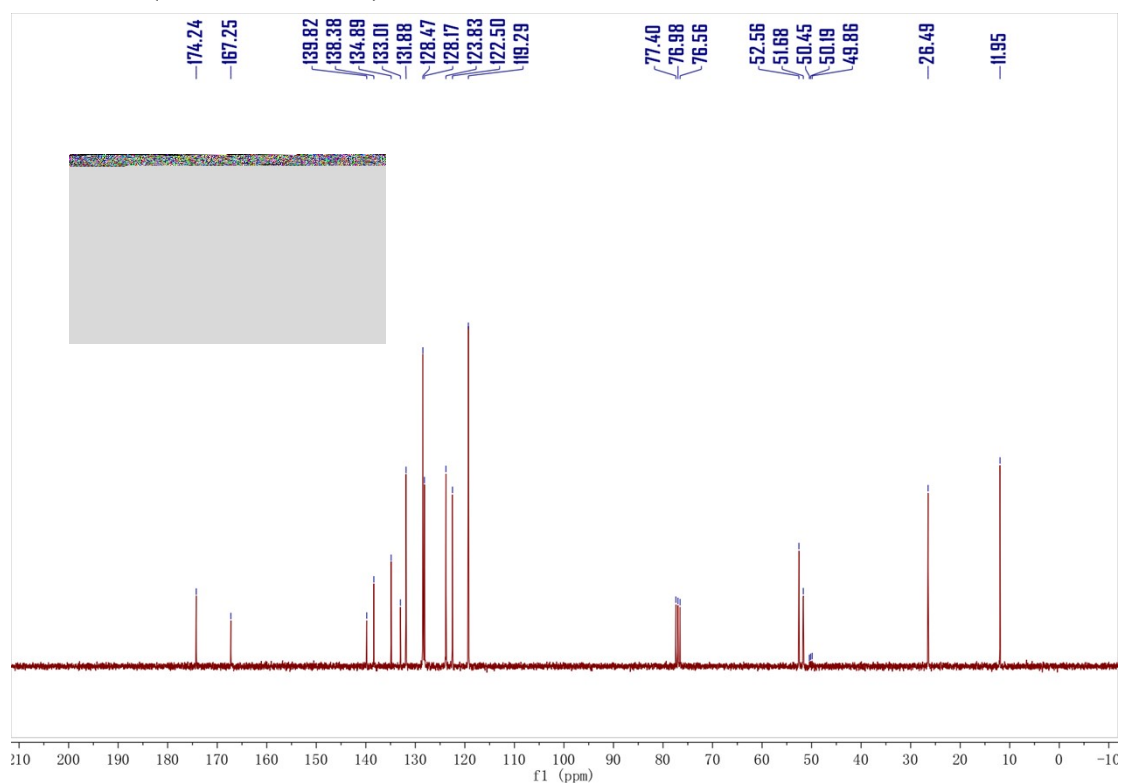
¹³C NMR (75 MHz, CDCl₃):



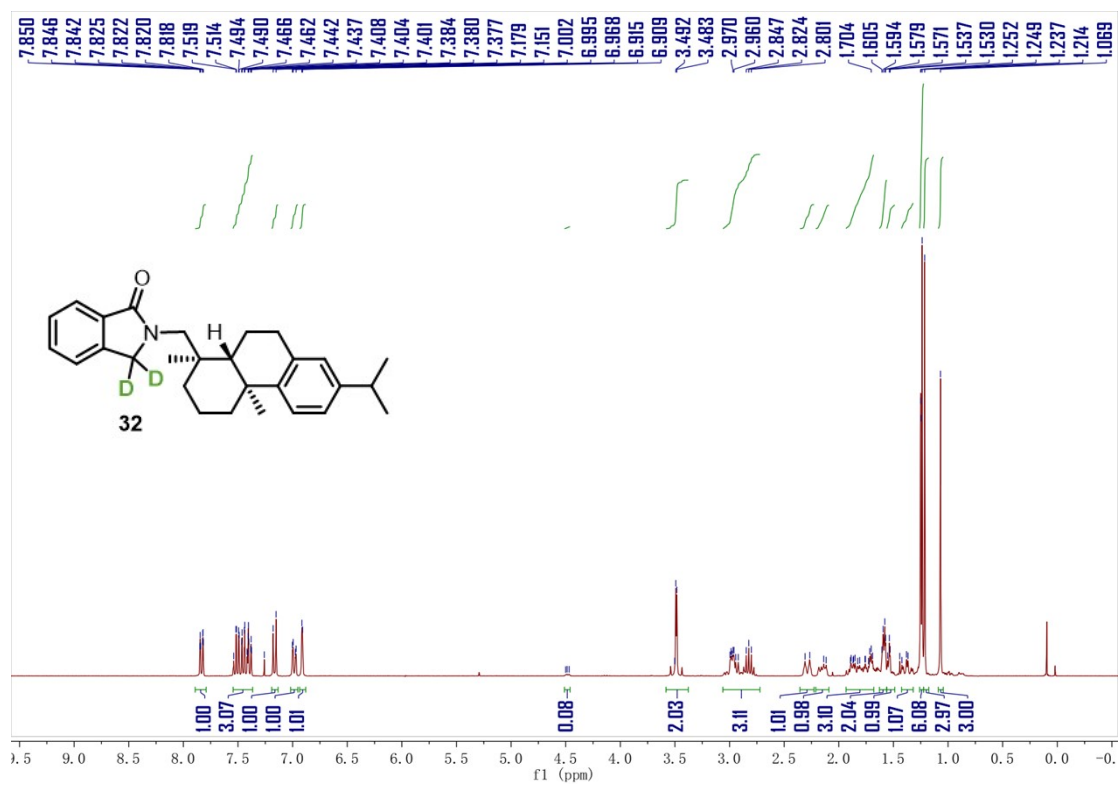
¹H NMR (300 MHz, CDCl₃):



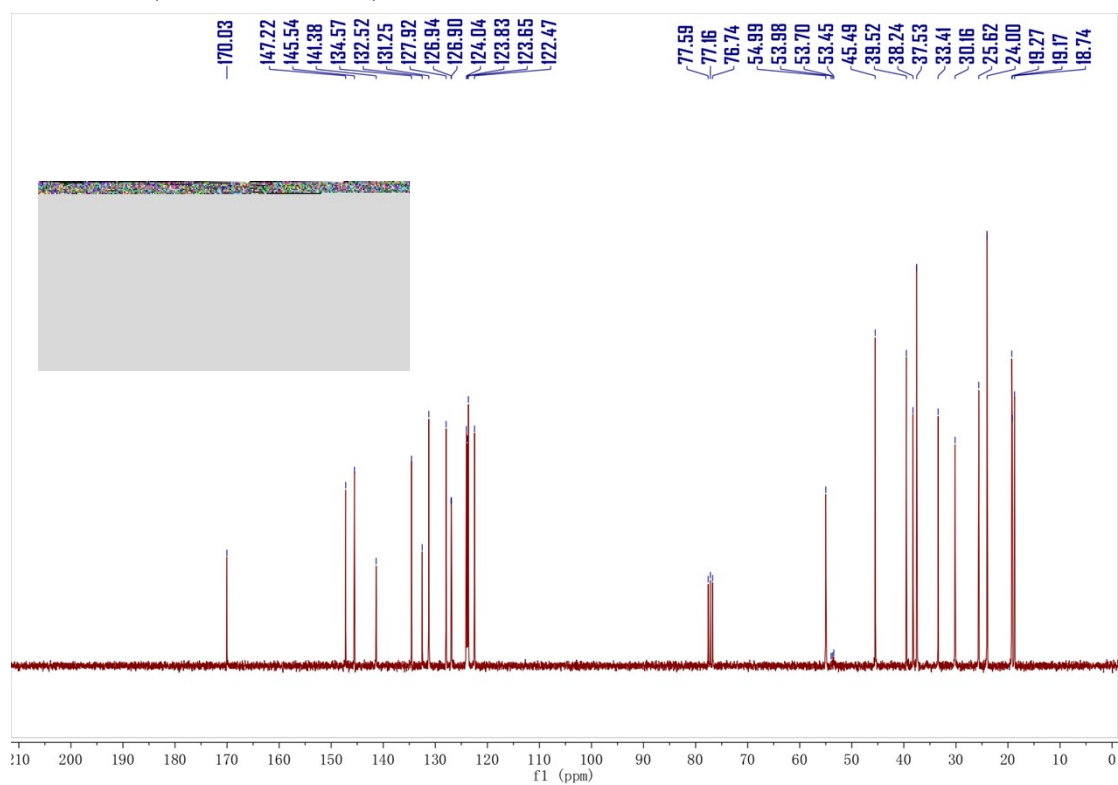
¹³C NMR (75 MHz, CDCl₃):



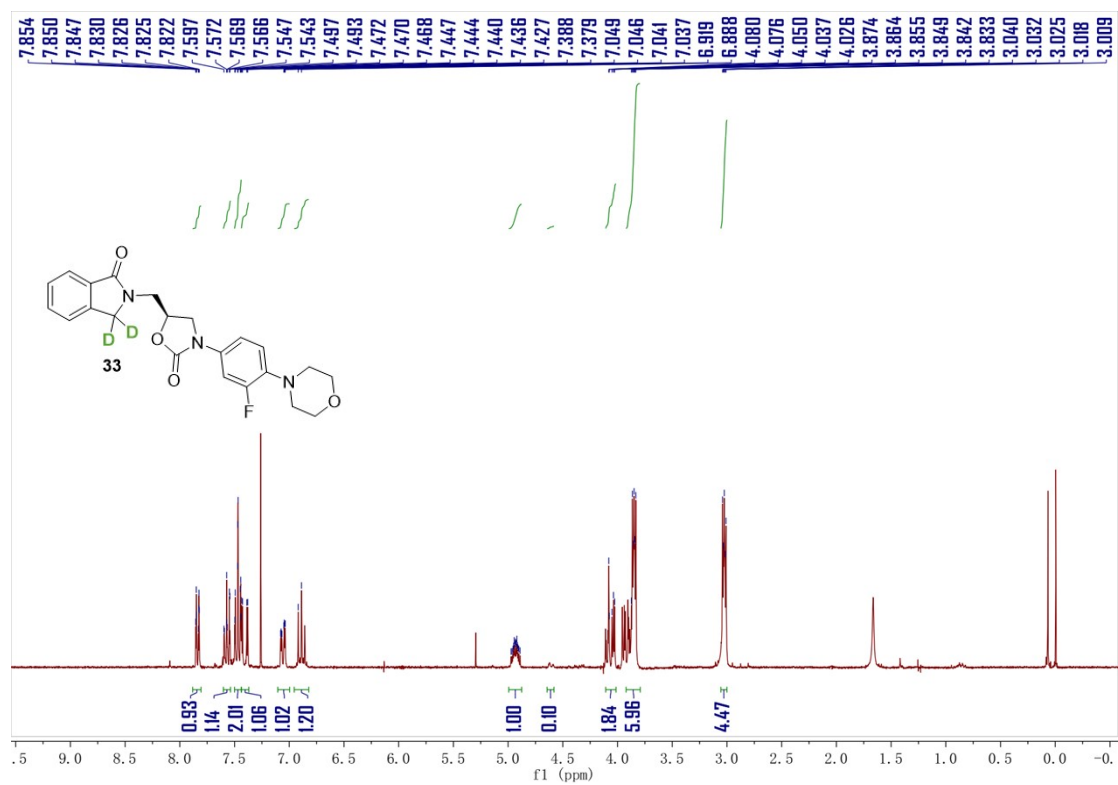
¹H NMR (300 MHz, CDCl₃):



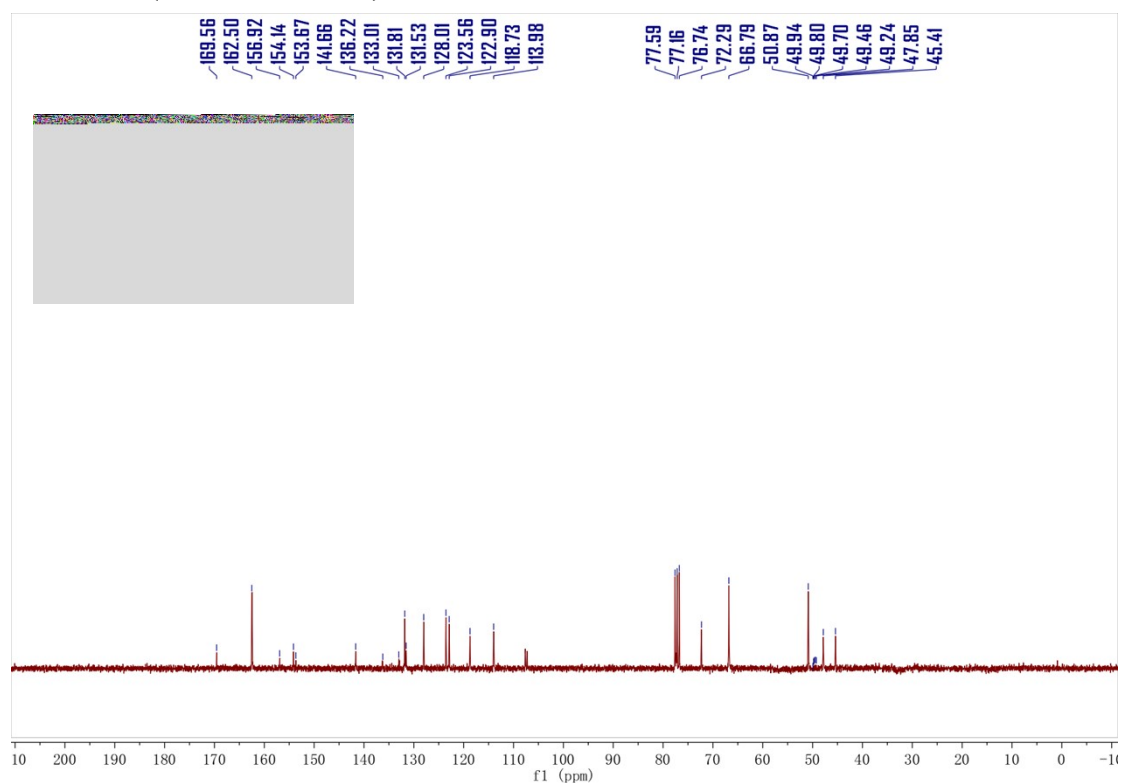
¹³C NMR (75 MHz, CDCl₃):



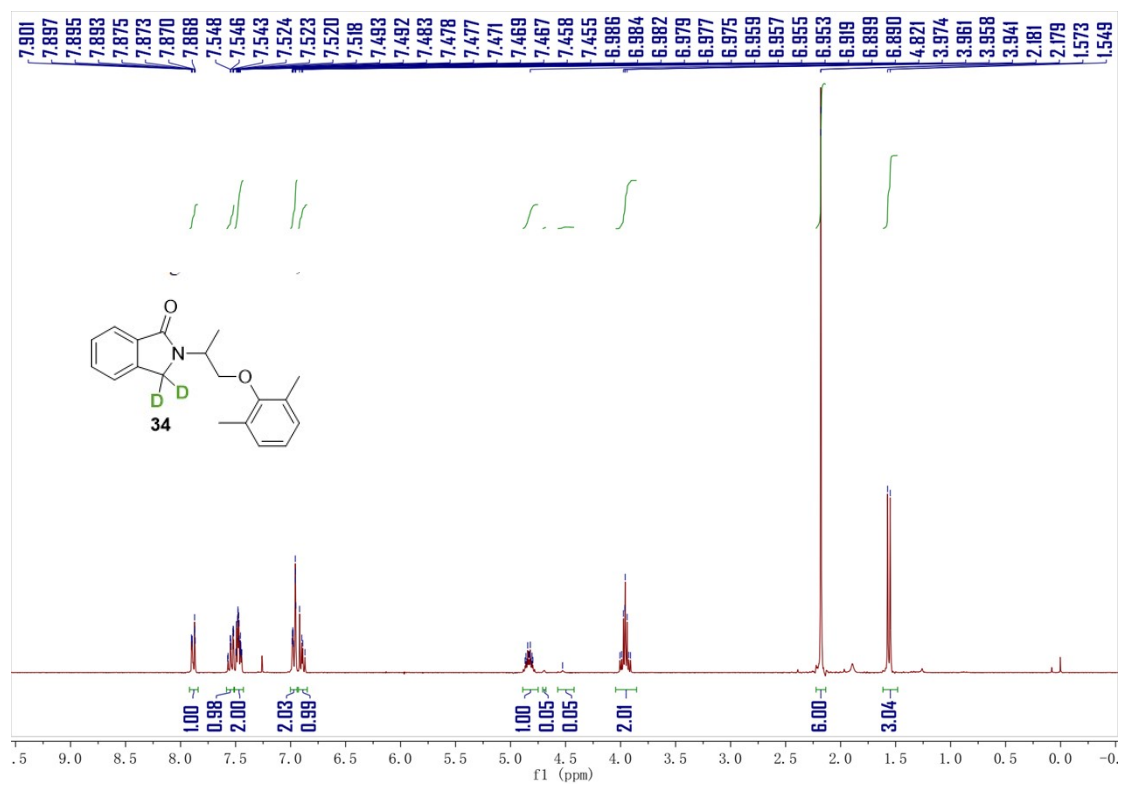
¹H NMR (300 MHz, CDCl₃):



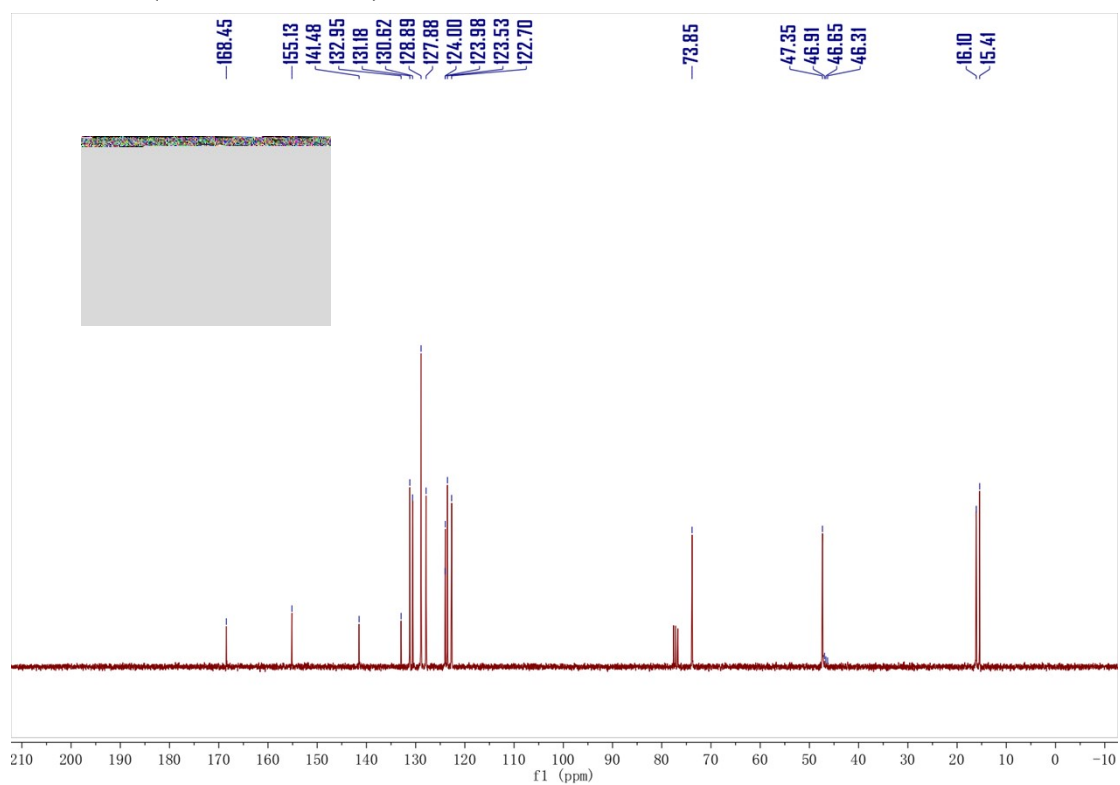
¹³C NMR (75 MHz, CDCl₃):



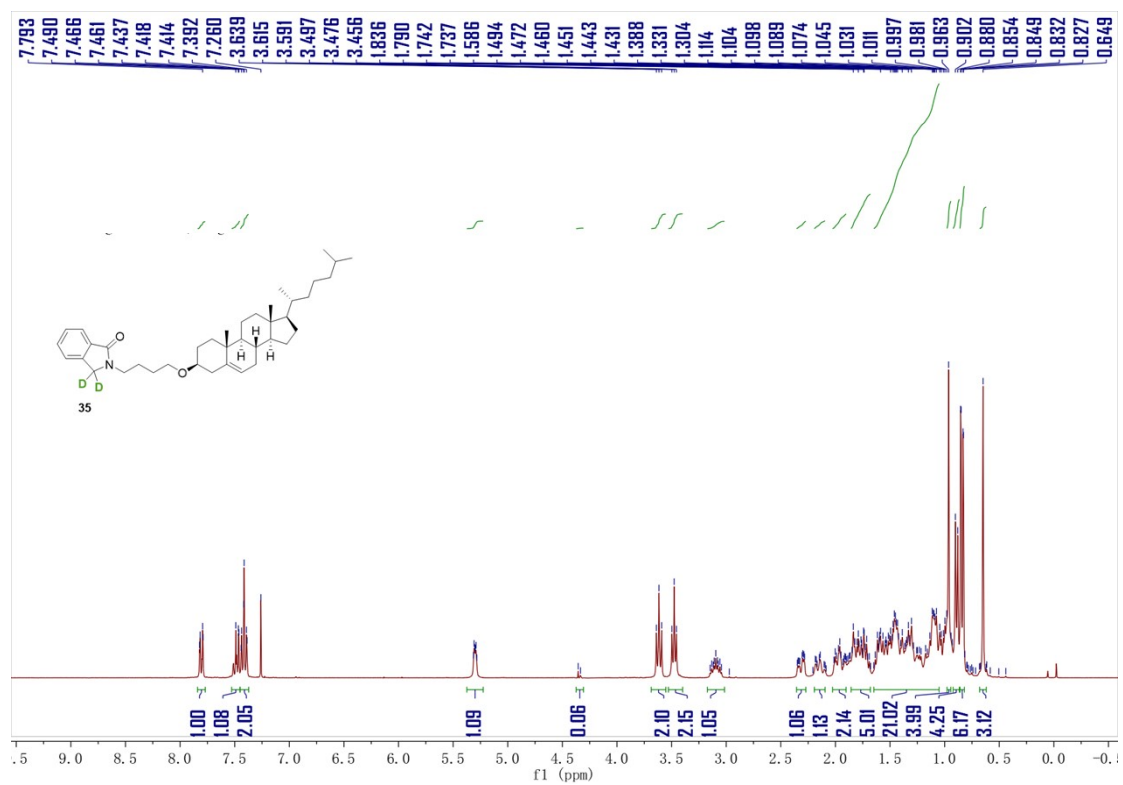
¹H NMR (300 MHz, CDCl₃):



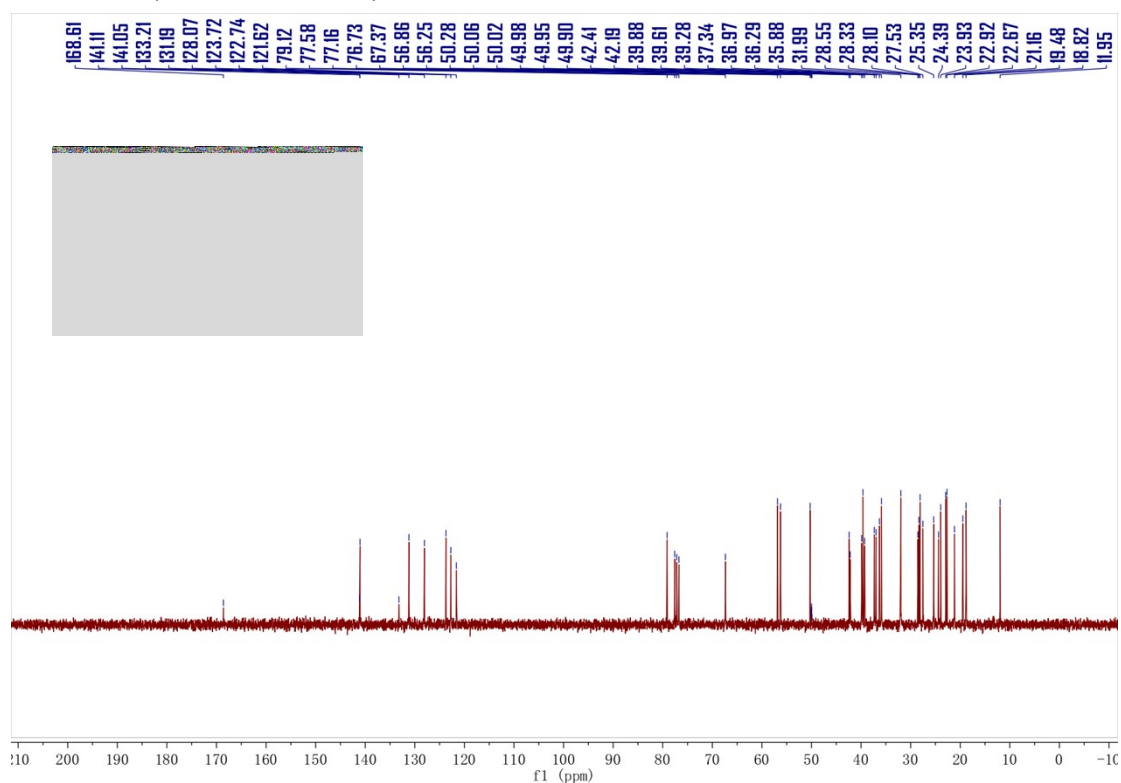
¹³C NMR (75 MHz, CDCl₃):



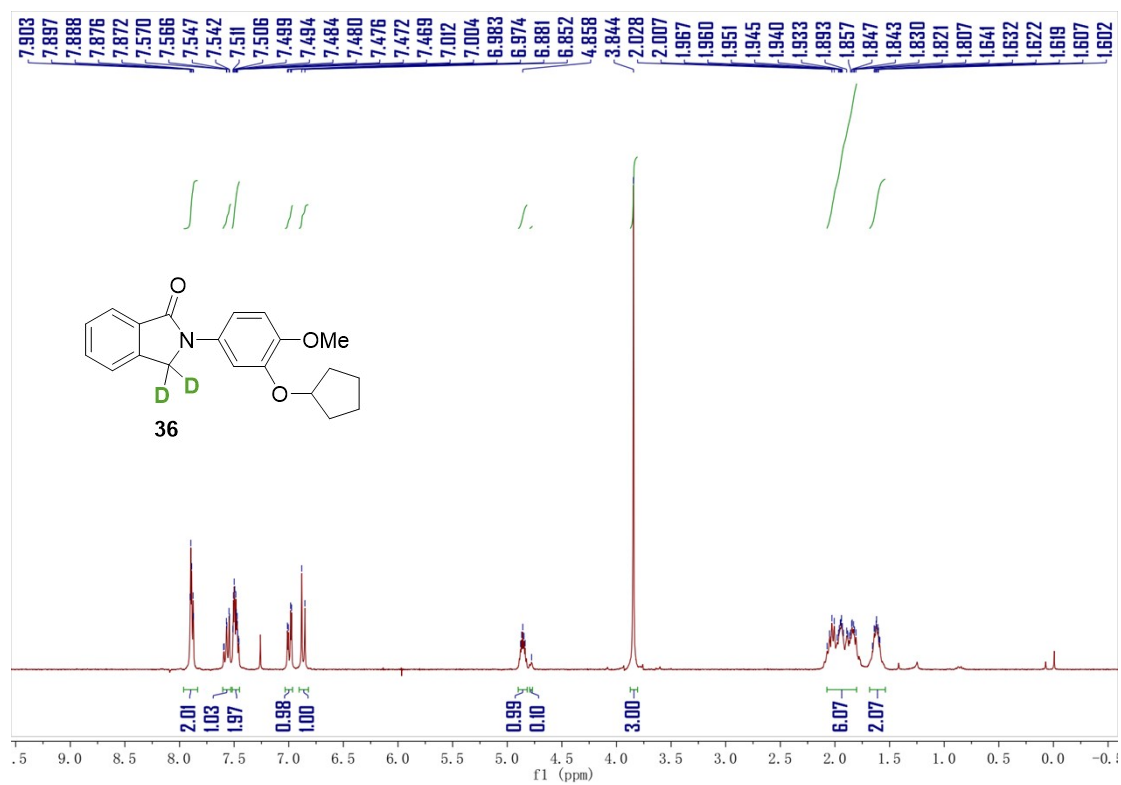
¹H NMR (300 MHz, CDCl₃):



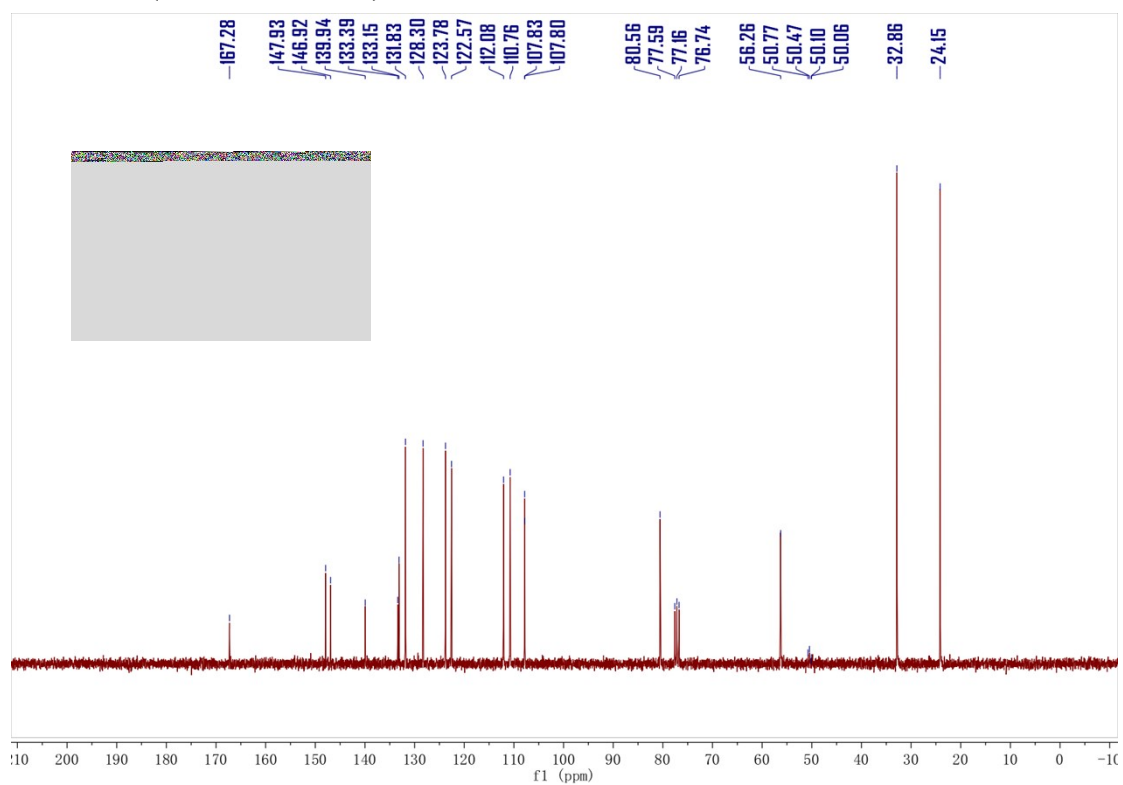
¹³C NMR (75 MHz, CDCl₃):



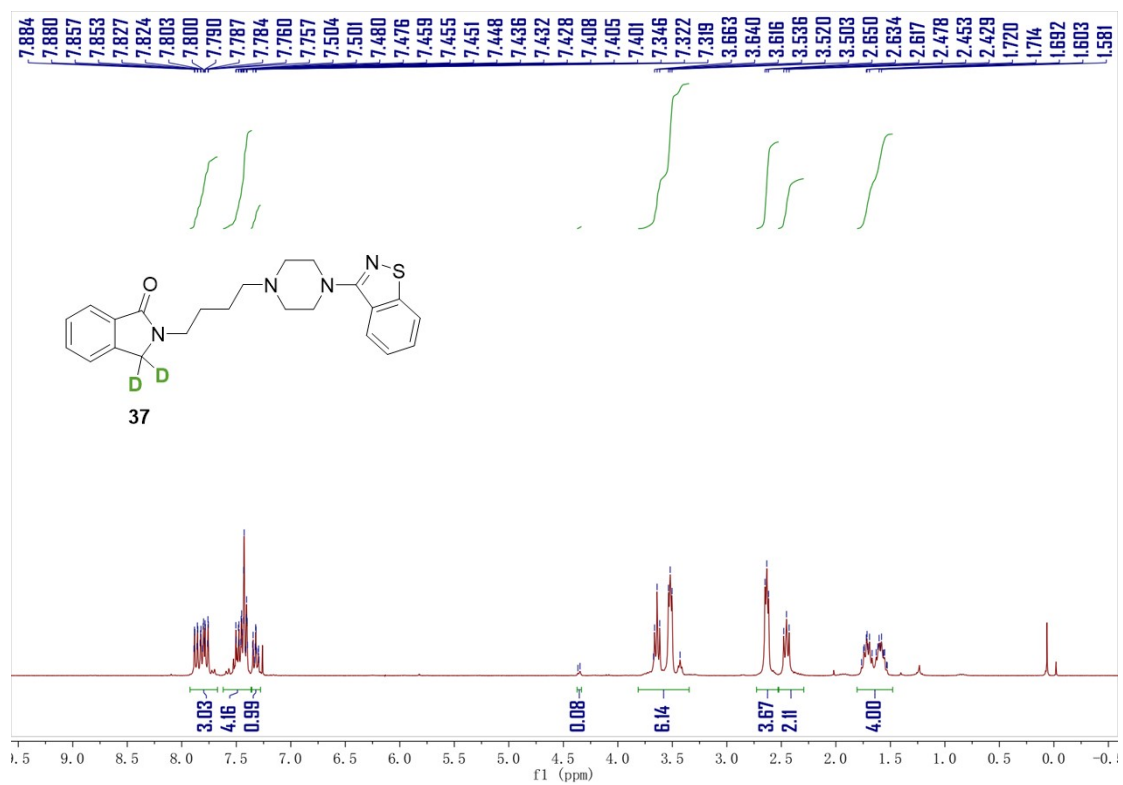
¹H NMR (300 MHz, CDCl₃):



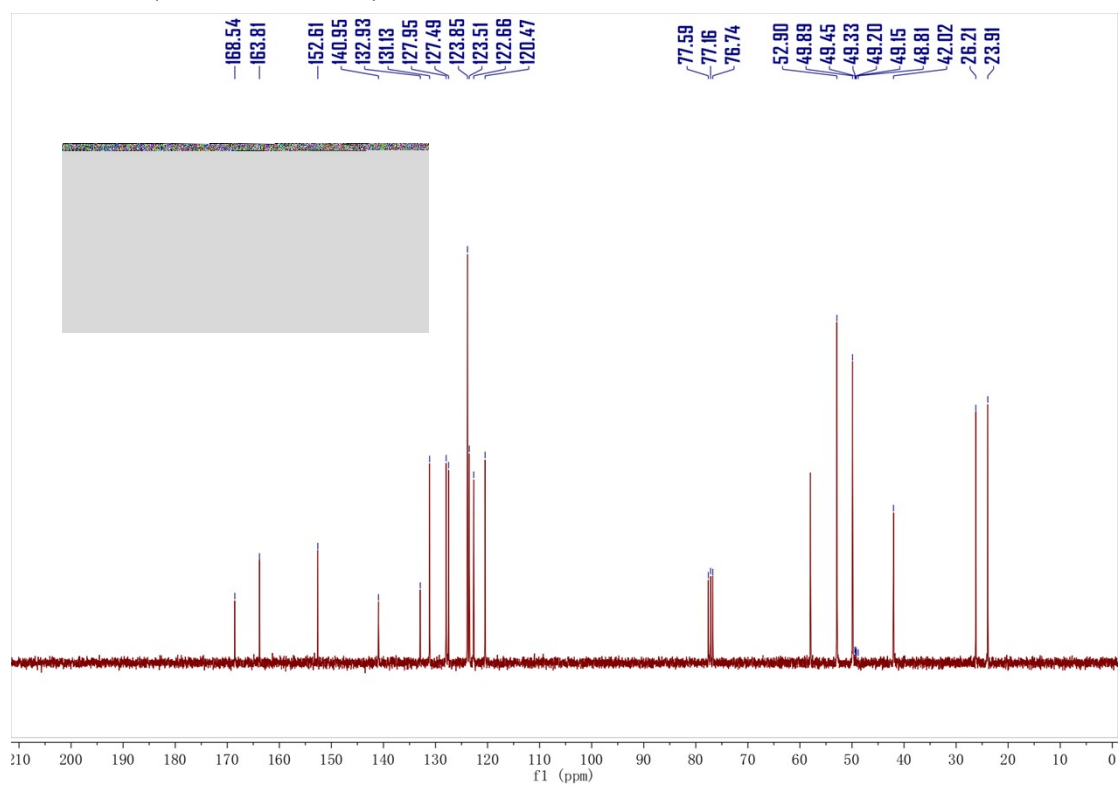
¹³C NMR (75 MHz, CDCl₃):



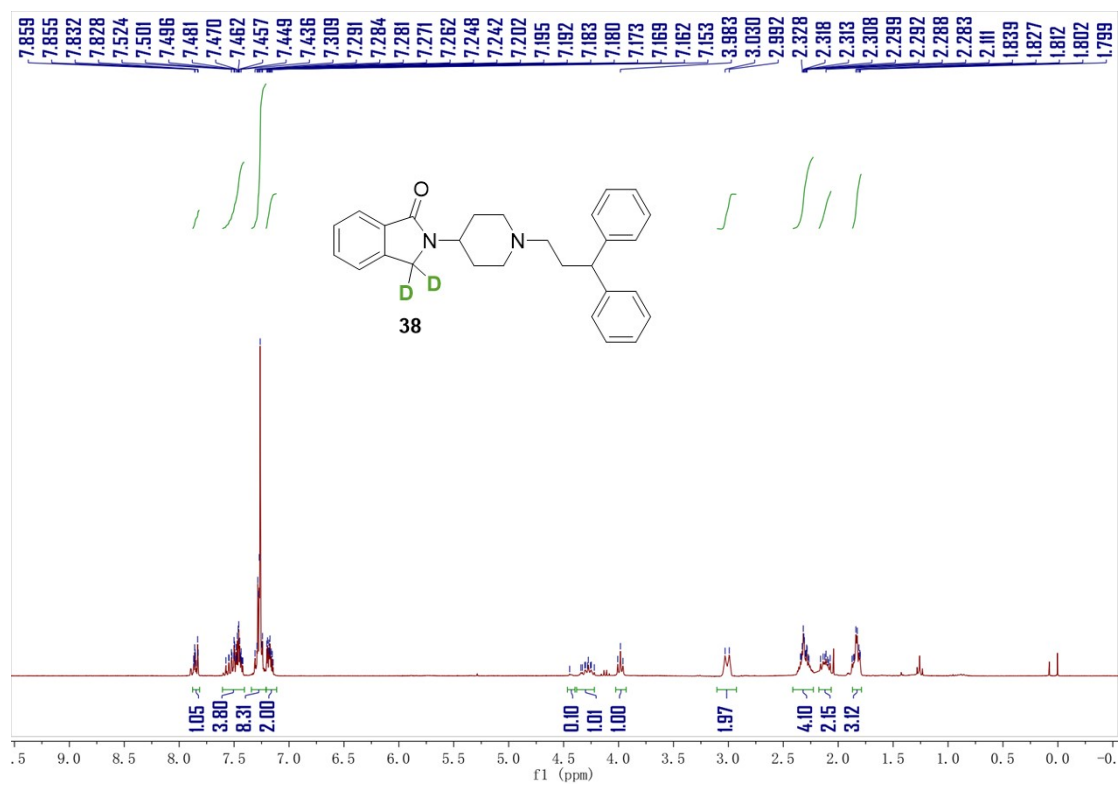
¹H NMR (300 MHz, CDCl₃):



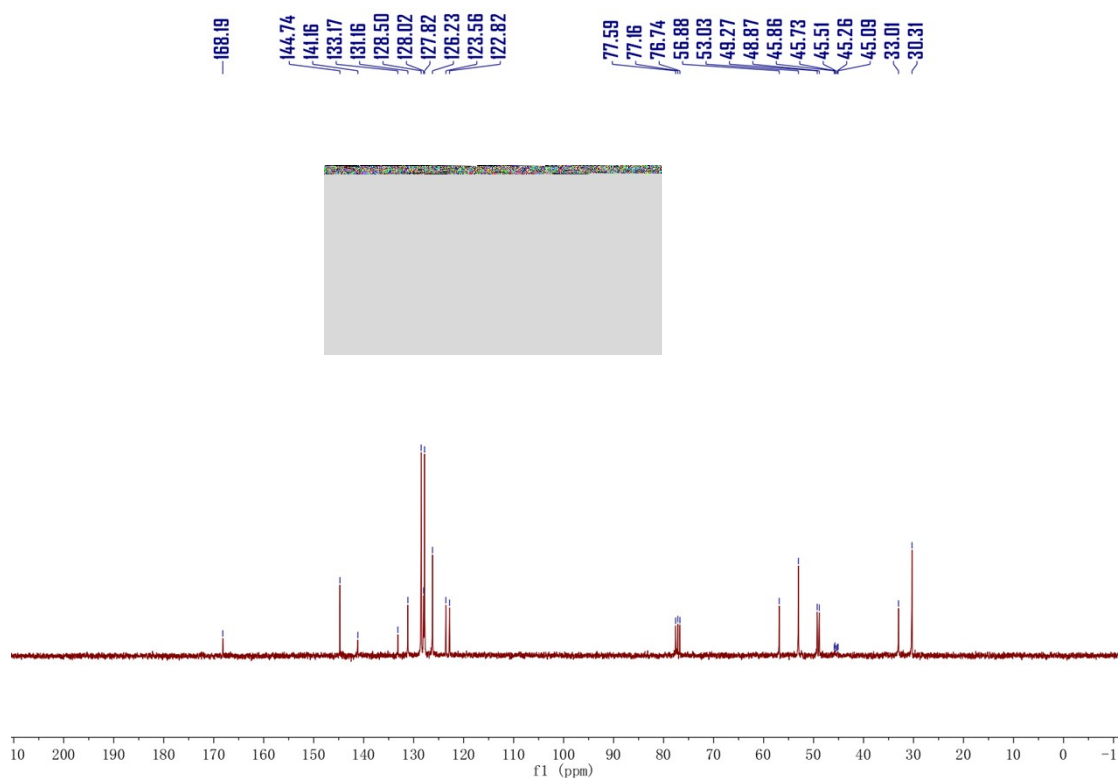
¹³C NMR (75 MHz, CDCl₃):



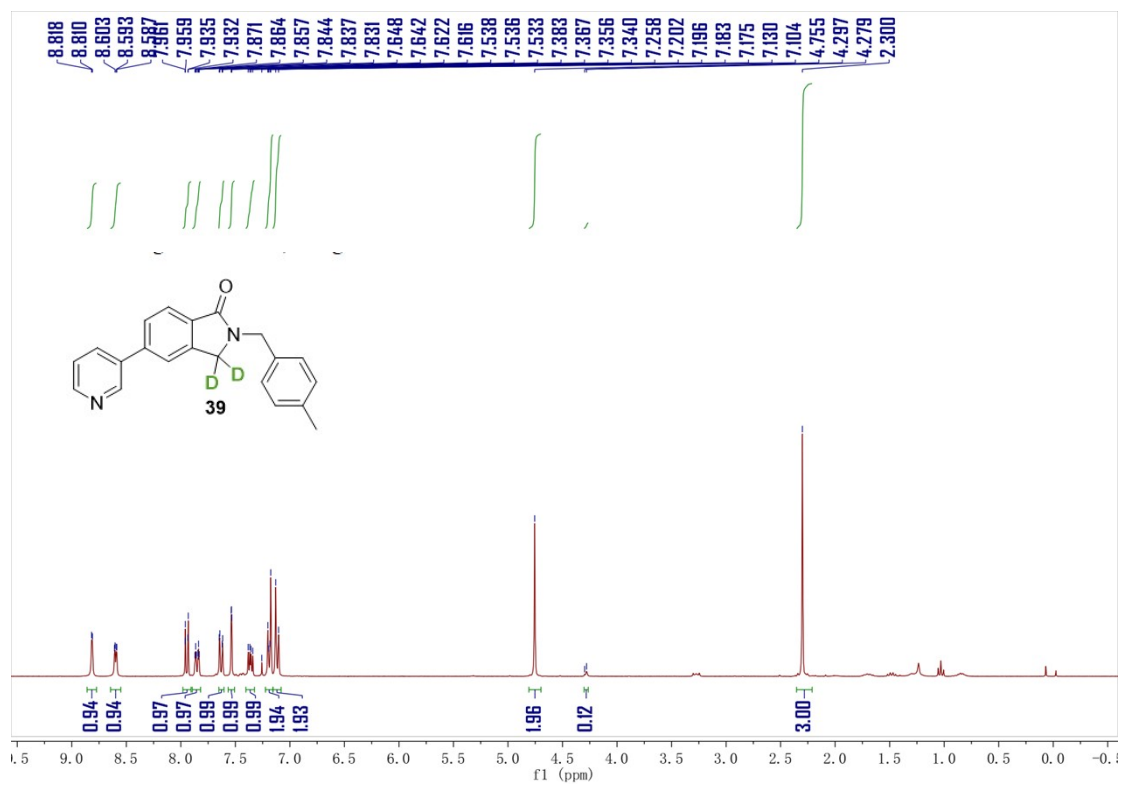
¹H NMR (300 MHz, CDCl₃):



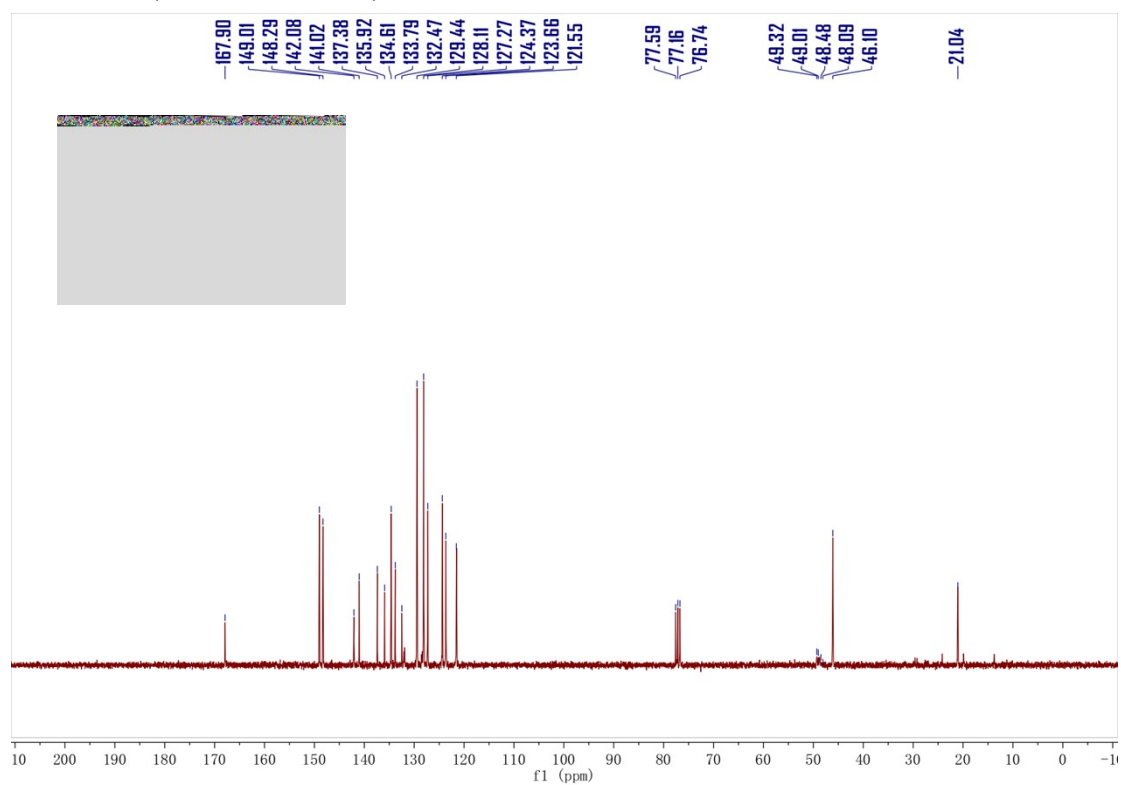
¹³C NMR (75 MHz, CDCl₃):



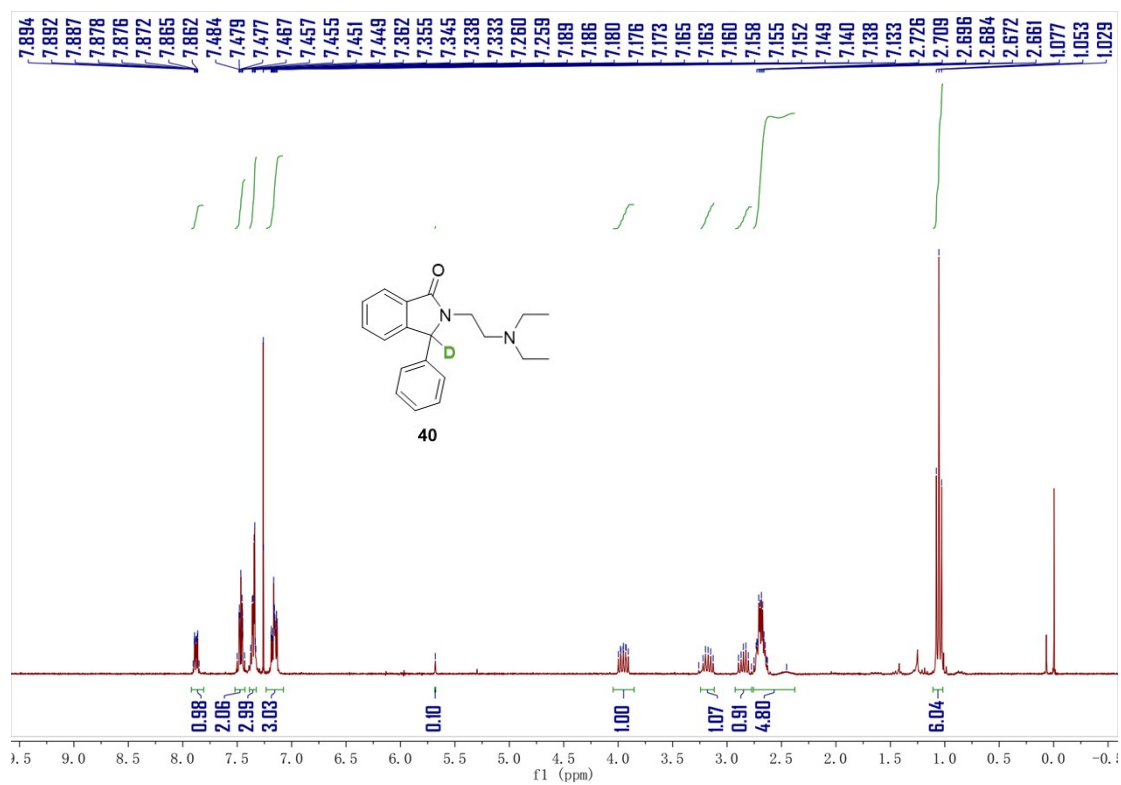
¹H NMR (300 MHz, CDCl₃):



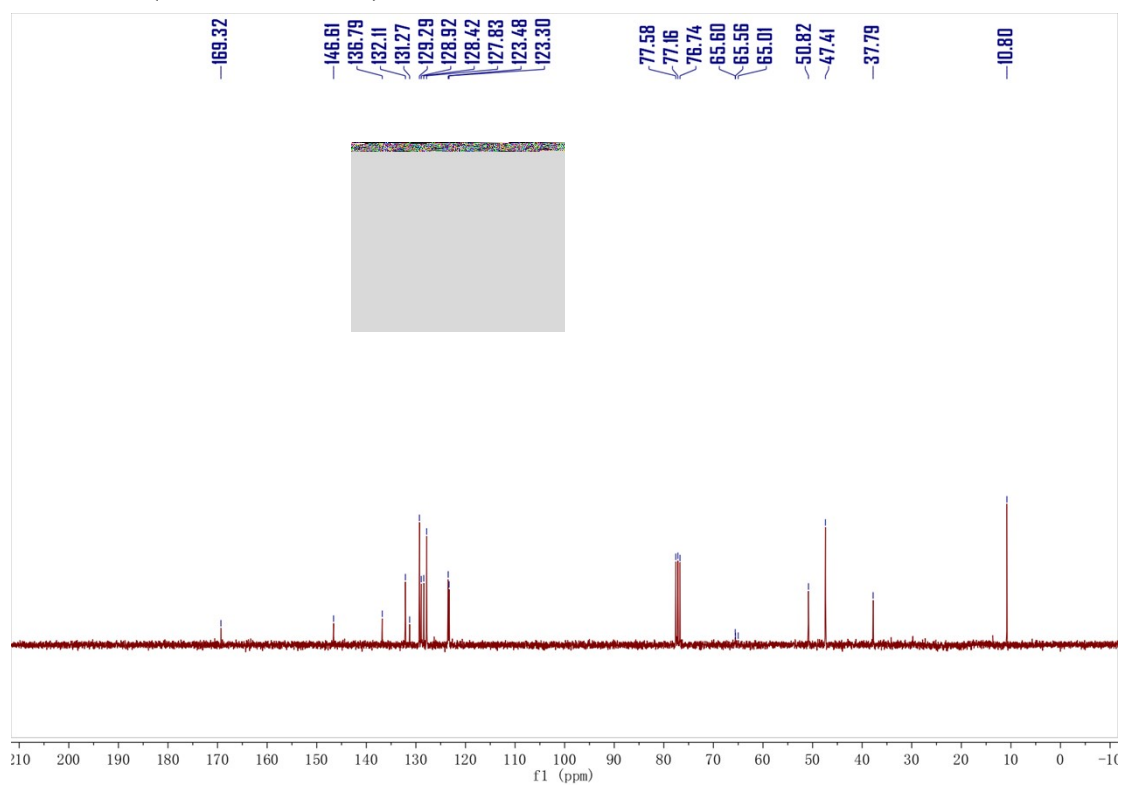
¹³C NMR (75 MHz, CDCl₃):



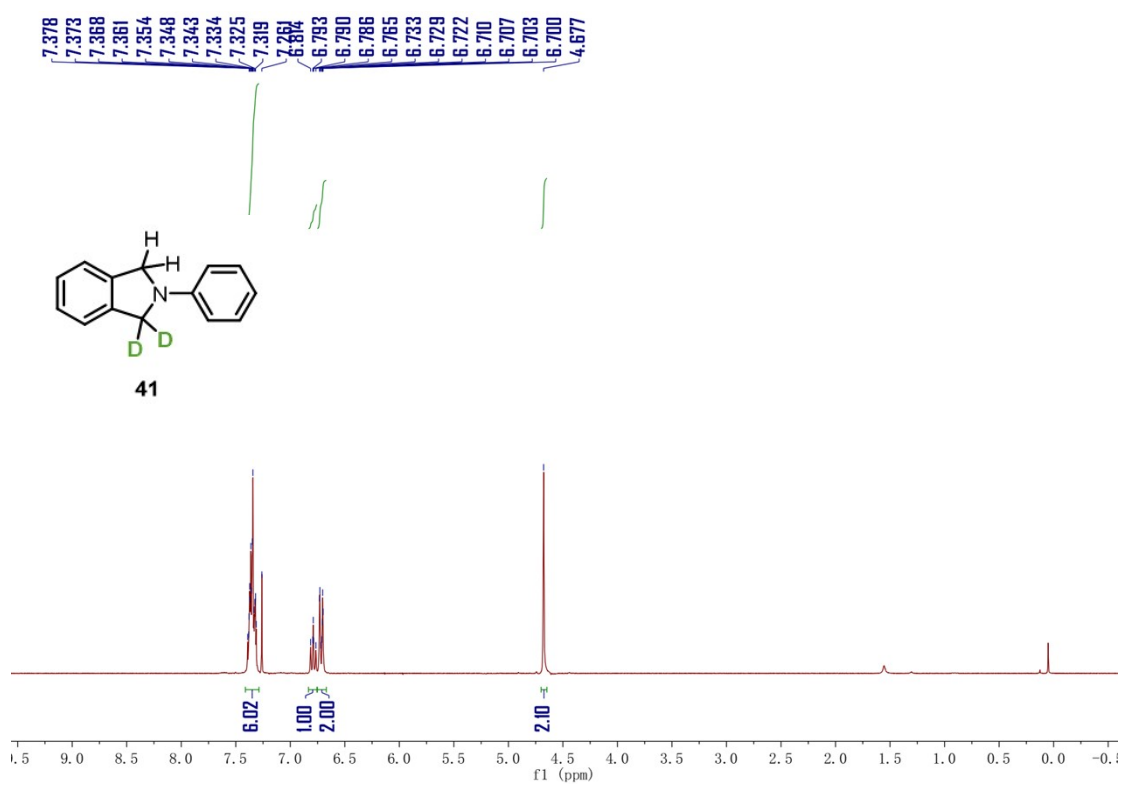
¹H NMR (300 MHz, CDCl₃):



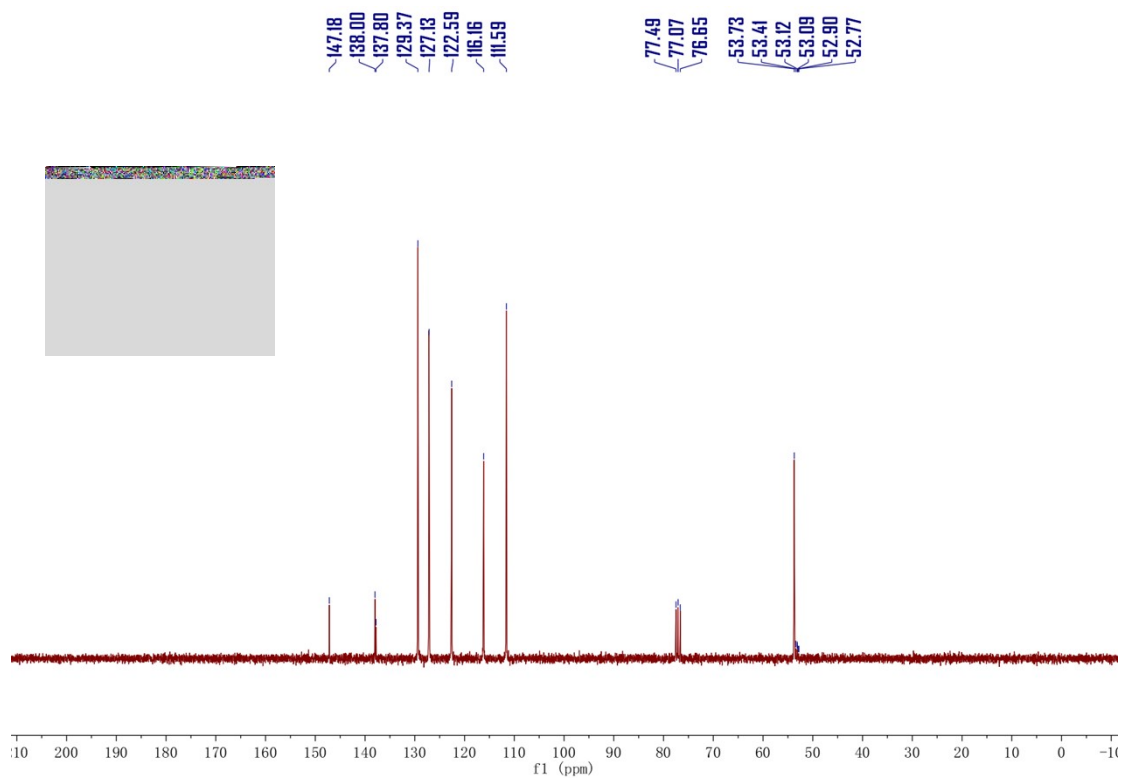
¹³C NMR (75 MHz, CDCl₃):



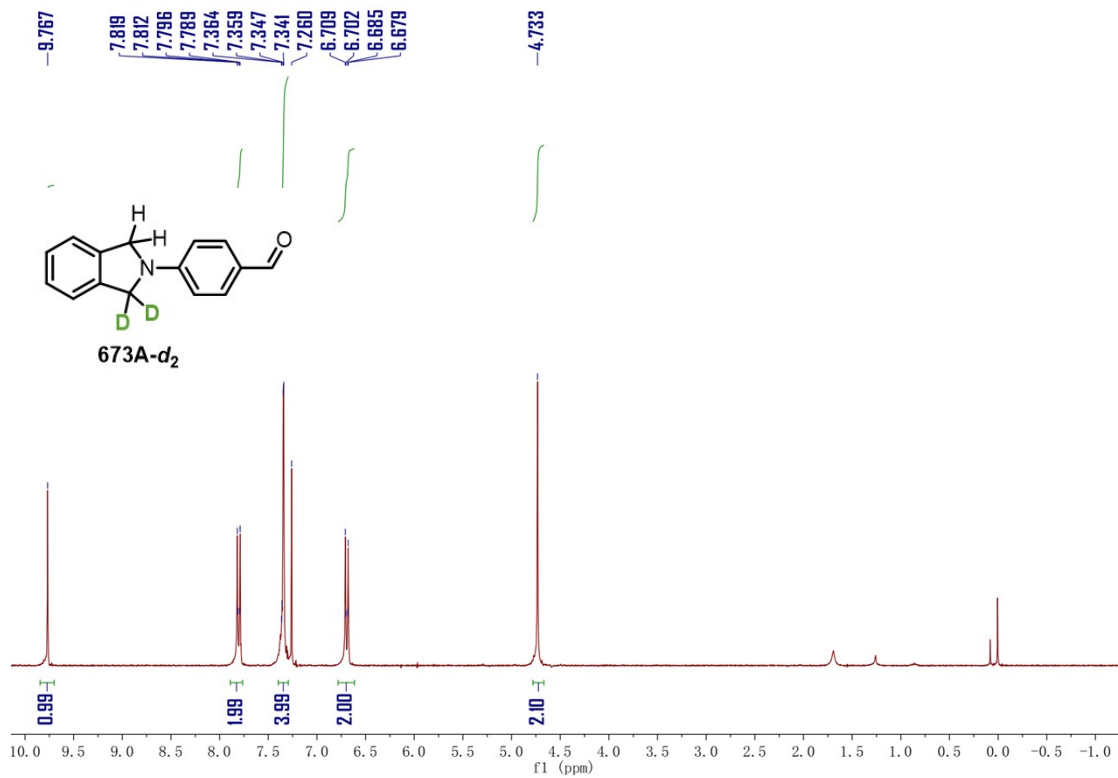
¹H NMR (300 MHz, CDCl₃):



^{13}C NMR (75 MHz, CDCl_3):



^1H NMR (300 MHz, CDCl_3):



^{13}C NMR (75 MHz, CDCl_3):

